Central and Peripheral Nervous System Complications in AFD

Subjects: Neurosciences

Contributor: Antonino Tuttolomondo, Irene Baglio, Renata Riolo, Federica Todaro, Gaspare Parrinello, Salvatore

Miceli, Irene Simonetta

Fabry disease (FD) is a recessive monogenic disease linked to chromosome X due to more than two hundred mutations in the alfa-galactosidase A (GLA) gene. In Anderson–Fabry disease (AFD), deficiency of the enzyme alfa-galactosidase A (α -GalA) leads to an abnormal buildup of globotriaosylceramide (Gb3), which is associated with end-organ damage, progressive organ failure, and subsequent clinical manifestations.

Anderson–Fabry disease

galactosidase-alfa

neurological

peripheral

1. Background

Anderson-Fabry Disease (AFD) represents the second most prevalent glycosphingolipid storage disorder (after Gaucher disease) with a frequency of 1 in 100,000 [1]. In this disease, a deficiency of a lysosomal hydrolase, alfagalactosidase A, causes the abnormal accumulation of uncleaved glycosphingolipids in lysosomes and other organules such as globotriosylceramide (Gb3) and its deacetylated form, globotriasylsphingosine (lyso-Gb3). AFD symptoms reflect the different organ profiles of the histopathological findings of lipid accumulation in the cardiac, renal, and peripheral and central nervous systems. Pain in Anderson-Fabry disease has been suggested as the result of degeneration of nerve fibers in the dorsal root ganglion cells with subsequent axonal degeneration of the tiny fibers involved in pain transmission patterns [2]3. The deposition of glycosphingolipids begins in the lysosomes and causes metabolic collapse of the cells, tissue compensatory hypertrophy, cell death, and organ failure. Lipid deposits are present in the endothelium, media of small vessels, renal tubules and glomeruli, cardiac muscle, conducting fibers, and autonomic ganglia. These histopathological findings have been reported as linked to the clinical results of the disease such as renal failure, cardiomyopathy, pain crisis, and multiple cerebrovascular accidents (CVAs) [4][5]. The accumulation of globotriaosylceramide (Gb3) has been reported as the vital link between pathology and clinical symptoms in most of the involved organs. In fact, Gb3 accumulation occurs in most non-neuronal tissues and body fluids. Only central nervous system symptoms seem to be not due to a direct neuron accumulation but to the epitheliopathy of the small cerebral vessels (SVDs). The natural history of neurological Fabry patients includes transitory cerebral ischemia and strokes, even in very young persons of both genders. The pathogenetic mechanism is due to vascular endothelial accumulation of Gb-3, causing ischemic stroke or white matter lesions (WMLs) [6]. Another potential factor influencing the alteration of endothelial function is associated with Nitric Oxide Synthase-3 genotypes. Nitric oxide, produced by the endothelium, is critical in regulating vessel wall function and maintaining cardiovascular homeostasis. Also, autopsy studies in AFD [7]8] have

reported the accumulation of neuronal globotriaosylceramide in specific cortical and brain stem regions, including the hippocampus. However, the clinical implications and relevance of these findings, as well as potential clinical surrogates, have not yet been explored. Despite this, the primary histological observation in AFD comprises small fiber neuropathy alongside cerebral micro- and macroangiopathy, leading to premature stroke. Cranial MRI demonstrates the presence of progressive white matter lesions (WMLs) at an early age, increased signal intensity in the pulvinar region, and the twisting and enlargement of larger blood vessels. Conventional MRI shows a gradual accumulation of white matter lesions (WMLs) resulting from cerebral vasculopathy during the progression of AFD. The peripheral neuropathy in Fabry disease causes neuropathic pain, reduced cold and warm sensation, and possibly gastrointestinal disturbances. Patients with Fabry disease suffer from pain crises from the end of the first decade of life or during puberty. In children, this pain is often associated with febrile illnesses and reduced heat and exercise tolerance. Patients describe the pain as burning, often associated with deep aches or paresthesia. Some patients also have joint pain. AFD patients may develop neuropsychiatric symptoms, such as depression and neuropsychological deficits. Due to both somatic and psychological impairment, health-related quality of life is considerably reduced in patients with Fabry disease. Targeted screening for Fabry disease among young individuals with stroke may help to reveal unrecognized cases. Furthermore, ischemic stroke is also related to inflammation and arterial stiffness, and no study has addressed this relationship in patients with AFD and cerebrovascular disease, so this topic could represent a possible future line of research [9][10].

2. Molecular Pathogenesis of Anderson–Fabry Disease

In Anderson–Fabry disease (AFD), deficiency of the enzyme alfa-galactosidase A (α-GalA) leads to an abnormal buildup of globotriaosylceramide (Gb3), which is associated with end-organ damage, progressive organ failure, and subsequent clinical manifestations. An important characteristic feature of AFD is the presence of distinct lipid deposits known as "zebra bodies", which are prominently observed in various cell types, particularly at endothelial levels [11]. Previous studies have identified the main constituent of these abnormal deposits as globoside globotriaosylceramide (Gb3), previously referred to as ceramidetrihexoside (CTH) [11]. Moreover, other abnormal glycosphingolipids such as galactosylceramide (Gb2) and blood group B, B1, and P1 antigens, sharing a terminal α -galactosyl moiety, have been described in end-organ damage of AFD patients $\frac{1}{2}$. The molecular pathogenesis of this lipid disorder has been investigated in a prior study, suggesting a potential causal relationship between the deficiency of lysosomal acid alfa-galactosidase activity and the impaired conversion of Gb3 to lactosylceramide (LacCer) [12]. Notably, α-GalA has been shown to play a crucial role in the degradation of the intermediate metabolite globoside Gb3, and the involvement of α-GalB in the metabolism of this metabolite has been reported in several studies $^{[7][13]}$. The α -GalA enzyme is derived from a precursor consisting of 429 amino acids, which transforms to form a homodimer with 398 amino acids $\frac{[8][14]}{14}$. The three N-linked glycans of α -GalA receive mannose-6-phosphate moieties collaborating with the enzyme's arrival to lysosomes by mannose-6-phosphate receptors. The activity of α-GalA toward the lipid substrate is increased by the activator protein saposin B and negatively charged lipids [3]. Over 1000 mutations have been identified in the GLA gene, primarily consisting of missense mutations. However, the complete pathogenic implications of several of these mutations remain unclear. Some α-GalA mutations do not appear to be associated with reduced α-galactosidase activity, leading to uncertainty about their actual pathogenic role. Over and above the cell Gb3-deposits, the end-organ damage in AFD may also be related to the immunoinflammatory mechanism (see Figure 1). Nevertheless, it has been reported that the accumulation of Gb3 [7][8][14] could be associated with specific molecular mechanisms, and early intervention through therapy may prevent the progression of organ failure. Valbuena et al. indicated a crucial pathogenetic role of the overloading of lysosomes with Gb3 and subsequent damage of cytoplasm and subsequent cell death [13]. Additionally, the Gb3 deposits and the resulting organ damage may also be influenced by inflammatory processes [7]. Recently, Gb3 has been reported as potentially identifiable as CD77 [8], which has a direct effect on apoptosis and necrosis [13]. Furthermore, according to Rozenfeld et al., individuals with Anderson-Fabry disease (AFD) exhibit disturbances in leukocyte function when compared to the progressive involvement of other immunocompetent cells, including lymphocytes, monocytes, CD8+ cells, B cells, and dendritic cells [14]. However, another study found no correlation between inflammatory biomarkers such as C-reactive protein and the Mainz Severity Score Index (MSSI), an index used to assess the clinical severity of AFD [15]. Some authors have suggested that there is an immune response against the enzyme, leading to alfa-galactosidase A deficiency. Moore et al. reported a higher degree of neuronal apoptosis due to the neutralization of anti-apoptotic molecules in pediatric AFD [16]. Finally, some authors indicated that Gb3 accumulation may enhance oxidative stress and the production of reactive oxygen species (ROS) [17]. Another interesting point of endothelial function may be due to the Nitric-Oxide-Synthase-3-genotypes. Endothelium-derived nitric oxide plays a crucial role in regulating vessel dilation and maintaining vascular homeostasis. There is a genetic variant of the Nitric-Oxide-Synthase-3 (NOS3) gene that has been identified as a potential factor in disrupting this homeostasis, and it appears to be associated with a reduced thickness of the posterior wall of the left ventricle [18]. This finding may offer insights into the pathogenesis of various cardiac phenotypes observed in Fabry disease. Additionally, Wang et al. reported the presence of Gb3 storage in pulmonary smooth muscle cells and the vascular endothelium of a female patient with Anderson-Fabry disease [19]. This observation further supports the involvement of Gb3 accumulation in the disease's pathogenesis, particularly in vessels. Deacylated globotriaosylceramide (lyso-globotriaosylceramide, lyso-Gb3) has been reported as increased in patients with AFD. Lyso-Gb3 abnormal deposits cause hypertrophy of smooth muscle cells in vitro and hyperplasia of the internal layer of arterioles [20]. The pathogenesis of Anderson-Fabry disease (AFD) involves various cell types, including endothelial and smooth muscle cells, cardiac cells at the myocardial and valvular level, tubular and glomerular cells, as well as podocytes and peripheral nervous cells [21]. Cerebrovascular involvement, particularly affecting perforating arterioles, is a determining factor in morbidity and mortality in individuals with AFD. The pathophysiological mechanisms underlying end-organ damage in AFD are intricate and challenging to describe due to their complexity. The initial clinical manifestations of cerebral AFD primarily involve the microvasculature. As indicated in the aging process, arterial remodeling and intima-media thickening in medium-to-large caliber vessels have been reported as an important step in cerebrovascular complications of AFD [22].

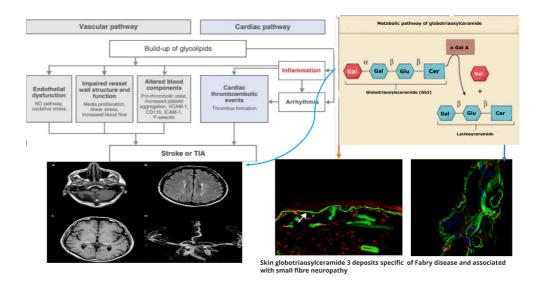


Figure 1. Pathogenetic mechanisms of central and peripheral nervous complications in Anderson-Fabry Disease.

Neurological symptoms in patients with Anderson–Fabry disease (AFD) can be attributed to the involvement of both large and small vessels within the central nervous system. Ischemic cerebral events resulting from large artery involvement may arise from thrombosis of the major intracranial vessels, cardioembolic mechanisms, or possibly atherothrombotic events [23]. In addition to the aforementioned mechanisms, another potential pathogenic factor contributing to cerebrovascular complications in AFD is the presence of distinctive small vessel disease. This implies that multiple pathways and vessel sizes are implicated in the development of neurological symptoms in AFD patients. The clinical pathways of cerebrovascular complications in AFD seem to be the small vessel disease and some neuroimaging patterns appearing as either subcortical stroke or the frequently asymptomatic white matter lesions (WMLs) and subcortical infarcts [23][24][25]. In patients with AFD, strokes occur in both the anterior and the posterior circulatory systems, as well as in cortical and subcortical locations. However, the mechanism and topography of such strokes represent not fully clarified issues because of the paucity of studies addressing this issue in all stroke subtypes as opposed to most studies of cryptogenic stroke

3. Molecular Pathogenesis of Central Nervous System Involvement in Anderson–Fabry Disease

Neurological symptoms associated with Fabry disease encompass both peripheral nervous system and central nervous system involvement. In AFD, peripheral nervous system involvement manifests as severe neuropathic pain, as depicted in **Figure 1** and **Figure 2**. Additionally, other non-central nervous clinical manifestations include autonomic dysfunction, characterized by symptoms such as reduced sweating (hypohidrosis), abdominal pain, intestinal dysmotility disorders, and arrhythmias [28][29]. Autoptic studies revealed Gb3 storage in autonomic ganglions using immunohistochemical staining [30]. At skin biopsy, some studies reported how Fabry patients are characterized by a progressive reduction in intra-epidermal innervation associated with small fiber sensory neuropathy [31]. Although the pathogenetic basis of peripheral neuronal pathology in Fabry disease is understood, mechanisms involving central neurons remain incompletely elucidated. While Gb3 deposits have been observed in ganglion locations, explaining the autonomic and peripheral nervous system involvement [32], no definitive evidence

of Gb3 deposits in central neurons has been reported. However, lyso-Gb3 levels are increased in the plasma and tissues of experimental rat models with AFD and the plasma of male subjects with classical pathogenetic mutations 13. The elevated levels of lyso-Gb3 are a potential pathogenic factor contributing to the pathology of AFD. In fact, lyso-Gb3 plays a role in the painful damage associated with deposits in dorsal root ganglia neurons. Further investigation is needed to understand the pathogenic mechanisms underlying central neuronal involvement in Fabry disease. Recently, Choi et al. reported how the administration of lyso-Gb3 caused a high degree of stimulation of pain-transmitting neurons of normal mice and that lyso-Gb3 caused an increase in Ca2 + influx in not-AFD root ganglion cells cultured from adult mice [33]. Furthermore, in other lysosomal storage disorders, such as Gaucher disease, some authors reported how glucosyl sphingosine (glucopsychosine), an analogue of lyso-Gb3, has a toxic effect on cultured neuronal cells $\frac{[34]}{}$. Patients with Anderson–Fabry disease (AFD) are susceptible to cerebrovascular disease, and this condition is observed more frequently in young individuals [34][35][36]. They may experience ischemic strokes at a higher rate compared to the general population of similar age groups. Furthermore, neuroimaging studies often reveal findings indicative of chronic cerebrovascular disease, which can subsequently lead to cognitive impairment [37][38]. This underscores the importance of monitoring and managing cerebrovascular complications in AFD patients to mitigate the risk of stroke and cognitive decline. Other neurological AFD symptoms due to peripheral and autonomic nervous involvement, such as typical pain, sensory disturbances, and hypohidrosis, have been reported. MRIs of the brain reported ischemic lesions with a higher frequency of cerebellum and brainstem localizations regardless of the presence of neurological signs, whereas T2 MRIs often reveal white matter lesions (WMLs) with hyperintensities which, according to some authors, resemble MRI brain findings characteristic of demyelinating diseases. Some studies have reported that sensory nerves from patients with AFD show several morphological and functional abnormalities [38][39][40], such as lower myelinated and unmyelinated fiber presence, lipid deposits in various cell types, myelin abnormality, and disorders of glial cells. Previous studies reported Gb3 accumulation and swelling of dorsal root ganglia (DRG) neurons in patients [39] and AFD rodent models (38). However, the extent of Gb3 accumulation or other pathologies of peripheral nerves in FD remains unclear. AFD rodent models are a powerful tool for the characterization of nerve pathology. If animal models reproduce human pathology, they could be utilized to understand pain mechanisms for patients with AFD [40][41][42]. In a study [43], the authors reported neuronal damage development of an in vitro model system with a useful model of neuronal functional disturbance in Fabry disease by using short-hairpin RNA to create a stable knock-down of AGA in the human cholinergic neuronal cell line, LA-N-2. The authors reported that these knockdown cellular lines show low levels of AGA activity and Gb3 accumulation. Furthermore, in experimental knock-out cells, the release of neurotransmitter acetylcholine appears to be significantly reduced. This confirms that this experimental model is adequate as a neuronal function model with a disturbance of neurotransmitter release possibly characteristic of AFD. The neuronal pathway involved in the pathogenesis of pain crisis and neuropathic disturbances in AFD is not due to the involvement of peripheral structures such as dorsal root ganglia (DRGs) as well as other nuclear regions of the CNS. In reverse, spinal and supraspinal nuclei and cerebral areas are involved in pain transmission and the anterior cingulate cortex in AFD $\frac{[41][42]}{}$. Gb3 accumulation was documented also in the central nervous system, particularly in the hippocampus and cortical layers [40], further confirmed in the AFD mouse model [41][42]. Moreover, abnormalities of gene expression in AFD have been reported in crucial brain regions that have an important role in the development of the AFD pain pathologic phenotypes, such as prefrontal and sensory

cortices, insular cortex, and basal ganglia circuits. These are all regions that have a direct role in the procession of abnormal pain signals involved in the pathogenesis of chronic neuropathic pain, and maybe also in AFD [40]. Cerebrovascular ischemic events in AFD are the result of cerebral microvessel occlusion, which is associated with progressive wall thickening caused by the accumulation of glycolipids, leading to both thrombotic and non-thrombotic lumen occlusion. Some researchers [44] have reported an abnormality in Gb3 metabolism within central nervous system (CNS) neurons. Additionally, these authors have observed that in AFD, neuronal swelling is likely due to disturbances linked to Gb3 accumulation, particularly in specific nuclei such as the amygdaloid body, the subiculum, and the dorsal vagus nucleus of the medulla oblongata [44]. These findings further support the notion that the pathogenesis of globotriaosylceramide deposits in AFD is not yet fully understood [44].

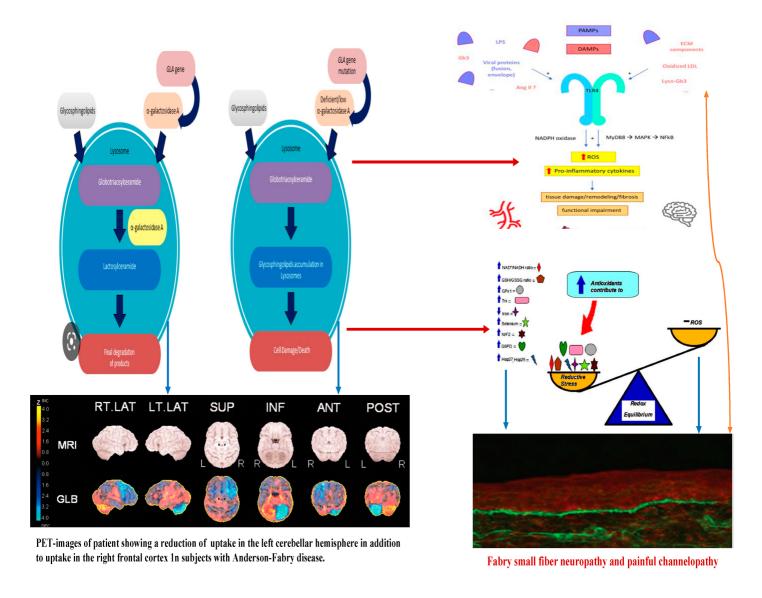


Figure 2. Linkage between metabolic pathogenesis of Anderson-Fabry disease and inflammatory and oxido-reductive pathogenesis of Gb3-related organ damage.

Concerning the pathology of cerebral vessels in AFD, the involvement has been observed [45][46][47][48] of the subarachnoidal arteries of medium size with narrowing of the lumen due to intimal fibrosis mixed with smooth muscle cells (SMC), with membrane abnormalities and stiffening of internal elastic tunica due to the total or partial

change of the medial SMC and subsequent fibrosis and adventitial fibrosis. AFD involves smooth muscle too, but it is not clear whether the first step in Fabry vasculopathy involves endothelial cells, with a subsequent prothrombotic state, or if it begins in the smooth muscle cells of the arterial media layer [45][46][47][48]. It has been reported that lyso-Gb3 plays a crucial role in the pathogenesis of Fabry vasculopathy, and it has been proposed that smooth muscle cells, rather than endothelial cells, represent the main target of cell accumulation. Smooth muscle cells exposed to lyso-Gb3 proliferate, and this proliferation has been reported as linked with the hypertrophy of arterial walls [46][47][48][49][50]. Accumulation of lyso-Gb3 within the media layer of the arteries may also promote cell proliferation, with the fibrotic remodeling of the arterial wall leading to arterial wall stiffness. Shear stress has also an important role in increasing the degree of exposition of angiotensin 1 and 2 receptors in endothelial cells; enhancing the production of reactive oxygen species, NF-κB, β-integrin, and cyclooxygenase 1 and 2 activity; and lowering nitric oxide synthesis (43). All these cellular and biochemical events seem to represent the candidate step to initiate an inflammatory cascade with prothrombotic and pro-inflammatory effects on leukocytes, endothelial cells, and vascular smooth muscle cells [47]. Indeed, inflammatory pathogenesis of Anderson–Fabry disease (AFD) complications in the central nervous system (CNS) has been previously documented [48]. It is important to note that the CNS serves as the primary target for numerous lysosomal storage disorders. In many lysosomal genetic diseases, inflammation mechanisms in the CNS involve microglial cells and astrocytes. Lysosomes, after cellular damage, produce Pathogen Associated Molecular Patterns (PAMPs) or Damage Associated Molecular Patterns (DAMPs) by the astrocytes and microglial cells, using Toll-like receptors T that enhance the cytokine release causing inflammation and cellular death. Ischemic stroke is considered one of the most extensively discussed potential complications of central nervous system (CNS) involvement in Fabry disease. Its pathogenesis involves inflammatory or degenerative occlusive processes affecting the arterial wall or (micro) embolic mechanisms. In the context of stroke pathogenesis, three vascular events related to cerebrovascular structures are influenced by inflammation. These events encompass endothelial cell dysfunction, impaired vessel wall structure and function, as well as alterations in blood components. Indeed, the progressive accumulation of Gb3 in the endothelial cells of intracranial blood vessels has been identified as the primary vasculopathy event associated with ischemic stroke pathogenesis in AFD [22]. Besides Gb3 accumulation, other pathogenetic factors may contribute to the development of ischemic stroke in AFD patients. These factors include possible acquired thrombophilic conditions, abnormalities in intravascular flow velocity (either impaired or increased), autonomic dysfunction [51], and oxidoreductive damage [52]. All these factors collectively play a role in the complex pathogenesis of ischemic strokes in individuals affected by AFD. The role of vascular or autonomic dysfunction as a pathogenic mechanism has been reported as able to impair cerebral blood flow velocities and cerebral autoregulation [51]. In their study, some authors [51] assessed transcranial Doppler sonography in Fabry patients and examined various parameters, including the resistance index, pulsatility index, cerebrovascular resistance, spectral powers of oscillations in RR intervals, mean blood pressure, and mean cerebral blood flow velocities. Their findings indicated a reduction in blood flow velocity, which was attributed to the involvement of certain branches of the middle cerebral artery caused by reduced sympathetic tone and/or progressive arterial stiffening. Additionally, abnormal blood flow oscillations were observed to impair the autoregulation of blood pressure directed to the brain. These observations highlight the potential impact of cerebrovascular changes on the pathophysiology of Fabry disease and its effects

on cerebral blood flow dynamics. Thus, both reduced cerebral blood flow velocities and impaired cerebral autoregulation are likely to be involved in the increased risk of cerebrovascular complications in AFD.

4. Molecular Pathogenesis of Peripheral Nerve Involvement in Anderson–Fabry Disease

Neuropathic pain represents a significant clinical aspect of Anderson–Fabry disease [49][50]. Patients with AFD often experience pain in their hands and feet, along with severe episodic pain attacks known as 'Fabry crises'. This pain is associated with the accumulation of Gb3 in pain-sensitive neurons of the dorsal root ganglia (DRG). The abnormal transmission of pain signals is linked to disturbances in ion channel function [50]. Among these channels, acid-sensing ion channels (ASIC) have been studied in connection with pain [53][54]. In the central nervous system (CNS), ASICs are located in areas highly involved in pain perception. Hyperalgesia, or increased sensitivity to pain, in AFD is attributed to an upregulation of ASIC activity, as observed in animal models and AFD patients [44]. ASIC channels act as proton sensors in the nervous system and play a crucial role in pain transmission. Furthermore, in AFD, elevated levels of Gb3 and lyso-Gb3 have been linked to chronic pain, and this association seems to be closely related to Trpy channels, and potassium, calcium, and sodium channels, which have been extensively studied in DRGs of AFD animal models [55]. In addition, some authors [56] analyzed the pathological nerve findings in AFD rat models [57][58]. They observed a pathological breakdown of Gb3 in lysosomes in AFD, and they correlated peripheral nerve pathology with the accumulation of Gb3 or lysosomes in the axons [57][58]. Morphological abnormalities in peripheral nerves have also been reported in patients with FD [57][58]. These findings shed light on the complex mechanisms underlying the neuropathic pain experienced by individuals with AFD and offer insights into potential targets for therapeutic interventions. Authors [59] who studied the saphenous nerve (sensory), the tibial nerve (mixed sensory/motor) at proximal and distal locations, and the femoral motor branch, reported a significant decrease in myelinated fiber frequency in the saphenous (sensory) and distal tibial nerves (mixed sensory/motor) of AFD rats. Also, a low degree of intra-epidermal nerve fiber density nerve fiber density (IENFD) in patients with AFD [59][60] has been reported. No abnormality in myelinated fiber density has been reported in AFD proximal tibial (mixed sensory/motor) or femoral motor branches [59][60], whereas anatomical abnormality of unmvelinated fiber has been observed in the tibial nerve (mixed sensory/motor) [61]. Authors have further shown abnormality in the density of unmyelinated fibers in the saphenous (sensory) and femoral motor branch nerves and a lower frequency of unmyelinated fiber density in the saphenous (sensory) but not femoral motor branch of AFD nerves. Indeed, a characteristic osmophilic accumulation in myelinated axons of the proximal tibial nerve has been reported [61]. In AFD, some abnormalities concerning myelinated A δ fiber conduction have been reported [61]. In rat models of AFD, 25% of myelinated axons showed significant lipid accumulation that may represent the pathogenetic explanation of myelinated A δ fiber dysfunction observed in patients with AFD [59][60][61]. C-fiber dysfunction has also been reported in patients with AFD with subsequent abnormalities in pain thresholds and heat and cold sensitivity [58]. The finding concerning the axon's diameter in abnormal function of unmyelinated fibers seems to be indicative of altered conduction in the C fibers of AFD peripheral nerves, and this finding is one of the pathogenetic bases of the AFD-characteristic pain crisis. Small sensory nerves, myelinated $A\delta$ fibers, play a main role in transmitting mechanical pain sensitivity, such as unmyelinated C fibers working with warm sensations

and pain sensitivity to heat. In AFD, small fiber disease involves A δ fibers $\frac{[62][63][64]}{[62]}$. Thermal sensation abnormality has been reported as mainly affecting the feet more than the hands with a progressive proximal sequential involvement. The first thermal abnormality involves cold perception (Aδ fibers) more than warmth sensitivity (C fibers), [65] indicating how the thinly myelinated Aδ fibers seem to be more prone to be involved in the Gb3 accumulation peripheral nerve damage [66]. Autonomic involvement in AFD has been described as the cause of gastrointestinal dysmotility (e.g., abdominal cramps, bloating, diarrhea, and nausea), hypohidrosis, abnormality of pupillary constriction, impaired tear and saliva formation, Raynaud phenomena, cardiac rhythm disturbances, and orthostatic hypotension [67][68]. Autonomic dysfunction also regards sudomotor nerve fibers and sweat gland function that have been reported as affected in AFD patients without treatment [69]. Sural nerve bioptic samples showed a characteristic reduction in small myelinated and unmyelinated nerve fibers [70][71]. Glycolipid deposits have been reported in the perineurium, sensory ganglia, vascular smooth muscle cells (SMCs), fibroblasts, and endothelial cells ^[69]. Additional bioptic studies have revealed a significant decline in nerve fibers as individuals age, systemic compromise, and kidney involvement [69]. The first pathogenetic hypothesis is based on the presence of Gb3 deposits in dorsal root ganglion (DRG) neurons driving neuronal damage with a subsequent ganglionopathy resulting in reduced intra-epidermal nerve fiber density (IENFD) [63][72][73]. Gb3 accumulation affects and impairs the function of cellular membrane proteins, such as ion channels, with subsequent abnormalities of excitability leading to cytotoxicity and nervous fiber dysfunction and damage. This hypothesis fits well with the reduction in intra-epidermal nerve fibers in patients with AFD, also found in the skin on the back, which is normally preserved from intra-epidermal fiber loss in length-dependent peripheral neuropathies $\frac{34}{2}$. Another pathogenetic hypothesis is microangiopathy of the vasa nervorum due to an ischemic mechanism caused by Gb3 deposition within the endothelial cells of the blood vessels [74][75]. Also, according to the literature, lyso-Gb3 seems to be a stimulus to SMC proliferation in vitro, and it is involved in the development of vascular pathology in AFD $^{[75]}$. Finally, a plausible hypothesis is linked to an aberration in excitation and signal transmission of neurites in pain-transmitting neurons due to myelin abnormalities [74][75] caused by nerve fiber reduction.

References

- 1. Meikle, P.J.; Hopwood, J.J.; Clague, A.E.; Carey, W.F. Prevalence of lysosomal storage disorders. JAMA 1999, 281, 249–254.
- 2. Kahn, P. Anderson-Fabry disease: A histopathological study of three cases with observations on the mechanism of production of pain. J. Neurol. Neurosurg. Psychiatry 1973, 36, 1053–1062.
- 3. Gemignani, F.; Marbini, A.; Bragaglia, M.M.; Govoni, E. Pathological study of the sural nerve in Fabry's disease. Eur. Neurol. 1984, 23, 173–181.
- 4. Desnick, R.J.; Ionnou, Y.; Eng, C.M. Fabry disease: Alpha galactosidase A deficiency. In The Metabolic and Molecular Bases of Inherited Disease; Scriver, C.H., Beaudet, A.L., Sly, W.S., Valle, D., Eds.; McGraw Hill: New York, NY, USA, 1995; pp. 2741–2784.

- 5. Wise, D.; Wallace, H.J.; Jellinek, E.H. Angiokeratoma corporis diVusum. Q. J. Med. 1962, XXXI, 177–212.
- 6. Desnick, R.J.; Ioannou, Y.A. α-Galactosidase a Deficiency. Fabry Disease. In The Metabolic and Molecular Bases of Inherited Disease, 8th ed.; Scriver, C.R., Beaudet, A.L., Sly, W.S., Valle, D., Eds.; McGraw-Hill: New York, NY, USA, 2001.
- 7. Safyan, R.; Whybra, C.; Beck, M.; Elstein, D.; Altarescu, G. An association study of inflammatory cytokine gene polymorphisms in Fabry disease. Eur. Cytokine Netw. 2006, 17, 271–275.
- 8. Thomaidis, T.; Relle, M.; Golbas, M.; Brochhausen, C.; Galle, P.R.; Beck, M.; Schwarting, A. Downregulation of alpha-galactosidase A upreg- ulates CD77: Functional impact for Fabry nephropathy. Kidney Int. 2009, 75, 399–407.
- 9. Tuttolomondo, A.; Pecoraro, R.; Simonetta, I.; Miceli, S.; Arnao, V.; Licata, G.; Pinto, A. Neurological complications of Anderson-Fabry disease. Curr. Pharm. Des. 2013, 19, 6014–6030.
- 10. Tran, N.; Garcia, T.; Aniqa, M.; Ali, S.; Ally, A.; Nauli, S.M. Endothelial Nitric Oxide Synthase (eNOS) and the Cardiovascular System: In Physiology and in Disease States. Am. J. Biomed. Sci. Res. 2022, 15, 153–177.
- 11. Sweeley, C.C.; Klionsky, B. Fabry's Disease: Classification as a sphingolipidosis and partial characterization of a novel glycolipid. J. Biol. Chem. 1963, 238, 3148–3150.
- 12. Brady, R.O.; Gal, A.E.; Bradley, R.M.; Martensson, E.; Warshaw, A.L.; Laster, L. Enzymatic Defect in Fabry's Disease. N. Engl. J. Med. 1967, 276, 1163–1167.
- 13. Valbuena, C.; Carvalho, E.; Bustorff, M.; Ganhao, M.; Relvas, S.; Nogueira, R.; Carneiro, F.; Oliveira, J.P. Kidney biopsy findings in heterozygous Fabry disease females with early nephropathy. Virchows Arch. 2008, 453, 329–338.
- 14. Rozenfeld, P.; Agriello, E.; De Francesco, N.; Martinez, P.; Fossati, C. Leukocyte perturbation associated with Fabry disease. J. Inherit. Metab. Dis. 2009, 32 (Suppl. S1), S67–S77.
- 15. Altarescu, G.; Chicco, G.; Whybra, C.; Delgado-Sanchez, S.; Sharon, N.; Beck, M.; Elstein, D. Correlation between interleukin-6 pro- moter and C-reactive protein (CRP) polymorphisms and CRP levels with the Mainz Severity Score Index for Fabry disease. J. Inherit. Metab. Dis. 2008, 31, 117–123.
- 16. Moore, D.F.; Goldin, E.; Gelderman, M.P.; Robinson, C.; Baer, J.; Ries, M.; Elkahloun, A.; Brady, R.O.; Schiffmann, R. Apoptotic abnormalities in differential gene expression in peripheral blood mononuclear cells from children with Fabry disease. Acta Paediatr. Suppl. 2008, 97, 48–52.
- 17. Shen, J.S.; Meng, X.L.; Moore, D.F.; Quirk, J.M.; Shayman, J.A.; Schiffmann, R.; Kaneski, C.R. Globotriaosylceramide induces oxidative stress and up-regulates cell adhesion molecule expression in Fabry disease endothelial cells. Mol. Genet. Metab. 2008, 95, 163–168.

- 18. Rohard, I.; Schaefer, E.; Kampmann, C.; Beck, M.; Gal, A. Association between polymorphisms of endothelial nitric oxide synthase gene (NOS3) and left posterior wall thickness (LPWT) of the heart in Fabry disease. J. Inherit. Metab. Dis. 2008, 31 (Suppl. S2), S349–S356.
- 19. Wang, R.Y.; Abe, J.T.; Cohen, A.H.; Wilcox, W.R. Enzyme replacement therapy stabilizes obstructive pulmonary Fabry disease associated with respiratory globotriaosylceramide storage. J. Inherit. Metab. Dis. 2008, 31 (Suppl. S2), S369–S374.
- 20. Aerts, J.M.; Groener, J.E.; Kuiper, S.; Donker-Koopman, W.E.; Strijland, A.; Ottenhoff, R.; van Roomen, C.; Mirzaian, M.; Wijburg, F.A.; Linthorst, G.E.; et al. Elevated globotriaosylsphingosine is a hallmark of abry disease. Proc. Natl. Acad. Sci. USA 2008, 105, 2812–2817.
- 21. Schiffmann, R. Fabry disease. Pharmacol. Ther. 2009, 122, 65-77.
- 22. Moore, D.F.; Kaneski, C.R.; Askari, H.; Schiffmann, R. The cerebral vasculopathy of Fabry disease. J. Neurol. Sci. 2007, 257, 258–263.
- 23. Rombach, S.M.; Twickler, T.B.; Aerts, J.M.; Linthorst, G.E.; Wijburg, F.A.; Hollak, C.E. Vasculopathy in patients with Fabry disease: Current controversies and research directions. Mol. Genet. Metab. 2010, 99, 99–108.
- 24. Namdar, M.; Gebhard, C.; Studiger, R.; Shi, Y.; Mocharla, P.; Schmied, C.; Brugada, P.; Lüscher, T.F.; Camici, G.G. Globotriaosylsphingosine accumulation and not alpha-galactosidase-A deficiency causes endothelial dysfunction in Fabry disease. PLoS ONE 2012, 7, e36373.
- 25. Sestito, S.; Ceravolo, F.; Concolino, D. Anderson-Fabry disease in children. Curr. Pharm. Des. 2013, 19, 6037–6045.
- 26. Sims, K.; Politei, J.; Banikazemi, M.; Lee, P. Stroke in Fabry disease frequently occurs before diagnosis and in the absence of other clinical events: Natural history data from the Fabry Registry. Stroke 2009, 40, 788–794.
- 27. Buechner, S.; Moretti, M.; Burlina, A.P.; Cei, G.; Manara, R.; Ricci, R.; Mignani, R.; Parini, R.; Di Vito, R.; Giordano, G.P.; et al. Central nervous system involvement in Anderson-Fabry disease: A clinical and MRI retrospective study. J. Neurol. Neurosurg. Psychiatry 2008, 79, 1249–1254.
- 28. Saito, S.; Ohno, K.; Sakuraba, H. Fabry-database.org: Database of the clinical phenotypes, genotypes and mutant α -galactosidase A structures in Fabry disease. J. Hum. Genet. 2011, 56, 467–468.
- 29. Cable, W.J.L.; Kolodny, E.H.; Adams, R.D. Fabry disease impaired autonomic function. Neurology 1982, 32, 498.
- 30. Tabira, T.; Goto, I.; Kuroiwa, Y.; Kikuchi, M. Neuropathological and biochemical studies in Fabry's disease. Acta Neuropathol. 1974, 30, 345–354.

- 31. Scott, L.J.C.; Griffin, J.W.; Luciano, C.; Barton, N.W.; Banerjee, T.; Crawford, T.; McArthur, J.C.; Tournay, A.; Schiffmann, R. Quantitative analysis of epidermal innervation in Fabry disease. Neurology 1999, 52, 1249.
- 32. Schiffmann, R. Neuropathy and Fabry disease: Pathogenesis and enzyme replacement therapy. Acta Neurol. Belg. 2006, 106, 61.
- 33. Choi, L.; Vernon, J.; Kopach, O.; Minett, M.S.; Mills, K.; Clayton, P.T.; Meert, T.; Wood, J.N. The Fabry disease-associated lipid Lyso-Gb3 enhances voltage-gated calcium currents in sensory neurons and causes pain. Neurosci. Lett. 2015, 594, 163–168.
- 34. Prado, V.F.; Roy, A.; Kolisnyk, B.; Gros, R.; Prado, M.A.M. Regulation of cholinergic activity by the vesicular acetylcholine transporter. Biochem. J. 2013, 450, 265–274.
- 35. Lücke, T. Fabry disease: Reduced activities of respiratory chain enzymes with decreased levels of energy-rich phosphates in fibroblasts. Mol. Genet. Metab. 2004, 82, 93–97.
- 36. Yamamoto, A.; Abuillan, W.; Burk, A.S.; Körner, A.; Ries, A.; Werz, D.B.; Demé, B.; Tanaka, M. Influence of length and conformation of saccharide head groups on the mechanics of glycolipid membranes: Unraveled by off-specular neutron scattering. J. Chem. Phys. 2015, 142, 154907.
- 37. Park, S.; Kim, J.A.; Joo, K.Y.; Choi, S.; Choi, E.N.; Shin, J.A.; Han, K.H.; Jung, S.C.; Suh, S.H. Globotriaosylceramide leads to KCa3.1 channel dysfunction: A new insight into endothelial dysfunction in Fabry disease. Cardiovasc. Res. 2011, 89, 290–299.
- 38. Schäfer, M.K.-H.; Eiden, L.E.; Weihe, E. Cholinergic neurons and terminal fields revealed by immunohistochemistry for the vesicular acetylcholine transporter. II. The peripheral nervous system. Neuroscience 1998, 84, 361–376.
- 39. Moore, A.M.; Wood, M.D.; Chenard, K.; Hunter, D.A.; Mackinnon, S.E.; Sakiyama-Elbert, S.E.; Borschel, G.H. Controlled delivery of glial cell line-derived neurotrophic factor enhances motor nerve regeneration. J. Hand Surg. Am. 2010, 35, 2008–2017.
- 40. Politei, J.M.; Bouhassira, D.; Germain, D.P.; Goizet, C.; Guerrero-Sola, A.; Hilz, M.J.; Hutton, E.J.; Karaa, A.; Liguori, R.; Üçeyler, N.; et al. Pain in Fabry Disease: Practical Recommendations for Diagnosis and Treatment. CNS Neurosci. Ther. 2016, 22, 568–576.
- 41. Miller, J.J.; Aoki, K.; Mascari, C.A.; Beltrame, A.K.; Sokumbi, O.; North, P.E.; Tiemeyer, M.; Kriegel, A.J.; Dahms, N.M. α-Galactosidase A-deficient rats accumulate glycosphingolipids and develop cardiorenal phenotypes of Fabry disease. FASEB J. 2018, 33, 418–429.
- 42. Burand, A.J., Jr.; Stucky, C.L. Fabry disease pain: Patient and preclinical parallels. Pain 2021, 162, 1305–1321.
- 43. Kaneski, C.R.; Brady, R.O.; Hanover, J.A.; Schueler, U.H. Development of a model system for neuronal dysfunction in Fabry disease. Mol. Genet. Metab. 2016, 119, 144–150.

- 44. Sluka, K.A.; Winter, O.C.; Wemmie, J.A. Acid-sensing ion channels: A new target for pain and CNS diseases. Curr. Opin. Drug Discov. Devel. 2009, 12, 693–704.
- 45. DeGraba, T.; Azhar, S.; Dignat-George, F.; Brown, E.; Boutière, B.; Altarescu, G.; McCarron, R.; Schiffmann, R. Profile of endothelial and leukocyte activation in Fabry patients. Ann. Neurol. 2000, 47, 229–233.
- 46. van Breemen, M.J.; Rombach, S.M.; Dekker, N.; Poorthuis, B.J.; Linthorst, G.E.; Zwinderman, A.H.; Breunig, F.; Wanner, C.; Aerts, J.M.; Hollak, C.E. Reduction of elevated plasma globotriaosylsphingosine in patients with classic Fabry disease following enzyme replacement therapy. Biochim. Biophys. Acta 2011, 1812, 70–76.
- 47. Zampetti, A.; Gnarra, M.; Borsini, W.; Giurdanella, F.; Antuzzi, D.; Piras, A.; Smaldone, C.; Pieroni, M.; Cadeddu, C.; de Waure, C.; et al. Vascular endothelial growth factor (VEGF-a) in Fabry disease: Association with cutaneous and systemic manifestations with vascular involvement. Cytokine 2013, 61, 933–939.
- 48. Klug, K.; Spitzel, M.; Hans, C.; Klein, A.; Schottmann, N.M.; Erbacher, C.; Üçeyler, N. Endothelial Cell Dysfunction and Hypoxia as Potential Mediators of Pain in Fabry Disease: A Human-Murine Translational Approach. Int. J. Mol. Sci. 2023, 24, 15422.
- 49. Biegstraaten, M.; Hollak, C.E.; Bakkers, M.; Faber, C.G.; Aerts, J.M.; van Schaik, I.N. Small fiber neuropathy in Fabry disease. Mol. Genet. Metab. 2012, 106, 135–141.
- 50. Schuller, Y.; Linthorst, G.E.; Hollak, C.E.; Van Schaik, I.N.; Biegstraaten, M. Pain management strategies for neuropathic pain in Fabry disease--a systematic review. BMC Neurol. 2016, 16, 25.
- 51. Hilz, M.J.; Kolodny, E.H.; Brys, M.; Stemper, B.; Haendl, T.; Marthol, H. Reduced cerebral blood flow velocity and impaired cerebral autoregulation in patients with Fabry disease. J. Neurol. 2004, 251, 564–570.
- 52. Moore, D.F.; Ye, F.; Brennan, M.; Gupta, S.; Barshop, B.A.; Steiner, R.D.; Rhead, W.J.; Brady, R.O.; Hazen, S.L.; Schiffmann, R. Ascorbate decreases fabry cerebral hyperperfusion suggesting a reactive oxygen species abnormality: An arterial spin tagging study. J. Magn. Reson. Imaging 2004, 20, 674–683.
- 53. Geevasinga, N.; Tchan, M.; Sillence, D.; Vucic, S. Upregulation of inward rectifying currents and Fabry disease neuropathy. J. Peripher. Nerv. Syst. 2012, 17, 399–406.
- 54. Castellanos, L.C.S.; Rozenfeld, P.; Gatto, R.G.; Reisin, R.C.; Uchitel, O.D.; Weissmann, C. Upregulation of ASIC1a channels in an in vitro model of Fabry disease. Neurochem. Int. 2020, 140, 104824.
- 55. Hofmann, L.; Hose, D.; Grießhammer, A.; Blum, R.; Döring, F.; Dib-Hajj, S.; Waxman, S.; Sommer, C.; Wischmeyer, E.; Üçeyler, N. Characterization of small fiber pathology in a mouse model of Fabry disease. Elife 2018, 7, e39300.

- 56. Waltz, T.B.; Burand, A.J., Jr.; Sadler, K.E.; Stucky, C.L. Sensory-specific peripheral nerve pathology in a rat model of Fabry disease. Neurobiol. Pain 2021, 10, 100074.
- 57. Kocen, R.S.; Thomas, P.K. Peripheral Nerve Involvement in Fabry's Disease. Arch. Neurol. 1970, 22, 81–88.
- 58. Torvin Møller, A.; Winther Bach, F.; Feldt-Rasmussen, U.; Rasmussen, A.; Hasholt, L.; Lan, H.; Sommer, C.; Kølvraa, S.; Ballegaard, M.; Staehelin Jensen, T. Functional and structural nerve fiber findings in heterozygote patients with Fabry disease. Pain 2009, 145, 237–245.
- 59. Uceyler, N.; Ganendiran, S.; Kramer, D.; Sommer, C. Characterization of pain in fabry disease. Clin. J. Pain 2014, 30, 915–920.
- 60. Burlina, A.P.; Sims, K.B.; Politei, J.M.; Bennett, G.J.; Baron, R.; Sommer, C.; Møller, A.T.; Hilz, M.J. Early diagnosis of peripheral nervous system involvement in Fabry disease and treatment of neuropathic pain: The report of an expert panel. BMC Neurol. 2011, 11, 61.
- 61. Üçeyler, N.; Kahn, A.K.; Kramer, D.; Zeller, D.; Casanova-Molla, J.; Wanner, C.; Weidemann, F.; Katsarava, Z.; Sommer, C. Impaired small fiber conduction in patients with Fabry disease: A neurophysiological case-control study. BMC Neurol. 2013, 13, 47.
- 62. Dütsch, M.; Marthol, H.; Stemper, B.; Brys, M.; Haendl, T.; Hilz, M.J. Small fiber dysfunction predominates in Fabry neuropathy. J. Clin. Neurophysiol. 2002, 19, 575–586.
- 63. Üçeyler, N.; He, L.; Schönfeld, D.; Kahn, A.K.; Reiners, K.; Hilz, M.J.; Breunig, F.; Sommer, C. Small fibers in Fabry disease: Baseline and follow-up data under enzyme replacement therapy. J. Peripher. Nerv. Syst. 2011, 16, 304–314.
- 64. Politei, J.M.; Pagano, M.A. Peripheral neuropathy in AndersonFabry disease: Its physiology, evaluation and treatment. Rev. Neurol. 2004, 38, 979–983.
- 65. Siedler, G.; Káhn, A.K.; Weidemann, F.; Wanner, C.; Sommer, C.; Üçeyler, N. Dyshidrosis is associated with reduced amplitudes in electrically evoked pain-related potentials in women with Fabry disease. Clin. Neurophysiol. 2019, 130, 528–536.
- 66. Schiffmann, R.; Scott, L.J. Pathophysiology and assessment of neuropathic pain in Fabry disease. Acta Paediatr. Suppl. 2002, 91, 48–52.
- 67. Hilz, M.J.; Koehn, J.; Kolodny, E.H.; Brys, M.; Moeller, S.; Stemper, B. Metronomic breathing shows altered parasympathetic baroreflex function in untreated Fabry patients and baroreflex improvement after enzyme replacement therapy. J. Hypertens. 2011, 29, 2387–2394.
- 68. deVeber, G.A.; Schwarting, G.A.; Kolodny, E.H.; Kowall, N.W. Fabry disease: Immunocytochemical characterization of neuronal involvement. Ann. Neurol. 1992, 31, 409–415.
- 69. Lao, L.-M.; Kumakiri, M.; Mima, H.; Kuwahara, H.; Ishida, H.; Ishiguro, K.; Fujita, T.; Ueda, K. The ultrastructural characteristics of eccrine sweat glands in a Fabry disease patient with

- hypohidrosis. J. Dermatol. Sci. 1998, 18, 109–117.
- 70. Lim, S.N.; Huang, C.C.; Kuo, H.C.; Hsieh, Y.C.; Chu, C.C. Subtle Changes in Cutaneous Nerves and Sural Nerve Biopsy in a Patient With Fabry's Disease. J. Clin. Neuromuscul. Dis. 2005, 7, 19–24.
- 71. Gayathri, N.; Yasha, T.; Kanjalkar, M.; Agarwal, S.; Sagar, B.C.; Santosh, V.; Shankar, S. Fabry's disease: An ultrastructural study of nerve biopsy. Ann. Indian Acad. Neurol. 2008, 11, 182–184.
- 72. Onishi, A.; Dyck, P.J. Loss of small peripheral sensory neurons in Fabry disease. Histologic and morphometric evaluation of cutaneous nerves, spinal ganglia, and posterior columns. Arch. Neurol. 1974, 31, 120–127.
- 73. Toyooka, K.; Said, G. Nerve biopsy findings in hemizygous and heterozygous patients with Fabry's disease. J. Neurol. 1997, 244, 464–468.
- 74. Brakch, N.; Dormond, O.; Bekri, S.; Golshayan, D.; Correvon, M.; Mazzolai, L.; Steinmann, B.; Barbey, F. Evidence for a role of sphingosine-1 phosphate in cardiovascular remodelling in Fabry disease. Eur. Heart J. 2010, 31, 67–76.
- 75. Tomé, F.M.; Fardeau, M.; Lenoir, G. Ultrastructure of muscle and sensory nerve in Fabry's disease. Acta Neuropathol. 1977, 38, 187–194.

Retrieved from https://encyclopedia.pub/entry/history/show/122589