Hydrogels for Oral Tissue Engineering

Subjects: Materials Science, Biomaterials

Contributor: Anfu Chen, Shuhua Deng, Jindi Lai, Jing Li, Weijia Chen, Swastina Nath Varma, Jingjing Zhang,

Caihong Lei , Chaozong Liu , Lijia Huang

Oral health is crucial to daily life, yet many people worldwide suffer from oral diseases. With the development of oral tissue engineering, there is a growing demand for dental biomaterials. Addressing oral diseases often requires a two-fold approach: fighting bacterial infections and promoting tissue growth. Hydrogels are promising tissue engineering biomaterials that show great potential for oral tissue regeneration and drug delivery.

tissue engineering

hydrogels

dental pulp

periodontium

mandible

1. Introduction

The oral cavity is an integral component of the digestive system, containing several significant anatomical components made up of various soft and hard tissues, such as teeth, oral mucosa, periodontal tissues, maxilla, and mandible. Additionally, the oral cavity is simultaneously populated largely by over 700 kinds of microorganisms, creating a complex ecological niche that directly impacts oral health [1]. Most researchers agree that dysbiosis of the microflora, rather than a particular species of bacteria, is the root cause of oral infectious disorders, including dental caries, periodontitis, peri-implantitis, and oral candidiasis [2]. Maintaining good oral health is critical in fending off periodontal diseases and dental caries, which can lead to more serious health problems, such as endocarditis, diabetes mellitus, and Alzheimer's disease. Therefore, the prevention and treatment of these oral illnesses have thus received significant attention [3]. Oral diseases are pathological changes that occur in the soft and hard structures of the oral cavity and maxillofacial region [4]. The most common oral diseases include dental caries, periodontitis, pulp necrosis, oral mucositis, and jaw abnormalities [5]. It is estimated that more than 3~5 billion people worldwide experience chronic oral disorders that progress over time, starting in early infancy and continuing throughout adolescence, adulthood, and later life [6]. In many countries, oral disorders place a significant health burden on individuals, causing various degrees of pain, discomfort, disfigurement, and even death in some cases. Given the complex and diverse nature of oral health issues, there seems to be an endless need for dental biomaterials that can effectively interact with a range of tissues, from soft gum tissue to hard bone tissue. Moreover, dental biomaterials must be able to withstand the challenges posed by the oral environment, including abrupt temperature changes, pH fluctuations caused by saliva and biofilms, and the presence of various types of bacteria . Consequently, there has been increasing research on developing effective treatments for oral illnesses. Bacterial infections frequently lead to oral disorders [8], and there are two main approaches for treating them. One aims to combat the infections, while the other aims to promote tissue regeneration. For repairing jawbone defects, autologous bone grafting is the most common treatment method currently used. However, autogenous bone is a limited source and cannot be reshaped to fit the defect. For periodontitis and oral mucositis, the prevention of bacterial infection is critical. Systemic administration has long been the primary treatment for the infectious diseases of the oral cavity. However, systemic administration may cause issues such as drug resistance and liver toxicity. For pulpal necrosis, regenerative root canal therapy is the mainstream treatment method, and the preferred scaffold for this therapy is an injectable biomaterial. However, there is no consensus on the most suitable injectable scaffold for endodontic treatment. Hydrogel, as a biocompatible material that can easily change shape, performs excellently in both drug delivery and tissue regeneration. Hydrogels usually possess a porous structure due to their internal network that forms many tiny pores and voids. The size and shape of these pores and voids can be controlled by regulating the conditions and preparation process during gel synthesis. By regulating the structure of the pores, it is possible to match them with surrounding tissues, promoting cell adhesion and growth [9][10]. Moreover, hydrogels exhibit excellent antimicrobial properties in oral clinical applications. Firstly, they physically isolate bacteria, preventing the invasion and spread of harmful microbes. Secondly, hydrogels act as effective drug release carriers, facilitating the delivery of various antimicrobial substances to achieve their desired effects. Recently, there has been considerable research on stimuli-responsive hydrogels. These hydrogels can respond to external physical, chemical, and biological stimuli, triggering the release of antimicrobial agents. Additionally, these hydrogels can exhibit antimicrobial properties through responsive physical and chemical properties, such as by modulating pore structure and regulating humidity. Therefore, hydrogel has become a promising material for the treatment of oral diseases.

2. Classification of Hydrogels

2.1. Natural Hydrogels

Hydrogels are polymer networks with a high water absorption capacity, formed by physical or chemical cross-linking. They can be categorized into two groups based on the source of their raw materials: natural hydrogels and synthetic hydrogels. Natural hydrogels include the collagen and gelatin extracted from animal proteins [11], the HA commonly found in animal epithelium and connective tissue [12], and alginate derived from the cytoplasm and cell wall of algae and seaweeds [13]. These biomaterials are non-toxic, highly biosafe, and biocompatible, making them suitable for various biomedical applications [14][15][16].

2.1.1. Collagen-Based Hydrogels

Collagen, the main component of the ECM in many mammalian tissues, provides support and protection to the organism and its organs due to its excellent coagulation effect. Collagen-based hydrogels have attracted significant attention from researchers, owing to their weak immunogenicity and good biocompatibility. These hydrogels have been widely applied in cartilage repair, dentistry, drug delivery, and corneal transplantation, with rapid development in recent years [17][18][19][20].

2.1.2. Hyaluronic Acid (HA) Hydrogels

HA, a nonsulfated glycosaminoglycan found in all connective tissue ECM, is one of the most commonly used natural polymers today [21][22]. Studies suggest that HA plays a crucial role in biological processes such as

angiogenesis, ECM structure, inflammation, and wound healing. Moreover, HA derivatives have been successfully employed as scaffold biomaterials for chondrocyte development, bone regeneration, and skin tissue regeneration [23][24]

2.1.3. Gelatin Hydrogels

Gelatin is a hydrophilic polymer that possesses excellent sol—gel transition properties and biocompatibility, making it a versatile material in the field of hydrogels [25]. Hydrogels made from gelatin as a matrix can mimic various tissue characteristics and allow for the tailoring of hydrogel properties, such as mechanics and degradation, to suit a wide range of biomedical applications [26][27]. Xie et al. prepared a self-shrinking wound dressing using the oxidized starch of shape memory hydrogels, which showed through ex vivo experiments that shape memory was activated at conditions similar to human physiological temperature, thus achieving atraumatic mouth closure [28]. Customized hydrogels that mimic natural cells have been found to enhance endodontic treatment, while gelatin methacrylate-based hydrogels with modifiable physical and mechanical properties have been identified as an effective strategy to promote endodontic regeneration [29][30].

2.1.4. Alginate Hydrogels

Alginate, a natural polysaccharide isolated from brown algae or bacteria, finds wide application in tissue engineering [31][32]. Its intrinsic structure is similar to natural ECM, making it an excellent choice for biocompatible scaffolds. Pan et al. successfully regenerated alveolar skeletal and soft tissues using a bionic polysaccharide hydrogel/hydroxyapatite composite scaffold, thus presenting a new approach for clinical bone defect repair [33].

2.1.5. Chitosan (CS) Hydrogels

CS, a long-chain cationic polysaccharide, is a deacetylated derivative of chitin [34]. Its wide range of biological properties, including antibacterial, anti-inflammatory, anticancer, and tissue repair properties, have been amply demonstrated. Additionally, CS has excellent drug-loading capacity in the form of nanoparticles and hydrogels. The reactive groups (e.g., –OH, –NH₂) on the chitosan backbone allow for the production of multiple derivatives with the same properties as the parent polymer, but with enhanced biocompatibility and non-toxicity [35][36][37].

2.2. Synthetic Hydrogels

Synthetic hydrogels are created through chemical reactions, as the name implies. Compared to natural hydrogels with poor mechanical properties, stability, and low bactericidal efficiency, synthetic polymer hydrogels have high molecular weight and stable mechanical properties. Moreover, the structural morphology of synthetic hydrogels can be tailored according to their intended functionality and degradability. Therefore, they can be designed with advantages such as low degradability, efficient gel formation, and a long service life. However, synthetic polymer hydrogels are typically not antimicrobial and require antimicrobial treatment [38]. Currently, synthetic hydrogels are usually made from synthetic polymers derived from natural resources, such as PLA, PEG, and GelMA [39].

2.2.1. Polylactic Acid (PLA)

PLA, a biodegradable synthetic polymer derived from abundant resources, is widely used in biomedical applications due to its excellent biocompatibility, low toxicity, biodegradability, ease of processing, and environmental friendliness [40][41]. PLA can be blended with other natural and synthetic polymers to create biodegradable PLA hydrogels [39].

2.2.2. Polyethylene Glycol (PEG)

PEG is a widely used synthetic polymer in biology due to its non-immunogenic, non-toxic, biodegradable, and highly hydrophilic nature. PEG-based hydrogels offer unique advantages in drug delivery and controlled release [42]. However, they do not provide optimal conditions for cell survival, adhesion, and development because they are inherently biologically inert.

2.2.3. Gelatin Methacryloyl (GelMA)

GelMA is a photocrosslinked hydrogel that combines the properties of both natural and synthetic biomaterials. It is produced by adding methacrylate groups to gelatin [43]. The synthesis process of GelMA is illustrated in **Figure 1**.

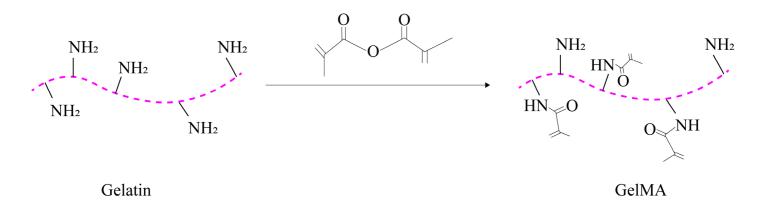


Figure 1. Synthesis process of GelMA.

3. Application of Hydrogels in Oral Tissue Engineering

3.1. Hydrogels for Dental Pulp Regeneration

Dental caries, trauma, and developmental malformations can lead to the irreversible destruction of the dental pulp, which plays a crucial role in maintaining immune defense, the sensory system, and regenerating the pulp—dentin complex [44][45][46]. The dental pulp is located inside the pulp cavity of the tooth and is protected by the non-resorbing dentin hard tissue. Following conventional root canal treatment for pulpal and periapical diseases, pulpless teeth lose their structural integrity, biological defense, and sensory capacity, which can result in the susceptibility of root fractures and poor long-term outcomes [47][48]. Thus, regenerating the dentin—pulp complex is of significant importance to restore the vitality of teeth, recover the biological function of teeth, and prolong the lifecycle of pulpless teeth.

Pulp regeneration, also known as regenerative endodontic treatment, was first proposed by Murray et al. in 2007. This ideal form of regenerative therapy aims to remove the diseased or necrotic pulp tissue and replace it with healthy, vital pulp tissue [49]. In 2016, the American Association of Endodontists formally defined pulp regeneration as the use of biological procedures to replace damaged tooth structures, including the pulp–dentin complex, root, and other structures, in order to form physiological and functional pulp-like tissue and restore biological function [50]

Biomaterial scaffolds play a crucial role in pulp regeneration by providing a 3D scaffold for stem cell adhesion, migration, proliferation, differentiation, and function. They not only regulate stem cell behaviors, intercellular and extracellular signaling, but also modulate the microenvironment, facilitating pulp—dentin complex regeneration. Recently, hydrogel-based scaffolds have been evaluated for tissue engineering-based pulp regeneration. These scaffolds have prominent biocompatibility, biodegradability, flexibility, elasticity, and ergonomic mechanical profiles, making them ideal candidates as cell or bioactive ingredient(s) delivery systems for promoting pulp—dentin complex regeneration. **Figure 2** provides a schematic illustration of the precise pulp regeneration procedure.

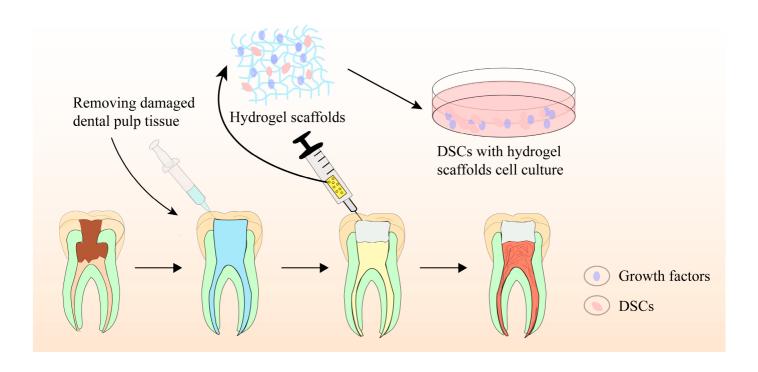


Figure 2. Schematic illustration of precise pulp regeneration procedure.

Hydrogels can be classified into two categories based on the source of polymeric chain: natural and synthetic hydrogels. Natural hydrogels mimic natural peptides and possess favorable biocompatibility, but their mechanical characteristics are poor. In contrast, synthetic hydrogels exhibit distinct mechanical features and tunable physiochemical properties, but they are deficient in biocompatibility and biodegradability capacity [51].

HA is a nonsulfated glycosaminoglycan component of the ECM of soft connective tissues [52]. As an outstanding candidate scaffold for pulp regeneration, HA has excellent biocompatibility, biodegradability, non-immunogenicity, and high water content [53].

The CS hydrogel is a promising biomaterial scaffold for regenerative endodontic treatment due to its bioactivity, biocompatibility, and capacity to blend with other bioactive ingredients. Bioactivity refers to properties of materials that trigger specific biological and chemical reactions, mainly at the interface between materials and biological tissues. Chitosan, for example, has the ability to promote cell adhesion, growth, proliferation, and differentiation. Additionally, chitosan possesses antibacterial activity. Several researchers have noted that CS-based hydrogels exhibit no biological toxicity toward various cell types and promote the proliferation and differentiation of stem cells, thereby facilitating the regeneration of pulp–dentin-like tissue [54][55].

Natural hydrogels are known for their ability to mimic natural ingredients and exhibit super biocompatibility, but they carry the risk of immune reactions and poor mechanical properties. Synthetic hydrogels, on the other hand, can be engineered to possess adjustable mechanical profiles and microstructures. When equipped with bioactive molecules and cell-binding peptides, synthetic hydrogels are optimal candidates for tissue engineering applications. For example, Kuang et al. fabricated biocompatible and biodegradable PLA-based scaffolds and tested their regulatory role in dentin–pulp complex regeneration. The PLA-based scaffolds significantly promoted the proliferation and odontogenic differentiation of human dental pulp stem cells (hDPSCs) by enhancing the expression of ALP, osteocalcin, bone sialoprotein, collagen 1, and dentin sialophosphoprotein genes in in vitro experiment. Histological analysis demonstrated superior dentin-like tissue formation in vivo [56].

3.2. Hydrogels for Periodontal Tissue Regeneration

Periodontitis is a well-known oral infectious disease characterized by inflammatory and the destruction of periodontal tissue, which can lead to an accelerated loss of alveolar bone and ultimately result in teeth loss [57][58]. In 2018, a new classification of periodontal disease and peri-implant disease was established at the Joint World Symposium in Chicago. The criteria for defining periodontal health are the absence of periodontitis in intact or reduced periodontal tissue, less than 10% bleeding on probes, and periodontal pockets less than or equal to 3 mm in depth. In the new classification, periodontitis previously categorized as "chronic" or "aggressive" is now reclassified as a single "periodontitis", with "stage" and "grade" classification. Systemic diseases and the status of affected periodontal tissue diseases have been updated. Moreover, for the first time, the classification includes peri-implant diseases and states.

Traditionally, mechanical debridement and flap surgery aiming at removing plaque have been effective in preventing the progress of inflammation and the destruction of periodontal tissue [59]. However, the reconstruction of both the structures and functions of periodontal tissue remains a great challenge and the ideal therapeutic objective of periodontal disease. Considering the physiological structure of periodontal tissues, the morphology and functional regeneration require the simultaneous or sequential repair of three components of periodontal tissue, including the periodontal ligament (PDL), which fixes the tooth, the cementum covering the root surface, and the

alveolar bone supporting the tooth [60][61]. Periodontal tissue engineering has emerged as a promising technique that combines stem cells, biological scaffolds, and growth factors to promote periodontal tissue regeneration [105]

Any biomaterial scaffold, whether biological or synthetic, must be biocompatible and biodegradable when applied to tissue regeneration. Hydrogels are widely used as regenerative scaffolds in periodontal tissue engineering [62]. Due to their key characteristics of porosity, stiffness, and viscoelasticity, hydrogels can mimic the microenvironment of the ECM and facilitate the regulation of cell adhesion, proliferation, and osteogenic differentiation [63]. Studies have reported that when combined with drugs, stem cells, or growth factors, hydrogels exhibit outstanding potential in the complex and sophisticated process of periodontal tissue regeneration [64][65]. **Figure 3** shows a schematic diagram of hydrogels applied in periodontal regeneration.

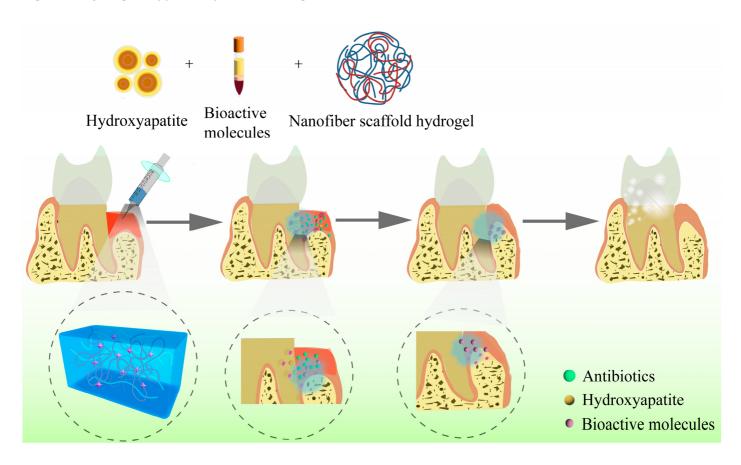


Figure 3. Schematic diagram of hydrogels applied in periodontal regeneration.

Collagen is the fundamental component of the ECM. It is composed of many specific cell signaling binding domains that facilitate cell adhesion, preserve cell phenotype, and guide cell growth, proliferation, and differentiation. For instance, Jung et al. reported that collagen hydrogels with different porosities can serve as a perfect scaffold for PDL repair and periodontal regeneration [66].

HA, another important component of the ECM of connective tissues and periodontal ligament matrix, has valuable potential in periodontal tissue regeneration [67][68][69][70]. Studies have shown that HA can modulate cell adhesion, migration, and differentiation by binding proteins and cell-surface receptors. When combined and modified with

other ingredients, HA hydrogels exhibit excellent mechanical properties, swelling, and low degradation speed in periodontal tissue engineering.

CS is a natural cationic polymer with a chemical structure and biological properties similar to glycosaminoglycan polysaccharide. Due to its excellent biocompatibility, biodegradability, and antimicrobial activity, CS is frequently used in periodontal therapy.

3.3. Hydrogels for Mandible Regeneration

The mandible is a crucial component of the human face, as it plays a vital role in mastication, pronunciation, and speaking, while also contributing to facial contour and shape [71]. However, loss of bone tissue in the oral cavity can occur due to various reasons, such as traumas, tumors, infections, functional atrophy, congenital disorders, and periodontitis, leading to different degrees of impact on the patient's facial appearance and oral function [72][73][74][75]. Certain drugs, such as isophosphonates, can also lead to jaw osteonecrosis, known as drug-related osteonecrosis of the jaw (MRONJ), as they are anti-angiogenic or anti-osteoporotic [76]. Even a short-term discontinuation of such drugs does not eliminate their effects on the jaws, and segmental resection becomes the only solution to remove necrotic bones in the late stage of MRONJ [77]. Although bones can heal themselves, repairing bone defects larger than a critical size and ununited fractures remains a challenge in clinical practice [78]. Moreover, self-repair is not feasible when the loss of the mandible exceeds 10% [79].

Figure 3 shows a schematic diagram of mandibular restoration using hydrogel biomaterials.

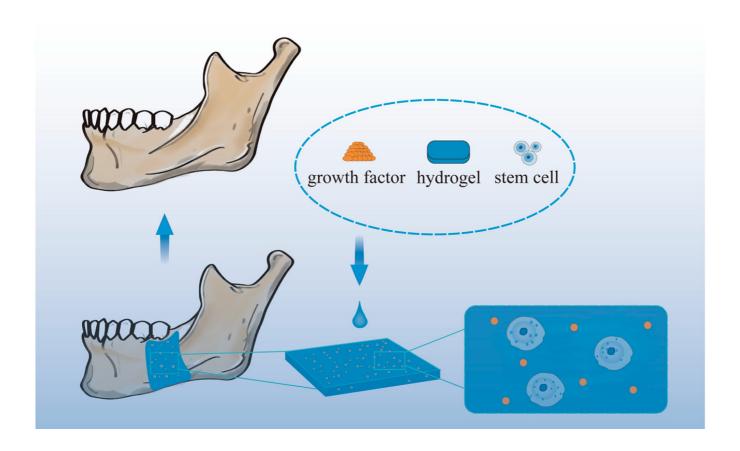


Figure 3. Schematic diagram of mandibular restoration using hydrogel biomaterials.

Hydrogel systems play a significant role in the treatment of infected jaw defects by facilitating antibiotic release. For instance, Sun et al. prepared vancomycin hydrochloride (Van)-SBA-15 using mesoporous silica (SBA-15) and encapsulated Van in a CS-sodium glycerophosphate-sodium alginate hydrogel. The hydrogel enabled the sustained release of Van and holds potential for the treatment of infected jaw defects [80]. In another study, clindamycin (CDM) was shown to penetrate human bone tissue effectively [81].

Furthermore, hydrogels have been used to mimic native bone and enhance bone regeneration. For example, Kumar et al. developed a bone tissue engineering biphasic construct loaded with bone BMP-2. They used a gelatin–HA hydrogel to bind to the osteogenic cue BMP-2, which was then loaded onto a PCL scaffold. The construct mimicked native bone and consisted of cortical bone and cancellous bone for vertical jawbone augmentation. In vitro studies showed that the cell viability of BMP-2 was maintained in the hydrogel for 21 days, and bone markers increased on the third and fourteenth days [82].

3.4. Hydrogels for Soft Tissue Healing

While dental and skeletal repair are the primary focus of craniofacial applications, the need for soft tissue regeneration is equally crucial [83]. Oral and maxillofacial soft tissues include various structures such as periodontal tissues, tongue, oral mucosa, muscles, skin. Soft tissue regeneration remains a major challenge in contemporary medicine and dentistry. Periodontitis, gingival recession, and chronic inflammation of the gums can lead to tooth loss. In the cases of long-term tooth loss, proper dental restoration and aesthetic outcomes heavily depend on critical soft tissues [84]. After dental implantation, the bacterial environment in the oral cavity can cause inflammation around the implant. Enhanced sealing of the soft tissues around the implant can improve the success of the implant [85].

Soft tissue engineering has emerged as a new approach to repair damaged or diseased soft tissues and organs [86]. Hydrogels, which are ideal biomaterials to mimic soft tissues [87], have been developed for this purpose. However, treating oral diseases can be challenging because high doses of topical medications are often required, and systemic administration is typically the main treatment option for small lesions of the oral mucosa. The moist, dynamic, and unstable environment of the oral cavity, including constant salivary flushing, and exposure to microorganisms, enzymes, and food and beverages, makes it difficult to maintain wound dressings. Furthermore, hydrogels must meet high demands for structural and mechanical stability [88].

4. Clinical Applications of Hydrogels in Oral Tissue Repair and Regeneration

There have been several clinical studies on the use of hydrogels for repairing oral tissues, and information about these studies is listed in **Table 1**. Most of these studies focused on the use of hydrogels for treating oral mucositis

and periodontitis, with one study investigating the treatment of oral bacterial infections and another exploring tooth loss.

Table 1. Clinical applications of hydrogels in oral tissue repair and regeneration [89].

Status	Study Title	Conditions	Interventions
Completed	Omega-3 hydrogel and prevention of oral mucositis	Mucositis oral	Drug: topical oral Omega-3 hydrogel; drug: conventional preventive treatment
Not yet recruiting	Efficacy of EGF-loaded self-healing gel in treating oral mucositis	Oral mucositis	Drug: EGF-loaded hydrogel; drug: hydrogel
Completed	A study to evaluate efficacy of MuGard for amelioration of oral mucositis in head and neck cancer patients	Oral mucositis	Device: MuGard; device: control rinse
Recruiting	MucoLox formulation to mitigate mucositis symptoms in head/neck cancer	Mucositis oral head and neck cancer	Other: MucoLox; other: sodium bicarbonate
Unknown	Topical chamomile in preventing chemotherapy-induced oral mucositis	Oral mucositis due to chemotherapy	Drug: chamomile topical oral gel; drug: miconazole topical gel; drug: BBC oral spray; drug: oracure gel
Completed	Impact of daily use of emanate tray adjunct to full mouth debridement compared to full mouth debridement alone	Wound heal mouth; wound periodontal inflammation	Device: emanate tray
Completed	Nitazoxanide as a new local adjunctive to nonsurgical treatment of moderate periodontitis	Periodontitis	Procedure: scaling and root planing; drug: nitazoxanide hydrogel
Completed	Efficacy of proanthocyanidins in nonsurgical periodontal therapy	Periodontitis, adult	Procedure: minimally invasive nonsurgical therapy; combination product: subgingival application of collagen hydrogels with proanthocyanidins; diagnostic test: collection of saliva samples
Completed	Does hyaluronic acid affect periodontal treatment?	Periodontitis	Procedure: scaling and root planing; drug: hyaluronic acid gel (HA) and SRP; drug: HA mouthrinse and SRP;

Status	Study Title	Conditions	Interventions
			drug: HA mouthrinse + gel and SRP
Completed	Use of adhesion molecule-loaded hydrogel with minimally invasive surgical technique in treating periodontal intrabony defects	Periodontitis	Drug: RGD peptide
Completed	PLGA nanoparticles entrapping ciprofloxacin to treat E-Fecalis infections in endodontics	Bacterial infections oral	Device: chitosan-coated PLGA nanoparticles entrapping ciprofloxacin incorporated in smart gels; device: ciprofloxacin paste and solution
Unknown	Hyaluronic acid effect on xenogenic bone healing	Bone resorption tooth loss	Procedure: ridge preservation: tooth extraction and immediate bone grafting in the socket

- 2. Lamont, R.J.; Koo, H.; Hajishengallis, G. The Oral Microbiota: Dynamic Communities and Host Interactions. Nat. Rev. Microbiol. 2018, 16, 745–759.
- 3. Fukushima-Nakayama, Y.; Ono, T.; Hayashi, M.; Inoue, M.; Wake, H.; Ono, T.; Nakashima, T. Reduced Mastication Impairs Memory Function. J. Dent. Res. 2017, 96, 1058–1066.
- 4. Gao, H.; Wu, N.; Wang, N.; Li, J.; Sun, J.; Peng, Q. Chitosan-Based Therapeutic Systems and Their Potentials in Treatment of Oral Diseases. Int. J. Biol. Macromol. 2022, 222, 3178–3194.
- 5. Ye, S.; Wei, B.; Zeng, L. Advances on Hydrogels for Oral Science Research. Gels 2022, 8, 302.
- 6. Peres, M.A.; Macpherson, L.M.; Weyant, R.J.; Daly, B.; Venturelli, R.; Mathur, M.R.; Listl, S.; Celeste, R.K.; Guarnizo-Herreño, C.C.; Kearns, C.; et al. Oral Diseases: A Global Public Health Challenge. Lancet 2019, 394, 249–260.
- 7. Fischer, N.G.; Münchow, E.A.; Tamerler, C.; Bottino, M.C.; Aparicio, C. Harnessing Biomolecules for Bioinspired Dental Biomaterials. J. Mater. Chem. B 2020, 8, 8713–8747.
- 8. Şenel, S.; Özdoğan, A.I.; Akca, G. Current Status and Future of Delivery Systems for Prevention and Treatment of Infections in the Oral Cavity. Drug Deliv. Transl. Res. 2021, 11, 1703–1734.
- 9. Yang, J.; Liu, F.; Zhou, C.; Li, H.; Yang, G.; Fang, S.; Lee, I.S.; Liu, Y.; Bai, H.; Chen, C. 3D Printed Porous Titanium Filled with Mineralized UV-Responsive Chitosan Hydrogel Promotes Cell Proliferation and Osteogenesis in vitro. J. Mater. Sci. Technol. 2023, 142, 34–44.
- 10. Hu, C.; Zhang, M.; Wu, J.; Cao, X.; Chen, L.; Yan, J.; Liang, G.; Tan, J. Bisphosphonate-Modified Functional Supramolecular Hydrogel Promotes Periodontal Bone Regeneration by Osteoclast Inhibition. ACS Appl. Mater. Interfaces 2023, 15, 9066–9079.

- 11. León-López, A.; Morales-Peñaloza, A.; Martínez-Juárez, V.M.; Vargas-Torres, A.; Zeugolis, D.I.; Aguirre-Álvarez, G. Hydrolyzed Collagen—Sources and Applications. Molecules 2019, 24, 4031.
- 12. Fallacara, A.; Baldini, E.; Manfredini, S.; Vertuani, S. Hyaluronic Acid in the Third Millennium. Polymers 2018, 10, 701.
- 13. Reakasame, S.; Boccaccini, A.R. Oxidized Alginate-Based Hydrogels for Tissue Engineering Applications: A Review. Biomacromolecules 2018, 19, 3–21.
- 14. Seliktar, D. Designing Cell-Compatible Hydrogels for Biomedical Applications. Science 2012, 336, 1124–1128.
- 15. Cao, H.; Duan, L.; Zhang, Y.; Cao, J.; Zhang, K. Current Hydrogel Advances in Physicochemical and Biological Response-Driven Biomedical Application Diversity. Signal Transduct. Target. Ther. 2021, 6, 426.
- 16. Nie, L.; Li, X.; Chang, P.; Liu, S.; Wei, Q.; Guo, Q.; Wu, Q.; Fan, L.; Okoro, O.V.; Shavandi, A. A Fast Method for in Vitro Biomineralization of PVA/Alginate/Biphasic Calcium Phosphate Hydrogel. Mater. Lett. 2022, 308, 131182.
- 17. Leucht, A.; Volz, A.C.; Rogal, J.; Borchers, K.; Kluger, P. Advanced Gelatin-Based Vascularization Bioinks for Extrusion-Based Bioprinting of Vascularized Bone Equivalents. Sci. Rep. 2020, 10, 5330.
- 18. Ikeda, Y.; Holcroft, J.; Ikeda, E.; Ganss, B. Amelotin Promotes Mineralization and Adhesion in Collagen-Based Systems. Cell. Mol. Bioeng. 2022, 15, 245–254.
- 19. Huang, Q.; Huang, X.; Gu, L. Periodontal Bifunctional Biomaterials: Progress and Perspectives. Materials 2021, 14, 7588.
- 20. Andonegi, M.; Las Heras, K.; Santos-Vizcaíno, E.; Igartua, M.; Hernandez, R.M.; de la Caba, K.; Guerrero, P. Structure-Properties Relationship of Chitosan/Collagen Films with Potential for Biomedical Applications. Carbohydr. Polym. 2020, 237, 116159.
- 21. Kwon, M.Y.; Wang, C.; Galarraga, J.H.; Puré, E.; Han, L.; Burdick, J.A. Influence of Hyaluronic Acid Modification on CD44 Binding towards the Design of Hydrogel Biomaterials. Biomaterials 2019, 222, 119451.
- 22. Passi, A.; Vigetti, D. Hyaluronan as Tunable Drug Delivery System. Adv. Drug Deliv. Rev. 2019, 146, 83–96.
- 23. Chircov, C.; Grumezescu, A.M.; Bejenaru, L.E. Hyaluronic Acid-Based Scaffolds for Tissue Engineering. Rom. J. Morphol. Embryol. 2018, 59, 71–76.
- 24. Graça, M.F.; Miguel, S.P.; Cabral, C.S.; Correia, I.J. Hyaluronic Acid—Based Wound Dressings: A Review. Carbohydr. Polym. 2020, 241, 116364.

- 25. Abbass, M.M.; El-Rashidy, A.A.; Sadek, K.M.; Moshy, S.E.; Radwan, I.A.; Rady, D.; Dörfer, C.E.; Fawzy El-Sayed, K.M. Hydrogels and Dentin–Pulp Complex Regeneration: From the Benchtop to Clinical Translation. Polymers 2020, 12, 2935.
- 26. Parthiban, S.P.; He, W.; Monteiro, N.; Athirasala, A.; França, C.M.; Bertassoni, L.E. Engineering Pericyte-Supported Microvascular Capillaries in Cell-Laden Hydrogels Using Stem Cells from the Bone Marrow, Dental Pulp and Dental Apical Papilla. Sci. Rep. 2020, 10, 21579.
- 27. Wang, Q.Q.; Liu, Y.; Zhang, C.J.; Zhang, C.; Zhu, P. Alginate/Gelatin Blended Hydrogel Fibers Cross-Linked by Ca2+ and Oxidized Starch: Preparation and Properties. Mater. Sci. Eng. C 2019, 99, 1469–1476.
- 28. Mao, Q.; Hoffmann, O.; Yu, K.; Lu, F.; Lan, G.; Dai, F.; Shang, S.; Xie, R. Self-Contracting Oxidized Starch/Gelatin Hydrogel for Noninvasive Wound Closure and Wound Healing. Mater. Des. 2020, 194, 108916.
- 29. Athirasala, A.; Lins, F.; Tahayeri, A.; Hinds, M.; Smith, A.J.; Sedgley, C.; Ferracane, J.; Bertassoni, L.E. A Novel Strategy to Engineer Pre-Vascularized Full-Length Dental Pulp-like Tissue Constructs. Sci. Rep. 2017, 7, 3323.
- 30. Park, J.H.; Gillispie, G.J.; Copus, J.S.; Zhang, W.; Atala, A.; Yoo, J.J.; Yelick, P.C.; Lee, S.J. The Effect of BMP-Mimetic Peptide Tethering Bioinks on the Differentiation of Dental Pulp Stem Cells (DPSCs) in 3D Bioprinted Dental Constructs. Biofabrication 2020, 12, 035029.
- 31. He, Y.; Zhao, W.; Dong, Z.; Ji, Y.; Li, M.; Hao, Y.; Zhang, D.; Yuan, C.; Deng, J.; Zhao, P.; et al. A Biodegradable Antibacterial Alginate/Carboxymethyl Chitosan/Kangfuxin Sponges for Promoting Blood Coagulation and Full-Thickness Wound Healing. Int. J. Biol. Macromol. 2021, 167, 182–192.
- 32. Abasalizadeh, F.; Moghaddam, S.V.; Alizadeh, E.; Akbari, E.; Kashani, E.; Fazljou, S.M.B.; Torbati, M.; Akbarzadeh, A. Alginate-Based Hydrogels as Drug Delivery Vehicles in Cancer Treatment and Their Applications in Wound Dressing and 3D Bioprinting. J. Biol. Eng. 2020, 14, 8.
- 33. Pan, Y.; Zhao, Y.; Kuang, R.; Liu, H.; Sun, D.; Mao, T.; Jiang, K.; Yang, X.; Watanabe, N.; Mayo, K.H.; et al. Injectable Hydrogel-Loaded Nano-Hydroxyapatite That Improves Bone Regeneration and Alveolar Ridge Promotion. Mater. Sci. Eng. C 2020, 116, 111158.
- 34. Kou, S.G.; Peters, L.M.; Mucalo, M.R. Chitosan: A Review of Sources and Preparation Methods. Int. J. Biol. Macromol. 2021, 169, 85–94.
- 35. Saeedi, M.; Vahidi, O.; Moghbeli, M.; Ahmadi, S.; Asadnia, M.; Akhavan, O.; Seidi, F.; Rabiee, M.; Saeb, M.R.; Webster, T.J.; et al. Customizing Nano-Chitosan for Sustainable Drug Delivery. J. Control. Release 2022, 350, 175–192.
- 36. Fabiano, A.; Beconcini, D.; Migone, C.; Piras, A.M.; Zambito, Y. Quaternary Ammonium Chitosans: The Importance of the Positive Fixed Charge of the Drug Delivery Systems. Int. J. Mol.

- Sci. 2020, 21, 6617.
- 37. Iftime, M.M.; Ailiesei, G.L.; Ungureanu, E.; Marin, L. Designing Chitosan Based Eco-Friendly Multifunctional Soil Conditioner Systems with Urea Controlled Release and Water Retention. Carbohydr. Polym. 2019, 223, 115040.
- 38. Wang, R.; Li, N.; Jiang, B.; Li, J.; Hong, W.; Jiao, T. Facile Preparation of Agar/Polyvinyl Alcohol-Based Triple-Network Composite Hydrogels with Excellent Mechanical Performances. Colloids Surf. Physicochem. Eng. Asp. 2021, 615, 126270.
- 39. Munim, S.A.; Raza, Z.A. Poly (Lactic Acid) Based Hydrogels: Formation, Characteristics and Biomedical Applications. J. Porous Mater. 2019, 26, 881–901.
- 40. Bhattarai, N.; Gunn, J.; Zhang, M. Chitosan-Based Hydrogels for Controlled, Localized Drug Delivery. Adv. Drug Deliv. Rev. 2010, 62, 83–99.
- 41. Ganji, F.; Abdekhodaie, M.J. Chitosan–g–PLGA Copolymer as a Thermosensitive Membrane. Carbohydr. Polym. 2010, 80, 740–746.
- 42. Fu, Y.; Ding, Y.; Zhang, L.; Zhang, Y.; Liu, J.; Yu, P. Poly Ethylene Glycol (PEG)-Related Controllable and Sustainable Antidiabetic Drug Delivery Systems. Eur. J. Med. Chem. 2021, 217, 113372.
- 43. Klotz, B.J.; Gawlitta, D.; Rosenberg, A.J.; Malda, J.; Melchels, F.P. Gelatin-Methacryloyl Hydrogels: Towards Biofabrication-Based Tissue Repair. Trends Biotechnol. 2016, 34, 394–407.
- 44. Fahmy, S.H.; Hassanien, E.E.S.; Nagy, M.M.; El Batouty, K.M.; Mekhemar, M.; Fawzy El Sayed, K.; Hassanein, E.H.; Wiltfang, J.; Dörfer, C. Investigation of the Regenerative Potential of Necrotic Mature Teeth Following Different Revascularisation Protocols. Aust. Endod. J. 2017, 43, 73–82.
- 45. Fawzy El-Sayed, K.M.; Elsalawy, R.; Ibrahim, N.; Gadalla, M.; Albargasy, H.; Zahra, N.; Mokhtar, S.; El Nahhas, N.; El Kaliouby, Y.; Dörfer, C.E. The Dental Pulp Stem/Progenitor Cells-Mediated Inflammatory-Regenerative Axis. Tissue Eng. Part B Rev. 2019, 25, 445–460.
- 46. El-Sayed, K.M.F.; Klingebiel, P.; Dörfer, C.E. Toll-like Receptor Expression Profile of Human Dental Pulp Stem/Progenitor Cells. J. Endod. 2016, 42, 413–417.
- 47. Jakovljevic, A.; Nikolic, N.; Jacimovic, J.; Pavlovic, O.; Milicic, B.; Beljic-Ivanovic, K.; Miletic, M.; Andric, M.; Milasin, J. Prevalence of Apical Periodontitis and Conventional Nonsurgical Root Canal Treatment in General Adult Population: An Updated Systematic Review and Meta-Analysis of Cross-Sectional Studies Published between 2012 and 2020. J. Endod. 2020, 46, 1371–1386.
- 48. Lempel, E.; Lovász, B.V.; Bihari, E.; Krajczár, K.; Jeges, S.; Tóth, Á.; Szalma, J. Long-Term Clinical Evaluation of Direct Resin Composite Restorations in Vital vs. Endodontically Treated Posterior Teeth—Retrospective Study up to 13 Years. Dent. Mater. 2019, 35, 1308–1318.

- 49. Murray, P.E.; Garcia-Godoy, F.; Hargreaves, K.M. Regenerative Endodontics: A Review of Current Status and a Call for Action. J. Endod. 2007, 33, 377–390.
- 50. Sui, B.; Chen, C.; Kou, X.; Li, B.; Xuan, K.; Shi, S.; Jin, Y. Pulp Stem Cell–Mediated Functional Pulp Regeneration. J. Dent. Res. 2019, 98, 27–35.
- 51. Ma, P.; Chen, Y.; Lai, X.; Zheng, J.; Ye, E.; Loh, X.J.; Zhao, Y.; Parikh, B.H.; Su, X.; You, M.; et al. The Translational Application of Hydrogel for Organoid Technology: Challenges and Future Perspectives. Macromol. Biosci. 2021, 21, 2100191.
- 52. Kim, H.; Koh, W.G.; Lee, H.G. Effects of Basic Fibroblast Growth Factor Combined with an Injectable in Situ Crosslinked Hyaluronic Acid Hydrogel for a Dermal Filler. React. Funct. Polym. 2021, 164, 104933.
- 53. Park, S.H.; Park, J.Y.; Ji, Y.B.; Ju, H.J.; Min, B.H.; Kim, M.S. An Injectable Click-Crosslinked Hyaluronic Acid Hydrogel Modified with a BMP-2 Mimetic Peptide as a Bone Tissue Engineering Scaffold. Acta Biomater. 2020, 117, 108–120.
- 54. El Ashiry, E.A.; Alamoudi, N.M.; El Ashiry, M.K.; Bastawy, H.A.; El Derwi, D.A.; Atta, H.M. Tissue Engineering of Necrotic Dental Pulp of Immature Teeth with Apical Periodontitis in Dogs: Radiographic and Histological Evaluation. J. Clin. Pediatr. Dent. 2018, 42, 373–382.
- 55. Park, S.J.; Li, Z.; Hwang, I.N.; Huh, K.M.; Min, K.S. Glycol Chitin–Based Thermoresponsive Hydrogel Scaffold Supplemented with Enamel Matrix Derivative Promotes Odontogenic Differentiation of Human Dental Pulp Cells. J. Endod. 2013, 39, 1001–1007.
- 56. Kuang, R.; Zhang, Z.; Jin, X.; Hu, J.; Gupte, M.J.; Ni, L.; Ma, P.X. Nanofibrous Spongy Microspheres Enhance Odontogenic Differentiation of Human Dental Pulp Stem Cells. Adv. Healthc. Mater. 2015, 4, 1993–2000.
- 57. Slots, J. Periodontitis: Facts, Fallacies and the Future. Periodontology 2017, 75, 7–23.
- 58. Wang, Y.; Wang, Z.; Dong, Y. Collagen-Based Biomaterials for Tissue Engineering. ACS Biomater. Sci. Eng. 2023, 9, 1132–1150.
- 59. Aljateeli, M.; Koticha, T.; Bashutski, J.; Sugai, J.V.; Braun, T.M.; Giannobile, W.V.; Wang, H.L. Surgical Periodontal Therapy with and without Initial Scaling and Root Planing in the Management of Chronic Periodontitis: A Randomized Clinical Trial. J. Clin. Periodontol. 2014, 41, 693–700.
- 60. Liu, J.; Ruan, J.; Weir, M.D.; Ren, K.; Schneider, A.; Wang, P.; Oates, T.W.; Chang, X.; Xu, H.H. Periodontal Bone-Ligament-Cementum Regeneration via Scaffolds and Stem Cells. Cells 2019, 8, 537.
- 61. Salar Amoli, M.; EzEldeen, M.; Jacobs, R.; Bloemen, V. Materials for Dentoalveolar Bioprinting: Current State of the Art. Biomedicines 2022, 10, 71.

- 62. Chen, F.M.; Jin, Y. Periodontal Tissue Engineering and Regeneration: Current Approaches and Expanding Opportunities. Tissue Eng. Part B Rev. 2010, 16, 219–255.
- 63. Chang, B.; Ahuja, N.; Ma, C.; Liu, X. Injectable Scaffolds: Preparation and Application in Dental and Craniofacial Regeneration. Mater. Sci. Eng. R Rep. 2017, 111, 1–26.
- 64. Fan, C.; Wang, D.A. Macroporous Hydrogel Scaffolds for Three-Dimensional Cell Culture and Tissue Engineering. Tissue Eng. Part B Rev. 2017, 23, 451–461.
- 65. Mantha, S.; Pillai, S.; Khayambashi, P.; Upadhyay, A.; Zhang, Y.; Tao, O.; Pham, H.M.; Tran, S.D. Smart Hydrogels in Tissue Engineering and Regenerative Medicine. Materials 2019, 12, 3323.
- 66. Yang, J.M.; Olanrele, O.S.; Zhang, X.; Hsu, C.C. Fabrication of Hydrogel Materials for Biomedical Applications. Nov. Biomater. Regen. Med. 2018, 1077, 197–224.
- 67. Jung, I.H.; Park, J.C.; Kim, J.C.; Jeon, D.W.; Choi, S.H.; Cho, K.S.; Im, G.I.; Kim, B.S.; Kim, C.S. Novel Application of Human Periodontal Ligament Stem Cells and Water-Soluble Chitin for Collagen Tissue Regeneration: In Vitro and In Vivo Investigations. Tissue Eng. Part A 2012, 18, 643–653.
- 68. Burdick, J.A.; Prestwich, G.D. Hyaluronic Acid Hydrogels for Biomedical Applications. Adv. Mater. 2011, 23, H41–H56.
- 69. Fraser, J.R.E.; Laurent, T.C.; Laurent, U. Hyaluronan: Its Nature, Distribution, Functions and Turnover. J. Intern. Med. 1997, 242, 27–33.
- 70. Oksala, O.; Salo, T.; Tammi, R.; Häkkinen, L.; Jalkanen, M.; Inki, P.; Larjava, H. Expression of Proteoglycans and Hyaluronan during Wound Healing. J. Histochem. Cytochem. 1995, 43, 125–135.
- 71. Rosaming, P.; Jirayupapong, J.; Thamnium, S.; Win, Y.Y.; Limprasutr, V.; Rodsiri, R.; Pavasant, P.; Luckanagul, J.A. Interpenetrating Low-Molecular Weight Hyaluronic Acid in Hyaluronic Acid-Based In Situ Hydrogel Scaffold for Periodontal and Oral Wound Applications. Polymers 2022, 14, 4986.
- 72. Kakarala, K.; Shnayder, Y.; Tsue, T.T.; Girod, D.A. Mandibular Reconstruction. Oral Oncol. 2018, 77, 111–117.
- 73. Zhang, Q.; Wu, W.; Qian, C.; Xiao, W.; Zhu, H.; Guo, J.; Meng, Z.; Zhu, J.; Ge, Z.; Cui, W. Advanced Biomaterials for Repairing and Reconstruction of Mandibular Defects. Mater. Sci. Eng. C 2019, 103, 109858.
- 74. Berg, B.I.; Juergens, P.; Soerensen, Y.; Savic, M.; Zeilhofer, H.F.; Schwenzer-Zimmerer, K. Traumatology of the Facial Skeleton in Octogenarian Patients: A Retrospective Analysis of 96 Cases. J. Cranio-Maxillofac. Surg. 2014, 42, 870–873.

- 75. Thariat, J.; Julieron, M.; Brouchet, A.; Italiano, A.; Schouman, T.; Marcy, P.Y.; Odin, G.; Lacout, A.; Dassonville, O.; Peyrottes-Birstwisles, I.; et al. Osteosarcomas of the Mandible: Are They Different from Other Tumor Sites? Crit. Rev. Oncol. Hematol. 2012, 82, 280–295.
- 76. Bede, S.Y.H.; Ismael, W.K.; Hashim, E.A. Reconstruction Plate-Related Complications in Mandibular Continuity Defects. Oral Maxillofac. Surg. 2019, 23, 193–199.
- 77. Diniz-Freitas, M.; Fernández-Feijoo, J.; Diz Dios, P.; Pousa, X.; Limeres, J. Denosumab-Related Osteonecrosis of the Jaw Following Non-Surgical Periodontal Therapy: A Case Report. J. Clin. Periodontol. 2018, 45, 570–577.
- 78. Joo, Y.H.; Cho, J.K.; Koo, B.S.; Kwon, M.; Kwon, S.K.; Kwon, S.Y.; Kim, M.S.; Kim, J.K.; Kim, H.; Nam, I.; et al. Guidelines for the Surgical Management of Oral Cancer: Korean Society of Thyroid-Head and Neck Surgery. Clin. Exp. Otorhinolaryngol. 2019, 12, 107–144.
- 79. Armiento, A.R.; Hatt, L.P.; Sanchez Rosenberg, G.; Thompson, K.; Stoddart, M.J. Functional Biomaterials for Bone Regeneration: A Lesson in Complex Biology. Adv. Funct. Mater. 2020, 30, 1909874.
- 80. Guo, J.; Yao, H.; Li, X.; Chang, L.; Wang, Z.; Zhu, W.; Su, Y.; Qin, L.; Xu, J. Advanced Hydrogel Systems for Mandibular Reconstruction. Bioact. Mater. 2023, 21, 175–193.
- 81. Sun, M.; Cheng, L.; Xu, Z.; Chen, L.; Liu, Y.; Xu, Y.; Zhou, D.; Zhang, X.; Zhou, Q.; Sun, J. Preparation and Characterization of Vancomycin Hydrochloride-Loaded Mesoporous Silica Composite Hydrogels. Front. Bioeng. Biotechnol. 2022, 10, 4.
- 82. Thabit, A.K.; Fatani, D.F.; Bamakhrama, M.S.; Barnawi, O.A.; Basudan, L.O.; Alhejaili, S.F. Antibiotic Penetration into Bone and Joints: An Updated Review. Int. J. Infect. Dis. 2019, 81, 128–136.
- 83. Kumar, P.S.; Hashimi, S.; Saifzadeh, S.; Ivanovski, S.; Vaquette, C. Additively Manufactured Biphasic Construct Loaded with BMP-2 for Vertical Bone Regeneration: A Pilot Study in Rabbit. Mater. Sci. Eng. C 2018, 92, 554–564.
- 84. Trubelja, A.; Kasper, F.K.; Farach-Carson, M.C.; Harrington, D.A. Bringing Hydrogel-Based Craniofacial Therapies to the Clinic. Acta Biomater. 2022, 138, 1–20.
- 85. Ali Salim, K.M.; Abd Jalil, A.; Radzi, Z.; Ismail, S.M.; Czernuszka, J.T.; Rahman, M.T. Inflammatory Responses in Oro-Maxillofacial Region Expanded Using Anisotropic Hydrogel Tissue Expander. Materials 2020, 13, 4436.
- 86. Su, T.; Zheng, A.; Cao, L.; Peng, L.; Wang, X.; Wang, J.; Xin, X.; Jiang, X. Adhesion-Enhancing Coating Embedded with Osteogenesis-Promoting PDA/HA Nanoparticles for Peri-Implant Soft Tissue Sealing and Osseointegration. Bio-Des. Manuf. 2022, 5, 233–248.

- 87. Pei, B.; Wang, W.; Fan, Y.; Wang, X.; Watari, F.; Li, X. Fiber-Reinforced Scaffolds in Soft Tissue Engineering. Regen. Biomater. 2017, 4, 257–268.
- 88. Kuth, S.; Karakaya, E.; Reiter, N.; Schmidt, L.; Paulsen, F.; Teßmar, J.; Budday, S.; Boccaccini, A.R. Oxidized Hyaluronic Acid-Gelatin-Based Hydrogels for Tissue Engineering and Soft Tissue Mimicking. Tissue Eng. Part C Methods 2022, 28, 301–313.
- 89. Wu, J.; Pan, Z.; Zhao, Z.Y.; Wang, M.H.; Dong, L.; Gao, H.L.; Liu, C.Y.; Zhou, P.; Chen, L.; Shi, C.J.; et al. Anti-Swelling, Robust, and Adhesive Extracellular Matrix-Mimicking Hydrogel Used as Intraoral Dressing. Adv. Mater. 2022, 34, 2200115.
- 90. ClinicalTrials.gov. Available online: https://clinicaltrials.gov/ct2/results? cond=oral&term=hydrogel&cntry=&state=&city=&dist= (accessed on 26 February 2023).

Retrieved from https://encyclopedia.pub/entry/history/show/99628