# *Cannabis sativa* Bioactive Compounds in Colorectal Cancer

Subjects: Chemistry, Medicinal Contributor: Rita Silva-Reis, Artur M. S. Silva, Paula A. Oliveira, Susana M. Cardoso

*Cannabis sativa* is a multipurpose plant that has been used in medicine for centuries. Considerable research has focused on the bioactive compounds of this plant, particularly cannabinoids and terpenes. Among other properties, these compounds exhibit antitumor effects in several cancer types, including colorectal cancer (CRC). Cannabinoids show positive effects in the treatment of CRC by inducing apoptosis, proliferation, metastasis, inflammation, angiogenesis, oxidative stress, and autophagy. Terpenes, such as  $\beta$ -caryophyllene, limonene, and myrcene, have also been reported to have potential antitumor effects on CRC through the induction of apoptosis, the inhibition of cell proliferation, and angiogenesis. In addition, synergy effects between cannabinoids and terpenes are believed to be important factors in the treatment of CRC.

Keywords: Cannabis sativa ; colorectal cancer ; cannabinoids ; terpenes ; apoptosis ; proliferation ; metastasis ; inflammation ; angiogenesis ; oxidative stress

# 1. Introduction

Cannabis is one of humanity's oldest plants, yet it has also been a topic of discussion throughout history <sup>[1]</sup>. This plant is psychotropic and includes about 500 distinct chemical components, the most important of which are cannabinoids. The *Cannabis indica* Lam, *Cannabis ruderalis* Janisch, and *Cannabis sativa* Linnaeus are the species with the most psychotropic secondary metabolites. *C. ruderalis* yields smaller amounts of  $\Delta^9$ -tetrahydrocannabinol ( $\Delta^9$ -THC) and is less used in medicine, while *C. indica* is typically employed as a sedative, and *C. Sativa* as a psychoactive <sup>[2]</sup>. Among them, *C. sativa* is the most widely cultivated and exploited for a variety of purposes <sup>[3]</sup>.

Cannabis for medicinal uses has recently been legalized in many countries. Plant material and extracts can be used to alleviate chronic pain and muscle spasms, reduce nausea during chemotherapy, improve appetite in HIV/AIDS patients, improve sleep, and reduce tics in Tourette's syndrome patients. Moreover, it can be used in extreme cases of anorexia, arthritis, glaucoma, and inflammatory bowel disease <sup>[4]</sup>. The bulk of medicinal compounds is found in feminine inflorescences, and medicinal properties are commonly attributed to cannabinoids, although other bioactive substances, including terpenes, may also contribute to the health effects of cannabis <sup>[5]</sup>. According to the entourage effect theory, the medicinal benefits of cannabis are increased when all the plant's constituents, such as terpenes, flavonoids, and cannabinoids, are present and interact with each other. Some researchers believe that the entourage effect may be especially important when it comes to the potential use of cannabis for cancer treatment <sup>[6]</sup>. In fact, cannabinoids and terpenes are known to have a whole range of potential health benefits, ranging from pain relief <sup>[7][8]</sup> to anti-inflammatory properties <sup>[9][10]</sup>. Thus, the high interest in these compounds is related to their numerous pharmacological properties. Regardless, this topic is far from being well understood.

# 2. Bioactive Compounds of Cannabis sativa

Among the multiple bioactive compounds found in *C. sativa*, the main ones are cannabinoids, terpenoids, flavonoids, stilbenoids, and alkaloids [11][12]. When consumed, these substances can induce a variety of beneficial health effects and are thought to contribute to the plant's therapeutic qualities. As for natural products in general, the phytochemical content of *C. sativa* varies, depending on distinct factors, including genetics, growing conditions, stage of growth, harvest time, processing, and storage, among others [13]. *C. ruderalis* and *C. indica* contain a smaller amount of cannabidiol (CBD) than *C. sativa*. In contrast, *C. indica* has the largest THC content compared to *C. sativa*, and *C. ruderalis* has the lowest [2].

#### 2.1. Cannabinoids

Cannabinoids are a type of terpenophenolic compounds with a C<sub>21</sub> backbone <sup>[14]</sup>. The diversity of chemical structures of phytocannabinoids is mainly due to the differences between the isoprenyl groups, the side chain and the resorcinyl core <sup>[15]</sup>. Cannabinoids, according to their chemical structure, can be classified into 11 different classes: cannabigerol (CBG), cannabidiol (CBD), cannabichromene (CBC),  $\Delta^9$ -THC,  $\Delta^8$ -THC, cannabicyclol (CBL), cannabinol (CBN), cannabinoidol (CBND), cannabitriol (CBT), and miscellaneous-type cannabinoids <sup>[14][16]</sup>.

Most cannabinoids originate from cannabigerolic acid (CBGA), as revealed in **Figure 1**. Its biosynthesis occurs with the production of CBGA from the alkylation of olivetolic acid with geranyl pyrophosphate (GPP). The primary cannabinoids, cannabichromenic acid (CBCA), cannabidiolic acid (CBDA), and  $\Delta^9$ -tetrahydrocannabinolic acid ( $\Delta^9$ -THCA), are synthesized in acid form by the enzymes CBCA synthase, CBDA synthase, and THCA synthase. Decarboxylation then converts the acidic forms of cannabinoids into neutral forms (THC, CBD, and CBC), which are recognized for having higher pharmacological potential <sup>[2][17][18][19]</sup>. This conversion typically occurs naturally as the plant matures and dries, or because of heat exposure during processing or cooking <sup>[20]</sup>.



**Figure 1.** Biosynthesis of cannabinoids in *Cannabis*. CBC, cannabichromene; CBCA, cannabichromenic acid; CBD, cannabidiol; CBDA, cannabidiolic acid; CBGA, cannabigerolic acid;  $\Delta^9$ -THCA,  $\Delta^9$ -tetrahydrocannabinolic acid;  $\Delta^9$ -THC,  $\Delta^9$ -tetrahydrocannabinol.

The phytocannabinoids are essentially produced in glandular trichomes, mainly in female inflorescences. *C. sativa* mostly synthetizes cannabinoids of the alkyl type, characterized by a pentyl (C<sub>5</sub>) side chain and an isoprenyl (C<sub>10</sub>) monoterpene component. In general,  $\Delta^9$ -THC and CBD are the prevalent components of this plant and, based on the proportion of these two, three *C. sativa* chemotypes are known: (i) drug type, with predominant  $\Delta^9$ -THC; (ii) intermediate type, with similar amounts of both cannabinoids; and (iii) fiber type, with predominant CBD. CBGA, CBDA, and their decarboxylated derivatives are the main cannabinoids found in the fiber type <sup>[12]</sup>.

#### 2.2. Terpenes

Terpenes, characterized by multiple five-carbon isoprene units linked together to form a chain of hydrocarbons, comprise the second-largest class of plant constituents (120 identified so far). These secondary metabolites are synthesized via the isoprenoid biosynthesis metabolic pathway (**Figure 2**). Isopentenyl pyrophosphate (IPP) and its isomer dimethylallyl pyrophosphate (DMAPP), which are generated through the mevalonic acid (MEV) or methylerythritol phosphate (MEP) pathway, serve as the precursor molecules for this pathway. Afterward, a series of enzyme-catalyzed processes transform the IPP and DMAPP into several terpene precursors, including GPP, farnesyl pyrophosphate (FPP), and geranylgeranyl pyrophosphate (GGPP). GPP is responsible for the production of terpene molecules, which are then used in the production of cannabinoids. Different terpene synthases can further modify these precursors to produce a variety of monoterpenes, sesquiterpenes, and diterpenes [21][22].

Acotyl CoA

 $P_{VPUVVato} \pm C2P$ 

**Figure 2.** Biosynthetic route of terpenes. CBGA, cannabigerolic acid; DMAPP, dimethylallyl pyrophosphate; FPP, farnesyl pyrophosphate; G3P, glyceraldehyde 3-phosphate; GGPP, geranylgeranyl diphosphate; GPP, geranyl pyrophosphate; IPP, isopentenyl pyrophosphate; MEP, methylerythritol phosphate; MEV, mevalonic acid; TPC, terpene synthase.

In general, these compounds are synthesized in different parts of the plant, such as the flowers, roots, leaves, and trichomes, with variable levels, which depend on abiotic and biotic factors <sup>[11]</sup>. Particularly in *C. sativa*, Chacon and coworkers found that terpenes may vary between 0.001 and 14.8 mg/g of the dry weight of the plant <sup>[22]</sup> and are in general rich in mono- and sesquiterpenes <sup>[23]</sup>. Monoterpenes, structurally characterized by a skeleton with two molecules of isoprene, are more volatile and contribute to the flavor and aroma of the plant, while sesquiterpenes (structure containing three molecules of isoprene) are more stable and contribute to the therapeutical benefits of the plant <sup>[24]</sup>. The monoterpene  $\beta$ -myrcene and the sesquiterpenes  $\beta$ -caryophyllene and  $\alpha$ -humulene have been identified in most cannabis species, including *C. sativa*. Other typical terpenes synthesized by this species include  $\alpha$ -pinene and limonene, among others (**Figure 3**) <sup>[25]</sup>. In addition, terpenoids (i.e., oxygenated terpenes) such as linalool,  $\alpha$ -terpineol, guaiol,  $\alpha$ - and  $\beta$ -bisabolol are often found in *C. sativa* <sup>[26]</sup>. Moreover, in a study by Janatová, among six genotypes of *C. sativa*, four were found to have pinocarveol in large amounts. Additionally, one of the genotypes contained significantly higher concentrations of the major terpenes (limonene, linalool, fenchol, and  $\alpha$ -terpineol) when compared to the other genotypes. As a result, depending on the substances of interest, the genotype selected, as well as the type of extraction, should always be adjusted.



Figure 3. Chemical structure of the main terpenes found in C. sativa.

# 3. Bioactive Compounds in the Prevention and Treatment of Colorectal Cancer

Studies on cancer prevention have typically focused on the inhibition of carcinogenic processes, such as the formation of aberrant crypt foci (ACF) <sup>[27]</sup> and CRC precursor lesions, as well as the inhibition of cancer cell proliferation <sup>[28]</sup>. On the other hand, studies on cancer therapy typically focus on inducing the apoptosis of cancer cells <sup>[29]</sup>, inhibiting angiogenesis and metastasis <sup>[30]</sup>, and reducing inflammation <sup>[31]</sup>. Cannabinoids and terpenes have been shown to be useful in both the chemoprevention and the treatment of CRC, making them an attractive therapeutic option.

#### 3.1. Effects of Cannabinoids in CRC-Associated Mechanisms

Over history, there has been increasing evidence of cannabis's beneficial effects on CRC. Several in vitro and in vivo experiments showed that natural phytocannabinoids from C. sativa can interact with some of the mechanisms inherent in cancer. Among them, apoptosis, autophagy, inflammation, migration, invasion, and metastasis are the target mechanisms that have demonstrated better outcomes in CRC carcinogenesis with cannabinoids. As mentioned earlier, cannabinoids can exert anticancer effects in part due to their interaction with the ECS. This is a complex cell signaling system present in all mammals, which is involved in regulating various physiological and cognitive processes, such as appetite, pain, mood, and memory. It is composed of three main components: endocannabinoids, receptors, and metabolic enzymes. Endocannabinoids are lipid-based neurotransmitters produced by the body, such as anandamide and 2arachidonoylglycerol (2-AG), which bind to cannabinoid receptors (CB1 and CB2) found on the surface of cells, triggering a cellular response. Cannabinoids then can act on the ECS by mimicking endocannabinoids and binding to CB1 and CB2 receptors. Other receptors interact with endocannabinoids and modulate the ECS, including G-protein-coupled receptor 55 (GPR55), transient receptor potential cation channel subfamily V member 1 (TRPV1), TRPV2, transient receptor potential cation channel subfamily M member 8 (TRPM8), and peroxisome proliferator-activated receptors (PPARs) (Figure 4) [32]. The role of ECS in CRC has been reviewed elsewhere [33]. However, it is important to note that CRC cells and tissues express both CB1 and CB2 receptors, and TRPM8, TRPA1, TRPV1, and TRPV2 receptors, to which cannabinoids can bind to exert biological effects on CRC [27][33].



**Figure 4.** Schematic representation of endocannabinoid system components. GPR55, G-protein-coupled receptor 55; PPARs, peroxisome proliferator-activated receptor; TRPM8, transient receptor potential cation channel subfamily M member 8; TRPV1, transient receptor potential cation channel subfamily V member 1. The figure was partly generated using Servier Medical Art, provided by Servier, licensed under a Creative Commons Attribution 3.0 unported license.

#### 3.1.1. CBD

Isolated CBD is the most extensively studied cannabinoid in cancer research due to its numerous health benefits and nonpsychoactive nature. This cannabinoid was described as cytotoxic in different types of CRC cells, inhibiting their viability, namely in HCT116, SW480, SW620, CACO-2, HT-29, and DLD-1 <sup>[29][34][35][36]</sup>. Lee et al. found that CBD suppressed cell viability through a mechanism dependent on the CB2 receptor and not on CB1. Additionally, normal human colon cells resisted CBD, establishing its safety for noncancer cells <sup>[34]</sup>. Furthermore, several CRC cell lines (CACO-2, HT-29, DLD-1, SW620, SW480, COLO205, and HCT116) suffered significant reductions in proliferation after treatment with CBD at different concentrations (2.5–15  $\mu$ M) <sup>[30][34][36][37][38][39]</sup>. In this way, its beneficial effects have become a target of study in different anticancer mechanisms.

Resistance to apoptosis (cell death) is a hallmark of all types of cancer, so several apoptosis-inducing drugs have been developed as cancer therapies, including for CRC <sup>[40]</sup>. Aviello et al. <sup>[38]</sup> corroborated that CACO-2 CRC cells, treated with CBD (10  $\mu$ M for 24 h), significantly decreased the expression of phospho-protein kinase B (Akt) (p < 0.001) and upregulated caspase-3 expression in colonic tissues of azoxymethane (AOM)-induced mice (CBD, 1 mg/kg). These data suggest that CBD can induce apoptosis involving the phosphoinositide 3-kinase (PI3K)/Akt signaling pathway, essential in the regulation of cell growth, migration, differentiation, and apoptosis. In other CRC cell lines, namely, HCT-116, SW480, and SW620, this mechanism occurred through the antagonism of several receptors, including CB1, TRPV1, and PPARy [34].

It is well established that oxidative stress and the generation of ROS are significant factors in apoptosis, and this cellular process can be delayed or prevented by some antioxidants <sup>[41]</sup>. A study confirmed that CBD caused oxidative stress in HT-29 cells, probably through the generation of ROS, which depleted glutathione (GSH) and inhibited the activities of catalase, glutathione reductase (GR), and glutathione peroxidase (GPx). These findings were supported by the observation that malondialdehyde (MDA) levels were markedly increased (p < 0.01) in cells treated with CBD (2.6 ± 0.18 nmol/mL) compared to control cells (1.6 ± 0.27 nmol/mL), while they were largely unaltered in HT-29 cells exposed to THC <sup>[36]</sup>. Combined with oxaliplatin, a chemotherapeutic drug for cancer, CBD (4 µM CBD for 24 h) also produced similar results in oxaliplatin-resistant DLD-1 and colo205 cells <sup>[39]</sup>.

Autophagy and apoptosis are linked and can interact with each other. Autophagy can prevent or delay apoptosis, and apoptosis can also trigger autophagy. Both processes help regulate cell survival and death, contributing to cellular homeostasis maintenance <sup>[42]</sup>. CBD has also been shown to induce autophagy in oxaliplatin-resistant DLD-1, colo205 cells, and tumor tissues of BALB/c nude mice (xenograft model, colo205) <sup>[39]</sup>. Microtubule-associated proteins light chain 3 (LC3) and p62 expression, which are commonly used autophagic biomarkers, were both considerably enhanced by the combination of oxaliplatin and CBD in CRC cells.

Angiogenesis, the recruitment of new blood vessels, is a therapeutic target and an integral component of tumor development, invasion, and metastasis. This process engages a variety of growth factors, including integrins, chemokines, vascular endothelial growth factor (VEGF), and fibroblast growth factor <sup>[43]</sup>. Honarmand et al. reported that

CBD treatments (1 and 5 mg/kg) decreased VEGF expression (p < 0.05) in a dose-dependent manner in tumor tissues of CRC-induced mice when compared to cancer control groups <sup>[44]</sup>.

As stated earlier, adhesion, migration, and invasion mechanisms are involved in metastasis and the spread of cancer to other tissues. In HCT116 CRC cells, CBD at 1 (p < 0.05) and 2.5  $\mu$ M (p < 0.001) caused a significant reduction in GPR55-dependent adhesion and migration to endothelial cells, suggesting that CBD's antagonistic activity in GPR55 plays a key role in the reduction in metastasis <sup>[45]</sup>.

#### 3.1.2. Δ<sup>9</sup>-THC

THC has also been reported to have good outcomes in inhibiting cell viability on CRC cells, namely, SW480, HCT-15, HT-29, and HCA7 <sup>[36][46]</sup>. In HT-29 cells, THC suppressed cell viability more than CB83 (a synthetic cannabinoid) after 24 h of treatment at 30  $\mu$ M <sup>[36]</sup>. Additionally, Shor and coworkers reported that  $\Delta^9$ -THC and  $\Delta^8$ -THC were more toxic to polypderived cells than other cannabinoids, namely, CBD, CBC, CBDV, THCA, CBG, and other minor cannabinoids <sup>[47]</sup>. Furthermore, THC at 30  $\mu$ M was shown to trigger significant inhibition of proliferation at 24 h (70.9 ± 5.59, *p* < 0.01) in HT-29 CRC cells, compared to untreated cells <sup>[36]</sup>.

The induction of apoptosis by THC through the activation of the CB1 receptor in CRC cells was first reported in 2007 by Greenhoug et al. <sup>[46]</sup>. The authors suggested that apoptosis was induced by the activation of the B-cell lymphoma-2 (Bcl-2) family member BAD protein dependent on the CB1-mediated Ras-mitogen-activated protein kinase and PI3K/Akt pathway inhibition. In reality, the inhibition of ERK and Akt activity by THC was complemented by activation of the proapoptotic BAD. Additionally, THC increased the simultaneous cleavage of the caspase-3 substrate PARP.

Optimizing the delivery method can improve the efficacy of THC treatment for CRC owing to its lipophilicity, tar-like viscosity, and instability. In fact, De la Ossa et al. suggested an alternative method of THC delivery. The oil-in-water emulsion solvent evaporation method used by the authors to encapsulate THC into biodegradable microspheres revealed its ability to prevent cancer cell proliferation in CACO-2 cells <sup>[48]</sup>.

#### 3.1.3. CBG

CBG, a non-psychoactive cannabinoid, has gained a lot of interest due to its ability to be a partial agonist of CB1 and CB2 receptors <sup>[49]</sup>. In addition to the main receptors of the endocannabinoid system, CBG also acts as an antagonist at TRPM8, TRPV4, and 5HT<sub>1A</sub> and an agonist at TRPA1, TRPV1, and TRPV2 receptors <sup>[50]</sup>.

In CRC, CBG treatment had positive results in cell survival, apoptosis, oxidative stress, and in vivo tumor growth. As for cell survival, CBG was reported to induce a significant decrease in the viability of CACO-2 cells at 30  $\mu$ M after 3 h of incubation (p < 0.001). However, after 48 h of incubation, CBG effects on cell viability were significantly decreased even for lower concentrations (3, 10, and 30  $\mu$ M). In HCT116 cells, a similar trend was observed, and this effect was repressed in cells where TRPM8 was suppressed <sup>[27]</sup>. In HCT116 cells, purified CBG exhibited antiproliferative potential at concentrations 2.5, 5, and 10  $\mu$ M <sup>[26]</sup>.

#### 3.1.4. Minor Cannabinoids

Despite being less studied, minor cannabinoids such as CBDV, CBL, CBGV, CBCA, THCV, and CBGA revealed positive effects in the cell survival and proliferation of CRC cells. Ben-Ami Shor et al.'s research <sup>[47]</sup> indicated that the combinations of CBCA (14.5 or 29  $\mu$ M), CBDV (23.5 or 47  $\mu$ M), THCV (20 or 40  $\mu$ M), and CBGA (25.6 or 51.2  $\mu$ M) triggered a decrease in the cell viability of cells derived from human polyps. The combination of CBCA and CBDV also had synergistic effects, as well as the combination of THCV and CBGA. In HCT116 cells, data revealed cell viability proportions of 93, 87, and 51% with CBDV; 69, 44, and 45% with CBL; and 93, 75, and 71% for CBGV treatments at 2.5, 5, and 10  $\mu$ M, suggesting that these cannabinoids exerted antiproliferative effects against CRC cells <sup>[34]</sup>. Despite the antiproliferative properties demonstrated for the cannabinoids CBCA, CBDV, THCV, and CBGA, the concentrations were relatively high compared to those of CBD, THC, and CBG, suggesting that their effects are not very strong.

#### 3.2. Effects of Terpenes on CRC

Terpenes have been screened for their potential to prevent CRC when used alone or, in the case of the *C. sativa* plant, to enhance the plant's medicinal properties. These compounds act by inhibiting the growth and proliferation of cancer cells, and by inducing apoptosis. In addition, there is evidence that some terpenoids may have similar effects to cannabinoids, such as THC, in terms of their ability to interact with the endocannabinoid system. This interaction can cause changes in the activity of certain receptors and enzymes, which in turn affect a wide range of physiological processes. Indeed,  $\beta$ -

caryophyllene was shown to interact with the CB2 receptor, acting as its full agonist, resulting in a reduction in pain and inflammation in male rats and mice <sup>[51]</sup>.

Inflammatory bowel diseases are closely associated with an increased risk of CRC <sup>[52]</sup>. In this sense, Bento et al. <sup>[53]</sup> emphasized that the preventive treatment of DSS-induced colitis CD1 mice with BCP (50 mg/kg) resulted in a significant decrease in TNF- $\alpha$ , IL-1 $\beta$ , keratinocyte-derived chemokine, and interferon- $\gamma$  protein levels (p < 0.05) in mice colon segments. These proteins are mediators involved in cellular migration and adhesion. The authors also revealed that the decrease in the inflammatory mediators was associated with the factor nuclear kappa B (NFkB) signaling pathway, as BCP decreased the activation of the p65 NFkB subunit.

#### 3.3. Effects of Cannabis sativa Extracts on CRC

Interactions between terpenes/terpenoids and cannabinoids may intensify their pharmacological effects. In reality, there is growing evidence that these substances interact more favorably when used in combination than when used alone. This is known as the "entourage effect" in modern parlance, an effect attributed to cannabis's medical properties <sup>[54]</sup>.

Janatová et al. <sup>[26]</sup> suggested that the compounds responsible for the cytotoxicity of *C. sativa* ethanolic (EtOH) extracts in CACO-2 and HT29 cell lines were THC and CBD. However, in the same work, a specific genotype containing high concentrations of myrcene,  $\beta$ -elemene,  $\beta$ -selinene, and  $\alpha$ -bisabolol oxide positively affected the selectivity of cytotoxic activity, due to synergistic effects. In another study conducted by Nallathambi and coworkers, the authors reported an increment in cytotoxic effects of a *C. sativa* EtOH extract in CRC cells (HCT 116, HT-29, and CACO-2) when this was combined with a THCA-rich fraction <sup>[28]</sup>. In addition to the EtOH extract, fractions obtained from it were also evaluated. The combination of a CBGA-rich fraction and a THCA-rich fraction resulted in a noticeable increase in apoptosis. The CBGA-rich fraction included other compounds such as CBN (3.67%), CBCA (3.53%), terpenes (0.72%), diterpenes (0.33%), and short free fatty acids (0.37%), which may have played a role in the outcome.

# 4. Conclusions

Data suggest that cannabinoids exert advantages in the treatment of CRC, mostly by inducing apoptosis, although some evidence also points out that they may target other key therapeutic events, such as proliferation, metastasis, inflammation, angiogenesis, oxidative stress, and autophagy. The currently available data on this subject refer mostly to the *C. sativa* major cannabinoids, i.e., CBD, THC, and CBG, but several pieces of evidence suggest that minor cannabinoids and other bioactive compounds such as terpenes also may hold potential as therapeutic agents for CRC. Data also suggest that certain combinations of cannabinoids and terpenes in *C. sativa* extracts can lead to a synergistic action known as the "entourage effect," which has been linked to certain pharmacological benefits. The potential therapeutic benefits of the cannabinoids and terpenes from this plant make them key candidates for further drug development.

#### References

- 1. Russo, E.B. History of Cannabis and Its Preparations in Saga, Science, and Sobriquet. Chem. Biodivers. 2007, 4, 1614 –1648.
- Ubeed, H.M.S.A.L.; Bhuyan, D.J.; Alsherbiny, M.A.; Basu, A.; Vuong, Q.V. A Comprehensive Review on the Techniques for Extraction of Bioactive Compounds from Medicinal Cannabis. Molecules 2022, 27, 604.
- 3. Duggan, P.J. The Chemistry of Cannabis and Cannabinoids. Aust. J. Chem. 2021, 74, 369–387.
- 4. Pratt, M.; Stevens, A.; Thuku, M.; Butler, C.; Skidmore, B.; Wieland, L.S.; Clemons, M.; Kanji, S.; Hutton, B. Benefits an d harms of medical cannabis: A scoping review of systematic reviews. Syst. Rev. 2019, 8, 320.
- Citti, C.; Braghiroli, D.; Vandelli, M.A.; Cannazza, G. Pharmaceutical and biomedical analysis of cannabinoids: A critical review. J. Pharm. Biomed. Anal. 2018, 147, 565–579.
- 6. Russo, E.B. Taming THC: Potential cannabis synergy and phytocannabinoid-terpenoid entourage effects. Br. J. Pharm acol. 2011, 163, 1344–1364.
- Vučković, S.; Srebro, D.; Vujović, K.S.; Vučetić, C.; Prostran, M. Cannabinoids and Pain: New Insights from Old Molecu les. Front. Pharmacol. 2018, 9, 1259.
- 8. Liktor-Busa, E.; Keresztes, A.; LaVigne, J.; Streicher, J.M.; Largent-Milnes, T.M. Analgesic Potential of Terpenes Derive d from Cannabis sativa. Pharmacol. Rev. 2021, 73, 1269–1297.

- McDougall, J.J.; McKenna, M.K. Anti-Inflammatory and Analgesic Properties of the Cannabis Terpene Myrcene in Rat A djuvant Monoarthritis. Int. J. Mol. Sci. 2022, 23, 7891.
- 10. Atalay, S.; Jarocka-Karpowicz, I.; Skrzydlewska, E. Antioxidative and Anti-Inflammatory Properties of Cannabidiol. Antio xidants 2020, 9, 21.
- 11. Liu, Y.; Liu, H.-Y.; Li, S.-H.; Ma, W.; Wu, D.-T.; Li, H.-B.; Xiao, A.-P.; Liu, L.-L.; Zhu, F.; Gan, R.-Y. Cannabis sativa bioact ive compounds and their extraction, separation, purification, and identification technologies: An updated review. TrAC T rends Anal. Chem. 2022, 149, 116554.
- 12. Kopustinskiene, D.M.; Masteikova, R.; Lazauskas, R.; Bernatoniene, J. Cannabis sativa L. Bioactive Compounds and T heir Protective Role in Oxidative Stress and Inflammation. Antioxidants 2022, 11, 660.
- 13. Andre, C.M.; Hausman, J.-F.; Guerriero, G. Cannabis sativa: The Plant of the Thousand and One Molecules. Front. Pla nt Sci. 2016, 7, 19.
- 14. Gülck, T.; Møller, B.L. Phytocannabinoids: Origins and Biosynthesis. Trends Plant Sci. 2020, 25, 985–1004.
- 15. Hanuš, L.O.; Meyer, S.M.; Muñoz, E.; Taglialatela-Scafati, O.; Appendino, G. Phytocannabinoids: A unified critical inven tory. Nat. Prod. Rep. 2016, 33, 1357–1392.
- Sommano, S.R.; Sunanta, P.; Leksawasdi, N.; Jantanasakulwong, K.; Rachtanapun, P.; Seesuriyachan, P.; Phimolsirip ol, Y.; Sringarm, K.; Ruksiriwanich, W.; Jantrawut, P.; et al. Mass Spectrometry-Based Metabolomics of Phytocannabino ids from Non-Cannabis Plant Origins. Molecules 2022, 27, 3301.
- 17. Mnekin, L.; Ripoll, L. Topical Use of Cannabis Sativa L. Biochemicals. Cosmetics 2021, 8, 85.
- 18. Odieka, A.E.; Obuzor, G.U.; Oyedeji, O.O.; Gondwe, M.; Hosu, Y.S.; Oyedeji, A.O. The Medicinal Natural Products of C annabis sativa Linn.: A Review. Molecules 2022, 27, 1689.
- Barrales-Cureño, H.J.; López-Valdez, L.G.; Reyes, C.; Cetina-Alcalá, V.M.; Vasquez-García, I.; Diaz-Lira, O.F.; Herrera-Cabrera, B.E. Chemical Characteristics, Therapeutic Uses, and Legal Aspects of the Cannabinoids of Cannabis sativa: A Review. Braz. Arch. Biol. Technol. 2020, 63.
- 20. Moreno, T.; Dyer, P.; Tallon, S. Cannabinoid Decarboxylation: A Comparative Kinetic Study. Ind. Eng. Chem. Res. 2020, 59, 20307–20315.
- Jin, D.; Dai, K.; Xie, Z.; Chen, J. Secondary Metabolites Profiled in Cannabis Inflorescences, Leaves, Stem Barks, and Roots for Medicinal Purposes. Sci. Rep. 2020, 10, 3309–3314.
- 22. Chacon, F.T.; Raup-Konsavage, W.M.; Vrana, K.E.; Kellogg, J.J. Secondary Terpenes in Cannabis sativa L.: Synthesis and Synergy. Biomedicines 2022, 10, 3142.
- Helcman, M.; Šmejkal, K. Biological activity of Cannabis compounds: A modern approach to the therapy of multiple dise ases. Phytochem. Rev. 2022, 21, 429–470.
- 24. Johnson, A.; Stewart, A.; El-Hakim, I.; Hamilton, T.J. Effects of super-class cannabis terpenes beta-caryophyllene and a lpha-pinene on zebrafish behavioural biomarkers. Sci. Rep. 2022, 12, 17250.
- 25. Booth, J.K.; Bohlmann, J. Terpenes in Cannabis sativa—From plant genome to humans. Plant Sci. 2019, 284, 67–72.
- Janatová, A.; Doskočil, I.; Božik, M.; Fraňková, A.; Tlustoš, P.; Klouček, P. The chemical composition of ethanolic extrac ts from six genotypes of medical cannabis (Cannabis sativa L.) and their selective cytotoxic activity. Chem. Interact. 20 22, 353, 109800.
- 27. Borrelli, F.; Pagano, E.; Romano, B.; Panzera, S.; Maiello, F.; Coppola, D.; De Petrocellis, L.; Buono, L.; Orlando, P.; Izz o, A.A. Colon carcinogenesis is inhibited by the TRPM8 antagonist cannabigerol, a Cannabis-derived non-psychotropic cannabinoid. Carcinogenesis 2014, 35, 2787–2797.
- 28. Nallathambi, R.; Mazuz, M.; Namdar, D.; Shik, M.; Namintzer, D.; Vinayaka, A.C.; Ion, A.; Faigenboim, A.; Nasser, A.; L aish, I.; et al. Identification of Synergistic Interaction between Cannabis-Derived Compounds for Cytotoxic Activity in Co lorectal Cancer Cell Lines and Colon Polyps That Induces Apoptosis-Related Cell Death and Distinct Gene Expression. Cannabis Cannabinoid Res. 2018, 3, 120–135.
- Jeong, S.; Yun, H.K.; Jeong, Y.A.; Jo, M.J.; Kang, S.H.; Kim, J.L.; Kim, D.Y.; Park, S.H.; Kim, B.R.; Na, Y.J.; et al. Cann abidiol-induced apoptosis is mediated by activation of Noxa in human colorectal cancer cells. Cancer Lett. 2019, 447, 1 2–23.
- 30. Feng, P.; Zhu, L.; Jie, J.; Yang, P.; Sheng, N.; Chen, X.; Chen, X. Cannabidiol inhibits invasion and metastasis in colore ctal cancer cells by reversing epithelial—Mesenchymal transition through the Wnt/β-catenin signaling pathway. J. Canc er Res. Clin. Oncol. 2022.
- 31. Becker, W.; Alrafas, H.R.; Wilson, K.; Miranda, K.; Culpepper, C.; Chatzistamou, I.; Cai, G.; Nagarkatti, M.; Nagarkatti, P.S. Activation of Cannabinoid Receptor 2 Prevents Colitis-Associated Colon Cancer through Myeloid Cell De-Activatio

n Upstream of IL-22 Production. iScience 2020, 23, 101504.

- 32. Khoury, M.; Cohen, I.; Bar-Sela, G. "The Two Sides of the Same Coin"—Medical Cannabis, Cannabinoids and Immunit y: Pros and Cons Explained. Pharmaceutics 2022, 14, 389.
- 33. Cherkasova, V.; Kovalchuk, O.; Kovalchuk, I. Cannabinoids and Endocannabinoid System Changes in Intestinal Inflam mation and Colorectal Cancer. Cancers 2021, 13, 4353.
- Lee, H.-S.; Tamia, G.; Song, H.-J.; Amarakoon, D.; Wei, C.-I.; Lee, S.-H. Cannabidiol exerts anti-proliferative activity via a cannabinoid receptor 2-dependent mechanism in human colorectal cancer cells. Int. Immunopharmacol. 2022, 108, 1 08865.
- 35. Kim, J.L.; Kim, B.R.; Kim, D.Y.; Jeong, Y.A.; Jeong, S.; Na, Y.J.; Park, S.H.; Yun, H.K.; Jo, M.J.; Kim, B.G.; et al. Canna bidiol Enhances the Therapeutic Effects of TRAIL by Upregulating DR5 in Colorectal Cancer. Cancers 2019, 11, 642.
- Cerretani, D.; Collodel, G.; Brizzi, A.; Fiaschi, A.I.; Menchiari, A.; Moretti, E.; Moltoni, L.; Micheli, L. Cytotoxic Effects of Cannabinoids on Human HT-29 Colorectal Adenocarcinoma Cells: Different Mechanisms of THC, CBD, and CB83. Int. J. Mol. Sci. 2020, 21, 5533.
- 37. Sreevalsan, S.; Joseph, S.; Jutooru, I.; Chadalapaka, G.; Safe, S.H. Induction of apoptosis by cannabinoids in prostate and colon cancer cells is phosphatase dependent. Anticancer Res. 2011, 31, 3799–3807.
- 38. Aviello, G.; Romano, B.; Borrelli, F.; Capasso, R.; Gallo, L.; Piscitelli, F.; Di Marzo, V.; Izzo, A.A. Chemopreventive effec t of the non-psychotropic phytocannabinoid cannabidiol on experimental colon cancer. J. Mol. Med. 2012, 90, 925–934.
- 39. Jeong, S.; Kim, B.G.; Kim, D.Y.; Kim, B.R.; Kim, J.L.; Park, S.H.; Na, Y.J.; Jo, M.J.; Yun, H.K.; Jeong, Y.A.; et al. Canna bidiol Overcomes Oxaliplatin Resistance by Enhancing NOS3- and SOD2-Induced Autophagy in Human Colorectal Ca ncer Cells. Cancers 2019, 11, 781.
- 40. Hanahan, D.; Weinberg, R.A. Hallmarks of cancer: The next generation. Cell 2011, 144, 646–674.
- 41. Kannan, K.; Jain, S.K. Oxidative stress and apoptosis. Pathophysiology 2000, 7, 153–163.
- 42. Lee, X.C.; Werner, E.; Falasca, M. Molecular Mechanism of Autophagy and Its Regulation by Cannabinoids in Cancer. Cancers 2021, 13, 1211.
- 43. Solinas, M.; Massi, P.; Cantelmo, A.; Cattaneo, M.; Cammarota, R.; Bartolini, D.; Cinquina, V.; Valenti, M.; Vicentini, L.; Noonan, D.; et al. Cannabidiol inhibits angiogenesis by multiple mechanisms. Br. J. Pharmacol. 2012, 167, 1218–1231.
- 44. Honarmand, M.; Namazi, F.; Mohammadi, A.; Nazifi, S. Can cannabidiol inhibit angiogenesis in colon cancer? Comp. Cl in. Pathol. 2019, 28, 165–172.
- 45. Kargl, J.; Andersen, L.; Hasenöhrl, C.; Feuersinger, D.; Stančić, A.; Fauland, A.; Magnes, C.; El-Heliebi, A.; Lax, S.; Ura nitsch, S.; et al. GPR55 promotes migration and adhesion of colon cancer cells indicating a role in metastasis. Br. J. Ph armacol. 2016, 173, 142–154.
- 46. Greenhough, A.; Patsos, H.A.; Williams, A.C.; Paraskeva, C. The cannabinoid δ 9-tetrahydrocannabinol inhibits RAS-M APK and PI3K-AKT survival signalling and induces BAD-mediated apoptosis in colorectal cancer cells. Int. J. Cancer 2 007, 121, 2172–2180.
- 47. Shor, D.B.-A.; Hochman, I.; Gluck, N.; Shibolet, O.; Scapa, E. The Cytotoxic Effect of Isolated Cannabinoid Extracts on Polypoid Colorectal Tissue. Int. J. Mol. Sci. 2022, 23, 11366.
- 48. De la Ossa, D.H.P.; Gil-Alegre, M.E.; Ligresti, A.; Aberturas, M.D.R.; Molpeceres, J.; Torres, A.I.; Di Marzo, V. Preparati on and characterization of Δ9-tetrahydrocannabinol-loaded biodegradable polymeric microparticles and their antitumor al efficacy on cancer cell lines. J. Drug Target. 2013, 21, 710–718.
- Navarro, G.; Varani, K.; Reyes-Resina, I.; Sánchez de Medina, V.; Rivas-Santisteban, R.; Sánchez-Carnerero Callado, C.; Vincenzi, F.; Casano, S.; Ferreiro-Vera, C.; Canela, E.I.; et al. Cannabigerol Action at Cannabinoid CB1 and CB2 R eceptors and at CB1-CB2 Heteroreceptor Complexes. Front. Pharmacol. 2018, 9, 632.
- Martínez, V.; Iriondo De-Hond, A.; Borrelli, F.; Capasso, R.; del Castillo, M.D.; Abalo, R. Cannabidiol and Other Non-Ps ychoactive Cannabinoids for Prevention and Treatment of Gastrointestinal Disorders: Useful Nutraceuticals? Int. J. Mol. Sci. 2020, 21, 3067.
- 51. Ceccarelli, I.; Fiorenzani, P.; Pessina, F.; Pinassi, J.; Aglianò, M.; Miragliotta, V.; Aloisi, A.M. The CB2 Agonist β-Caryop hyllene in Male and Female Rats Exposed to a Model of Persistent Inflammatory Pain. Front. Neurosci. 2020, 14, 850.
- 52. Lucafò, M.; Curci, D.; Franzin, M.; Decorti, G.; Stocco, G. Inflammatory Bowel Disease and Risk of Colorectal Cancer: An Overview from Pathophysiology to Pharmacological Prevention. Front. Pharmacol. 2021, 12, 772101.
- 53. Bento, A.F.; Marcon, R.; Dutra, R.C.; Claudino, R.F.; Cola, M.; Leite, D.F.P.; Calixto, J.B. β-Caryophyllene Inhibits Dextr an Sulfate Sodium-Induced Colitis in Mice through CB2 Receptor Activation and PPARy Pathway. Am. J. Pathol. 2011, 178, 1153–1166.

54. Gonçalves, E.C.D.; Baldasso, G.M.; Bicca, M.A.; Paes, R.S.; Capasso, R.; Dutra, R.C. Terpenoids, Cannabimimetic Lig ands, beyond the Cannabis Plant. Molecules 2020, 25, 1567.

Retrieved from https://encyclopedia.pub/entry/history/show/100515