Metschnikowia pulcherrima

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Yeasts affiliated with the *Metschnikowia pulcherrima* clade (subclade) of the large ascomycetous genus *Metschnikowia* frequently turn out to produce the characteristic maroon-red pulcherrimin when tested for pigment production and prove to exert antagonistic effects on many types of microorganisms. The determination of the exact taxonomic position of the strains is hampered by the shortage of distinctive morphological and physiological properties of the species of the clade and the lack of rDNA barcode gaps. The rDNA repeats of the type strains of the species are not homogenized and are assumed to evolve by a birth-and-death mechanism combined with reticulation. The taxonomic division is further hampered by the incomplete biological (reproductive) isolation of the species: certain type strains can be hybridized and genome sequencing revealed chimeric genome structures in certain strains that might have evolved from interspecies hybrids (alloploid genome duplication). Various mechanisms have been proposed for the antimicrobial antagonism. One is related to pulcherrimin production. The diffusible precursor of pulcherrimin, the pulcherriminic acid is secreted by the cells into the environment where it forms the insoluble pulcherrimin with the ferric ions. The lack of free iron caused by the immobilization of ferric ions inhibits the growth of many microorganisms.

Keywords: yeast ; antimicrobial antagonism ; bioprotection ; wine ; pulcherrimin ; rDNA reticulation ; iron immobilisation

1. Definition

Metschnikowia Kamienski (1899) is a large ascomycetous genus currently comprising 79 species (Mycobank, 04. 2020) but the number or species is continuously growing. The *M. pulcherrima* clade of the genus contains seven validly described species that share the ability to produce pulcherrimin, a maroon-red pigment (reviewed in ^{[1][2]}). These species and the strains closely related to them have broad biotechnological potential for application in various industrial processes. In wine fermentation, these yeasts can modulate the population dynamics of the fermenting yeast communities and produce enzymes and a broad range of compounds that improve the aromatic complexity of the wine (for a review, see ^[3]).

2. Introduction

Since the mechanisms underlying their antagonistic effect are not associated with the production of toxic compounds, these strains can safely be used as bioprotective agents to curb the invasion of pathogenic and rotting (saprophytic) microorganisms (Figure 1D) and/or additives in food technologies to modulate the dynamics of microbial populations. Numerous technological innovations involving antagonistic *Metschnikowia* strains have been patented (e.g., JPH01117778A, 1989; US6991930B1, 2006; NZ528225A, 2008; P0800775, 2008; ITTO20070655A1, 2009; WO2010149370, 2010; WO2010149369, 2010; CN101946805A 2011; CN103642705A, 2014; EP3266305A1, 2018; CN107904180A, 2018; CN110684678A, 2020;) and several *Metschnikowia*-based products have been commercialiZed [Excellence Bio-Nature (Lamothe-Abiet), Flavia and Gaïa (Lallemand), Shemer (Bayer, Koppert Biological Systems) Zymaflore Egide (Laffort)] as ADYs (Active Dry Yeast) for inoculated fermentation or as biocontrol agents for application against plant pathogens and post-harvest plant diseases.

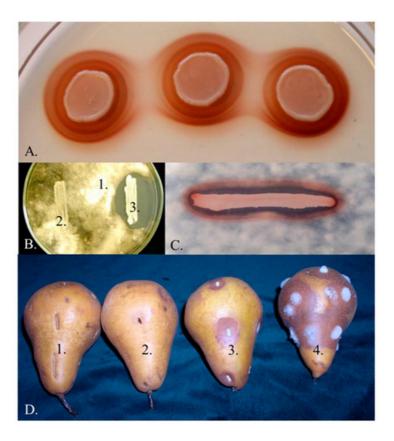


Figure 1. Pigment production and antagonism. (**A**) Pigmented halos around the colonies of the *Metschnikowia* strain 02.11.1.21. (**B**) Inhibition zone around the colony of the same strain (1: *Botrytis cinerea* 3318, 2: *Saccharomyces cerevisiae* S288c, 3: *Metschnikowia* 02.11.1.21). (**C**) Coincidence of pigmented halo and inhibition zone around the colony of *Metschnikowia* 02.11.1.21 on a medium supplemented with 0.005 mg/mL FeCl₃ and flooded with conidia of *B. cinerea* 3318. (**D**) Inhibition of rotting caused by *Botrytis* (1: untreated, 2: dipped in a suspension *Metschnikowia* 02.11.1.21 cells, 3: dipped in a mixed suspension of *B. cinerea* 3318 conidia and *Metschnikowia* 02.11.1.21 cells, 4: dipped in a suspension of *B. cinerea* 3318 conidia). See reference ^[4] for the description of the strains.

Over the past two decades, large numbers of strains isolated from various substrates have been assigned to one or the other of these species (preferentially to M. pulcherrima) on the basis of barcode sequence (preferentially the D1/D2 domains of the LSU rRNA genes and the ITS1-5.8S-ITS2 segments of the rDNA repeats) identities/similarities. The general practice of sequence-based strain identification is a search with the sequence of the strain in nucleotide databases for identical/similar sequences and assigning the strain to the species whose database sequence is found most similar. The sequence of the strain is then routinely deposited in the database (most journals request accession numbers) under this taxonomic name usually without an expert taxonomic verification. Since small sequence differences are usually tolerated during identification, the new entries will gradually fill up the barcode gaps separating the closely related species; the species boundaries gradually become fuzzy. Thus, the taxonomic identification of new isolates by comparing their rDNA sequences with those deposited in databases can easily lead to false results. In addition to this general problem, other difficulties can also arise when pulcherrimin-producing Metschnikowia strains are to be identified taxonomically. The results of the in-depth analyses of the rDNA repeats of certain type strains [5][6], the hybridization of type strains with eachother ^[6] and the analysis of genome sequences (e.g., [2][8][9]) raised doubts as to whether the taxonomic division of the *M*. pulcherrima clade is correct at all. The incongruences around the mechanism of the antimicrobial antagonism pose another problem. While many researchers associate it with iron depletion, others prefer the view that non-iron-related mechanisms similar to those known in other antagonistic microorganisms are also involved or even play the major role.

3. The M. pulcherrima Clade

Pulcherrimin-producing *Metschnikowia* strains are common components of the yeast communities that colonise ripening fruits, flowers (nectar), tree sap fluxes and also frequently occur in fruit juices and fermenting wine (e.g., $^{[2][3][4][10][11][12][13]}$ [14][15][16][17][18][19][20][21][22][23][24][25][26]). New yeast isolates producing the characteristic maroon-red pulcherrimin halos around their colonies (Figure 1A) are frequently declared to belong to *M. pulcherrima* without taking into account that *M. pulcherrima* is not the only pigment-producing *Metschnikowia* species. Over the past two decades, five additional species (*M. andauensis, M. rubicola, M. shanxiensis, M. sinensis, M. zizyphicola*) were validly described and *M. fructicola*, originally described as a pigment-less species has also turned out to produce pulcherrimin (for a review, see ^[1]). The

intensity of pigment production is variable and highly dependent on the culturing conditions ^{[1][33][34]} and probably also on ploidy. The phylogenetic analysis of the barcode sequences of the type strains of the genus clustered these species in a group designated *M. pulcherrima* clade ^{[1][2]}.

Recently, two additional species, M. persimmonesis and M. citriensis were proposed to accommodate pulcherriminproducing strains. The taxonomic name *M. persimmonesis* was proposed for a single Korean isolate but without providing a complete taxonomic description [27]. The phylogenetic position of the strain is uncertain because its different rDNA barcode sequences (D1/D2, ITS and 18S) show the highest similarities to sequences of the type strains of different species. *M. citriensis* is based on two strains isolated from citrus leaves ^[30]. The taxonomic position of these strains is also somewhat obscure, because the authors found them closely related to M. koreensis based on the neighbour-joining analysis of the D1/D2 domains of the 26S rRNA genes, but the M. koreenesis sequence used in the analysis was a direct GenBank submission amplified from a strain for which no taxonomic description is available. The other closest relatives were strains of three pigment-producing members of the *M. pulcherrima* clade and the non-pigmented *M. chrysoperlae*, but only one sequence used in the phylogenetic analysis represented a type strain. As previous analyses found M. koreensis separated by a large phylogenetic distance from the *M. pulcherrima* clade ^{[1][33]}, the proposed simultaneous close relationship to M. koreensis and the M. pulcherrima clade needs to be revised or reinforced by the analysis or more sequences. Interestingly, when the ITS sequences are examined, the M. citriensis type strain differ more from the other M. citriensis strain than from the *M. persimmonesis* type strain. Besides, both the D1/D2 and the ITS sequences were cloned, and a phylogenetic analysis based on cloned sequences can easily be misleading in this group of yeast species because of the very high intragenomic diversity of the rDNA repeats (see below). The formation of spheroidal ascospores is another problematic feature of these isolates. M. pulcherrima and its relatives have needle-shape spores [33]. Because of these uncertainties, further examination is required to validate the status of *M. persimmonesis* and *M. citriensis* as distinct species. Nevertheless, most properties of their strains and the results of the sequence analyses indicate taxonomic affinity with the M. pulcherrima clade. Many pulcherrimin-producing isolates were not identified at the species level or could not be assigned to any species and were therefore only classified as Metschnikowia sp., M. aff. pulcherrima or M. aff. fructicola. On the other hand, many strains have been classified into these species without presenting sufficient taxonomic evidence. Since pigmentation is an irrelevant property in most biotechnological processes, the strains isolated for industrial purposes are normally not tested for pulcherrimin production. Therefore and because of the sensitivity of pulcherrimin synthesis to the culturing conditions, it is unknown whether pigmentation is a general ability of all strains of the clade.

References

- 1. Lachance, M.A. Metschnikowia: Half tetrads, a regicide and the fountain of youth. Yeast 2016, 33, 563–574.
- Kurtzman, C.P.; Robnett, C.J.; Basehoar, E.; Ward, T.J. Four new species of Metschnikowia and the transfer of seven Candida species to Metschnikowia and Clavispora as new combinations. Antonie Van Leeuwenhoek 2018, 111, 2017– 2035.
- 3. Morata, A.; Loira, I.; Escott, C.; del Fresno, J.M.; Bañuelos, M.A.; Suárez-Lepe, J.A. Applications of Metschnikowia pulcherrima in wine biotechnology. Fermentation 2019, 5, 63.
- 4. Sipiczki, M. Metschnikowia strains isolated from botrytized grapes antagonize fungal and bacterial growth by iron depletion. Appl. Environ. Microbiol. 2006, 72, 6716–6724.
- 5. Sipiczki, M.; Pfliegler, W.P.; Holb, I.J. Metschnikowia species share a pool of diverse rRNA genes differing in regions that determine hairpin-loop structures and evolve by reticulation. PLoS ONE 2013, 8, e67384.
- Sipiczki, M.; Horvath, E.; Pfliegler, W.P. Birth-and-death evolution and reticulation of ITS segments of Metschnikowia andauensis and Metschnikowia fructicola rDNA repeats. Front. Microbiol. 2018, 9, 1193.
- Piombo, E.; Sela, N.; Wisniewski, M.; Hoffmann, M.; Gullino, M.L.; Allard, M.W.; Levin, E.; Spadaro, D.; Droby, S. Genome sequence, assembly and characterization of two Metschnikowia fructicola strains used as biocontrol agents of postharvest diseases. Front. Microbiol. 2018, 9, 593.
- Venkatesh, A.; Murray, A.L.; Boyle, A.B.; Quinn Farrington, L.; Maher, T.J.; Ó'Gaora, P.; Wolfe, K.H.; O'Brien, C.E.; Butler, G. Draft genome sequence of a highly heterozygous yeast strain from the Metschnikowia pulcherrima subclade, UCD127. Genome Announc. 2018, 6, e00550-18.
- Gore-Lloyd, D.; Sumann, I.; Brachmann, A.O.; Schneeberger, K.; Ortiz-Merino, R.A.; Moreno-Beltrán, M.; Schläfli, M.; Kirner, P.; Santos Kron, A.; Rueda-Mejia, M.P.; et al. Snf2 controls pulcherriminic acid biosynthesis and antifungal activity of the biocontrol yeast Metschnikowia pulcherrima. Mol. Microbiol. 2019, 112, 317–332.

- 10. Kántor, A.; Hutková, J.; Petrová, J.; Hleba, L.; Kačániová, M. Antimicrobial activity of pulcherrimin pigment produced by Metschnikowia pulcherrima against various yeast species. J. Microbiol. Biotechnol. Food Sci. 2015, 5, 282–285.
- 11. Pawlikowska, E.; James, S.A.; Breierova, E.; Antolak, H.; Kregiel, D. Biocontrol capability of local Metschnikowia sp. isolates. Antonie Van Leeuwenhoek 2019, 112, 1425–1445.
- Molnar, O.; Prillinger, H. Analysis of yeast isolates related to Metschnikowia pulcherrima using the partial sequences of the large subunit rDNA and the actin gene; description of Metschnikowia andauensis sp. nov. Syst. Appl. Microbiol. 2005, 28, 717–726.
- Xue, M.L.; Zhang, L.Q.; Wang, Q.M.; Zhang, J.S.; Bai, F.Y. Metschnikowia sinensis sp. nov., Metschnikowia zizyphicola sp. nov. and Metschnikowia shanxiensis sp. nov., novel yeast species from jujube fruit. Int. J. Syst. Evol. Microbiol. 2006, 56, 2245–2250.
- 14. Sipiczki, M. Overwintering of vineyard yeasts: Survival of interacting yeast communities in grapes mummified on vines. Front. Microbiol. 2016, 7, 212.
- 15. Türkel, S.; Ener, B. Isolation and characterization of new Metschnikowia pulcherrima strains as producers of the antimicrobial pigment pulcherrimin. Z. Naturforsch. C J. Biosci. 2009, 64, 405–410.
- 16. Zhang, D.; Spadaro, D.; Garibaldi, A.; Gullino, M.L. Selection and evaluation of new antagonists for their efficacy against postharvest brown rot of peaches. Postharvest Biol. Technol. 2009, 55, 174–181.
- 17. Vadkertiova, R.; Molnarova, J.; Vranova, D.; Slavikova, E. Yeasts and yeast-like organisms associated with fruits and blossoms of different fruit trees. Can. J. Microbiol. 2012, 58, 1344–1352.
- 18. Csutak, O.; Vassu, T.; Cornea, P.; Grebenisan, I. Genetic characterization of two new Metschnikowia strains with antifungal activity. Rom. Biotech. Lett. 2007, 12, 3175–3182.
- 19. Csutak, O.; Vassu, T.; Sarbu, I.; Stoica, I.; Cornea, P. Antagonistic activity of three newly isolated yeast strains from the surface of fruits. Food Technol. Biotechnol. 2013, 51, 70–77.
- 20. Hadwiger, L.A.; McDonel, H.; Glawe, D. Wild yeast strains as prospective candidates to induce resistance against potato late blight (Phytophthora infestans). Am. J. Potato Res. 2015, 92, 379–386.
- 21. Kang, Y.M.; Choi, J.E.; Komakech, R.; Park, J.H.; Kim, D.W.; Cho, K.M.; Kang, S.M.; Choi, S.H.; Song, K.C.; Ryu, C.M.; et al. Characterization of a novel yeast species Metschnikowia persimmonesis KCTC 12991BP (KIOM G15050 type strain) isolated from a medicinal plant, Korean persimmon calyx (Diospyros kaki Thumb). AMB Express 2017, 7, 199.
- 22. Pawlikowska, E.; Kręgiel, D. Enzymatic profiles and antimicrobial activity of the yeast Metschnikowia pulcherrima. Acta Innov. 2017, 23, 17–24.
- 23. Liu, Y.; Wang, W.; Zhou, Y.; Yaoa, S.; Denga, L.; Zeng, K. Isolation, identification and in vitro screening of Chongqing orangery yeasts for the biocontrol of Penicillium digitatum on citrus fruit. Biol. Control 2017, 110, 18–24.
- 24. Liu, Y.; Yao, S.; Deng, L.; Ming, J.; Zeng, K. Metschnikowia citriensis sp. nov., a novel yeast species isolated from leaves with potential for biocontrol of postharvest fruit rot. Biol. Control 2018, 125, 15–19.
- 25. Tian, Y.Q.; Li, W.; Jiang, Z.T.; Jing, M.M.; Shao, Y.Z. The preservation effect of Metschnikowia pulcherrima yeast on anthracnose of postharvest mango fruits and the possible mechanism. Food. Sci. Biotechnol. 2017, 27, 95–105.
- 26. Barbosa, C.; Lage, P.; Esteves, M.; Chambel, L.; Mendes-Faia, A.; Mendes-Ferreira, A. Molecular and phenotypic characterization of Metschnikowia pulcherrima strains from Douro wine region. Fermentation 2018, 4, 8.
- 27. Lachance, M.A. Metschnikowia Kamienski (1899). In The Yeasts. A Taxonomic Study; Kurtzman, C.P., Fell, J.W., Boekhout, T., Eds.; Elsevier: Amsterdam, The Netherlands, 2011; pp. 575–620.
- Genç, T.T. Effects of various environmental conditions on pulcherrimin production and extracellular enzyme profiles of Metschnikowia pulcherrima. SAR J. 2020, 3, 10–16.
- 29. Liu, Y.; Wang, W.; Zhou, Y.; Yaoa, S.; Denga, L.; Zeng, K. Isolation, identification and in vitro screening of Chongqing orangery yeasts for the biocontrol of Penicillium digitatum on citrus fruit. Biol. Control 2017, 110, 18–24.
- 30. Liu, Y.; Yao, S.; Deng, L.; Ming, J.; Zeng, K. Metschnikowia citriensis sp. nov., a novel yeast species isolated from leaves with potential for biocontrol of postharvest fruit rot. Biol. Control 2018, 125, 15–19.
- 31. Tian, Y.Q.; Li, W.; Jiang, Z.T.; Jing, M.M.; Shao, Y.Z. The preservation effect of Metschnikowia pulcherrima yeast on anthracnose of postharvest mango fruits and the possible mechanism. Food. Sci. Biotechnol. 2017, 27, 95–105.
- 32. Barbosa, C.; Lage, P.; Esteves, M.; Chambel, L.; Mendes-Faia, A.; Mendes-Ferreira, A. Molecular and phenotypic characterization of Metschnikowia pulcherrima strains from Douro wine region. Fermentation 2018, 4, 8.

- 33. Lachance, M.A. Metschnikowia Kamienski (1899). In The Yeasts. A Taxonomic Study; Kurtzman, C.P., Fell, J.W., Boekhout, T., Eds.; Elsevier: Amsterdam, The Netherlands, 2011; pp. 575–620.
- 34. Genç, T.T. Effects of various environmental conditions on pulcherrimin production and extracellular enzyme profiles of Metschnikowia pulcherrima. SAR J. 2020, 3, 10–16.

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