

Anatomical Defects Associated with Repeat Breeder Cow Syndrome

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Repeat breeder cow (RBC) syndrome encompasses cows failing three or more times to become pregnant, but with the special characteristic that their estrous cycles have a normal duration. Farmers and technicians commonly feel incapable of addressing this reproductive issue, and after numerous unsuccessful attempts to conceive, it is often necessary to cull cows. Important economic losses are linked to RBC syndrome due to diagnosis, therapy, or reduced milk production, among others.

Keywords: cattle ; estrus ; repeat breeding ; cow

1. Introduction

Nowadays, dairy and beef cattle are being developed under demanding production systems due to increased competitiveness in their markets, amid an uncertain economic world situation characterized by high food and fuel prices, as well as the urgent climate emergency on our planet. In this context, farms need to be highly efficient and profitable, and the identification of non-productive animals is a priority to prevent economic losses. For many years, researchers and technicians have focused on the Repeat Breeder Cow (RBC) syndrome, in which cows exhibit estrus signs at apparently normal intervals (i.e., estrous cycles between 17 and 25 days) but repeatedly fail to be pregnant after at least three attempts, despite the absence of apparent anatomical abnormalities or infectious diseases ^{[1][2][3][4]}. While many diagnostic tools for identifying reproductive pathologies are available, it is not practical or feasible to employ all of them, as they can be expensive or technically unavailable. Consequently, cows may experience reproductive imbalances or diseases that are not correctly diagnosed. On some occasions, these subfertile cows may eventually conceive after three or more cycles, but the economic impact of delayed conception often necessitates culling the animals because they become unprofitable and, in the case of dairy cows, produce less milk than the cost of their maintenance.

This problem has been extensively studied in various countries and regions, and researchers widely agree on its significant impact on the farm economy. By reviewing different studies, the frequency of RBC syndrome has been reported as approximately 5% in Jordan ^[5], 9% in the UK ^[6], 10% in Sweden ^[3], 11% in Bangladesh ^[7], 14% in Japan ^[8], 22% in the United States ^[9], 23% in Australia ^[10] and India ^[11], 25% in Spain ^[12], 36% in Cuba ^[13] and Ethiopia ^[14], and even as high as 62% in Indonesia ^{[14][15]}. The vast variation between countries can be attributed to various management practices, including the aptitude, production system, number of cows per farm, or genetic merit (mainly based on milk yield), among others. Studies have shown higher percentages of RBC syndrome in tie-stall barns ^[15], where hormonal treatment for reproduction control was not used ^{[14][15]}, under tropical conditions ^{[11][13][15]}, or in farms with high culling rate ^[12], which could be associated with poor animal welfare ^[16]. This global perspective suggests that the incidence of RBC syndrome could potentially be reduced and controlled.

The majority of studies on RBC syndrome have focused on dairy cows because their reproductive management allows for better monitoring of animals, making it easier to track estrus signs and insemination records. In contrast, beef cattle are typically raised under extensive systems, which makes obtaining such records challenging. However, the implementation of new precision livestock farming technologies, such as GPS collars, accelerometers, or proximity tags, among others, is expected to improve animal data collection and management. These advancements hold the potential to aid in the identification of subfertile cows, including RBCs, which is especially relevant in the case of extensive beef cattle farming due to the difficulty of obtaining reproductive records, as previously mentioned.

The subfertility observed in RBCs can be attributed to various causes, not only of maternal origin but also related to bull defects (as subfertility, anatomical defects of reproductive organs, low frozen-thawed semen quality, difficulties for mounting due to low libido or lameness, high hierarchy of infertile bulls) or management mistakes (wrong site or timing of semen deposition by technician, inadequate heat detection). Multiple factors have been reported as contributing to the

occurrence of RBC syndrome, including subclinical endometritis [17], nutritional deficiencies [18], abnormal heat behavior or improper heat detection [19][20], mismanagement in artificial insemination (AI) [21], or endocrine dysfunctions [20][22]. In relation to hormonal alterations, factors such as elevated progesterone levels, abnormal follicular dynamics, delayed ovulation, and lower oocyte quality have been identified as responsible for subfertility in repeat breeder heifers [23]. Additionally, these causes are influenced by various risk factors that contribute to the occurrence of imbalances and the incidence of RBC syndrome, including factors such as age, parity, body condition, milk yield, environmental conditions, and peri- and postpartum imbalances, among others.

Successful reproductive performance in cows relies on the delicate interplay of hormone patterns, ovarian dynamics, estrus behavior, uterine functions, and mating or AI [23]. Several endogenous factors operating at the ovary, oviduct, or uterus can affect oocyte and embryo quality, thereby influencing the incidence of repeat breeding in dairy cows.

2. Anatomical Defects Associated with Repeat Breeder Cow Syndrome

Oviductal defects, such as tubal stenosis or occlusion, have been associated with the RBC syndrome [24]. These conditions prevent the movement of oocytes and spermatozoa through the oviducts, and if fertilization occurs, the passage of the embryo into the uterus will be difficult. In cows with oviductal alterations, their estrous cycle remains unaffected, showing a normal duration. Sometimes, these issues can be resolved spontaneously. Confirming oviductal patency is challenging, but various methods have been used in studies, including examination at slaughterhouses [25], ultrasonographic contrast liquid [26][27], and dyes (phenol-sulphon-phthalein) [24] or granules (starch) [25]. Tubal inflammation is commonly associated with endometritis, and it reduces sperm motility and impairs sperm–oviduct interaction. The transport of early embryos can also be compromised due to mucus accumulation and reduced ciliary beat frequency in the oviduct. Research has indicated that around 20% of RBCs show bilateral oviduct occlusion or stenosis, while unilateral defects are reported in 24% of cows [24], highlighting the significance of these defects in the etiology of RBC syndrome.

In the search for other causes of RBC syndrome, studies analyze cervical consistency and patency [28]. It is reported that RBCs with normal uterine findings may exhibit cervicitis, which is independent of endometritis [28]. This suggests the importance of conducting a comprehensive breeding soundness examination that includes evaluating the external genitalia, perineal/vulvar conformation, and internal genitalia via vaginocopy in RBC cases. Cervicitis is characterized by a prolapsed and swollen portio vaginalis cervicis with reddening and can negatively impact conception rates and the number of pregnant cows at 200 days into lactation [28]. Additionally, identifying abnormal vulvar conformation in cows is crucial, as it is associated with pneumovagina, vaginitis, or endometritis (or SCE). According to the definition of RBC syndrome, when reproductive inflammatory alterations are present in cows, they should not be considered RBCs [29]; however, certain inflammatory disorders cannot be correctly diagnosed. In other instances, cows may experience SCE, which negatively affects reproductive performance and can be included in the definition of RBC syndrome.

References

1. Casida, L.E. Present status of the repeat-breeder cow problem. *J. Dairy Sci.* 1961, 44, 2323–2329.
2. Heuwieser, W.; Oltenacu, P.A.; Lednor, A.J.; Foote, R.H. Evaluation of different protocols for prostaglandin synchronization to improve reproductive performance in dairy herds with low estrus detection efficiency. *J. Dairy Sci.* 1997, 80, 2766–2774.
3. Gustafsson, H.; Emanuelson, U. Characterisation of the repeat breeding syndrome in Swedish dairy cattle. *Acta Vet. Scand.* 2002, 43, 115–125.
4. Pérez-Marín, C.C.; España, F. Oestrus expression and ovarian function in repeat breeder cows, monitored by ultrasonography and progesterone assay. *Reprod. Dom. Anim.* 2007, 42, 449–456.
5. Ayalon, N. The repeat breeder problem. In *Proceedings of the 10th International Congress Animal Reproduction and AI, Urbana, IL, USA, 10–14 June 1984; Volume 4*, pp. 111–141.
6. Bartlett, P.C.; Kirk, J.H.; Mather, E.C. Repeated insemination in Michigan Holstein-Friesian cattle: Incidence, descriptive epidemiology and estimated economic impact. *Theriogenology* 1986, 26, 309–322.
7. Asaduzzaman, K.M.; Bhuiyan, M.M.U.; Rahman, M.M.; Bhattacharjee, J. Prevalence of repeat breeding and its effective treatment in cows at selected areas of Bangladesh. *Bangladesh J. Vet. Med.* 2016, 14, 183–190.
8. Yusuf, M.; Nakao, T.; Ranasinghe, R.B.; Gautam, G.; Long, S.T.; Yoshida, C.; Koike, K.; Hayashi, A. Reproductive performance of repeat breeders in dairy herds. *Theriogenology* 2010, 73, 1220–1229.

9. Bullman, D.C.; Lamming, G.E. Milk progesterone levels in relation to conception, repeat breeding and factors influencing acyclicity in dairy cows. *J. Reprod. Fert.* 1978, 54, 447–458.
10. Moss, N.; Lean, I.J.; Reid, S.W.J.; Hodgson, D.R. Risk factors for repeat-breeder syndrome in New South Wales dairy cows. *Prev. Vet. Med.* 2002, 54, 91–103.
11. Deka, R.P.; Magnusson, U.; Grace, D.; Randolph, T.F.; Shome, R.; Lindahl, J.F. Estimates of the economic cost caused by five major reproductive problems in dairy animals in Assam and Bihar, India. *Animals* 2021, 11, 3116.
12. García-Ispuerto, I.; López-Gatius, F. Progesterone supplementation in the early luteal phase after artificial insemination improves conception rates in high-producing dairy cows. *Theriogenology* 2017, 90, 20–24.
13. Zambrano, A.; Avila, O.; Albi, E. La repetición de servicio de IA en el ganado vacuno y su repercusión económica. In *Proceedings of the ISCAB, La Habana, Cuba, 1982*.
14. Eshete, T.; Demisse, T.; Yilma, T.; Tamir, B. Repeat breeding and its' associated risk factors in crossbred dairy cattle in Northern Central Highlands of Ethiopia. *Vet. Med. Int.* 2023, 2023, 1176924.
15. Yusuf, M.; Rahim, L.; Asja, M.A.; Wahyudi, A. The incidence of repeat breeding in dairy cows under tropical condition. *Media Peternak.* 2012, 35, 28–31.
16. De Vries, M.; Bokkers, E.A.; Dijkstra, T.; Van Schaik, G.; De Boer, I.J. Invited review: Associations between variables of routine herd data and dairy cattle welfare indicators. *J. Dairy Sci.* 2011, 94, 3213–3228.
17. Salasel, B.; Mokhtari, A.; Taktaz, T. Prevalence, risk factors for and impact of subclinical endometritis in repeat breeder dairy cows. *Theriogenology* 2010, 74, 1271–1278.
18. Talukdar, D.J.; Papori, T.; Ahmed, K. Minerals and its impact on fertility of livestock: A review. *Agr. Rev.* 2016, 4, 333–337.
19. Cummins, S.B.; Lonergan, P.; Evans, A.C.; Butler, S.T. Genetic merit for fertility traits in Holstein cows: II. Ovarian follicular and corpus luteum dynamics, reproductive hormones, and estrus behavior. *J. Dairy Sci.* 2012, 95, 3698–3710.
20. Sood, P.; Zachut, M.; Dube, H.; Moallem, U. Behavioral and hormonal pattern of repeat breeder cows around estrus. *Reproduction* 2015, 149, 545–554.
21. Walsh, S.W.; Williams, E.J.; Evans, A.C. A review of the causes of poor fertility in high milk producing dairy cows. *Anim. Reprod. Sci.* 2011, 123, 127–138.
22. Kafi, M.; Azari, M.; Chashnigir, O.; Gharibzadeh, S.; Aghabozorgi, Z.; Asaadi, A.; Divar, M.R. Inherent inferior quality of follicular fluid in repeat breeder heifers as evidenced by low rates of in vitro production of bovine embryos. *Theriogenology* 2017, 102, 29–34.
23. Bage, R.; Gustafsson, H.; Larsson, B.; Forsberg, M.; Rodriguez-Martinez, H. Repeat breeding in dairy heifers: Follicular dynamics and estrous cycle characteristics in relation to sexual hormone patterns. *Theriogenology* 2002, 57, 2257–2269.
24. Garrido, M.R.; Peña, A.I.; Herradon, P.G.; Becerra, J.J.; Sande, J.; Quintela, L.A. Evaluation of tubal patency in repeat breeder Holstein cows. *Span. J. Agric. Res.* 2020, 17, e04SC02.
25. Kessy, B.M.; Noakes, D.E. The use of the starch grain and phenolsulphonphthalein tests to investigate infertile cows. *Vet. Rec.* 1979, 24, 489–491.
26. Kauffold, J.; Groeger, S.; Bergmann, K.; Wehrend, A. Use of contrast sonography to test for tubal patency in dairy cows. *J. Reprod. Dev.* 2009, 55, 335–338.
27. Itoh, K.; Endo, N.; Kataoka, S.I.; Tanaka, T. Assessment of tubal patency by hysterosalpingo–contrast sonography in cow. *J. Anim. Sci.* 2016, 94, 56–57.
28. Hartmann, D.; Rohkohl, J.; Merbach, S.; Heilkenbrinker, T.; Klindworth, H.P.; Schoon, H.A.; Hoedemaker, M. Prevalence of cervicitis in dairy cows and its effect on reproduction. *Theriogenology* 2016, 85, 247–253.
29. Dhara, S.; Mahajan, M.; Arya, D.; Kumar, S. Surgical management of pneumovagina in a cross bred cow: A case report. *Indian J. Anim. Health* 2022, 61, 368–370.