

# Polycaprolactone-Based Biocomposites

Subjects: [Materials Science](#), [Biomaterials](#)

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The developments within the topic of biomaterials has taken hold of researchers due to the mounting concern of current environmental pollution as well as scarcity resources. Amongst all compatible biomaterials, polycaprolactone (PCL) is deemed to be a great potential biomaterial, especially to the tissue engineering sector, due to its advantages, including its biocompatibility and low bioactivity exhibition. The commercialization of PCL is deemed as infant technology despite of all its advantages. This contributed to the disadvantages of PCL, including expensive, toxic, and complex.

polycaprolactone

green biocomposites

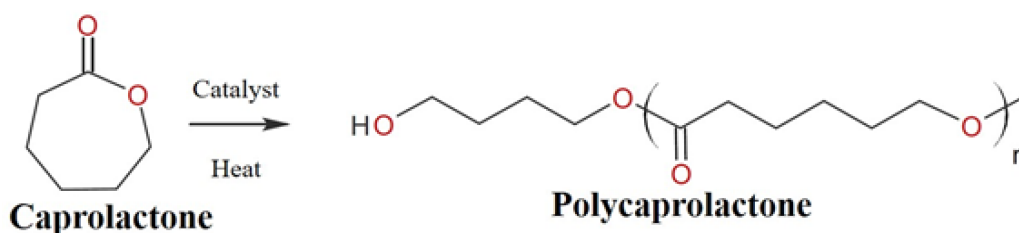
hybrid biocomposites

mechanical properties

thermal properties

## 1. Introduction

Among several types of biopolymers, polycaprolactone (PCL) has received a lot of attention due to its several advantages, such as its biodegradability, high strength, and biocompatibility. It can withstand water, oil, solvents, and chlorine. PCL is a semicrystalline ester polymer that is derived from ring-opening polymerization of  $\epsilon$ -caprolactone monomers, as shown in **Figure 1**. PCL consists of a glass transition temperature ( $T_g$ ) of around 60 °C and a melting point ranging between 59–64 °C, dictated by the crystalline nature of PCL which enables easy formability at relatively low temperatures [\[1\]](#).



**Figure 1.** Synthesis of PCL.

In contrast, the average molecular weight of PCL samples can range from 3000 to 90,000 g/mol, and they can be classified based on their molecular weight. With increasing molecular weight, its crystallinity decreases. High solubility, a low melting point, and exceptional blend compatibility have sparked a lot of interest in its potential biomedical applications [\[2\]](#). **Table 1** summarizes the effect of PCL molecular weight on their properties [\[3\]](#).

**Table 1.** Properties of PCL with different molecular weights.

Molecular Weight	Melting Point, °C	Tensile Stress, N/m <sup>2</sup>	Elongation at Break, %
37,000	58–60	$1.37 \times 10^7$	660
50,000	58–60	$3.53 \times 10^7$	800
80,000	60–62	$5.69 \times 10^7$	900

PCL also shows great electrospinning properties, as it can be spun into fibers at temperatures around 200 °C, without undergoing thermal degradation. Since PCL is a synthetic material, it is possible to achieve high material purity. PCL also has relatively long biodegradable time. According to the literature, PCL can biodegrade in a few months to many years, depending on its molecular weight, degree of crystallinity, shape, porosity, sample thickness, and the surrounding environment [4], unlike traditional plastics, such as polypropylene (PP) and polyethylene (PE), which take hundreds or even thousands of years to fully decay.

Recently, due to the environmental concerns, as well as energy and cost considerations, increased research efforts have been directed to the creation of bio-based materials. Bio-based polymers, which minimize reliance on petrochemical-based synthetic polymers, contribute significantly to global environmental sustainability. Recently, Gokhan et al. (2020) [5] has synthesized bio-based PCL from soybean oil-derived polyol via ring-opening polymerization. He found that the soybean-based PCL containing higher PCL molar ratio has poorer biodegradability but higher hydrophobicity and thermal characteristics compared to the others. Nonetheless, biobased polymers possess several weaknesses, including inferior mechanical properties, insufficient heat tolerance, and high moisture sensitivity relative to petroleum-derived polymers [6].

As mentioned in a previous statement, this polymer is well known for being biodegradable and have notably lower degradation. Due to its slow degradation rate, PCL is often used as a material in drug delivery devices that is active for a long time, with a period of over one year [7]. The material is also noted to be potentially used as a bioscaffold [8][9]. In terms of its biodegradability, permeability, and inability to establish an acidic environment, PCL is a better polymer than PLA or PGA in terms of possible biomedical uses [10]. Furthermore, because of the low rate of PCL degradation compared to PLA and PGA, it can be utilized to release medications over longer periods of time, even a year. The hydrolytic cleavage of ester groups is involved in PCL degradation. It is easy to predict that inserting different co-monomer units in the PCL chain will result in diverse properties, including changes in biodegradability, allowing for biodegradability control through targeted introduction of such groups [11]. Manivasagam et al. (2019) [12] have mentioned the application of PCL in dentistry by modifying the material structure with other materials (such as hydrophilic polyethylene glycol (PEG) and ceramic) and introducing co-polymers (such as polylactic acid (PLA) or polyglycolic acid (PGA)). The pKa of the degradation products; the primary mechanisms of the degradation; and the in vivo degradation rate of PCL, PLA, and PGA are summarized in **Table 2**.

**Table 2.** The degradation behaviour of the biodegradable polyesters. Reproduce from ref. [13].

Polyester	Degradation By-Products (pKa)	In Vivo Degradation Rate	Degradation Mechanism
PCL	Caproic acid (4.88)	50% in 4 years 1% in 6 months	Hydrolytic degradation
PLA	Lactic acid (3.85) Lactic acid (3.08)	50% in 1–2 years 98% in 12 months 100% in >12 months 100% in 12–16 month	Hydrolysis through the action of enzymes
PGA	Glycolic acid (3.83)	100% in 2–3 month 100% in 6–12 months	Both enzymatic and non- enzymatic hydrolysis

Unfortunately, because PCL manufacture is both complex and expensive, its wider commercialization has been limited. In addition, the material adheres poorly to cells due to its hydrophobic surface. PCL solvents are also known to be toxic, which can potentially harm human beings. The relatively low melting point is also demonstrates another drawback, since it hinders the material from being applied at higher temperatures [14][15][16]. **Table 3** shows the summary of advantages and disadvantages of PCL.

**Table 3.** Advantages and disadvantages of PCL.

Advantages	Disadvantages
High biocompatibility	Adheres poorly to cells
Highly biodegradable	Toxic solvent
Great electrospinning properties	Low melting point
Long biodegradable time	Complex and expensive production
High material purity	

Nonetheless, those limitations of PCL can potentially be overcome by the use of PCL-based biocomposites. Extensive efforts have been made over the last decade in the development of PCL-based biocomposites. The properties of PCL can be improved by blending with other polymers or fibres, allowing more people to benefit from its excellent properties [17][18][19][20]. Mechanical and thermal properties are the most influential, affected by the addition of fillers to the PCL [21][22][23]. Several studies have shown that the PCL-based biocomposites can be benefited for several applications, especially in the biomedical field [15][24][25][26][27][28].

## 2. Applications of Polycaprolactone-Based Biocomposites

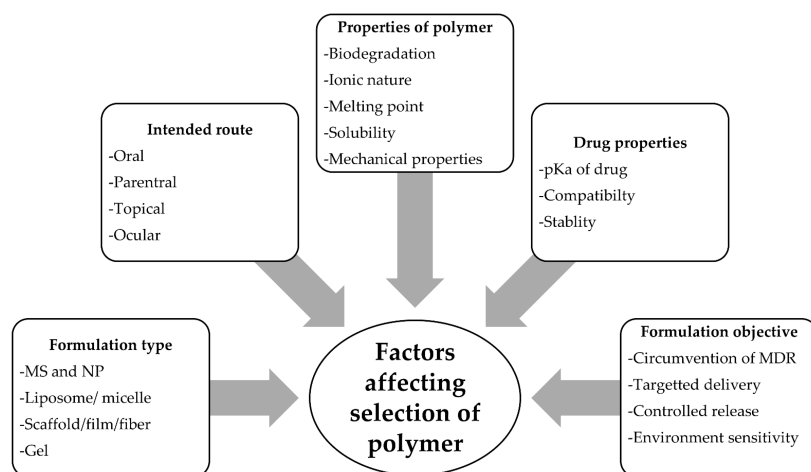
PCL has been widely applied in biomedical fields, especially in tissue engineering and medical implants, because they are biodegradable and have high biocompatibility. Manivasagam et al. (2019) [12] mentioned that polycaprolactones are being explored extensively in bone tissue engineering due to a lack of bioactivity and high degradation rates. Miller et al. (2011) [9] have stated that PCL can potentially be used as a bioscaffold.

Nevertheless, when it comes to the application of tissue engineering, PCL suffers from some shortcomings, such as poor mechanical properties, slow degradation rate, and low cell adhesion. The reinforcement of bioactive glasses and calcium phosphate-based ceramics within PCL has created a class of hybrid biomaterials with improved controllable degradation rates, good mechanical properties, and enhanced bioactivity, making them suitable for bone tissue engineering [29]. PCL green biocomposites with superior mechanical properties can potentially be used as orthopaedic implants, as briefly outlined as.

The Food and Medication Administration (FDA) has approved PCL for use in specified applications in the human body as a suture, drug delivery device, or adhesion barrier [7][30][31][32]. Following the recent launch of a PCL-based microsphere dermal filler belonging to the collagen stimulator class (Ellansé), PCL is being employed in the field of human aesthetics [33][34]. PCL-based products have also been utilized to treat facial ageing indications, such as contour laxity and volume loss, by stimulating collagen formation, resulting in an immediate and long-lasting natural impact [30][34][35][36][37][38]. In addition, it is being studied as a scaffold for tissue engineering-mediated injury repair using the guided bone regeneration (GBR) membrane, which has been extensively reported by numerous researchers [39][40][41][42][43][44][45]. It has been utilized as a hydrophobic block in amphiphilic synthetic block copolymers that are used to build the vesicle membrane of polymersomes [46][47].

Recently, PCL green biocomposites have been developed and commercialized. The majority of PCL green biocomposites are currently in the research and development stage. To make green biocomposites at a cheaper cost, new processing techniques and technologies are being developed [48]. Hao and co-workers found that the twin screw extrusion was used to make PCL–CNC nanocomposites. Microcellular nanocomposite samples were created utilising microcellular injection moulding and a physical blowing agent of carbon dioxide (CO<sub>2</sub>). The biocompatibility of the material was examined. The green hue in the fluorescence photographs symbolises live cells, whereas the red colour depicts dead cells. In comparison to bare dead cells, the clean PCL sample exhibits a substantial number of living cells. It was also found that 0.5% CNC and 1% CNC samples had mostly live cells, thus indicating good compatibility between the substrate and the cells.

PCL beads have been used to encapsulate a range of medicines for controlled re-release and targeted drug delivery [49][50][51][52][53][54][55][56][57][58][59]. In addition to polymeric properties several other factors, such as the type and objective of formulation, the route of administration, drug or polymeric properties etc., also affect the selection of the polymer (**Figure 2**). Hence, selection of a polymer is an important step for the development of a successful drug delivery system or device [60]. PCL-based modified polymers demonstrated so far predominantly involve its copolymers with several other polymers in different forms. Nevertheless, other modifications (blends and composite) were also reported in different formulations [61][62].



**Figure 2.** Factors affecting polymer selection.

Among biodegradable polymers modified for amelioration of properties, a special focus is made on PCL mainly due to its broad spectrum of compatibility with a wide range of other polymers. Its versatile nature, ease of fabrication, and biocompatibility establish it to be the polymer of interest by investigators worldwide for drug delivery and tissue engineering applications [63][64][65][66][67]. However, when scientists consider properties of unmodified PCL, there are considerable restrictions for its use. For example, its hydrophobic nature does not allow the facile release of hydrophobic drugs (from prepared formulations) and micelle formation. Furthermore, long-term degradation (ranging from weeks to months) slows down tissue replacement in the case of scaffolds, mechanical property limits its application to hard tissue engineering only, and nonreactivity is unsuitable for preparation of NC. Therefore, attempts have been made to overcome these undesirable properties by various types of modification, for successful application in pharmaceutical formulations. To great excitement, the use of modified PCL dominated over the past decade by virtue of which PCL was demonstrated in almost all novel formulations overcoming the above-mentioned restrictions. Additionally, functionalization as a result of PCL modifications is a featured advantage and is considered as another cause for this typical preference for modified PCL.

Additionally, the possibility of using PCL to be used as implants for targeted drug delivery has been explored. Boia et al. (2019) [68] have developed a novel way of using porous polycaprolactone as an intraocular implant to deliver dexamethasone to replace eye drops or intravitreal injections. The implants were made by using green supercritical carbon dioxide foaming or mixing methods, resulting in the implant having high porosity and high surface area. This, in turn, will cause a higher degradation rate than typical PCL-based implants, which have a relatively slower degradation rate. They are then inserted into adult rats for further observation. The implant is shown to have good biocompatibility, since it does not cause cell death or reduce the number of neurons [68]. In contrast, Hivechi et al. (2019) [69] investigated the regulated release of tetracycline hydrochloride using CNC-reinforced PCL nanofibers. The amount of CNC in the PCL nanofibers was increased, which resulted in a delayed release of a medication.

### 3. Conclusions

PCL is truly a promising material for the future to replace the current materials that are impossible and expensive to reuse. The aim is to investigate the potential of PCL biocomposites reinforced with natural fibres to enhance the quality of the produced biocomposites. Blending and processing of PCL into biocomposites has improved the materials' properties, thus providing more area to exploit these excellent properties. Herein, green biocomposites and hybrid biocomposites of PCL were evaluated in terms of their mechanical and thermal properties. The characteristics and properties of natural fibres can greatly influence the final properties of PCL-based biocomposites. Most of the studies reported that the performance of the PCL biocomposites improved, as compared to the neat PCL after blended with natural fibres. Moreover, emerging uses of nanofillers, such as nanocellulose and MMT, also managed to greatly improve the performance of PCL-based green and hybrid biocomposites. Besides that, PC-based biocomposites have huge utilization and potential within drug delivery devices, medical devices, and tissue engineering.

## References

1. Sisson, A.L.; Ekinici, D.; Lendlein, A. The contemporary role of  $\epsilon$ -caprolactone chemistry to create advanced polymer architectures. *Polymer* 2013, 54, 4333–4350.
2. Mohamed, R.M.; Yusoh, K. A Review on the Recent Research of Polycaprolactone (PCL). *Adv. Mater. Res.* 2015, 1134, 249–255.
3. Jiang, L.; Zhang, J. *7 Biodegradable and Biobased Polymers*, 2nd ed.; Elsevier Inc.: Amsterdam, The Netherlands, 2017; ISBN 9780323390408.
4. Nevoralová, M.; Koutný, M.; Ujčič, A.; Starý, Z.; Šerá, J.; Vlková, H.; Šlouf, M.; Fortelný, I.; Kruliš, Z. Structure Characterization and Biodegradation Rate of Poly( $\epsilon$ -caprolactone)/Starch Blends. *Front. Mater.* 2020, 7, 141.
5. Acik, G. Bio-based Poly( $\epsilon$ -caprolactone) from Soybean-Oil Derived Polyol via Ring-Opening Polymerization. *J. Polym. Environ.* 2020, 28, 668–675.
6. Johansson, C.; Bras, J.; Mondragon, I.; Nechita, P.; Plackett, D.; Šimon, P.; Svetec, D.G.; Virtanen, S.; Baschetti, M.G.; Breen, C.; et al. Renewable fibers and bio-based materials for packaging applications—A review of recent developments. *BioResources* 2012, 7, 2506–2552.
7. Woodruff, M.A.; Hutmacher, D.W. The return of a forgotten polymer—Polycaprolactone in the 21st century. *Prog. Polym. Sci.* 2010, 35, 1217–1256.
8. Patr, T.; Glória, A. Mechanical and biological behaviour of PCL and PCL/PLA scaffolds for tissue engineering applications. *Chem. Eng. Trans.* 2013, 32, 1645–1650.
9. Miller, K.; Hsu, J.E.; Soslowsky, L.J. *Materials in Tendon and Ligament Repair*; Elsevier Ltd.: Amsterdam, The Netherlands, 2011; Volume 6, ISBN 9780080552941.

10. Malikmammadov, E.; Tanir, T.E.; Kiziltay, A.; Hasirci, V.; Hasirci, N. PCL and PCL-based materials in biomedical applications. *J. Biomater. Sci. Polym. Ed.* 2018, 29, 863–893.
11. Todea, A.; Bîtcă, I.; Aparaschivei, D.; Păușescu, I.; Badea, V.; Péter, F.; Gherman, V.D.; Rusu, G.; Nagy, L.; Kéki, S. Biodegradable Oligoesters of  $\epsilon$ -Caprolactone and 5-Hydroxymethyl-2-Furancarboxylic Acid Synthesized by Immobilized Lipases. *Polymer* 2019, 11, 1402.
12. Manivasagam, G.; Reddy, A.; Sen, D.; Nayak, S.; Mathew, M.T.; Rajamanikam, A. *Dentistry: Restorative and Regenerative Approaches*; Elsevier: Amsterdam, The Netherlands, 2019; Volume 1–3, ISBN 9780128051443.
13. Manavitehrani, I.; Fathi, A.; Badr, H.; Daly, S.; Shirazi, A.N.; Dehghani, F. Biomedical applications of biodegradable polyesters. *Polymers* 2016, 8, 20.
14. Arunagiri, V.; Prasannan, A.; Udomsin, J.; Lai, J.-Y.; Wang, C.-F.; Hong, P.-D.; Tsai, H.C. Facile fabrication of eco-friendly polycaprolactone (PCL)/Poly-D, L-Lactic acid (PDLLA) modified melamine sorbent for oil-spill cleaning and water/oil (W/O) emulsion separation. *Sep. Purif. Technol.* 2021, 259, 118081.
15. Kim, S.; Gwon, Y.; Park, S.; Kim, W.; Jeon, Y.; Han, T.; Jeong, H.E.; Kim, J. Synergistic effects of gelatin and nanotopographical patterns on biomedical PCL patches for enhanced mechanical and adhesion properties. *J. Mech. Behav. Biomed. Mater.* 2021, 114, 104167.
16. Valerini, D.; Tamaro, L.; Vitali, R.; Guillot, G.; Rinaldi, A. Sputter-Deposited Ag Nanoparticles on Electrospun PCL Scaffolds: Morphology, Wettability and Antibacterial Activity. *Coatings* 2021, 11, 345.
17. Ebrahimifar, M.; Taherimehr, M. Evaluation of in-vitro drug release of polyvinylcyclohexane carbonate as a CO<sub>2</sub>-derived degradable polymer blended with PLA and PCL as drug carriers. *J. Drug Deliv. Sci. Technol.* 2021, 63, 102491.
18. El-Naggar, M.E.; Shalaby, E.S.; Abd-Al-Aleem, A.H.; Abu-Saied, M.A.; Youssef, A.M. Synthesis of environmentally benign antimicrobial dressing nanofibers based on polycaprolactone blended with gold nanoparticles and spearmint oil nanoemulsion. *J. Mater. Res. Technol.* 2021, 15, 3447–3460.
19. Herrero-Herrero, M.; Alberdi-Torres, S.; González-Fernández, M.L.; Vilariño-Feltrer, G.; Rodríguez-Hernández, J.C.; Vallés-Lluch, A.; Villar-Suárez, V. Influence of chemistry and fiber diameter of electrospun PLA, PCL and their blend membranes, intended as cell supports, on their biological behavior. *Polym. Test.* 2021, 103, 107364.
20. Doganci, M.D. Effects of star-shaped PCL having different numbers of arms on the mechanical, morphological, and thermal properties of PLA/PCL blends. *J. Polym. Res.* 2021, 28, 11.
21. Ouled Ltaief, A.; Ghorbel, N.; Benhamou, K.; Arous, M.; Kaddami, H.; Kallel, A. Impact of cellulose nanocrystals reinforcement on molecular dynamics and dielectric properties of PCL-based polyurethane. *Polym. Compos.* 2021, 42, 2737–2750.

22. dos Santos Filho, E.A.; Siqueira, D.D.; Araújo, E.M.; Luna, C.B.B.; de Medeiros, E.P. The Impact of the Macaíba Components Addition on the Biodegradation Acceleration of Poly ( $\epsilon$ -Caprolactone) (PCL). *J. Polym. Environ.* 2021, 1–18.
23. Reis, R.S.; Souza, D. de H.S.; Marques, M. de F.V.; da Luz, F.S.; Monteiro, S.N. Novel bionanocomposite of polycaprolactone reinforced with steam-exploded microfibrillated cellulose modified with ZnO. *J. Mater. Res. Technol.* 2021, 13, 1324–1335.
24. Ramamoorthy, R.; Andiappan, M.; Muthalagu, M. Preparation and characterization of Terminalia bellerica loaded PCL nanofibrous mats for biomedical applications. *Mater. Today Proc.* 2021, 45, 7247–7252.
25. Balan, R.; Gayathri, V. In-vitro and antibacterial activities of novel POT/TiO<sub>2</sub>/PCL composites for tissue engineering and biomedical applications. *Polym. Bull.* 2021, 1–18.
26. El Fawal, G.; Hong, H.; Mo, X.; Wang, H. Fabrication of scaffold based on gelatin and polycaprolactone (PCL) for wound dressing application. *J. Drug Deliv. Sci. Technol.* 2021, 63, 102501.
27. Ma, S.; Jiang, Z.; Wang, M.; Zhang, L.; Liang, Y.; Zhang, Z.; Ren, L.; Ren, L. 4D printing of PLA/PCL shape memory composites with controllable sequential deformation. *Bio-Design Manuf.* 2021, 4, 867–878.
28. Al-Kaabi, W.J.; Albukhaty, S.; Al-Fartosy, A.J.M.; Al-Karagoly, H.K.; Al-Musawi, S.; Sulaiman, G.M.; Dewir, Y.H.; Alwahibi, M.S.; Soliman, D.A. Development of Inula graveolens (L.) Plant Extract Electrospun/Polycaprolactone Nanofibers: A Novel Material for Biomedical Application. *Appl. Sci.* 2021, 11, 828.
29. Hajiali, F.; Tajbakhsh, S.; Shojaei, A. Fabrication and Properties of Polycaprolactone Composites Containing Calcium Phosphate-Based Ceramics and Bioactive Glasses in Bone Tissue Engineering: A Review. *Polym. Rev.* 2018, 58, 164–207.
30. Moers-Carpi, M.M.; Sherwood, S. Polycaprolactone for the Correction of Nasolabial Folds: A 24-Month, Prospective, Randomized, Controlled Clinical Trial. *Dermatol. Surg.* 2013, 39, 457–463.
31. Hao, Y.; Chen, Y.; He, X.; Yang, F.; Han, R.; Yang, C.; Li, W.; Qian, Z. Near-infrared responsive 5-fluorouracil and indocyanine green loaded MPEG-PCL nanoparticle integrated with dissolvable microneedle for skin cancer therapy. *Bioact. Mater.* 2020, 5, 542–552.
32. Peng, W.; Jiang, X.; Zhu, Y.; Omari-Siaw, E.; Deng, W.; Yu, J.; Xu, X.; Zhang, W. Oral delivery of capsaicin using MPEG-PCL nanoparticles. *Acta Pharmacol. Sin.* 2015, 36, 139–148.
33. Kim, J.A.; Van Abel, D. Neocollagenesis in human tissue injected with a polycaprolactone-based dermal filler. *J. Cosmet. Laser Ther.* 2015, 17, 99–101.



34. Christen, M.-O.; Vercesi, F. Polycaprolactone: How a Well-Known and Futuristic Polymer Has Become an Innovative Collagen-Stimulator in Esthetics. *Clin. Cosmet. Investig. Dermatol.* 2020, 13, 31–48.
35. de Melo, F.; Carrijo, A.; Hong, K.; Trumbic, B.; Vercesi, F.; Waldorf, H.A.; Zenker, S. Minimally Invasive Aesthetic Treatment of the Face and Neck Using Combinations of a PCL-Based Collagen Stimulator, PLLA/PLGA Suspension Sutures, and Cross-Linked Hyaluronic Acid. *Clin. Cosmet. Investig. Dermatol.* 2020, 13, 333–344.
36. de Melo, F.; Nicolau, P.; Piovano, L.; Lin, S.-L.; Baptista-Fernandes, T.; King, M.I.; Camporese, A.; Hong, K.; Khattar, M.; Christen, M.-O. Recommendations for volume augmentation and rejuvenation of the face and hands with the new generation polycaprolactone-based collagen stimulator (Ellansé®). *Clin. Cosmet. Investig. Dermatol.* 2017, 10, 431–440.
37. Morera Serna, E.; Serna Benbassat, M.; Terré Falcón, R.; Murillo Martín, J. Anatomy and Aging of the Perioral Region. *Facial Plast. Surg.* 2021, 37, 176–193.
38. Goodwin, P. Collagen stimulation with a range of polycaprolactone dermal fillers. *J. Aesthetic Nurs.* 2018, 7, 22–28.
39. Fujihara, K.; Kotaki, M.; Ramakrishna, S. Guided bone regeneration membrane made of polycaprolactone/calcium carbonate composite nano-fibers. *Biomaterials* 2005, 26, 4139–4147.
40. Won, J.-Y.; Park, C.-Y.; Bae, J.-H.; Ahn, G.; Kim, C.; Lim, D.-H.; Cho, D.-W.; Yun, W.-S.; Shim, J.-H.; Huh, J.-B. Evaluation of 3D printed PCL/PLGA/  $\beta$ -TCP versus collagen membranes for guided bone regeneration in a beagle implant model. *Biomed. Mater.* 2016, 11, 055013.
41. Ji, W.; Yang, F.; Ma, J.; Bouma, M.J.; Boerman, O.C.; Chen, Z.; van den Beucken, J.J.J.P.; Jansen, J.A. Incorporation of stromal cell-derived factor-1 $\alpha$  in PCL/gelatin electrospun membranes for guided bone regeneration. *Biomaterials* 2013, 34, 735–745.
42. Shim, J.-H.; Yoon, M.-C.; Jeong, C.-M.; Jang, J.; Jeong, S.-I.; Cho, D.-W.; Huh, J.-B. Efficacy of rhBMP-2 loaded PCL/PLGA/  $\beta$ -TCP guided bone regeneration membrane fabricated by 3D printing technology for reconstruction of calvaria defects in rabbit. *Biomed. Mater.* 2014, 9, 065006.
43. Türkkan, S.; Pazarçeviren, A.E.; Keskin, D.; Machin, N.E.; Duygulu, Ö.; Tezcaner, A. Nanosized CaP-silk fibroin-PCL-PEG-PCL/PCL based bilayer membranes for guided bone regeneration. *Mater. Sci. Eng. C* 2017, 80, 484–493.
44. Ren, K.; Wang, Y.; Sun, T.; Yue, W.; Zhang, H. Electrospun PCL/gelatin composite nanofiber structures for effective guided bone regeneration membranes. *Mater. Sci. Eng. C* 2017, 78, 324–332.
45. Castro, A.G.B.; Diba, M.; Kersten, M.; Jansen, J.A.; van den Beucken, J.J.J.P.; Yang, F. Development of a PCL-silica nanoparticles composite membrane for Guided Bone Regeneration.

- Mater. Sci. Eng. C 2018, 85, 154–161.
46. Ahmed, F.; Discher, D.E. Self-porating polymersomes of PEG–PLA and PEG–PCL: Hydrolysis-triggered controlled release vesicles. *J. Control. Release* 2004, 96, 37–53.
  47. Köthe, T.; Martin, S.; Reich, G.; Fricker, G. Dual asymmetric centrifugation as a novel method to prepare highly concentrated dispersions of PEG-b-PCL polymersomes as drug carriers. *Int. J. Pharm.* 2020, 579, 119087.
  48. Zwawi, M. A Review on Natural Fiber Bio-Composites, Surface Modifications and Applications. *Molecules* 2021, 26, 404.
  49. Grossen, P.; Witzigmann, D.; Sieber, S.; Huwyler, J. PEG-PCL-based nanomedicines: A biodegradable drug delivery system and its application. *J. Control. Release* 2017, 260, 46–60.
  50. Rai, B.; Teoh, S.H.; Hutmacher, D.W.; Cao, T.; Ho, K.H. Novel PCL-based honeycomb scaffolds as drug delivery systems for rhBMP-2. *Biomaterials* 2005, 26, 3739–3748.
  51. Nabid, M.R.; Tabatabaei Rezaei, S.J.; Sedghi, R.; Niknejad, H.; Entezami, A.A.; Oskooie, H.A.; Heravi, M.M. Self-assembled micelles of well-defined pentaerythritol-centered amphiphilic A4B8 star-block copolymers based on PCL and PEG for hydrophobic drug delivery. *Polymer* 2011, 52, 2799–2809.
  52. Holländer, J.; Genina, N.; Jukarainen, H.; Khajeheian, M.; Rosling, A.; Mäkilä, E.; Sandler, N. Three-Dimensional Printed PCL-Based Implantable Prototypes of Medical Devices for Controlled Drug Delivery. *J. Pharm. Sci.* 2016, 105, 2665–2676.
  53. Gong, C.; Shi, S.; Wu, L.; Gou, M.; Yin, Q.; Guo, Q.; Dong, P.; Zhang, F.; Luo, F.; Zhao, X.; et al. Biodegradable in situ gel-forming controlled drug delivery system based on thermosensitive PCL–PEG–PCL hydrogel. Part 2: Sol–gel–sol transition and drug delivery behavior. *Acta Biomater.* 2009, 5, 3358–3370.
  54. Coombes, A.G.A.; Rizzi, S.C.; Williamson, M.; Barralet, J.E.; Downes, S.; Wallace, W.A. Precipitation casting of polycaprolactone for applications in tissue engineering and drug delivery. *Biomaterials* 2004, 25, 315–325.
  55. Chang, C.; Wei, H.; Quan, C.-Y.; Li, Y.-Y.; Liu, J.; Wang, Z.-C.; Cheng, S.-X.; Zhang, X.-Z.; Zhuo, R.-X. Fabrication of thermosensitive PCL-PNIPAAm-PCL triblock copolymeric micelles for drug delivery. *J. Polym. Sci. Part A Polym. Chem.* 2008, 46, 3048–3057.
  56. Danafar, H. MPEG–PCL copolymeric nanoparticles in drug delivery systems. *Cogent Med.* 2016, 3, 1142411.
  57. Zhang, J.; Zhao, S.; Zhu, M.; Zhu, Y.; Zhang, Y.; Liu, Z.; Zhang, C. 3D-printed magnetic Fe<sub>3</sub>O<sub>4</sub>/MBG/PCL composite scaffolds with multifunctionality of bone regeneration, local anticancer drug delivery and hyperthermia. *J. Mater. Chem. B* 2014, 2, 7583–7595.

58. Deng, H.; Dong, A.; Song, J.; Chen, X. Injectable thermosensitive hydrogel systems based on functional PEG/PCL block polymer for local drug delivery. *J. Control. Release* 2019, 297, 60–70.
59. Zhao, P.; Liu, L.; Feng, X.; Wang, C.; Shuai, X.; Chen, Y. Molecular Nanoworm with PCL Core and PEO Shell as a Non-spherical Carrier for Drug Delivery. *Macromol. Rapid Commun.* 2012, 33, 1351–1355.
60. Dash, T.K.; Konkimalla, V.B. Polymeric Modification and Its Implication in Drug Delivery: Poly- $\epsilon$ -caprolactone (PCL) as a Model Polymer. *Mol. Pharm.* 2012, 9, 2365–2379.
61. Hosseinkazemi, H.; Biazar, E.; Bonakdar, S.; Ebadi, M.-T.; Shokrgozar, M.-A.; Rabiee, M. Modification of PCL Electrospun Nanofibrous Mat With *Calendula officinalis* Extract for Improved Interaction With Cells. *Int. J. Polym. Mater. Polym. Biomater.* 2015, 64, 459–464.
62. Hosseini, Y.; Emadi, R.; Kharaziha, M. Surface modification of PCL-diopside fibrous membrane via gelatin immobilization for bone tissue engineering. *Mater. Chem. Phys.* 2017, 194, 356–366.
63. Kweon, H. A novel degradable polycaprolactone networks for tissue engineering. *Biomaterials* 2003, 24, 801–808.
64. Siddiqui, N.; Asawa, S.; Birru, B.; Baadhe, R.; Rao, S. PCL-Based Composite Scaffold Matrices for Tissue Engineering Applications. *Mol. Biotechnol.* 2018, 60, 506–532.
65. Kundu, J.; Shim, J.-H.; Jang, J.; Kim, S.-W.; Cho, D.-W. An additive manufacturing-based PCL-alginate-chondrocyte bioprinted scaffold for cartilage tissue engineering. *J. Tissue Eng. Regen. Med.* 2015, 9, 1286–1297.
66. Patrício, T.; Domingos, M.; Gloria, A.; Bártoło, P. Characterisation of PCL and PCL/PLA Scaffolds for Tissue Engineering. *Procedia CIRP* 2013, 5, 110–114.
67. Gautam, S.; Dinda, A.K.; Mishra, N.C. Fabrication and characterization of PCL/gelatin composite nanofibrous scaffold for tissue engineering applications by electrospinning method. *Mater. Sci. Eng. C* 2013, 33, 1228–1235.
68. Boia, R.; Dias, P.A.N.; Martins, J.M.; Galindo-Romero, C.; Aires, I.D.; Vidal-Sanz, M.; Agudo-Barriuso, M.; de Sousa, H.C.; Ambrósio, A.F.; Braga, M.E.M.; et al. Porous poly( $\epsilon$ -caprolactone) implants: A novel strategy for efficient intraocular drug delivery. *J. Control. Release* 2019, 316, 331–348.
69. Hivechi, A.; Bahrami, S.H.; Siegel, R.A. Drug release and biodegradability of electrospun cellulose nanocrystal reinforced polycaprolactone. *Mater. Sci. Eng. C* 2019, 94, 929–937.

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