The Angiogenic Balance

Subjects: Cardiac & Cardiovascular Systems

Contributor: Cătălina Ionescu, Bogdan Oprea, Georgeta Ciobanu, Milena Georgescu, Ramona Bică, Garofiţa-Olivia Mateescu, Fidan Huseynova, Veronique Barragan-Montero

Angiogenesis is the process of developing new blood vessels from pre-existing ones.

Keywords: angiogenesis ; activators ; inhibitors

1. Formation and Remodeling of Blood Vessels

Blood vessels that supply the body with oxygen and nutrients, are formed through three different mechanisms (vasculogenesis, angiogenesis, arterio-genesis) united under the name of neovascularization.

Vasculo-genesis. Blood vessels are formed in the early stages of embryo development through vasculo-genesis when the embryonic mesenchymal cells differentiate into endothelial cells and a "primary capillary plexus" is formed. At present, it is known that, besides its role in embryonic development, vasculo-genesis is also induced after birth. In this case, bone marrow-derived endothelial progenitor cells mediate the cases of physiological and pathological neovascularization, for example, in the cases of wound healing or cardiac ischemia [112][3].

Angiogenesis is the main process in post-natal neovascularization and represents the process of new vessel formation from pre-existing ones $^{[4]}$, through two mechanisms: intussusception $^{[5][6]}$ and sprouting $^{[7]}$. Intussusception represents the "splitting" of an existing vessel into two new vessels, with interior reconstruction of novel blood vessel walls. Leading to a rise in the number of vessels without augmentation of the number of endothelial cells, intussusception angiogenesis is involved in the remodeling of existing vessels, for example, in capillary formation starting from the primary plexus in embryo development, but it can also be seen in post-natal development. Sprouting angiogenesis is a more complicated process. It is regulated by different angiogenic factors and involves several steps: degradation of the basement membrane of the vessel under enzymatic conditions, endothelial cells activation, proliferation and migration, formation of a new lumen and pericyte stabilization [Z|[8][9][10]. There is much evidence in different studies that a tumor's vascularization is associated with the metastatic risk and negatively influences the survival rate. The micro-density of a tumor's vascularization is especially used for the follow-up of anti-angiogenetic therapy $[11]$.

Arterio-genesis represents the formation of new blood vessels from co-existing, co-lateral vessels, with the participation of smooth muscle cells, as an adaptive process to an arterial occlusion $[12]$ [13], being rather a re-modeling process. In the proximity of an arterial stenosis, the substitution network is architectured through both arterio-genesis (in near regions, unleashed by mechanical constraints and cytokines) and angiogenesis (in distal sites, where hypoxia would generate new vessel sproutings) [14][15].

2. The Angiogenic Balance: Synthetic and Endogenous Regulators

2.1. Synthetic Modulators of Angiogenesis

Most of the endogenous molecules playing a role in angiogenesis modulation have protein structure, but their usage in therapy is delicate, because of the high cost of their production in large quantities and because of the difficulty in penetrating tissues.

For this reason, more attention has been shown to the preparation and investigation of certain compounds impacting the angiogenesis, such as the polypeptides with therapeutic effect $^{[16]}$. In the same terms, relating to angiogenesis modeling, some antibiotics, polysaccharides, steroids, and other synthetic small-molecular compounds have been reported [17][18].

As these contents are represented by carbohydrates, this type of compounds used as angiogenesis modulators is introduced in detail here. Many carbohydrate-binding proteins are involved in angiogenesis; therefore, carbohydrates and their analogues may be important factors for angiogenesis regulation $[19]$.

One example is represented by galectin-3 and MCP (modified citrus pectin). Galectin-3 is a β-galactoside-binding lectin, which mediates endothelial cell morphogenesis in vitro and angiogenesis in vivo ^[20].

Nangia-Makker et al. proved that it is able to tightly bind to galectin-3, via recognition of its carbohydrate recognition domain, and to inhibit angiogenesis and tumor growth ^[21]. Johnston at al. synthesized and studied heparan sulfate mimetics, represented by a series of poly-sulfated penta- and tetra-saccharide glycosides containing alpha(1→3)/alpha(1→2)-linked mannose residues. They found that the investigated mimetics bound tightly to angiogenic growth factors and exhibited potent activity indicative of angiogenesis; they strongly inhibited heparanase activity and also showed good antitumor activity [22].

One research group has proved that synthetic mannose-6-phosphate analogues can act as angiogenesis activators or inhibitors, depending on the structure of the chemical group functionalizing the C6 position of mannose ^{[23][24]}. In another study, researchers prepared gold nanoparticles decorated with various mannose derivatives functionalized in the C6 position and they proved to be effective over angiogenesis ^[25]. The cation-independent mannose-6-phosphate receptor was previously indicated as inducing angiogenesis through several possible mechanisms $[26]$, but this was the first time that mono-carbohydrates have directly been indicated as an agent possessing angiogenic activities.

Since then, it has been proven that 1,2,3,4,6-penta-O-galloyl-β-d-glucopyranose (PGG) has antiangiogenic activity in vitro and in vivo. Derivatives of PGG with different sugar cores and phenolic substituents have been tested and they are also angiogenesis inhibitors [27][28][29]

2.2. Endogenous Regulators of Angiogenesis

Besides mechanical (shear stress and blood flow augmentation) and chemical (hypoxia and nitric oxide increase) influences, angiogenesis is regulated by molecular influences, among which the most important are the angiogenic growth factors: fibroblast growth factor (FGF), vascular endothelial growth factor (VEGF), "platelet-derived endothelial cell growth factor" (PD-ECGF), angiopoietin; angiogenin, etc. ^{[30][31][32]}. The vascular endothelial growth factor (VEGF) action is closely related to the stimulation of angiogenesis ^{[33][34]}. Angiogenesis is, on the other hand, inhibited by anti-angiogenic factors, such as angiostatin, endostatin, thrombospondin-1 (TSP-1), heparinases, etc. ^[35]. When the balance between angiogenesis activators and inhibitors loses equilibrium, abnormal (either insufficient or excessive) angiogenesis occurs and various diseases appear or degenerate ^[36].

The main endogenous angiogenesis activators are summarized in **Table 1**. The most commonly studied endogenous angiogenesis activators are the FGF and VEGF families. bFGF is the first angiogenic factor that has been purified, in 1975 $[32]$, and FGFs are the first angiogenic factors that were sequenced, in 1985 $[38]$. VEGF has been identified in 1983 as a vascular permeability factor ^[39], and only later, in 1989, has it been shown to possess angiogenic action ^[40]. **Table 2** summarizes the main endogenous angiogenesis inhibitors. Thrombospondin-1 is the first protein observed to possess naturally occurring antiangiogenic properties $\frac{[41][42]}{2}$. Since then, the number of angiogenesis regulators have grown and, besides endogenous regulators, synthetic molecules with effect on angiogenesis have been obtained and tested, some of them already on the market and available for treatment.

Table 1. Endogenous angiogenesis activators.

Abbreviations: FGF—fibroblast growth factor; aFGF—acidic FGF; bFGF—basic FGF; FGFR—FGF receptor; HSPG heparan sulfate proteoglycans; VEGF—vascular endothelial growth factor; VEGFR—vascular endothelial growth factor receptor; PlGF—placental growth factor; Ang 1—Angiopoietin 1; Ang 2—Angiopoietin 2; TIE2—tyrosine kinase with immunoglobulin and epidermal growth factor homology domain 2; Ephs—ephrin receptors; RTKs—receptor protein tyrosine-kinase; MMPs—Matrix metalloproteinases; PAR—Protease-activated receptor.

Table 2. Endogenous Angiogenesis inhibitors.

Abbreviations: MMPs-Matrix metalloproteinases; EC-endothelial cell; ATP-adenosine triphosphate; TSPs-Thrombospondins; EGF—Epidermal growth factor; COMP—cartilage oligomeric matrix protein; 2-ME—2- Methoxyestradiol.

There are studies that prove the binding of growth factors to the cell surface, serving as target, receptors, or even as a storage mechanism. This seems to be valid for the inductor FGF which induces the activation of VEGF.

The class of VEGF proteins consists of several derivatives such as VEGF-A, VEGF-B, VEGF-C, VEGF-D, VEGF-E (encoded-virus) and VEGF-F (derived from snake venom) and placental growth factor (PIGF) [33][74][75]. Vascular permeability and inflammation, angiogenesis and apoptosis, lymphangio-genesis and fibrogenesis can be adjusted by the VEGF family $^{[75]}$. From all the VEGF class components, VEGF-A is the most individualized, representing a substantial angiogenesis promotor and consequently designed as an objective for the study of certain anti-cancer therapies [74][76]. According to the chain length, different VEGF-A isoforms have resulted after the splicing of alternative VEGF mRNA ^[ZS],

and have further been referred to as VEGF_{XXX}, where "XXX" indicates the number of amino acids from the final protein chain $^{[74]}$. The most well-known subtypes are VEGF $_{111}$, VEGF $_{121}$, VEGF $_{145}$, VEGF $_{165}$, VEGF $_{189}$ and VEGF $_{206}$ $^{[33]}$.

Human VEGF-A contains eight exons separated by seven introns, all their subtypes presenting similar regions, namely exons 1–5 and 8 $^{[75]}$. The longer VEGF isoforms containing both exons 6a and 7, such as VEGF₁₄₅, VEGF₁₈₉ and VEGF₂₀₆ have high affinity for heparin sulphate glycoproteins $^{[33][74][77][78]}$. VEGF₁₆₂ is a VEGF isoform, whose protein sequence has exons $1-5$, 6a, 6b and 8. proliferating the angiogenesis in vivo, while VEGF₁₆₅ is the most potent endothelial cells proliferation agent. The VEGF shorter isoforms, such as VEGF $_{111}$ and VEGF $_{121}$, do not have exons 6 and 7, are highly diffusible and. therefore, cannot connect to the extracellular matrix [33].

The main endogenous activators and inhibitors of angiogenesis are summarized in **Table 1** and **Table 2**.

Certain studies have demonstrated that the thickness of the capillary's basal lamina may lead to insufficient oxygen diffusion, limiting the elimination of some metabolites, eventually leading to the increase of different diseases' severity ^[79] $[80]$. Minchenko et al. $[81]$ suggested that hypoxia is both inductor and stimulator of VEGF expression in vivo, along with the increase of glycemia [82].

VEGF also increases the microvascular permeability that precedes and accompanies angiogenesis, playing a central role in its process, acting as an anti-apoptotic factor for endothelial cells in newly formed blood vessels.

Many studies have highlighted the histological structural changes in the blood vessels of periodontopathic diabetic patients, but less data were reported on the number of abnormally evolution vessels (MVD) and/or their tissue distribution process, which can lead to tumor growth and the development of metastases (neo-angiogenesis).

Thus, in addition to the structural changes visible with light microscopy for the gingival blood capillaries, the number of blood vessels from the gingiva of the periodontal patients with diabetes mellitus (DM) can be displayed by quantitative immunohistochemical method and vascular markers.

It can be believed that the best immunohistochemical marker to highlight the newly formed blood vessels is the CD31+CD34 antibody cocktail which may show the entire micro-vascular network, as shown in **Figure 1**.

Figure 1. Small caliper blood vessels immunostained for CD34 antibody (DAB ×200).

3. Physiological and Pathological Angiogenesis

The balance of activators and inhibitors is shown in **Figure 2**. Besides its role in embryonic development, angiogenesis appears as a normal process in adults, in female reproductive functions [83][84][85][86][87] and in tissue regeneration (ex. wounds healing) ^[88] (Figure 2B). Under certain conditions, it can appear as a disequilibrium in the synthesis of the endogenous factors that control angiogenesis, and an abnormal angiogenesis can occur. It can be either insufficient,

leading to the impossibility of the body healing wounds or participating in normal organ regeneration, or it can lead to the ischemia of a part of the body, such as myocardial, peripheral of intestinal ischemia. Other complications include ulcers, infertility and hair loss (**Figure 2A**).

As mentioned above, excessive angiogenesis can favor tumor growth and metastasis appearance or it can induce the evolution of diseases such as rheumatoid arthritis, psoriasis, etc. (Figure 2C) ^{[89][90]}.

Excessive angiogenesis is also linked to a series of eye diseases, which can lead to blindness. It is the cause of visual loss in the case of age-related macular degeneration. Patients who suffer from proliferative diabetic retinopathy are known to have higher VEGF levels when compared to healthy persons ^[91]. Increased VEGF levels cause uncontrolled angiogenesis in these patients.

Figure 2. The balance of activators and inhibitors in: (**A**)—insufficient angiogenesis; (**B**)—normal angiogenesis; (**C**) excessive angiogenesis. Abbreviations: FGF: fibroblast growth factor, VEGF: vascular endothelial growth factor, PD-ECGF: platelet-derived endothelial cell growth factor, TSP-1: thrombospondin-1. Normal angiogenesis is the balanced action of angiogenesis activators and inhibitors. In the case of preponderant action of angiogenesis inhibitors, insufficient angiogenesis appears, leading to diseases such as chronic wounds, cardiovascular diseases, neuropathies, ulcers, hair loss, and infertility (**Figure 2A**). Physiological angiogenesis appears as a normal process in female reproductive functions and in tissue regeneration (**Figure 2B**). On the contrary, if the action of angiogenesis activators prevails, complications in diseases such as cancer, diabetic retinopathy, rheumatoid arthritis, AIDS, psoriasis, osteomyelitis or uterine bleeding may appear (**Figure 2C**).

References

- 1. Eguchi, M.; Masuda, H.; Asahara, T. Endothelial progenitor cells for postnatal vasculogenesis. Clin. Exp. Nephrol. 200 7, 11, 18–25.
- 2. Murasawa, S.; Asahara, T. Endothelial Progenitor Cells for Vasculogenesis. Physiology 2005, 20, 36–42.
- 3. Asahara, T.; Masuda, H.; Takahashi, T.; Kalka, C.; Pastore, C.; Silver, M.; Kearne, M.; Magner, M.; Isner, J.M. Bone mar row origin of endothelial progenitor cells responsible for postnatal vasculogenesis in physiological and pathological neo vascularization. Circ. Res. 1999, 85, 221–228.
- 4. Carmeliet, P. Angiogenesis in health and disease. Nat. Med. 2003, 9, 653–660.
- 5. Djonov, V.; Baum, O.; Burri, P.H. Vascular remodeling by intussusceptive angiogenesis. Cell Tissue Res. 2003, 314, 10 7–117.
- 6. Djonov, V.; Schmid, M.; Tschanz, S.A.; Burri, P.H. Intussusceptive Angiogenesis: Its Role in Embryonic Vascular Networ k Formation. Circ. Res. 2000, 86, 286–292.
- 7. Tomanek, R.J.; Schatteman, G.C. Angiogenesis: New insights and therapeutic potential. Anat. Rec. 2000, 261, 126–13 5.
- 8. Couffinhal, T.; Dufourcq, P.; Daret, D.; Duplaa, C. The mechanisms of angiogenesis. Medical and therapeutic applicatio ns. Rev. Méd. Interne 2001, 22, 1064–1082.
- 9. Bussolino, F.; Mantovani, A.; Persico, G. Molecular mechanisms of blood vessel formation. Trends Biochem. Sci. 1997, 22, 251–256.
- 10. Ribatti, D.; Crivellato, E. "Sprouting angiogenesis", a reappraisal. Dev. Biol. 2012, 372, 157–165.
- 11. Tanigawa, N.; Amaya, H.; Matsumura, M.; Shimomatsuya, T.; Horiuchi, T.; Muraoka, R.; Iki, M. Extent of tumor vasculari zation correlates with prognosis and hematogenous metastasis in gastric carcinomas. Cancer Res. 1996, 56, 2671–26 76.
- 12. Scholz, D.; Cai, W.J.; Schaper, W. Arteriogenesis, a new concept of vascular adaptation in occlusive disease. Angiogen esis 2001, 4, 247–257.
- 13. Heil, M.; Wagner, S.; Schaper, W. Arterial regeneration by collateral artery growth (arteriogenesis). Drug Discov. Today Dis. Models 2004, 1, 265–271.
- 14. Stephan, D.; Weltin, D.; Zaric, V.; Chapelon, D.; Da Silva, A.; Lugnier, C. Angiogenèse: De la physiologie à la thérapeuti que. Réanim. Urgences 2000, 9, 534–544.
- 15. Levy, A.P.; Levy, N.S.; Goldberg, N.A. Post-transcriptional Regulation of Vascular Endothelial Growth Factor by Hypoxi a. J. Biol. Chem. 1996, 271, 2746–2753.
- 16. Rosca, E.V.; Koskimaki, J.E.; Rivera, C.G.; Pandey, N.B.; Tamiz, A.P.; Popel, A.S. Anti-angiogenic peptides for cancer t herapeutics. Curr. Pharm. Biotechnol. 2011, 12, 1101–1116.
- 17. Fan, T.P.; Jaggar, R.; Bicknell, R. Controlling the vasculature: Angiogenesis, anti-angiogenesis and vascular targeting o f gene therapy. Trends Pharmacol. Sci. 1995, 16, 57–66.
- 18. Quesada, A.R.; Munoz-Chapuli, R.; Medina, M.A. Anti-Angiogenic Drugs: From Bench to Clinical Trials. Med. Res. Rev. 2006, 26, 483–530.
- 19. Nangia-Makker, P.; Baccarini, S.; Raz, A. Carbohydrate-recognition and angiogenesis. Cancer Metastasis Rev. 2000, 1 9, 51–57.
- 20. Funasaka, T.; Raz, A.; Nangia-Makker, P. Galectin-3 in angiogenesis and metastasis. Glycobiology 2014, 24, 886–891.
- 21. Nangia-Makker, P.; Hogan, V.; Honjo, Y.; Baccarini, S.; Tait, L.; Bresalier, R.; Raz, A. Inhibition of human cancer cell gro wth and metastasis in nude mice by oral intake of modified citrus pectin. J. Natl. Cancer. Inst. 2002, 94, 1854–1862.
- 22. Johnstone, K.D.; Karoli, T.; Liu, L.; Dredge, K.; Copeman, E.; Li, C.P.; Davis, K.; Hammond, E.; Bytheway, I.; Kostewicz, E.; et al. Synthesis and biological evaluation of polysulfated oligosaccharide glycosides as inhibitors of angiogenesis an d tumor growth. J. Med. Chem. 2010, 53, 1686–1699.
- 23. Barragan-Montero, V.; Awwad, A.; Combemale, S.; de Santa Barbara, P.; Jover, B.; Molès, J.P.; Montero, J.L. Synthesis of Mannose-6-Phosphate Analogues and their Utility as Angiogenesis Regulators. ChemMedChem 2011, 6, 1771–177 4.
- 24. Ionescu, C.; Sippelli, S.; Toupet, L.; Barragan-Montero, V. New mannose derivatives: The tetrazole analogue of manno se-6-phosphate as angiogenesis inhibitor. Bioorg. Med. Chem. Lett. 2016, 26, 636–639.
- 25. Combemale, S.; Assam-Evoung, J.N.; Houaidji, S.; Bibi, R.; Barragan-Montero, V. Gold Nanoparticles Decorated with Mannose-6-phosphate Analogues. Molecules 2014, 19, 1120–1149.
- 26. Volpert, O.; Jackson, D.; Bouck, N.; Linzer, D.I. The insulin-like growth factor II/Mannose 6 -phosphate receptor is requi red for proliferin-induced angiogenesis. Endocrinology 1996, 137, 3871–3876.
- 27. Doyagüez, G.E.; Carrero, P.; Madrona, A.; Rodriguez-Salamanca, P.; Martínez-Gualda, B.; Camarasa, M.J.; Jimeno, M. L.; Bennallack, P.R.; Finnell, J.G.; Tsang, T.M.; et al. Galloyl Carbohydrates with Antiangiogenic Activity Mediated by Ca pillary Morphogenesis Gene 2 (CMG2) Protein Binding. J. Med. Chem. 2019, 62, 3958–3970.
- 28. Cryan, L.M.; Bazinet, L.; Habeshian, K.A.; Cao, S.; Clardy, J.; Christensen, K.A.; Rogers, M.S. 1,2,3,4,6-Penta-O-galloy l-β-D-glucopyranose inhibits angiogenesis via inhibition of capillary morphogenesis gene 2. J. Med. Chem. 2013, 56, 1 940–1945.
- 29. Huh, J.E.; Lee, E.O.; Kim, M.S.; Kang, K.S.; Kim, C.H.; Cha, B.C.; Surh, Y.J.; Kim, S.H. Penta-O-galloyl-beta-D-glucose suppresses tumor growth via inhibition of angiogenesis and stimulation of apoptosis: Roles of cyclooxygenase-2 and m

itogen-activated protein kinase pathways. Carcinogenesis 2005, 26, 1436–1445.

- 30. Al Sabti, H. Therapeutic angiogenesis in cardiovascular disease. J. Cardiothorac. Surg. 2007, 2, 49.
- 31. Pandya, N.M.; Dhalla, N.S.; Santani, D.D. Angiogenesis—A new target for future therapy. Vascul. Pharmacol. 2006, 44, 265–274.
- 32. Bouıs, D.; Kusumanto, Y.; Meijer, C.; Mulder, N.H.; Hospers, G.A. A review on pro- and anti-angiogenic factors as target s of clinical intervention. Pharmacol. Res. 2006, 53, 89–103.
- 33. Al Kawas, H.; Saaid, I.; Jank, P.; Westhoff, C.C.; Denkert, C.; Pross, T.; Weiler, K.B.S.; Karsten, M.M. How VEGF-A and its splice variants affect breast cancer development—clinical implications. Cell Oncol. 2022, 45, 227–239.
- 34. Lopes-Coelho, F.; Martins, F.; Pereira, S.A.; Jacinta Serpa, J. Anti-Angiogenic Therapy: Current Challenges and Future Perspectives. Int. J. Mol. Sci. 2021, 22, 3765.
- 35. Nyberg, P.; Xie, L.; Kalluri, R. Endogenous Inhibitors of Angiogenesis. Cancer Res. 2005, 65, 3967–3979.
- 36. Iruela-Arispe, M.L.; Dvorak, H.F. Angiogenesis: A dynamic balance of stimulators and inhibitors. Thromb. Haemost. 199 7, 78, 672–677.
- 37. Gospodarowicz, D. Purification of a fibroblast growth factor from bovine pituitary. J. Biol. Chem. 1975, 250, 2515–2520.
- 38. Esch, F.; Baird, A.; Ling, N.; Ueno, N.; Hill, F.; Denoroy, L.; Klepper, R.; Gospodarowicz, D.; Böhlen, P.; Guillemin, R. Pri mary structure of bovine pituitary basic fibroblast growth factor (FGF) and comparison with the amino-terminal sequenc e of bovine brain acidic FGF. Proc. Natl. Acad. Sci. USA 1985, 82, 6507–6511.
- 39. Senger, D.R.; Galli, S.J.; Dvorak, A.M.; Perruzzi, C.A.; Harvey, V.S.; Dvorak, H.F. Tumor cells secrete a vascular perme ability factor that promotes accumulation of ascites fluid. Science 1983, 219, 983–9855.
- 40. Leung, D.W.; Cachianes, G.; Kuang, W.J.; Goeddel, D.V.; Ferrara, N. Vascular endothelial growth factor is a secreted a ngiogenic mitogen. Science 1989, 246, 1306–1309.
- 41. Guo, N.; Krutzsch, H.C.; Inman, J.K.; Roberts, D.D. Thrombospondin 1 and type I repeat peptides of thrombospondin 1 specifically induce apoptosis of endothelial cells. Cancer Res. 1997, 57, 1735–1742.
- 42. Good, D.J.; Polverini, P.J.; Rastinejad, F.; Le Beau, M.M.; Lemons, R.S.; Frazier, W.A.; Bouck, N.P. A tumor suppressor -dependent inhibitor of angiogenesis is immunologically and functionally indistinguishable from a fragment of thrombos pondin. Proc. Natl. Acad. Sci. USA 1990, 87, 6624–6628.
- 43. Manetti, F.; Corelli, F.; Botta, M. Fibroblast growth factors and their inhibitors. Curr. Pharm. Des. 2000, 6, 1897–1924.
- 44. Zakrzewska, M.; Marcinkowska, E.; Wiedlocha, A. FGF-1: From biology through engineering to potential medical applic ations. Crit. Rev. Clin. Lab. Sci. 2008, 45, 91–135.
- 45. Yayon, A.; Klagsbrun, M.; Esko, J.D.; Leder, P.; Omitz, D.M. Cell surface, heparin-like molecules are required for bindin g of basic fibroblast growth factor to its high affinity receptor. Cell 1991, 64, 841–848.
- 46. Roghani, M.; Mansukhani, A.; Dell'Era, P.; Bellosta, P.; Basilico, C.; Rifkin, D.B.; Moscatelli, D. Heparin increases the aff inity of basic fibroblast growth factor for its receptor but is not required for binding. J. Biol. Chem. 1994, 269, 3976–398 4.
- 47. Friesel, R.E.; Maciag, T. Molecular mechanisms of angiogenesis: Fibroblast growth factor signal transduction. FASEB J. 1995, 9, 919–925.
- 48. Hagedorn, M.; Bikfalvi, A. Target molecules for anti-angiogenic therapy: From basic research to clinical trials. Crit. Rev. Oncol. Hematol. 2000, 34, 89–110.
- 49. Meyer, M.; Clauss, M.; Lepple-Wienhues, A.; Waltenberger, J.; Augustin, H.G.; Ziche, M.; Lanz, C.; Büttner, M.; Rziha, H.J.; Dehio, C. A novel vascular endothelial growth factor encoded by Orf virus, VEGF-E, mediates angiogenesis via si gnalling through VEGFR-2 (KDR) but not VEGFR-1 (Flt-1) receptor tyrosine kinases. EMBO J. 1999, 18, 363–374.
- 50. Veikkola, T.; Alitalo, K. VEGFs, receptors and angiogenesis. Semin. Cancer Bio. 1999, 9, 211–220.
- 51. Koblizek, T.I.; Weiss, C.; Yancopoulos, G.D.; Deutsch, U.; Risau, W. Angiopoietin-1 induces sprouting angiogenesis in v itro. Curr. Biol. 1998, 8, 529–532.
- 52. Maisonpierre, P.C.; Suri, C.; Jones, P.F.; Bartunkova, S.; Wiegand, S.J.; Radziejewski, C.; Compton, D.; McClain, J.; Al drich, T.H.; Papadopoulos, N.; et al. Angiopoietin-2, a natural antagonist for Tie2 that disrupts in vivo angiogenesis. Sci ence 1997, 277, 55–60.
- 53. Yu, X.; Seegar, T.M.C.; Dalton, A.C.; Tzvetkova-Robev, D.; Goldgur, Y.; Rajashankar, K.R.; Nikolov, D.B.; Barton, W.A. Structural basis for angiopoietin-1–mediated signaling initiation. Proc. Natl. Acad. Sci. USA 2013, 110, 7205–7210.
- 54. Fagiani, E.; Christofori, G. Angiopoietins in angiogenesis. Cancer Lett. 2013, 328, 18–26.
- 55. Cheng, N.; Brantley, D.M.; Chen, J. The ephrins and Eph receptors in angiogenesis. Cytokine Growth Factor Rev. 200 2, 13, 75–85.
- 56. Salvucci, O.; Tosato, G. Essential roles of EphB receptors and EphrinB ligands in endothelial cell function and angioge nesis. Adv. Cancer Res. 2012, 114, 21–57.
- 57. Kullander, K.; Klein, R. Mechanisms and functions of Eph and ephrin signalling. Nat. Rev. Mol. Cell Biol. 2002, 3, 475–4 86.
- 58. Zitka, O.; Kukacka, J.; Krizkova, S.; Huska, D.; Adam, V.; Masarik, M.; Prusa, R.; Kizek, R. Matrix metalloproteinases. Curr. Med. Chem. 2010, 17, 3751–3768.
- 59. Johnson, L.L.; Dyer, R.; Hupe, D.J. Matrix metalloproteinases. Curr. Opin. Chem. Biol. 1998, 2, 466–471.
- 60. Boire, A.; Covic, L.; Agarwal, A.; Jacques, S.; Sherifi, S.; Kuliopulos, A. PAR1 is a matrix metalloprotease-1 receptor tha t promotes invasion and tumorigenesis of breast cancer cells. Cell 2005, 120, 303–313.
- 61. Sasaki, T.; Larsson, H.; Kreuger, J.; Salmivirta, M.; Claesson-Welsh, L.; Lindahl, U.; Hohenester, E.; Timpl, R. Structura l basis and potential role of heparin/heparan sulfate binding to the angiogenesis inhibitor endostatin. EMBO J. 1999, 1 8, 6240–6248.
- 62. O'Reilly, M.S.; Boehm, T.; Shing, Y.; Fukai, N.; Vasios, G.; Lane, W.S.; Flynn, E.; Birkhead, J.R.; Olsen, B.R.; Folkman, J. Endostatin: An endogenous inhibitor of angiogenesis and tumor growth. Cell 1997, 88, 277–285.
- 63. Kim, Y.M.; Jang, J.W.; Lee, O.H.; Yeon, J.; Choi, E.Y.; Kim, K.W.; Lee, S.-T.; Kwon, Y.G. Endostatin inhibits endothelial and tumor cellular invasion by blocking the activation and catalytic activity of matrix metalloproteinase. Cancer Res. 20 00, 60, 5410–5413.
- 64. Sudhakar, A.; Sugimoto, H.; Yang, C.; Lively, J.; Zeisberg, M.; Kalluri, R. Human tumstatin and human endostatin exhibi t distinct antiangiogenic activities mediated by αvβ3 and α5β1 integrins. Proc. Natl. Acad. Sci. USA 2003, 100, 4766–47 71.
- 65. O'Reilly, M.S.; Holmgren, L.; Shing, Y.; Chen, C.; Rosenthal, R.A.; Moses, M.; Lane, W.S.; Cao, Y.; Sage, E.; Folkman, J. Angiostatin: A novel angiogenesis inhibitor that mediates the suppression of metastases by a Lewis lung carcinoma. Cell 1994, 79, 315–328.
- 66. Geiger, J.H.; Cnudde, S.E. What the structure of angiostatin may tell us about its mechanism of action. J. Thromb. Hae most. 2004, 2, 23–34.
- 67. Troyanovsky, B.; Levchenko, T.; Mansson, G.; Matvijenko, O.; Holmgren, L. Angiomotin: An angiostatin binding protein t hat regulates endothelial cell migration and tube formation. J. Cell Biol. 2001, 152, 1247–1254.
- 68. Iruela-Arispe, M.L.; Lombardo, M.; Krutzsch, H.C.; Lawler, J.; Roberts, D.D. Inhibition of angiogenesis by thrombospon din-1 is mediated by 2 independent regions within the type 1 repeats. Circulation 1999, 100, 1423–1431.
- 69. Adams, J.; Lawler, J. The thrombospondin family. Curr. Biol. 1993, 3, 188–190.
- 70. Bornstein, P. Thrombospondins function as regulators of angiogenesis. J. Cell Commun. Signal 2009, 3, 189–200.
- 71. Mirochnik, Y.; Kwiatek, A.; Volpert, O.V. Thrombospondin and apoptosis: Molecular mechanisms and use for design of c omplementation treatments. Curr. Drug Targets 2008, 9, 851–862.
- 72. Pribluda, V.S.; Gubish, E.R., Jr.; Lavallee, T.M.; Treston, A.; Swartz, G.M.; Green, S.J. 2-Methoxyestradiol: An endogen ous antiangiogenic and antiproliferative drug candidate. Cancer Metastasis Rev. 2000, 19, 173–179.
- 73. Yue, T.L.; Wang, X.; Louden, C.S.; Gupta, S.; Pillarisetti, K.; Gu, J.L.; Hart, T.K.; Lysko, P.G.; Feuerstein, G.Z. 2-Methox yestradiol, an endogenous estrogen metabolite, induces apoptosis in endothelial cells and inhibits angiogenesis: Possi ble role for stress-activated protein kinase signaling pathway and Fas expression. Mol. Pharmacol. 1997, 51, 951–962.
- 74. Peach, C.J.; Mignone, V.W.; Arruda, M.A.; Diana, C.; Alcobia, D.C.; Stephen, J.; Hill, S.J.; Kilpatrick, L.E.; Woolard, J. Molecular Pharmacology of VEGF-A Isoforms: Binding and Signalling at VEGFR2. Int. J. Mol. Sci. 2018, 19, 1264.
- 75. Zhou, Y.; Zhu, X.; Cui, H.; Shi, J.; Yuan, G.; Shi, S.; Hu, Y. The Role of the VEGF Family in Coronary Heart Disease. Fr ont. Cardiovasc. Med. 2021, 8, 738325.
- 76. Ferrara, N.; Adamis, A.P. Ten years of anti-vascular endothelial growth factor therapy. Nat. Rev. Drug Discov. 2016, 15, 385–403.
- 77. Mateescu, G.O.; Comanescu, M.; Mehedinţi, R.; Niculescu, Z.; Bold, A.; Panduru, L.; Cernea, D. Immunohistochemical expression of growth factors in the exocrine pancreas of patients with chronic liver diseases. Rom. J. Morphol. Embryo l. 2010, 51, 303–307.
- 78. Braile, M.; Marcella, S.; Cristinziano, L.; Galdiero, M.R.; Luca Modestino, L.; Ferrara, A.L.; Varricchi, G.; Marone, G.; Lo ffredo, S. VEGF-A in Cardiomyocytes and Heart Diseases. Int. J. Mol. Sci. 2020, 21, 5294.
- 79. Oliver, R.C.; Tervonen, T. Diabetes-a risk factor for periodontitis in adults? J. Periodontol. 1994, 65, 530–538.
- 80. Yalda, B.; Offenbacher, S.; Collins, J.G. Diabetes as a modifier of periodontal disease expression. Periodontology 2000 1994, 6, 37–49.
- 81. Minchenko, A.; Bauer, T.; Salceda, S.; Caro, J. Hypoxic stimulation of vascular endothelial growth factor expression in v itro and in vivo. Lab. Investig. 1994, 71, 374–379. Available online: https://pubmed.ncbi.nlm.nih.gov/7933988/ (accesse d on 4 July 2022).
- 82. Teshima-Kondo, S.; Kondo, K.; Prado-Lourenco, L.; Gonzalez-Herrera, I.; Rokutan, K.; Bayard, F.; Arnal, J.F.; Prats, A. C. Hyperglycemia up-regulates translation of the fibroblast growth factor 2 mRNA in mouse aorta via internal ribosome entry site. FASEB J. 2004, 18, 1583–1585.
- 83. Groothius, P.G. Angiogenesis and vascular remodelling in female reproductive organs. Angiogensis 2005, 8, 87–88.
- 84. Gargett, C.E.; Rogers, P.A.W. Human endometrial angiogenesis. Reproduction 2001, 121, 181–186.
- 85. Okada, H.; Tsuzuki, T.; Shindoh, H.; Nishigaki, A.; Yasuda, K.; Kanzaki, H. Regulation of decidualization and angiogene sis in the human endometrium: Mini review. J. Obstet. Gynaecol. Res. 2014, 40, 1180–1187.
- 86. Chung, A.S.; Lee, J.; Ferrara, N. Targeting the tumour vasculature: Insights from physiological angiogenesis. Nat. Rev. Cancer 2010, 10, 505–514.
- 87. Boldeanu, L.; Dijmărescu, A.L.; Radu, M.; Siloşi, C.A.; Popescu-Drigă, M.V.; Poenariu, I.S.; Siloşi, I.; Boldeanu, M.V.; N ovac, M.B.; Novac, L.V. The role of mediating factors involved in angiogenesis during implantation. Rom. J. Morphol. E mbryol. 2020, 61, 665–672.
- 88. Singer, A.J.; Clark, R.A. Cutaneous Wound Healing. N. Engl. J. Med. 1999, 341, 738–746.
- 89. Pandya, N.M.; Dhalla, N.S.; Santani, D.D. Angiogenesis—A new target for future therapy. Vascul. Pharmacol. 2006, 44, 265–274.
- 90. Stephan, D.; Weltin, D.; Zaric, V.; Chapelon, D.; Da Silva, A.; Lugnier, C. Angiogenèse: De la physiologie à la thérapeuti que. Réanim. Urgences 2000, 9, 534–544.
- 91. Adamis, A.P.; Miller, J.W.; Bernal, M.T. Increased vascular endothelial growth factor levels in the vitreous of eyes with p roliferative diabetic retinopathy. Am. J. Ophthalmol. 1994, 118, 445–450.

Retrieved from https://encyclopedia.pub/entry/history/show/61887