## Therapeutic Approaches Utilizing Macrophage-Derived iNOS/NO in Cancer

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Nitric oxide and its production by iNOS is an established mechanism critical to tumor promotion or suppression. Macrophages have important roles in immunity, development, and progression of cancer and have a controversial role in pro-and anti-tumoral effects. The tumor microenvironment consists of tumor-associated macrophages (TAM), among other cell types that influence the fate of the growing tumor. Depending on the microenvironment and various cues, macrophages polarize into a continuum represented by the M1-like pro-inflammatory phenotype or the anti-inflammatory M2-like phenotype; these two are predominant, while there are subsets and intermediates. Manipulating their plasticity through programming or reprogramming of M2-like to M1-like phenotypes presents the opportunity to maximize tumoricidal defenses. The dual role of iNOS derived NO also influences TAM activity by repolarization to tumoricidal M1-type phenotype. Regulatory pathways and immunomodulation achieve this through miRNA that may inhibit the immunosuppressive tumor microenvironment.

Keywords: iNOS/NO ; cancer ; tumor-associated macrophage ; M1-M2 ; miRNA ; arginase ; cancer progression

### 1. Introduction

The tumor and the immune cell interaction are critical areas of study in cancer growth and progression. Macrophages have a controversial role in the tumor microenvironment with both anti- and pro- tumoral effects <sup>[1]</sup>. The gaseous transmitter and signaling molecule nitric oxide (NO) and the expression of inducible nitric oxide synthase (iNOS) that produces NO are noted to have dual roles in cancer, which is to either promote or inhibit tumor growth <sup>[2][3]</sup>. Therefore, NO shows potential either as a therapeutic agent in its own right or as a target molecule in cancer therapies.

NO is a small signaling molecule that is synthesized by three NO synthases (NOS) isoforms. The two isoforms neuronal NOS (nNOS or NOS I), and endothelial NOS (eNOS or NOS III) are constitutive. The third NOS isoform (NOS 2 or inducible NOS, iNOS) is negligible in resting cells and is induced by cytokines and bacterial lipopolysaccharide (LPS) <sup>[3][4]</sup>. NO plays an important role in cell growth, differentiation, and apoptosis <sup>[3][5]</sup>. The enzyme iNOS produces NO through the conversion of L-arginine into citrulline utilizing NADPH and oxygen. NO may also be associated with resistance to apoptosis <sup>[6]</sup> and immune escape <sup>[Z]</sup>. iNOS is induced by inflammatory cytokines <sup>[8]</sup> and is transcriptionally regulated <sup>[9]</sup>. It is well established and acknowledged that NO's role in cancer depends on its concentration, exposure duration in cells, cell-specific sensitivities, iNOS localization in tissues, and extracellular conditions <sup>[10][11][12]</sup>.

Antitumor effects of NO have been demonstrated even though its pro-tumor effects have largely dominated the scenarios. In terms of the tumorigenic effects, NO contributes to tumor growth and metastasis, regulates metabolism by the Warburg effect, and promotes cancer growth via high glycolytic activity <sup>[13]</sup>. Furthermore, antitumor iNOS activity is related to its cytotoxicity and immunogenic effects <sup>[14]</sup>. Therefore, NO-releasing hybrids are subjects of intensive investigations as potential anticancer drugs, either as single cytotoxic agents or in combination with standard radio- and chemotherapy <sup>[13]</sup>.

More recently, studies relate the chemo- and immunoresistance [I] in cancer cells with NO as a mediator for the events in the tumor microenvironment (TME) and as a bonafide molecular target [15][16].

# 2. Therapeutic Approaches Utilizing Macrophage-Derived iNOS/NO in Cancer

Antitumor strategies targeting TAMs include potential mechanisms of lowering TAM survival, reducing macrophage recruitment, and switching M2-like TAMs into an M1-like phenotype <sup>[17]</sup>. Utilizing the iNOS derived-NO or exogenous NO delivery, there has been some success with nano-therapeutics. In the tumor microenvironment, the nanoparticles of self-assembled poly(L-arginine) are taken up by the activated macrophages, followed by the hydrolytic release of L-arginine,

and conversion to NO by the iNOS of the TAM <sup>[18]</sup>. In low doses, the NO produced by this mechanism in tumor-bearing mice increased the angiogenesis of the tumor tissues, whereas the high doses led to tumor volume reduction and apoptotic tumor cell death <sup>[18]</sup>.

Immunotherapeutic approaches have demonstrated reprogramming M2 to M1 macrophages  $\frac{19|[20]}{19|[20]}$ . As stated earlier, induction of the innate immune response is initiated by activating the macrophage to M1-type, which produces NO/RNS, secrete TNF- $\alpha$ , IL-1 $\beta$ , and IL-6 with pro-inflammatory cytokines proteases such as MMP-9. Higher NO production activates downstream signaling pathways that perform a critical role in the cytotoxic activity of immune cells against tumor cells  $\frac{[12][21]}{[12]}$ . Furthermore, among other immune cells, NO synthesis in NK cells was shown to regulate their tumoricidal activity to some extent  $\frac{[22]}{2}$ .

#### 2.1. iNOS Inhibitors

In colon, breast, gastric, hepatocellular carcinoma, melanoma, ovarian, leukemia, gastric, prostate, esophageal, and cervical cancers, high iNOS expression has correlated relatively well with poor patient survival <sup>[23][24]</sup>. Thus, iNOS expression may be used as a biomarker of poor patient prognosis and perhaps survival <sup>[25][26]</sup>. In contrast, a favorable prognosis has been associated with high iNOS expression in ovarian <sup>[27]</sup> and non-small cell lung cancers <sup>[28]</sup>.

When cell lines expressing high levels of iNOS, such as that of triple-negative breast cancers, were treated with 1400W, which is a highly selective iNOS inhibitor, or L-NAME, which is a relatively selective eNOS inhibitor, or L-NMMA, which is pan-NOS inhibitor, they all reduced cell proliferation, migration, and mammosphere formation <sup>[29]</sup>. In a xenograft model of TNBC, treatment of mice with L-NAME, and L-NMMA significantly reduced tumor growth <sup>[29]</sup>. Administration of AG, another iNOS specific inhibitor to athymic nude mice bearing TNBC xenografts, abated tumor growth and metastatic burden <sup>[30]</sup>. Further, the growth of glioma <sup>[31]</sup> or melanoma <sup>[32]</sup> cells in xenografts was significantly reduced when iNOS was silenced in these cells before they were implanted. The overarching data from all of these studies is the observation that the enhanced growth of the iNOS-overexpressing tumors appears to be due to enhanced angiogenesis, reviewed in <sup>[3]</sup>.

iNOS deficient mice exhibited enhanced M1 macrophage polarization with no significant effects on M2 macrophages. L-NIL, an iNOS selective inhibitor, significantly enhanced M1 macrophage polarization in cell cultures from wild-type (WT) mice. Whereas the NO donor SNAP, suppressed M1 macrophage differentiation in WT and *iNOS<sup>-/-</sup>* cell cultures <sup>[33]</sup>.

#### 2.2. NO and Curcumin: A Natural Dietary Compound

Natural dietary compounds modified or formulated as nanoparticles induce iNOS in TAMs and show promise in reprogramming M2 to M1 states to produce antitumor effects <sup>[34]</sup>. A curcumin formulation containing two additional natural polyphenols, aptly named TriCurin, produced repolarization of M2 TAM that had higher Arg-1 expression into the M1 TAM population with higher iNOS expression. The underlying mechanism appears to be the suppression of activated STAT3 in M2 type TAM, which causes activation of STAT1, leading to the M1 phenotype. Co-activated transcription factors STAT1 and NF-kB initiate the expression of the iNOS and NO in M1 cells leading to tumor elimination <sup>[35][36]</sup>. Of note, these M1 phenotypes are low-IL10 and high-IL-12 and showed anti-glioblastoma activity <sup>[37]</sup>. Specifically, for TriCurin, the M1 TAM-derived IL-12 that was induced was responsible for the recruitment of NK cells and cytotoxic T lymphocytes, leading to the reduction in cervical cancer cells in xenograft tumors <sup>[36]</sup>.

#### 2.3. NO and Immunomodulation with microRNAs

MicroRNA-mediated regulation modulates macrophage states to the M2 or M1 phenotype <sup>[38]</sup>. Delivery with nanomaterials has been developed wherein a marked increase in iNOS expression occurs for inducing or reprogramming the TAMs to M1-state or repolarization from M2- to M1-state, which provides an antitumor immune response (**Table 1**). For example, Zhang et al. <sup>[35]</sup> designed lipid-coated calcium phosphonate nanoparticles which were further conjugated with mannose for specific delivery of miR155 to TAMs, which altered phenotype successfully from pro-tumor M2-like TAMs to antitumor M1-like TAMs, and therefore produced a potent antitumor immune response and inhibiting tumor growth, reviewed in <sup>[39]</sup>. Layered double hydroxides NPs that are miR-155 loaded are taken up by TAMs, increase their iNOS expression, decrease the expression level of phosphorylated STAT3 and ERK1/2 and activate NF-κB expression. Combined therapies showed improvement also, for example, these nanoparticles with carboplatin improved TC-1 tumor recession in animal models and prolonged overall survival <sup>[40]</sup>. Similarly, Parayath et al. <sup>[41]</sup> developed a CD44 targeting hyaluronic acid-poly(ethylenimine) (HA-PEI)-based nanoparticle for the delivery of miR-125b to peritoneal macrophages to promote M1-like TAMs activation in the lungs. In vivo results found a more than 6-fold increase in the ratio of M1 to M2 TAMs and a 300-fold increase in iNOS to Arg-1 in TAMs after treatment with HA-PEI-125b nanoparticles. The continued success of

inducing M1-like TAMs polarization opens many avenues in anticancer immunotherapy via NO and enhancement of ROS release (Figure 1).



**Figure 1.** Exogenous NO or iNOS-derived NO modulate the macrophage status. M2-type macrophages may be repolarized into M1 phenotype via regulatory miRNA. ROS-generated oxidative stress may produce a cytocidal profile and reverse tumor progression.

#### 2.4. NO-Releasing Nanoparticles

Synthetic compounds and various carriers also hold promise considering that years of iNOS focus have created avenues to mimic iNOS/ NO activity in various ways <sup>[1]</sup>. Nanoparticles are demonstrated to preferentially accumulate in macrophages after systemic administration <sup>[42]</sup>. Nanoparticles made to release NO produce cytotoxicity depending on the nanocarrier's chemical nature, the concentration of NO released and the cell type. Low concentrations of NO released will have a proliferative effect on tumor cells, whereas high NO flux is expected to have toxic effects. There is considerable interest in identifying delivery methods to modulate TAM polarization for cancer treatment. TAMs overexpress the macrophage mannose receptor and therefore, mannose functionalized nanoparticles are used for recognition and internalization <sup>[43]</sup>. Affinity to TAMs was improved via a mannose- conjugate modified on lipid-coated calcium phosphonate nanoparticles which delivered miRNA into TAMs in vitro <sup>[44]</sup>. Some NO-releasing nanoparticles and materials with potential use in cancer treatment are presented in **Table 1** and principal actions are discussed. Reviews of various types of nanoparticles and effects on TME are found in, <sup>[42][45][46]</sup>.

Certain NO-releasing nanoparticles have been designed for photo-release. Nanoparticles of supramolecular assemblies of cyclodextrin-based polymer contain a NO photo donor and a fluorophore/photochrome dyad with an average size of 30 nm releases NO by light input <sup>[47]</sup>. Photogenerated NO in human melanoma cancer cells showed cytotoxicity with light stimulation and low levels of cytotoxicity in the dark. Similarly, internalized nanoparticles NONOate into the endosomes and lysosomes of a cell, and cytotoxicity from NO-mediated apoptosis was found with NONOate modified silica nanoparticles which produced high NO flux <sup>[48]</sup>. However, the desired sustained NO release akin to endogenous production by iNOS/ NO was not obtained.

Gold nanoparticles with 2-mercapto-5-nitrobenzimidazole also photo release NO and a low dose has similar antitumor and cytotoxicity effects as cisplatin in HeLa, Siha (cervical cancer cell lines), MCF-7 (breast cancer cell lines), and A549 (lung cancer lines); wherein 80% lower dose of Gold nanoparticles was found to produce cytotoxicity as that of 10 g/mL of cisplatin <sup>[49]</sup>.

Nanoparticles and Effect on iNOS or NO	Model System or Cell Type	Effect	Reference
CD44 coated HA-PEI based NPs, miR-125b loaded, iNOS increased	Naïve and KRAS/p53 double mutant nonsmall cell lung cancer (NSCLC) mouse model	Specifically target peritoneal macrophages which reprogram lung TAMs into M1 type	[50]
Layered double hydroxides NPs, miR-155 loaded, acidity sensitive, taken up by TAM iNOS increased	TC-1 mouse tumor model Uptake by TAM Repolarize TAM into M1	Synergistic enhancement of therapeutic effects with programmed cell death-1 antibody (α-PD-1) antibody	[40]

Table 1. Nanoparticles releasing NO or inducing iNOS and effect on macrophages or TAM.

Nanoparticles and Effect on iNOS or NO	Model System or Cell Type	Effect	Reference
Lipid-coated calcium phosphonate, miR-155 conjugated mannose, iNOS increased	S180 mouse sarcoma model	Repolarize M2 into M1 TAMs Significant antitumor effect	[44]
Gold nanoparticles, Photo release of NO	HeLa	Low doses of Gold nanoparticles were found to produce cytotoxicity as that of 10 g/mL of cisplatin	[49]
Poly(D,L-lactic-co-glycolic) acid (PLGA), loaded with ruthenium nitrosyl compounds, NO releasing upon light irradiation	Melanoma B16-F10 cells	In vitro cytotoxicity assays showed cell death	[51]
Cyclodextrin and NO photorelease by a donor	HeLa, Melanoma, A431- Human squamous carcinoma, Melanoma	Phototoxicity cell mortality	(52) (53) (54)
Polymeric, NO-releasing	BE(2)-C, Neuroblastoma cell line	Cisplatin in combination with nanoparticles produced synergistic cytotoxicity	[55]
4-arm branched polymer, NO- releasing	Human head and neck cancer cell line human breast cancer cell lines	Improved cell mortality	[ <u>56</u> ]
Liposome, NO-releasing	Breast cancer cell lines MDA-MB- 231 and MDAMB-468	Improved cell mortality	[57]

In addition to NONOates, S-nitrosothiols (RSNOs) such as S-nitrosoglutathione (GSNO) were encapsulated into polymeric nanoparticles. GSNO was incorporated into polymeric nanoparticles consisting of diblock copolymers, which extended the RSNO stability. Because the combination of NO donors with classical chemotherapy agents is of considerable interest, GSNO-containing polymeric nanoparticles and cisplatin were used in in vitro experiments <sup>[55]</sup>. No-polymeric nanoparticles showed enhanced NO stability in aqueous media, were non-toxic and could efficiently release NO intracellularly <sup>[55]</sup>. Neuroblastoma cell lines treated with GSNO-containing polymeric nanoparticles followed by cisplatin provided sensitization of cells and lower IC50 of cisplatin. NONOate-multiarm polymer nanocarriers to tumor-bearing nude mice inhibited tumor growth and extended the average survival of the animals in 7 weeks compared with intravenous administration of the classical NO-donor prodrug JS-K, and owed to a steady NO release profile. In vivo models may be reexamined for effects on macrophages <sup>[55]</sup>.

It is important to note that nanoparticle-based studies on the release of NO and the effects on the TME or TAMs should use appropriate reference treatment conditions for accurate comparisons of the efficiency of nanoparticle-induced effects. Nanoparticle-induced macrophage programming effects are generally compared to small molecule-induced effects. Some anticancer drugs may suppress immune activity within tumors and promote tumor growth and continue to be used in the clinical landscape. Therefore, it is imperative to compare the effect of nanoparticle-treated macrophages to biomolecule-treated macrophages, for example, with biomolecules such as IFN-y, IL-4, IL-10, or LPS, or drug-treated macrophages for advancing the field immunotherapy based on NO.

#### References

- 1. Vannini, F.; Kashfi, K.; Nath, N. The dual role of iNOS in cancer. Redox Biol. 2015, 6, 334–343.
- 2. Kashfi, K. Anti-inflammatory agents as cancer therapeutics. Adv. Pharmacol. 2009, 57, 31-89.
- Kashfi, K. The dichotomous role of H(2)S in cancer cell biology? Déjà vu all over again. Biochem. Pharmacol. 2018, 149, 205–223.
- 4. Murphy, M.P. Nitric oxide and cell death. Biochim. Biophys. Acta 1999, 1411, 401–414.
- 5. Kashfi, K. Nitric oxide in cancer and beyond. Biochem. Pharmacol. 2020, 176, 114006.
- Engels, K.; Knauer, S.; Loibl, S.; Fetz, V.; Harter, P.; Schweitzer, A.; Fisseler-Eckhoff, A.; Kommoss, F.; Hanker, L.; Nekljudova, V.; et al. NO signaling confers cytoprotectivity through the survivin network in ovarian carcinomas. Cancer Res. 2008, 68, 5159–5166.

- Bailey, P.; Chang, D.K.; Forget, M.-A.; Lucas, F.A.S.; Alvarez, H.A.; Haymaker, C.; Chattopadhyay, C.; Kim, S.-H.; Ekmekcioglu, S.; Grimm, E.A.; et al. Exploiting the neoantigen landscape for immunotherapy of pancreatic ductal adenocarcinoma. Sci. Rep. 2016, 6, 35848.
- 8. Fukumura, D.; Kashiwagi, S.; Jain, R.K. The role of nitric oxide in tumour progression. Nat. Rev. Cancer 2006, 6, 521–534.
- 9. Nathan, C.; Xie, Q.W. Regulation of biosynthesis of nitric oxide. J. Biol. Chem. 1994, 269, 13725–13728.
- Mocellin, S.; Bronte, V.; Nitti, D. Nitric oxide, a double edged sword in cancer biology: Searching for therapeutic opportunities. Med. Res. Rev. 2007, 27, 317–352.
- McGinity, C.L.; Palmieri, E.; Somasundaram, V.; Bhattacharyya, D.; Ridnour, L.; Cheng, R.; Ryan, A.; Glynn, S.; Thomas, D.; Miranda, K.; et al. Nitric Oxide Modulates Metabolic Processes in the Tumor Immune Microenvironment. Int. J. Mol. Sci. 2021, 22, 7068.
- 12. Khan, F.H.; Dervan, E.; Bhattacharyya, D.D.; McAuliffe, J.D.; Miranda, K.M.; Glynn, S.A. The Role of Nitric Oxide in Cancer: Master Regulator or Not? Int. J. Mol. Sci. 2020, 21, 9393.
- Coulter, J.A.; McCarthy, H.O.; Xiang, J.; Roedl, W.; Wagner, E.; Robson, T.; Hirst, D.G. Nitric oxide—A novel therapeutic for cancer. Nitric Oxide 2008, 19, 192–198.
- De Boo, S.; Kopecka, J.; Brusa, D.; Gazzano, E.; Matera, L.; Ghigo, D.; Bosia, A.; Riganti, C. iNOS activity is necessary for the cytotoxic and immunogenic effects of doxorubicin in human colon cancer cells. Mol. Cancer 2009, 8, 108.
- 15. Lee, M.; Rey, K.; Besler, K.; Wang, C.; Choy, J. Immunobiology of Nitric Oxide and Regulation of Inducible Nitric Oxide Synthase. Results Probl. Cell Differ. 2017, 62, 181–207.
- 16. Mintz, J.; Vedenko, A.; Rosete, O.; Shah, K.; Goldstein, G.; Hare, J.; Ramasamay, R.; Arora, H. Current Advances of Nitric Oxide in Cancer and Anticancer Therapeutics. Vaccines 2021, 9, 94.
- Zheng, X.; Turkowski, K.; Mora, J.; Brüne, B.; Seeger, W.; Weigert, A.; Savai, R. Redirecting tumor-associated macrophages to become tumoricidal effectors as a novel strategy for cancer therapy. Oncotarget 2017, 8, 48436– 48452.
- Kudo, S.; Nagasaki, Y. A novel nitric oxide-based anticancer therapeutics by macrophage-targeted poly(l-arginine)based nanoparticles. J. Control. Release 2015, 217, 256–262.
- 19. Guiducci, C.; Vicari, A.P.; Sangaletti, S.; Trinchieri, G.; Colombo, M.P. Redirecting In vivo Elicited Tumor Infiltrating Macrophages and Dendritic Cells towards Tumor Rejection. Cancer Res. 2005, 65, 3437–3446.
- Sinha, P.; Clements, V.K.; Ostrand-Rosenberg, S. Reduction of myeloid-derived suppressor cells and induction of M1 macrophages facilitate the rejection of established metastatic disease. J. Immunol. 2005, 174, 636–645.
- 21. Zuo, S.; Song, J.; Zhang, J.; He, Z.; Sun, B.; Sun, J. Nano-immunotherapy for each stage of cancer cellular immunity: Which, why, and what? Theranostics 2021, 11, 7471–7487.
- 22. Hu, W.; Wang, G.; Huang, D.; Sui, M.; Xu, Y. Cancer Immunotherapy Based on Natural Killer Cells: Current Progress and New Opportunities. Front. Immunol. 2019, 10, 1205.
- Cheng, R.; Ridnour, L.A.; Glynn, S.A.; Switzer, C.H.; Flores-Santana, W.; Hussain, P.; Thomas, D.D.; Ambs, S.; Harris, C.C.; Wink, D.A. Nitric Oxide and Cancer: An Overview. In Nitric Oxide (NO) and Cancer: Prognosis, Prevention, and Therapy; Bonavida, B., Ed.; Springer: Berlin/Heidelberg, Germany, 2010; pp. 3–20.
- 24. Saied, E.M.; El-Etreby, N.M. The role and prognostic value of inducible nitric oxide synthase (iNOS) and interleukin-33 (IL-33) in serous and mucinous epithelial ovarian tumours. Ann. Diagn. Pathol. 2017, 27, 62–68.
- De Oliveira, G.A.; Cheng, R.Y.; Ridnour, L.A.; Basudhar, D.; Somasundaram, V.; McVicar, D.W.; Monteiro, H.P.; Wink, D.A. Inducible Nitric Oxide Synthase in the Carcinogenesis of Gastrointestinal Cancers. Antioxidants Redox Signal. 2017, 26, 1059–1077.
- Switzer, C.H.; Glynn, S.; Ridnour, L.A.; Cheng, R.; Vitek, M.P.; Ambs, S.; Wink, D.A. Nitric oxide and protein phosphatase 2A provide novel therapeutic opportunities in ER-negative breast cancer. Trends Pharmacol. Sci. 2011, 32, 644–651.
- 27. Anttila, M.A.; Voutilainen, K.; Merivalo, S.; Saarikoski, S.; Kosma, V.-M. Prognostic significance of iNOS in epithelial ovarian cancer. Gynecol. Oncol. 2007, 105, 97–103.
- 28. Puhakka, A.; Kinnula, V.; Napankangas, U.; Saily, M.; Koistinen, P.; Paakko, P.; Soini, Y. High expression of nitric oxide synthases is a favorable prognostic sign in non-small cell lung carcinoma. APMIS 2003, 111, 1137–1146.
- 29. Granados-Principal, S.; Liu, Y.; Guevara, M.L.; Blanco, E.; Choi, D.S.; Qian, W.; Patel, T.; A Rodriguez, A.; Cusimano, J.; Weiss, H.L.; et al. Inhibition of iNOS as a novel effective targeted therapy against triple-negative breast cancer.

Breast Cancer Res. 2015, 17, 1-16.

- Heinecke, J.L.; Ridnour, L.A.; Cheng, R.Y.S.; Switzer, C.H.; Lizardo, M.M.; Khanna, C.; Glynn, S.A.; Hussain, S.P.; Young, H.A.; Ambs, S.; et al. Tumor microenvironment-based feed-forward regulation of NOS2 in breast cancer progression. Proc. Natl. Acad. Sci. USA 2014, 111, 6323–6328.
- 31. Kostourou, V.; Cartwright, J.; Johnstone, A.P.; Boult, J.K.R.; Cullis, E.R.; Whitley, G.; Robinson, S.P. The role of tumourderived iNOS in tumour progression and angiogenesis. Br. J. Cancer 2010, 104, 83–90.
- 32. Sikora, A.G.; Gelbard, A.; Davies, M.A.; Sano, D.; Ekmekcioglu, S.; Kwon, J.; Hailemichael, Y.; Jayaraman, P.; Myers, J.N.; Grimm, E.A.; et al. Targeted Inhibition of Inducible Nitric Oxide Synthase Inhibits Growth of Human Melanoma In vivo and Synergizes with Chemotherapy. Clin. Cancer Res. 2010, 16, 1834–1844.
- 33. Lu, G.; Zhang, R.; Geng, S.; Peng, L.; Jayaraman, P.; Chen, C.; Xu, F.; Yang, J.; Li, Q.; Zheng, H.; et al. Myeloid cellderived inducible nitric oxide synthase suppresses M1 macrophage polarization. Nat. Commun. 2015, 6, 6676.
- 34. Piao, L.; Mukherjee, S.; Chang, Q.; Xie, X.; Li, H.; Castellanos, M.R.; Banerjee, P.; Iqbal, H.; Ivancic, R.; Wang, X.; et al. TriCurin, a novel formulation of curcumin, epicatechin gallate, and resveratrol, inhibits the tumorigenicity of human papillomavirus-positive head and neck squamous cell carcinoma. Oncotarget 2016, 8, 60025–60035.
- Zhang, X.; Tian, W.; Cai, X.; Wang, X.; Dang, W.; Tang, H.; Cao, H.; Wang, L.; Chen, T. Hydrazinocurcumin Encapsuled Nanoparticles "Re-Educate" Tumor-Associated Macrophages and Exhibit Anti-Tumor Effects on Breast Cancer Following STAT3 Suppression. PLoS ONE 2013, 8, e65896.
- 36. Mukherjee, S.; Hussaini, R.; White, R.; Atwi, D.; Fried, A.; Sampat, S.; Piao, L.; Pan, Q.; Banerjee, P. TriCurin, a synergistic formulation of curcumin, resveratrol, and epicatechin gallate, repolarizes tumor-associated macrophages and triggers an immune response to cause suppression of HPV+ tumors. Cancer Immunol. Immunother. 2018, 67, 761–774.
- Mukherjee, S.; Baidoo, J.; Fried, A.; Atwi, D.; Dolai, S.; Boockvar, J.; Symons, M.; Ruggieri, R.; Raja, K.; Banerjee, P. Curcumin changes the polarity of tumor-associated microglia and eliminates glioblastoma. Int. J. Cancer 2016, 139, 2838–2849.
- Squadrito, M.L.; Etzrodt, M.; De Palma, M.; Pittet, M.J. MicroRNA-mediated control of macrophages and its implications for cancer. Trends Immunol. 2013, 34, 350–359.
- 39. Lin, Y.-X.; Wang, Y.; Blake, S.; Yu, M.; Mei, L.; Wang, H.; Shi, J. RNA Nanotechnology-Mediated Cancer Immunotherapy. Theranostics 2020, 10, 281–299.
- Yang, L.; Sun, J.; Liu, Q.; Zhu, R.; Yang, Q.; Hua, J.; Zheng, L.; Li, K.; Wang, S.; Li, A. Synergetic Functional Nanocomposites Enhance Immunotherapy in Solid Tumors by Remodeling the Immunoenvironment. Adv. Sci. 2019, 6, 1802012.
- Parayath, N.N.; Gandham, S.K.; Leslie, F.; Amiji, M.M. Improved anti-tumor efficacy of paclitaxel in combination with MicroRNA-125b-based tumor-associated macrophage repolarization in epithelial ovarian cancer. Cancer Lett. 2019, 461, 1–9.
- 42. Reichel, D.; Tripathi, M.; Perez, J.M. Biological Effects of Nanoparticles on Macrophage Polarization in the Tumor Microenvironment. Nanotheranostics 2019, 3, 66–88.
- 43. Scodeller, P.; Gracia, L.S.; Kopanchuk, S.; Tobi, A.; Kilk, K.; Säälik, P.; Kurm, K.; Squadrito, M.L.; Kotamraju, V.R.; Rinken, A.; et al. Precision Targeting of Tumor Macrophages with a CD206 Binding Peptide. Sci. Rep. 2017, 7, 1–12.
- 44. Zang, X.; Zhang, X.; Zhao, X.; Hu, H.; Qiao, M.; Deng, Y.; Chen, D. Targeted Delivery of miRNA 155 to Tumor Associated Macrophages for Tumor Immunotherapy. Mol. Pharm. 2019, 16, 1714–1722.
- 45. Seabra, A.B.; de Lima, R.; Calderón, M. Nitric oxide releasing nanomaterials for cancer treatment: Current status and perspectives. Curr. Top. Med. Chem. 2015, 15, 298–308.
- 46. Yang, M.; Li, J.; Gu, P.; Fan, X. The application of nanoparticles in cancer immunotherapy: Targeting tumor microenvironment. Bioact. Mater. 2020, 6, 1973–1987.
- 47. Kandoth, N.; Vittorino, E.; Sciortino, M.T.; Parisi, T.; Colao, I.; Mazzaglia, A.; Sortino, S. A Cyclodextrin-Based Nanoassembly with Bimodal Photodynamic Action. Chem. A Eur. J. 2011, 18, 1684–1690.
- 48. Stevens, E.V.; Carpenter, A.W.; Shin, J.H.; Liu, J.; Der, C.J.; Schoenfisch, M.H. Nitric Oxide-Releasing Silica Nanoparticle Inhibition of Ovarian Cancer Cell Growth. Mol. Pharm. 2010, 7, 775–785.
- 49. Sudhesh, P.; Tamilarasan, K.; Arumugam, P.; Berchmans, S. Nitric Oxide Releasing Photoresponsive Nanohybrids As Excellent Therapeutic Agent for Cervical Cancer Cell Lines. ACS Appl. Mater. Interfaces 2013, 5, 8263–8266.
- 50. Parayath, N.N.; Parikh, A.; Amiji, M.M. Repolarization of Tumor-Associated Macrophages in a Genetically Engineered Nonsmall Cell Lung Cancer Model by Intraperitoneal Administration of Hyaluronic Acid-Based Nanoparticles

Encapsulating MicroRNA-125b. Nano Lett. 2018, 18, 3571-3579.

- Gomes, A.J.; Espreafico, E.M.; Tfouni, E. trans-(PF6)2 and incorporated in PLGA nanoparticles for the delivery of nitric oxide to B16-F10 cells: Cytotoxicity and phototoxicity. Mol. Pharm. 2013, 10, 3544–3554.
- Deniz, E.; Kandoth, N.; Fraix, A.; Cardile, V.; Graziano, A.C.E.; Furno, D.L.; Gref, R.; Raymo, F.M.; Sortino, S. Photoinduced Fluorescence Activation and Nitric Oxide Release with Biocompatible Polymer Nanoparticles. Chem.-A Eur. J. 2012, 18, 15782–15787.
- 53. Kandoth, N.; Kirejev, V.; Monti, S.; Gref, R.; Ericson, M.; Sortino, S. Two-Photon Fluorescence Imaging and Bimodal Phototherapy of Epidermal Cancer Cells with Biocompatible Self-Assembled Polymer Nanoparticles. Biomacromolecules 2014, 15, 1768–1776.
- Fraix, A.; Kandoth, N.; Manet, I.; Cardile, V.; Graziano, A.C.E.; Gref, R.; Sortino, S. An engineered nanoplatform for bimodal anticancer phototherapy with dual-color fluorescence detection of sensitizers. Chem. Commun. 2013, 49, 4459–4461.
- 55. Duong, H.T.T.; Kamarudin, Z.M.; Erlich, R.B.; Li, Y.; Jones, M.W.; Kavallaris, M.; Boyer, C.; Davis, T.P. Intracellular nitric oxide delivery from stable NO-polymeric nanoparticle carriers. Chem. Commun. 2012, 49, 4190–4192.
- 56. Duan, S.; Cai, S.; Yang, Q.; Forrest, M.L. Multi-arm polymeric nanocarrier as a nitric oxide delivery platform for chemotherapy of head and neck squamous cell carcinoma. Biomaterials 2012, 33, 3243–3253.
- 57. Lee, S.Y.; Rim, Y.; McPherson, D.D.; Huang, S.-L.; Kim, H. A novel liposomal nanomedicine for nitric oxide delivery and breast cancer treatment. Bio-Med. Mater. Eng. 2014, 24, 61–67.

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