Endothelial Dysfunction after Hematopoietic Stem Cell Transplantation

Subjects: Hematology | Transplantation Contributor: Giuseppe Milone

Endothelial dysfunction (ED) is frequently encountered in transplant medicine. After hematopoietic stem cell transplantation (HSCT), ED participates in the pathogenesis of various complications such as sinusoidal obstruction syndrome/veno-occlusive disease (SOS/VOD), graft-versus-host disease (GVHD), transplant-associated thrombotic microangiopathy (TA-TMA), idiopathic pneumonia syndrome (IPS), capillary leak syndrome (CLS), and engraftment syndrome (ES).

endothelial dysfunction hematopoietic stem cell transplantation (HSCT)

Therapeutic Interventions

Clinical

1. Introduction

The endothelium is a thin structure composed of a monolayer of flattened cells, covering the inner part of blood vessels and playing a pivotal role in vascular homeostasis ^[1]. The endothelium is provided with molecules and mechanisms with antithrombotic and anti-inflammatory functions and a set of molecules that retain prothrombotic properties. A balance among them maintains normal antithrombotic and anti-inflammatory status ^[2].

The shift from the basal state to the procoagulant and inflammatory condition is referred to as endothelial activation; prolonged endothelium activation will lead to endothelium dysfunction (ED).

ED refers to the inability of endothelium cells (ECs) to determine vasodilatation of the vessel wall. ED is associated with reduced nitric oxide (NO) production, increased adhesiveness of leukocytes and platelets, increased endothelium permeability, and finally, apoptosis of EC ^[3]. ED is believed to play an essential role in cardiovascular diseases, renal diseases, infections, liver diseases, and multiorgan failure ^[4]. More widely, dysfunction of the vascular endothelium has been considered a hallmark of human diseases ^[1]. Notably, an endothelial activation induced by cytokines may contribute to the pathophysiology of COVID 19 disease ^{[5][6][7]}.

ED is also frequently found in many complications arising in transplant medicine ^[8]. After allogeneic hematopoietic stem cell transplantation (HSCT), many clinical factors play pathogenetic roles in ED. Some are common to other clinical settings such as advanced age, diabetes, hypertension ^{[9][10]}. However, others such as alloreactivity, infections, immunosuppressive agents, and, in HSCT, pretransplant conditioning are specific to the transplanted patients ^{[11][12]}.

The term alloreactivity is widely accepted and used. Immunological reactions after an allogeneic HSCT are heterogeneous and may determine ED by different mechanisms. Class I and II histocompatibility antigens, expressed on EC, may be the targets of immunologic attack ^[13]. The expression of class II antigens in EC is induced by y interferon and down modulated by fluvastatin and everolimus ^[14].

After solid organ transplantation, antibody-mediated rejection is believed to represent antibody and complementdependent injury to the microvasculature. When rejection is diagnosed after solid organ transplantation, the alloimmune reaction is readily apparent from the histopathology of the transplanted organ (leukocytes infiltrate, vascular damage, complement deposition, thrombosis) ^{[15][16]}. It results in allograft dysfunction, allograft loss, and accelerated graft vasculopathy ^[17]. However, in the rejection setting, the immunological mechanisms may also involve a cytotoxic T-cell response or NK response ^{[16][18]}.

After allogeneic HSCT, the target of an alloimmune attack can be, at least theoretically, the entire vascular tree of the recipient. For instance, and as proof, graft-versus-host disease (GVHD) is associated with endothelium damage characterized, at the immunohistochemistry level, by perivascular infiltrate of activated lymphocyte and by an increased level of von Willebrand factor (v-WF) ^[19]. However, at gross histopathology, the evidence of alloimmune reaction is scarce. The reasons for that paucity have not yet been clarified ^[20].

Although alloimmunity can be the initiating trigger, other mechanisms besides direct cytotoxicity may occur, and innate immunity may take part in tissue damage without any histologically visible cellular effector mechanism ^[21]. In the context of innate immunity, the release of cytokine, along with the activation of complement and Toll-like receptors, is a potent mediator of tissue damage. In determining ED, infections or administration of pharmacologic agents may also act as important cofactors.

A three steps model has been proposed ^[22]. Predisposition for ED may be the first step. Conditioning and tissue damage (second steps) act on this baseline status to determine subclinical ED. Finally, as the third step, alloimmunity or infections or pharmacological agents may further increase the prothrombotic/proinflammatory status, causing the full-blown clinical picture.

2. Clinical Pictures of Endothelial Dysfunction after Allogeneic HSCT

From the clinician's point of view, the involvement of endothelium after allogeneic HSCT is frequent and may manifest in practice, with different clinical pictures. This issue has been the object of several reviews during the last decade ^{[12][23][24]}. A number of organ-specific diseases such as SOS/VOD, IPS, CLS, ES, and TA-TMA have their pathogenesis in EC dysfunction. All these diseases may terminate in multiorgan failures (MOFs). However, there is no agreement on which clinical picture has to be considered as derived from systemic endothelial dysfunction ^[24]. Primarily, venous-occlusive disease, capillary leak syndrome, and engraftment syndrome are not considered by all researchers as dependent on a systemic endothelial dysfunction ^[24]. In contrast, some researchers retain that EC has a relevant role in corticosteroid-refractory acute graft-versus-host disease.

2.1. SOS/VOD

SOS/VOD is characterized by increased bilirubin, body weight increase due to liquid retention, and painful liver enlargement. Conditioning intensity and conditioning type play significant roles, together with age, underlying diagnosis, and the state of liver parenchyma ^[25]. The main histopathological findings are round-up of EC, EC detachment, and downstream embolization of EC, together with hemorrhage in Disse space and narrowing of the centrum-lobular vein ^[26]. The increase in body weight, which can reach 10–20% of the basal value, and the lack of response to diuretic treatment, demonstrate that, in this disease, endothelium damage is systemic. Severe forms of SOS/VOD may progress to multiorgan failure with renal, lung, or CNS toxicity, thus confirming the systemic nature of this disease. SOS/VOD is more frequent in conditions of increased HLA distance between donor and recipient, but no concomitant and overt a-GVHD is evident in most of these cases.

In SOS/VOD, nitric oxide synthase activity is reduced in liver cells ^[27]. HMGB-1 has been found to be involved in models of VOD induced experimentally by monocrotaline ^[28]. An upsurge in the level of HMGB-1 follows the administration of Monocrotaline. Activation of endothelium cells in VOD is demonstrated by an increased level of vWF, ICAM1, VLA4, and Ang-2 ^[29].

2.2. Capillary Leak Syndrome (CLS)

An increase in body weight, blood pressure reduction, tachycardia, and sudden decrease in serum albumin is the cluster of clinical abnormalities found in capillary leak syndrome. In the idiopathic form, a monoclonal immunoglobulin is frequently present in the plasma. Secondary CLS may be associated with severe infections or with the administration of pharmacological agents, such as interleukin-2, GM-CSF, gemcitabine, and monoclonal antibodies anti-CD19 and anti-CD22. High serum levels of Ang-2 and VCAM1 have been found in patients affected by idiopathic CLS.

A significant increase in body weight (>2.5%) has been reported after allogeneic HSCT in 20–30% of all patients ^[30]. Severe hydric retention is associated with reduced survival and a higher risk of severe GVHD ^[31]. Patients at risk for CLS at the start of conditioning may be identified as having a high EASIX score ^[32].

2.3. Idiopathic Pneumonia Syndrome (IPS)

IPS criteria include evidence of widespread alveolar injury with symptoms and signs of pneumonia in the absence of active lower respiratory tract infection. Diagnosis is made after the exclusion of commonly found pulmonary infections. It requires an intensive diagnostic workup, including at least a bronchoalveolar lavage. Alloreactivity toward lung tissue after HSCT in SCID mice is accompanied by signs of activation of lung EC ^[33]. Further, in the development of experimental IPS, an injury to the vascular endothelium has been observed ^[34]. Vessels in the lung are surrounded by a dense mononuclear cell infiltrate. There is apoptosis of ECs, presence of activated cytoplasmic caspase 3, and TUNEL positivity of nuclei. Cytotoxicity via the Fas-FasL pathway contributes to the development of experimental IPS. A role for TNF alpha has been hypothesized ^[35]. The expression of ICAM-1, VCAM-1, and eNOS are increased in lung biopsies of patients developing IPS ^[36].

However, over half of the patients diagnosed with IPS have a virus detected in bronchoalveolar lavage (BAL) samples ^[37]. The significance of these viruses in the pathogenicity of pneumonia remains unclear, although emerging evidence suggests that at least in the case of human herpesvirus 6 (HHV-6), these viruses may lead to lung injury and raises plausible concern that IPS may have been misdiagnosed in earlier studies. Alloimmune reactions toward lung tissue and infections may interact and be cofactors.

An imbalance between Ang-1 Ang-2 may also have a role in IPS. This imbalance has been found in ARDS ^[38]. A four-endothelial biomarker panel, including elevated angiopoietin-2/angiopoietin-1 ratio, vascular cell-adhesion molecule, and von Willebrand factor, is useful in identifying acute respiratory distress syndrome ^[39].

2.4. Engraftment Syndrome (ES)

Diagnostic criteria for engraftment syndrome, according to T. Spitzer, include major criteria (non-infectious fever, skin rash, and non-cardiogenic pulmonary edema) and minor criteria (weight gain, hepatic/renal dysfunction, or transient encephalopathy) ^[40]. Diagnosis is reached with the development of two or more of the previously cited symptoms within 96 h of the start of neutrophil recovery (absolute neutrophil count > 100). According to Maiolino, diarrhea is a further criterion of this syndrome. In the time frame of peri-engraftment, it is possible also to observe lung abnormalities such as diffuse ground-glass opacities, often with septal thickening and small pleural effusions ^[41]. ES has been described after autologous transplantation in patients mainly affected by multiple myeloma, POEMS syndrome, amyloidosis, and autoimmune diseases ^{[42][43]}. ES is, however, possible in patients affected by other underlying diagnoses.

The distinction between ES and autologous GVHD is difficult since both syndromes may present the involvement of the skin and diarrhea. Indeed, the relationship between autologous GVHD and ES is still debated, and these two clinical pictures may represent the same disease. ES has overlapping signs also with capillary leak syndrome.

ES is frequent after syngeneic HSCT ^[44]. ES has also been described after allogeneic HSCT. Indeed, not-infectious fever and manifestations of a vascular leak (edema with weight gain) may occur during granulocyte recovery in both the auto- and allotransplantation settings ^[45]. ES may precede a-GVHD, these two diseases being temporally associated, or ES may represent an initial stage of a-GVHD ^[46]. Therefore, in the allogeneic setting, the distinction of ES and a-GVHD is a matter of debate.

2.5. Transplant-Associated Thrombotic Microangiopathy (TA-TMA)

Micro-angiopathic anemia is a predominating feature. Clinical and laboratory signs are anemia, schistocytes, hemolysis, increase in LDH and decreased haptoglobin, hypertension, fever, decreased renal function, and proteinuria ^{[47][48][49]}. It may involve the kidney, the central nervous system, and the intestinal tract, and can be associated with pulmonary hypertension and serosal surface effusions.

According to Jodelle, it may present in a severe form in up to 18% of all patients in a pediatric population. TRM in patients having signs of the disease may be as high as 48%.

TA-TMA has been considered an endothelial form of a-GVHD. In most of these cases, a previous a-GVHD episode is present, or a-GVHD at the time of TA-TMA diagnosis is still ongoing, although with apparently minimal clinical signs ^[50]. However, TA-TMA may also appear after autologous HSCT. The pretransplantation patient's risk factors are a-GVHD, previous transplantation, MUD donor, and myeloablative conditioning ^{[51][52]}.

In some cases, calcineurin inhibitors are significant contributors, and regression of signs has been reported after their discontinuation ^[48], although this remains a controversial issue ^[52]. In some other cases, infections may play essential roles as cofactors (aspergillus, HHV6, BK, adenovirus, CMV).

Intestinal TA-TMA is characterized by abdominal pain and may cause significant gastrointestinal bleeding. Differential diagnosis from intestinal a-GVHD may be problematic. Nishida et al. showed that intestinal thrombotic microangiopathy might mimic clinically a progressive a-GVHD ^[53]. Histopathology of intestinal biopsy shows microangiopathic changes in the gastrointestinal vasculature. Histologic features include endothelial cell swelling, endothelial cell separation, perivascular mucosal hemorrhage, intraluminal schistocytes, intraluminal fibrin, intraluminal microthrombi, loss of glands, and total denudation of mucosa ^{[54][55]}.

Laskin et al. have reported complement activation after TA-TMA in HSC transplantation. These researchers found in renal biopsy C4d in the vessel ^[56]. Indeed, complement activation has been demonstrated in this type of TMA with an increase in C5b-9 ^{[57][58]}. Subsequently, similar complement regulatory defects to those found in a-HUS were identified in a small series of pediatric patients affected by TA-TMA ^[59].

Complement activation due to genetic deletion of CFI and CFH has also been demonstrated in children suffering transplant-associated thrombotic microangiopathy ^[60]. However, further confirmations of this physiopathological view are needed, in children as well as in adults.

A two-step pathogenetic process of TA-TMA may be hypothesized. Genetic abnormalities such as deficiency of complement regulatory proteins (CFI, CFH, complement genes CFHR1-CFHR3) may be predisposing factors. Subsequently, high dose chemotherapy, a-GVHD, factor H autoantibody, or infections will lead to endothelium damage and complement activation ^{[61][52][59][62]}.

A reduced level of NO has also been found in TA-TMA ^[63]. High levels of NET predict TA-TMA; NETosi can determine complement activation and deposition of C5b-9. In TA-TMA, the immunosuppressive agents and infections are suspected of contributing as copathogens ^{[64][65]}.

The concentration of soluble products derived from complement activation (C5b-9, C3a) may be measured in plasma. It could help in predicting the diagnosis and severity of TA-TMA ^{[57][66][67][68]}.

2.6. Endothelial Dysfunction and GVHD

Conditioning and a-GVHD are linked to cytokine secretion and endothelium dysfunction. vWF levels measured after conditioning anticipate and predict GVHD ^[69]. A high level of Ang-2 at the start of conditioning and later early

after transplantation also predicts GVHD and TRM. Ang-2 levels since admission are higher in patients who will develop a-GVHD. Ang-2 remains high in patients affected by corticosteroid-refractory GVHD, while it is reduced in patients responding to treatment of GVHD. Endothelium damage is an important mechanism in the physiopathology of the disease, and it is FAS driven. There are some situations in which ED manifest during severe and protracted a-GVHD.

Patients with corticosteroid-refractory a-GVHD exhibited elevated serum levels of Ang-2, sTM, HGF, and IL-8 posttransplantation, compared with patients with sensitive a-GVHD and patients without a-GVHD (Dietrich et al. 2013). A high level of Ang-2 persisting after first-line therapy is a marker of corticosteroid-refractory GVHD ^[70], and ED may explain gastrointestinal signs and symptoms in these patients. Luft hypothesized that endothelial cell vulnerability and dysfunction, rather than refractory T-cell activity, drive the pathophysiology of corticosteroidrefractory GVHD ^[22]^[70].

A double hit has been proposed. In patients having a high Ang-2 level at pretransplantation (first hit), the occurrence of severe GVHD (second hit) will be followed by a risk of high NRM and a poor prognosis ^[71]. These data underline that endothelium damage may have an essential role in GVHD pathophysiology, especially in corticosteroid-refractory patients.

3. Therapeutic Interventions for EC Dysfunction

3.1. Defibrotide

Defibrotide is indicated in the severe form of SOS/VOD. In the INT study, the complete remission rate that can be achieved will depend on the age and the presence of MOF, and it varies between 40% and 70% ^[72]. Very severe forms of SOS/VOD have an unsatisfactory outcome ^[73].

In vitro activity on endothelial cells has been extensively studied, and it can reduce endothelial activation by lowering adhesion molecules expression and leukocyte adherence in several experimental models ^[74].

Defibrotide use has been explored in TM-TMA; in a limited number of patients affected with TM-TMA, the administration of a low dosage of defibrotide induced disease remission in all cases ^[75]. In a small study (<u>ClinicalTrials.gov</u> NCT03384693), administration of defibrotide as prophylaxis of TA-TMA resulted in very low NRM. The use of defibrotide as prophylaxis for acute GVHD in patients showing high risk has been proposed since a pediatric study found a reduction in transplantation toxicity and a reduction in severe GVHD ^[76]. In an experimental mouse model, prophylaxis with defibrotide reduces acute-GVHD and improves survival ^[77].

3.2. Anti-complement Agents

Blocking the complement system with eculizumab is currently the most effective treatment to circumvent the poor outcome in patients with severe TA-TMA ^[78].

In 2014, Jodelle reported that six patients were treated using eculizumab and 4/6 responded ^[79]. A single-center study reported eculizumab effective in 50% of adult patients, and a-GVHD was the only factor associated with inferior results ^[80].

At MD Anderson, in 5 years, 10 patients received eculizumab in an uncontrolled and retrospective study ^[81]. The anti-complement agent was associated with a change in immunosuppression. The OS in the eculizumab-treated cohort was better, compared with patients not receiving eculizumab. After transplantation, patients require modification of the dose according to CH50.

A recent meta-analysis on 116 patients suggests that eculizumab improves overall survival and response rate in patients with TA-TMA ^[82]. However, randomized, controlled trials and prospective studies are needed.

Narsoplimab is a monoclonal antibody able to inhibit MASP-2. It has been found effective in thrombotic microangiopathy of COVID-19 patients ^[83]. Narsoplimab has been studied in the treatment of IgA nephropathy, a disease in which the lectin pathway of complement is involved; narsoplimab reduced the progression of this kidney disease ^{[84][85]}. Narsoplimab has also been studied in TA-TMA, and preliminary results have been reported (EHA 2018).

Since these anti-complement agents have high costs, there is a need for diagnostic tests that can guide after allogeneic HSCT the selection of patients to be treated.

3.3. Anti-CD20 (Rituximab)

This agent is currently employed in cases of TMA not responsive to plasma exchange [86].

Moreover, it is used in the setting of TMA associated with LES ^[87]. Some cases of patients affected by TA-TMA and improvement after treatment with anti-CD20 have been reported ^{[88][89][90]}. In recipients after transplantation, the development of antibodies against factor H ^[59] might be an indication of anti-CD20 treatment.

3.4. Withdrawal of Calcineurin Inhibitors

Cyclosporin (CSA) modifies the endothelium, and it increases the synthesis of thromboxane A2 while decreasing the production of prostacyclin (PGI-2) [91][92].

CSA inhibits NOS ^[93]. Disturbances in constitutive and inducible NOS in the vascular wall may predispose to vasospasm, contributing to hypertension and vascular diseases. CSA inhibits angiogenesis induced by vascular endothelial growth factor (VEGF); VEGF activates the transcription of COX2, and CSA ^[69] inhibits this effect of VEGF on cyclooxygenase (Cox)-2 ^[94]. On this basis, withdrawal of CSA or switching to tacrolimus has been advocated in patients affected by TA-TMA ^[95]. However, other data do not support the usefulness of this practice ^[52].

3.5. Therapeutic Plasma Exchange

Therapeutic plasma exchange (TPE) has been widely used in treating TA-TMA since the procedure is active in TTP disease (Moschowitz's disease). However, its efficacy in the TA-TMA setting is limited, with response varying in the literature from 25% to 75% ^{[96][97][98]}. Clinically, the improvement is observed on the serum level of LDH and transfusion requirement, but survival remains very poor ^[98]. In TA-TMA patients, TPE is not able to prevent evolution into chronic kidney disease ^[99]. Moreover, the rate of complication of TPE is significant ^{[100][101][102]}.

3.6. Thrombomodulin

TM has been found clinically helpful as a therapy for DIC in children ^{[103][104][105]} and for ARDS ^[106]. After allogeneic hematopoietic stem cell transplantation, a study reported that TM administration significantly reduced a-GVHD and ameliorated OS ^[107]. This result has been confirmed in a subsequent study ^[108]. TM has been found effective, after allogeneic HSCT, in cases of TA-TMA, SOS/VOD, and ES ^{[109][110][111][112][113][114][115]}.

3.7. Statins

They are active in various diseases based on endothelial dysfunctions, such as cardiovascular disease and rheumatoid arthritis ^[116]. Statins have pleiotropic effects and can increase NO production in EC ^{[117][118]}. In a mice model, statins ameliorate the histopathologic signs of GVHD injury ^[83]. Statins increase the levels of Ang-1 ^[119]. In humans, prophylaxis using pravastatin after allogeneic HSCT reduces the incidence of SOS/VOD ^[120].

3.8. Angiopoietin1 Mimetics

Vasculotides is an Ang-1 mimetic provided with an anti-inflammatory effect. In animal models, it has been found to be helpful in various conditions such as hemorrhagic shock, pneumonia ^{[121][122]}, strokes ^{[123][124]}, and in preventing pathologic vascular leakage ^{[125][126]}.

3.9. Alpha-1Anti-Trypsin (A1AT)

A1AT has an inhibitory effect on the expression of genes induced by TNF-alpha in endothelial cells, thereby reducing endothelial cell activation ^[127]. A1AT is also able to reduce the harmful effects of heme on EC ^[128]. Additionally, A1AT has immunoregulatory effects and decreases the production of IL-8, IL-6, TNF-a, and IL-1b. It promotes the differentiation and expansion of FoxP3+ regulatory T cells (Tregs).

A1AT has a demonstrated role in treating corticosteroid-refractory acute GVHD. Their anti-inflammatory and immunoregulatory effects merit further studies in treating complications based on ED in the transplantation setting [129].

References

- 1. Rajendran, P.; Rengarajan, T.; Thangavel, J.; Nishigaki, Y.; Sakthisekaran, D.; Sethi, G.; Nishigaki, I. The vascular endothelium and human diseases. Int. J. Biol. Sci. 2013, 9, 1057–1069.
- 2. Neubauer, K.; Zieger, B. Endothelial cells and coagulation. Cell Tissue Res. 2021.
- Endemann, D.H.; Schiffrin, E.L. Endothelial dysfunction. J. Am. Soc. Nephrol. 2004, 15, 1983– 1992.
- 4. De Backer, D.; Orbegozo Cortes, D.; Donadello, K.; Vincent, J.-L. Pathophysiology of microcirculatory dysfunction and the pathogenesis of septic shock. Virulence 2014, 5, 73–79.
- 5. Martinod, K.; Wagner, D.D. Thrombosis: Tangled up in NETs. Blood 2014, 123, 2768–2776.
- Barnes, B.J.; Adrover, J.M.; Baxter-Stoltzfus, A.; Borczuk, A.; Cools-Lartigue, J.; Crawford, J.M.; Daßler-Plenker, J.; Guerci, P.; Huynh, C.; Knight, J.S.; et al. Targeting potential drivers of COVID-19: Neutrophil extracellular traps. J. Exp. Med. 2020, 217, e20200652.
- 7. Zuo, Y.; Yalavarthi, S.; Shi, H.; Gockman, K.; Zuo, M.; Madison, J.A.; Blair, C.; Weber, A.; Barnes, B.J.; Egeblad, M.; et al. Neutrophil extracellular traps in COVID-19. JCI Insight 2020, 5, e138999.
- 8. Penack, O.; Luft, T. Editorial: Endothelial Dysfunction During Inflammation and Alloimmunity. Front. Immunol. 2018, 9, 2886.
- 9. Higashi, Y.; Kihara, Y.; Noma, K. Endothelial dysfunction and hypertension in aging. Hypertens. Res. 2012, 35, 1039–1047.
- 10. Avogaro, A.; Albiero, M.; Menegazzo, L.; De Kreutzenberg, S.; Fadini, G.P. Endothelial dysfunction in diabetes: The role of reparatory mechanisms. Diabetes Care 2011, 34, 285–290.
- 11. Lia, G.; Giaccone, L.; Leone, S.; Bruno, B. Biomarkers for Early Complications of Endothelial Origin After Allogeneic Hematopoietic Stem Cell Transplantation: Do They Have a Potential Clinical Role? Front. Immunol. 2021, 12, 1869.
- 12. Carreras, E.; Diaz-Ricart, M. The role of the endothelium in the short-term complications of hematopoietic SCT. Bone Marrow Transplant. 2011, 46, 1495–1502.
- Suitters, A.; Rose, M.; Higgins, A.; Yacoub, M.H. MHC antigen expression in sequential biopsies from cardiac transplant patients--correlation with rejection. Clin. Exp. Immunol. 1987, 69, 575– 583.
- Maenaka, A.; Kenta, I.; Ota, A.; Miwa, Y.; Ohashi, W.; Horimi, K.; Matsuoka, Y.; Ohnishi, M.; Uchida, K.; Kobayashi, T. Interferon-γ-induced HLA Class II expression on endothelial cells is decreased by inhibition of mTOR and HMG-CoA reductase. FEBS Open Bio 2020, 10, 927–936.
- González-Molina, M.; Ruiz-Esteban, P.; Caballero, A.; Burgos, D.; Cabello, M.; Leon, M.; Fuentes, L.; Hernandez, D. Immune response and histology of humoral rejection in kidney transplantation. Nefrologia 2016, 36, 354–367.

- Loupy, A.; Haas, M.; Solez, K.; Racusen, L.; Glotz, D.; Seron, D.; Nankivell, B.J.; Colvin, R.B.; Afrouzian, M.; Akalin, E.; et al. The Banff 2015 Kidney Meeting Report: Current Challenges in Rejection Classification and Prospects for Adopting Molecular Pathology. Am. J. Transplant. 2017, 17, 28–41.
- Hammond, M.E.H.; Revelo, M.P.; Miller, D.V.; Snow, G.L.; Budge, D.; Stehlik, J.; Molina, K.M.; Selzman, C.H.; Drakos, S.G.; Rami, A.A.; et al. ISHLT pathology antibody mediated rejection score correlates with increased risk of cardiovascular mortality: A retrospective validation analysis. J. Heart Lung Transplant. 2016, 35, 320–325.
- 18. Zhang, X.; Reed, E.F. Effect of Antibodies on Endothelium. Am. J. Transplant. 2009, 9, 2459–2465.
- 19. Biedermann, B.C.; Sahner, S.; Gregor, M.; Tsakiris, D.A.; Jeanneret, C.; Pober, J.S.; Gratwohl, A. Endothelial injury mediated by cytotoxic T lymphocytes and loss of microvessels in chronic graft versus host disease. Lancet 2002, 359, 2078–2083.
- 20. Biedermann, B.C. Vascular endothelium and graft-versus-host disease. Best Pract. Res. Clin. Haematol. 2008, 21, 129–138.
- 21. Maeda, Y. Pathogenesis of graft-versus-host disease: Innate immunity amplifying acute alloimmune responses. Int. J. Hematol. 2013, 98, 293–299.
- 22. Luft, T.; Dreger, P.; Radujkovic, A. Endothelial cell dysfunction: A key determinant for the outcome of allogeneic stem cell transplantation. Bone Marrow Transplant. 2021, 56, 2326–2335.
- 23. Hildebrandt, G.C.; Chao, N. Endothelial cell function and endothelial-related disorders following haematopoietic cell transplantation. Br. J. Haematol. 2020, 190, 508–519.
- Pagliuca, S.; Michonneau, D.; Sicre de Fontbrune, F.; Sutra del Galy, A.; Xhaard, A.; Robin, M.; Peffault de Latour, R.; Socie, G. Allogeneic reactivity–mediated endothelial cell complications after HSCT: A plea for consensual definitions. Blood Adv. 2019, 3, 2424–2435.
- 25. Dalle, J.-H.; Giralt, S.A. Hepatic Veno-Occlusive Disease after Hematopoietic Stem Cell Transplantation: Risk Factors and Stratification, Prophylaxis, and Treatment. Biol. Blood Marrow Transplant. 2016, 22, 400–409.
- DeLeve, L.D.; McCuskey, R.S.; Wang, X.; Hu, L.; McCuskey, M.K.; Epstein, R.B.; Kanel, G.C. Characterization of a reproducible rat model of hepatic veno-occlusive disease. Hepatology 1999, 29, 1779–1791.
- DeLeve, L.D.; Wang, X.; Kanel, G.C.; Ito, Y.; Bethea, N.W.; McCuskey, M.K.; Tokes, Z.A.; Tsai, J.; McCuskey, R.S. Decreased hepatic nitric oxide production contributes to the development of rat sinusoidal obstruction syndrome. Hepatology 2003, 38, 900–908.

- 28. Huang, Z.; Chen, M.; Wei, M.; Lu, B.; Wu, X.; Wang, Z.; Ji, L. Liver Inflammatory Injury Initiated by DAMPs-TLR4-MyD88/TRIF-NFκB Signaling Pathway Is Involved in Monocrotaline-Induced HSOS. Toxicol. Sci. 2019, 172, 385–397.
- Akil, A.; Zhang, Q.; Mumaw, C.L.; Raiker, N.; Yu, J.; Velez de Mendizabal, N.; Haneline, L.S.; Robertson, K.A.; Skiles, J.; Diaz-Ricart, M.; et al. Biomarkers for Diagnosis and Prognosis of Sinusoidal Obstruction Syndrome after Hematopoietic Cell Transplantation. Biol. Blood Marrow Transplant. 2015, 21, 1739–1745.
- Rondón, G.; Saliba, R.M.; Chen, J.; Ledesma, C.; Alousi, A.M.; Oran, B.; Hosing, C.M.; Kebriaei, P.; Khouri, I.F.; Shpall, E.J.; et al. Impact of Fluid Overload as New Toxicity Category on Hematopoietic Stem Cell Transplantation Outcomes. Biol. Blood Marrow Transplant. 2017, 23, 2166–2171.
- Choi, S.-J.; Lee, K.-H.; Lee, J.-H.; Lee, J.-H.; Kim, S.; Seol, M.; Lee, Y.-S.; Kim, W.-K.; Park, C.-J.; Chi, H.-S.; et al. Peri-engraftment clinical abnormalities following allogeneic hematopoietic cell transplantation: A retrospective review of 216 patients. Bone Marrow Transplant. 2003, 32, 809– 813.
- Varma, A.; Rondon, G.; Srour, S.A.; Chen, J.; Ledesma, C.; Champlin, R.E.; Ciurea, S.O.; Saliba, R.M. Endothelial Activation and Stress Index (EASIX) at Admission Predicts Fluid Overload in Recipients of Allogeneic Stem Cell Transplantation. Biol. Blood Marrow Transplant. 2020, 26, 1013–1020.
- Janin, A.; Deschaumes, C.; Daneshpouy, M.; Estaquier, J.; Micic-Polianski, J.; Rajagopalan-Levasseur, P.; Akarid, K.; Mounier, N.; Gluckman, E.; Socié, G.; et al. CD95 engagement induces disseminated endothelial cell apoptosis in vivo: Immunopathologic implications. Blood 2002, 99, 2940–2947.
- 34. Gerbitz, A.; Nickoloff, B.J.; Olkiewicz, K.; Willmarth, N.E.; Hildebrandt, G.; Liu, C.; Kobzik, L.; Eissner, G.; Holler, E.; Ferrara, J.L.M.; et al. A role for tumor necrosis factor-alpha-mediated endothelial apoptosis in the development of experimental idiopathic pneumonia syndrome. Transplantation 2004, 78, 494–502.
- 35. Hildebrandt, G.C.; Olkiewicz, K.M.; Corrion, L.A.; Chang, Y.; Clouthier, S.G.; Liu, C.; Cooke, K.R. Donor-derived TNF-α regulates pulmonary chemokine expression and the development of idiopathic pneumonia syndrome after allogeneic bone marrow transplantation. Blood 2004, 104, 586–593.
- Altmann, T.; Slack, J.; Slatter, M.A.; O'Brien, C.; Cant, A.; Thomas, M.; Brodlie, M.; Annavarapu, S.; Gennery, A.R. Endothelial cell damage in idiopathic pneumonia syndrome. Bone Marrow Transplant. 2018, 53, 515–518.
- 37. Seo, S.; Renaud, C.; Kuypers, J.M.; Chiu, C.Y.; Huang, M.-L.; Samayoa, E.; Xie, H.; Yu, G.; Fisher, C.E.; Gooley, T.A.; et al. Idiopathic pneumonia syndrome after hematopoietic cell

transplantation: Evidence of occult infectious etiologies. Blood 2015, 125, 3789-3797.

- Parikh, S.M.; Mammoto, T.; Schultz, A.; Yuan, H.-T.; Christiani, D.; Karumanchi, S.A.; Sukhatme, V.P. Excess circulating angiopoietin-2 may contribute to pulmonary vascular leak in sepsis in humans. PLoS Med. 2006, 3, e46.
- 39. Whitney, J.E.; Feng, R.; Koterba, N.; Chen, F.; Bush, J.; Graham, K.; Lacey, S.F.; Melenhorst, J.J.; Parikh, S.M.; Weiss, S.L.; et al. Endothelial Biomarkers Are Associated With Indirect Lung Injury in Sepsis-Associated Pediatric Acute Respiratory Distress Syndrome. Crit. Care Explor. 2020, 2, e0295.
- 40. Spitzer, T.R. Engraftment syndrome: Double-edged sword of hematopoietic cell transplants. Bone Marrow Transplant. 2015, 50, 469–475.
- Maiolino, A.; Biasoli, I.; Lima, J.; Portugal, A.C.; Pulcheri, W.; Nucci, M. Engraftment syndrome following autologous hematopoietic stem cell transplantation: Definition of diagnostic criteria. Bone Marrow Transplant. 2003, 31, 393–397.
- 42. Oyama, Y.; Cohen, B.; Traynor, A.; Brush, M.; Rodriguez, J.; Burt, R.K. Engraftment syndrome: A common cause for rash and fever following autologous hematopoietic stem cell transplantation for multiple sclerosis. Bone Marrow Transplant. 2002, 29, 81–85.
- Dispenzieri, A.; Lacy, M.Q.; Hayman, S.R.; Kumar, S.K.; Buadi, F.; Dingli, D.; Litzow, M.R.; Gastineau, D.A.; Inwards, D.J.; Elliott, M.A.; et al. Peripheral blood stem cell transplant for POEMS syndrome is associated with high rates of engraftment syndrome. Eur. J. Haematol. 2008, 80, 397–406.
- 44. Koreth, J.; Biernacki, M.; Aldridge, J.; Kim, H.T.; Alyea, E.P., 3rd; Armand, P.; Cutler, C.; Ho, V.T.; Wu, C.J.; Antin, J.H.; et al. Syngeneic donor hematopoietic stem cell transplantation is associated with high rates of engraftment syndrome. Biol. Blood Marrow Transplant. 2011, 17, 421–428.
- 45. Omer, A.K.; Kim, H.T.; Yalamarti, B.; McAfee, S.L.; Dey, B.R.; Ballen, K.K.; Attar, E.; Chen, Y.-B.; Spitzer, T.R. Engraftment syndrome after allogeneic hematopoietic cell transplantation in adults. Am. J. Hematol. 2014, 89, 698–705.
- 46. Chang, L.; Frame, D.; Braun, T.; Gatza, E.; Hanauer, D.A.; Zhao, S.; Magenau, J.M.; Schultz, K.; Tokala, H.; Ferrara, J.L.M.; et al. Engraftment syndrome after allogeneic hematopoietic cell transplantation predicts poor outcomes. Biol. Blood Marrow Transplant. 2014, 20, 1407–1417.
- Ruutu, T.; Barosi, G.; Benjamin, R.J.; Clark, R.E.; George, J.N.; Gratwohl, A.; Holler, E.; Iacobelli, M.; Kentouche, K.; Lämmle, B.; et al. Diagnostic criteria for hematopoietic stem cell transplantassociated microangiopathy: Results of a consensus process by an International Working Group. Haematologica 2007, 92, 95–100.
- 48. Ho, V.T.; Cutler, C.; Carter, S.; Martin, P.; Adams, R.; Horowitz, M.; Ferrara, J.; Soiffer, R.; Giralt, S. Blood and marrow transplant clinical trials network toxicity committee consensus summary:

Thrombotic microangiopathy after hematopoietic stem cell transplantation. Biol. Blood Marrow Transplant. 2005, 11, 571–575.

- 49. Jodele, S.; Laskin, B.L.; Dandoy, C.E.; Myers, K.C.; El-Bietar, J.; Davies, S.M.; Goebel, J.; Dixon, B.P. A new paradigm: Diagnosis and management of HSCT-associated thrombotic microangiopathy as multi-system endothelial injury. Blood Rev. 2015, 29, 191–204.
- 50. Tichelli, A.; Gratwohl, A. Vascular endothelium as "novel" target of graft-versus-host disease. Best Pract. Res. Clin. Haematol. 2008, 21, 139–148.
- 51. Kraft, S.; Bollinger, N.; Bodenmann, B.; Heim, D.; Bucher, C.; Lengerke, C.; Kleber, M.; Tsakiris, D.A.; Passweg, J.; Tzankov, A.; et al. High mortality in hematopoietic stem cell transplant-associated thrombotic microangiopathy with and without concomitant acute graft-versus-host disease. Bone Marrow Transplant. 2019, 54, 540–548.
- Li, A.; Wu, Q.; Davis, C.; Kirtane, K.S.; Pham, P.D.; Sorror, M.L.; Lee, S.J.; Gopal, A.K.; Dong, J.-F.; Garcia, D.A.; et al. Transplant-Associated Thrombotic Microangiopathy Is a Multifactorial Disease Unresponsive to Immunosuppressant Withdrawal. Biol. Blood Marrow Transplant. 2019, 25, 570–576.
- 53. Nishida, T.; Hamaguchi, M.; Hirabayashi, N.; Haneda, M.; Terakura, S.; Atsuta, Y.; Imagama, S.; Kanie, T.; Murata, M.; Taji, H.; et al. Intestinal thrombotic microangiopathy after allogeneic bone marrow transplantation: A clinical imitator of acute enteric graft-versus-host disease. Bone Marrow Transplant. 2004, 33, 1143–1150.
- El-Bietar, J.; Warren, M.; Dandoy, C.; Myers, K.C.; Lane, A.; Wallace, G.; Davies, S.M.; Jodele, S. Histologic Features of Intestinal Thrombotic Microangiopathy in Pediatric and Young Adult Patients after Hematopoietic Stem Cell Transplantation. Biol. Blood Marrow Transplant. 2015, 21, 1994–2001.
- 55. Warren, M.; Jodele, S.; Dandoy, C.; Myers, K.C.; Wallace, G.; Nelson, A.; El-Bietar, J. A Complete Histologic Approach to Gastrointestinal Biopsy From Hematopoietic Stem Cell Transplant Patients With Evidence of Transplant-Associated Gastrointestinal Thrombotic Microangiopathy. Arch. Pathol. Lab. Med. 2017, 141, 1558–1566.
- 56. Laskin, B.L.; Maisel, J.; Goebel, J.; Yin, H.J.; Luo, G.; Khoury, J.C.; Davies, S.M.; Jodele, S. Renal arteriolar C4d deposition: A novel characteristic of hematopoietic stem cell transplantationassociated thrombotic microangiopathy. Transplantation 2013, 96, 217–223.
- 57. Jodele, S.; Davies, S.M.; Lane, A.; Khoury, J.; Dandoy, C.; Goebel, J.; Myers, K.; Grimley, M.; Bleesing, J.; El-Bietar, J.; et al. Diagnostic and risk criteria for HSCT-associated thrombotic microangiopathy: A study in children and young adults. Blood 2014, 124, 645–653.
- 58. Qi, J.; Wang, J.; Chen, J.; Su, J.; Tang, Y.; Wu, X.; Ma, X.; Chen, F.; Ruan, C.; Zheng, X.L.; et al. Plasma levels of complement activation fragments C3b and sC5b-9 significantly increased in

patients with thrombotic microangiopathy after allogeneic stem cell transplantation. Ann. Hematol. 2017, 96, 1849–1855.

- 59. Jodele, S.; Licht, C.; Goebel, J.; Dixon, B.P.; Zhang, K.; Sivakumaran, T.A.; Davies, S.M.; Pluthero, F.G.; Lu, L.; Laskin, B.L. Abnormalities in the alternative pathway of complement in children with hematopoietic stem cell transplant-associated thrombotic microangiopathy. Blood 2013, 122, 2003–2007.
- Gavriilaki, E.; Sakellari, I.; Chatzikonstantinou, T.; Mallouri, D.; Batsis, I.; Vardi, A.; Bousiou, Z.; Koravou, E.-E.; Masmanidou, M.; Touloumenidou, T.; et al. Endothelial and Complement Activation As Predictors of Survival in Adult Allogeneic Hematopoietic Cell Transplantation. HemaSphere 2021, 5, e487.
- 61. Noris, M.; Mescia, F.; Remuzzi, G. STEC-HUS, atypical HUS and TTP are all diseases of complement activation. Nat. Rev. Nephrol. 2012, 8, 622–633.
- 62. Jodele, S.; Dandoy, C.E.; Myers, K.C.; El-Bietar, J.; Nelson, A.; Wallace, G.; Laskin, B.L. New approaches in the diagnosis, pathophysiology, and treatment of pediatric hematopoietic stem cell transplantation-associated thrombotic microangiopathy. Transfus. Apher. Sci. 2016, 54, 181–190.
- 63. Thachil, J. Nitric oxide in transplantation-related thrombotic microangiopathy. Bone Marrow Transplant. 2009, 43, 513–514.
- 64. Trapp, A.; Weis, M. The impact of immunosuppression on endothelial function. J. Cardiovasc. Pharmacol. 2005, 45, 81–87.
- Keller, T.T.; Mairuhu, A.T.A.; de Kruif, M.D.; Klein, S.K.; Gerdes, V.E.A.; ten Cate, H.; Brandjes, D.P.M.; Levi, M.; van Gorp, E.C.M. Infections and endothelial cells. Cardiovasc. Res. 2003, 60, 40–48.
- 66. Mezö, B.; Horváth, O.; Sinkovits, G.; Veszeli, N.; Kriván, G.; Prohászka, Z. Validation of Early Increase in Complement Activation Marker sC5b-9 as a Predictive Biomarker for the Development of Thrombotic Microangiopathy After Stem Cell Transplantation. Front. Med. 2020, 7, 646.
- Okamura, H.; Nakamae, H.; Shindo, T.; Ohtani, K.; Hidaka, Y.; Ohtsuka, Y.; Makuuchi, Y.; Kuno, M.; Takakuwa, T.; Harada, N.; et al. Early Elevation of Complement Factor Ba Is a Predictive Biomarker for Transplant-Associated Thrombotic Microangiopathy. Front. Immunol. 2021, 12, 2669.
- Horváth, O.; Kállay, K.; Csuka, D.; Mező, B.; Sinkovits, G.; Kassa, C.; Stréhn, A.; Csordás, K.; Sinkó, J.; Prohászka, Z.; et al. Early Increase in Complement Terminal Pathway Activation Marker sC5b-9 Is Predictive for the Development of Thrombotic Microangiopathy after Stem Cell Transplantation. Biol. Blood Marrow Transplant. 2018, 24, 989–996.
- 69. Mir, E.; Palomo, M.; Rovira, M.; Pereira, A.; Escolar, G.; Penack, O.; Holler, E.; Carreras, E.; Diaz-Ricart, M. Endothelial damage is aggravated in acute GvHD and could predict its development.

Bone Marrow Transplant. 2017, 52, 1317-1325.

- Luft, T.; Dietrich, S.; Falk, C.; Conzelmann, M.; Hess, M.; Benner, A.; Neumann, F.; Isermann, B.; Hegenbart, U.; Ho, A.D.; et al. Steroid-refractory GVHD: T-cell attack within a vulnerable endothelial system. Blood 2011, 118, 1685–1692.
- Dietrich, S.; Falk, C.S.; Benner, A.; Karamustafa, S.; Hahn, E.; Andrulis, M.; Hegenbart, U.; Ho, A.D.; Dreger, P.; Luft, T. Endothelial Vulnerability and Endothelial Damage Are Associated with Risk of Graft-versus-Host Disease and Response to Steroid Treatment. Biol. Blood Marrow Transplant. 2013, 19, 22–27.
- Kernan, N.A.; Grupp, S.; Smith, A.R.; Arai, S.; Triplett, B.; Antin, J.H.; Lehmann, L.; Shore, T.; Ho, V.T.; Bunin, N.; et al. Final results from a defibrotide treatment-IND study for patients with hepatic veno-occlusive disease/sinusoidal obstruction syndrome. Br. J. Haematol. 2018, 181, 816–827.
- Richardson, P.; Aggarwal, S.; Topaloglu, O.; Villa, K.F.; Corbacioglu, S. Systematic review of defibrotide studies in the treatment of veno-occlusive disease/sinusoidal obstruction syndrome (VOD/SOS). Bone Marrow Transplant. 2019, 54, 1951–1962.
- 74. Pescador, R.; Capuzzi, L.; Mantovani, M.; Fulgenzi, A.; Ferrero, M.E. Defibrotide: Properties and clinical use of an old/new drug. Vascul. Pharmacol. 2013, 59, 1–10.
- 75. Devadas, S.K.; Toshniwal, M.; Bagal, B.; Khattry, N. Successful Treatment of Transplant Associated Thrombotic Microangiopathy (TA-TMA) with Low Dose Defibrotide. Indian J. Hematol. Blood Transfus. 2018, 34, 469–473.
- 76. Richardson, P.G.; Soiffer, R.J.; Antin, J.H.; Uno, H.; Jin, Z.; Kurtzberg, J.; Martin, P.L.; Steinbach, G.; Murray, K.F.; Vogelsang, G.B.; et al. Defibrotide for the Treatment of Severe Hepatic Veno-Occlusive Disease and Multiorgan Failure after Stem Cell Transplantation: A Multicenter, Randomized, Dose-Finding Trial. Biol. Blood Marrow Transplant. 2010, 16, 1005–1017.
- García-Bernal, D.; Palomo, M.; Martínez, C.M.; Millán-Rivero, J.E.; García-Guillén, A.I.; Blanquer, M.; Díaz-Ricart, M.; Sackstein, R.; Carreras, E.; Moraleda, J.M. Defibrotide inhibits donor leucocyte-endothelial interactions and protects against acute graft-versus-host disease. J. Cell. Mol. Med. 2020, 24, 8031–8044.
- 78. Rosenthal, J. Hematopoietic cell transplantation-associated thrombotic microangiopathy: A review of pathophysiology, diagnosis, and treatment. J. Blood Med. 2016, 7, 181–186.
- Jodele, S.; Fukuda, T.; Vinks, A.; Mizuno, K.; Laskin, B.L.; Goebel, J.; Dixon, B.P.; Teusink, A.; Pluthero, F.G.; Lu, L.; et al. Eculizumab therapy in children with severe hematopoietic stem cell transplantation-associated thrombotic microangiopathy. Biol. Blood Marrow Transplant. 2014, 20, 518–525.
- 80. De Fontbrune, F.S.; Galambrun, C.; Sirvent, A.; Huynh, A.; Faguer, S.; Nguyen, S.; Bay, J.-O.; Neven, B.; Moussi, J.; Simon, L.; et al. Use of Eculizumab in Patients With Allogeneic Stem Cell

Transplant-Associated Thrombotic Microangiopathy: A Study From the SFGM-TC. Transplantation 2015, 99, 1953–1959.

- Jan, A.S.; Hosing, C.; Aung, F.; Yeh, J. Approaching treatment of transplant-associated thrombotic Microangiopathy from two directions with Eculizumab and transitioning from Tacrolimus to Sirolimus. Transfusion 2019, 59, 3519–3524.
- Zhang, R.; Zhou, M.; Qi, J.; Miao, W.; Zhang, Z.; Wu, D.; Han, Y. Efficacy and Safety of Eculizumab in the Treatment of Transplant-Associated Thrombotic Microangiopathy: A Systematic Review and Meta-Analysis. Front. Immunol. 2021, 11, 3486.
- Rambaldi, A.; Gritti, G.; Micò, M.C.; Frigeni, M.; Borleri, G.; Salvi, A.; Landi, F.; Pavoni, C.; Sonzogni, A.; Gianatti, A.; et al. Endothelial injury and thrombotic microangiopathy in COVID-19: Treatment with the lectin-pathway inhibitor narsoplimab. Immunobiology 2020, 225, 152001.
- 84. Poppelaars, F.; Faria, B.; Schwaeble, W.; Daha, M.R. The contribution of complement to the pathogenesis of IgA nephropathy: Are complement-targeted therapies moving from rare disorders to more common diseases? J. Clin. Med. 2021, 10, 4715.
- 85. Lafayette, R.A.; Rovin, B.H.; Reich, H.N.; Tumlin, J.A.; Floege, J.; Barratt, J. Safety, Tolerability and Efficacy of Narsoplimab, a Novel MASP-2 Inhibitor for the Treatment of IgA Nephropathy. Kidney Int. Rep. 2020, 5, 2032–2041.
- 86. Chen, J.; Jin, J.-X.; Xu, X.-F.; Zhang, X.-X.; Ye, X.-N.; Huang, J. Successful treatment of plasma exchange-refractory thrombotic thrombocytopenic purpura with rituximab: A case report. World J. Clin. Cases 2020, 8, 2617–2622.
- Niewold, T.B.; Alpert, D.; Scanzello, C.R.; Paget, S.A. Rituximab treatment of thrombotic thrombocytopenic purpura in the setting of connective tissue disease. J. Rheumatol. 2006, 33, 1194–1196.
- Marr, H.; McDonald, E.-J.; Merriman, E.; Smith, M.; Mangos, H.; Stoddart, C.; Ganly, P. Successful treatment of transplant-associated microangiopathy with rituximab. N. Z. Med. J. 2009, 122, 72–74.
- Ostronoff, M.; Ostronoff, F.; Calixto, R.; Florêncio, R.; Florêncio, M.; Domingues, M.C.; Souto Maior, A.P.; Sucupira, A.; Tagliari, C. Life-threatening hemolytic-uremic syndrome treated with rituximab in an allogeneic bone marrow transplant recipient. Bone Marrow Transplant. 2007, 39, 649–651.
- Gallerani, E.; Lerch, E.; Romagnani, E.; Stathis, A.; Giardelli, G.; Zwhalen, H.; Marone, C.; Cavalli, F. Thrombotic thrombocytopenic purpura associated with renal failure after autologous transplantation for multiple myeloma successfully treated with rituximab. Eur. J. Haematol. 2006, 77, 527–529.

- 91. Rosenthal, R.A.; Chukwuogo, N.A.; Ocasio, V.H.; Kahng, K.U. Cyclosporine inhibits endothelial cell prostacyclin production. J. Surg. Res. 1989, 46, 593–596.
- Voss, B.L.; Hamilton, K.K.; Samara, E.N.; McKee, P.A. Cyclosporine suppression of endothelial prostacyclin generation. A possible mechanism for nephrotoxicity. Transplantation 1988, 45, 793– 796.
- 93. Conde, M.; Andrade, J.; Bedoya, F.J.; Santa Maria, C.; Sobrino, F. Inhibitory effect of cyclosporin A and FK506 on nitric oxide production by cultured macrophages. Evidence of a direct effect on nitric oxide synthase activity. Immunology 1995, 84, 476–481.
- 94. Hernández, G.L.; Volpert, O.V.; Iñiguez, M.A.; Lorenzo, E.; Martínez-Martínez, S.; Grau, R.; Fresno, M.; Redondo, J.M. Selective inhibition of vascular endothelial growth factor-mediated angiogenesis by cyclosporin A: Roles of the nuclear factor of activated T cells and cyclooxygenase 2. J. Exp. Med. 2001, 193, 607–620.
- 95. Wolff, D.; Wilhelm, S.; Hahn, J.; Gentilini, C.; Hilgendorf, I.; Steiner, B.; Kahl, C.; Junghanss, C.; Hartung, G.; Casper, J.; et al. Replacement of calcineurin inhibitors with daclizumab in patients with transplantation-associated microangiopathy or renal insufficiency associated with graftversus-host disease. Bone Marrow Transplant. 2006, 38, 445–451.
- Laskin, B.L.; Goebel, J.; Davies, S.M.; Jodele, S. Small vessels, big trouble in the kidneys and beyond: Hematopoietic stem cell transplantation-associated thrombotic microangiopathy. Blood 2011, 118, 1452–1462.
- 97. Jodele, S.; Laskin, B.L.; Goebel, J.; Khoury, J.C.; Pinkard, S.L.; Carey, P.M.; Davies, S.M. Does early initiation of therapeutic plasma exchange improve outcome in pediatric stem cell transplant-associated thrombotic microangiopathy? Transfusion 2013, 53, 661–667.
- 98. Daly, A.S.; Xenocostas, A.; Lipton, J.H. Transplantation-associated thrombotic microangiopathy: Twenty-two years later. Bone Marrow Transplant. 2002, 30, 709–715.
- Sartain, S.; Shubert, S.; Wu, M.-F.; Srivaths, P.; Teruya, J.; Krance, R.; Martinez, C. Therapeutic Plasma Exchange does not Improve Renal Function in Hematopoietic Stem Cell Transplantation-Associated Thrombotic Microangiopathy: An Institutional Experience. Biol. Blood Marrow Transplant. 2019, 25, 157–162.
- 100. Rizvi, M.A.; Vesely, S.K.; George, J.N.; Chandler, L.; Duvall, D.; Smith, J.W.; Gilcher, R.O. Complications of plasma exchange in 71 consecutive patients treated for clinically suspected thrombotic thrombocytopenic purpura-hemolytic-uremic syndrome. Transfusion 2000, 40, 896– 901.
- 101. Nguyen, L.; Terrell, D.R.; Duvall, D.; Vesely, S.K.; George, J.N. Complications of plasma exchange in patients treated for thrombotic thrombocytopenic purpura. IV. An additional study of 43 consecutive patients, 2005 to 2008. Transfusion 2009, 49, 392–394.

- 102. Howard, M.A.; Williams, L.A.; Terrell, D.R.; Duvall, D.; Vesely, S.K.; George, J.N. Complications of plasma exchange in patients treated for clinically suspected thrombotic thrombocytopenic purpura-hemolytic uremic syndrome. Transfusion 2006, 46, 154–156.
- 103. Saito, H.; Maruyama, I.; Shimazaki, S.; Yamamoto, Y.; Aikawa, N.; Ohno, R.; Hirayama, A.; Matsuda, T.; Asakura, H.; Nakashima, M.; et al. Efficacy and safety of recombinant human soluble thrombomodulin (ART-123) in disseminated intravascular coagulation: Results of a phase III, randomized, double-blind clinical trial. J. Thromb. Haemost. 2007, 5, 31–41.
- 104. Shirahata, A.; Mimuro, J.; Takahashi, H.; Tsuji, H.; Kitajima, I.; Matsushita, T.; Eguchi, Y.; Kitamura, N.; Honda, G.; Sakata, Y. Postmarketing Surveillance of Recombinant Human Soluble Thrombomodulin (Thrombomodulin α) in Pediatric Patients With Disseminated Intravascular Coagulation. Clin. Appl. Thromb./Hemost. 2014, 20, 465–472.
- 105. Shirahata, A.; Mimuro, J.; Takahashi, H.; Kitajima, I.; Tsuji, H.; Eguchi, Y.; Matsushita, T.; Kajiki, M.; Honda, G.; Sakata, Y. Recombinant soluble human thrombomodulin (thrombomodulin alfa) in the treatment of neonatal disseminated intravascular coagulation. Eur. J. Pediatr. 2014, 173, 303–311.
- 106. Hirata, N.; Ngo, D.T.; Phan, P.H.; Ainai, A.; Phung, T.T.B.; Ta, T.A.; Takasaki, J.; Kawachi, S.; Nunoi, H.; Nakajima, N.; et al. Recombinant human thrombomodulin for pneumonia-induced severe ARDS complicated by DIC in children: A preliminary study. J. Anesth. 2021, 35, 638–645.
- 107. Nomura, S.; Konishi, A.; Tsubokura, Y.; Azuma, Y.; Hotta, M.; Yoshimura, H.; Nakanishi, T.; Fujita, S.; Satake, A.; Katayama, Y.; et al. Effects of recombinant thrombomodulin on long-term prognosis after allogeneic hematopoietic stem cell transplantation. Transpl. Immunol. 2019, 57, 101247.
- 108. Yamamoto, S.; Toyama, D.; Sugishita, Y.; Kaneko, R.; Okamoto, N.; Koganesawa, M.; Fujita, S.; Akiyama, K.; Matsuno, R.; Isoyama, K. Prophylactic recombinant thrombomodulin treatment prevents hepatic sinusoidal obstruction syndrome in high-risk pediatric patients that undergo hematopoietic stem cell transplants. Pediatr. Transplant. 2018, 22, e13269.
- 109. Sakai, M.; Ikezoe, T.; Bandobashi, K.; Togitani, K.; Yokoyama, A. Successful treatment of transplantation-associated thrombotic microangiopathy with recombinant human soluble thrombomodulin. Bone Marrow Transplant. 2010, 45, 803–805.
- 110. Ito, D.; Akamatsu, N.; Ichida, A.; Kaneko, J.; Arita, J.; Hasegawa, K.; Sakamoto, Y.; Kokudo, N. Possible efficacy of recombinant human soluble thrombomodulin for the treatment of thrombotic microangiopathy after liver transplantation. Liver Transplant. 2016, 22, 689–692.
- 111. Fujiwara, H.; Maeda, Y.; Sando, Y.; Nakamura, M.; Tani, K.; Ishikawa, T.; Nishimori, H.; Matsuoka, K.-I.; Fujii, N.; Kondo, E.; et al. Treatment of thrombotic microangiopathy after hematopoietic stem cell transplantation with recombinant human soluble thrombomodulin. Transfusion 2016, 56, 886–892.

- 112. Otsuka, Y.; Kondo, T.; Nomura, R.; Yamashita, K.; Takaori-Kondo, A. Successful treatment with recombinant thrombomodulin in transplant-associated thrombotic microangiopathy following HLA-haploidentical transplantation. Rinsho. Ketsueki. 2019, 60, 1560–1566.
- 113. Ikezoe, T.; Takeuchi, A.; Taniguchi, A.; Togitani, K.; Yokoyama, A. Recombinant human soluble thrombomodulin counteracts capillary leakage associated with engraftment syndrome. Bone Marrow Transplant. 2011, 46, 616–618.
- 114. Ikezoe, T.; Togitani, K.; Komatsu, N.; Isaka, M.; Yokoyama, A. Successful treatment of sinusoidal obstructive syndrome after hematopoietic stem cell transplantation with recombinant human soluble thrombomodulin. Bone Marrow Transplant. 2010, 45, 783–785.
- 115. Inagaki, J.; Kurauchi, K.; Fukano, R.; Noguchi, M.; Okamura, J. Heterogeneous response to recombinant thrombomodulin by grade of sinusoidal obstructive syndrome after pediatric stem cell transplantation. Bone Marrow Transplant. 2016, 51, 1543–1545.
- 116. Hermann, F.; Forster, A.; Chenevard, R.; Enseleit, F.; Hürlimann, D.; Corti, R.; Spieker, L.E.; Frey, D.; Hermann, M.; Riesen, W.; et al. Simvastatin improves endothelial function in patients with rheumatoid arthritis. J. Am. Coll. Cardiol. 2005, 45, 461–464.
- 117. Reriani, M.K.; Dunlay, S.M.; Gupta, B.; West, C.P.; Rihal, C.S.; Lerman, L.O.; Lerman, A. Effects of statins on coronary and peripheral endothelial function in humans: A systematic review and meta-analysis of randomized controlled trials. Eur. J. Cardiovasc. Prev. Rehabil. 2011, 18, 704– 716.
- 118. Ii, M.; Losordo, D.W. Statins and the endothelium. Vascul. Pharmacol. 2007, 46, 1–9.
- 119. Zheng, P.; Wu, Q.-L.; Li, B.-B.; Chen, P.; Nie, D.-M.; Zhang, R.; Fang, J.; Xia, L.-H.; Hong, M. Simvastatin ameliorates graft-vs-host disease by regulating angiopoietin-1 and angiopoietin-2 in a murine model. Leuk. Res. 2017, 55, 49–54.
- 120. Jiang, S.; Penack, O.; Terzer, T.; Schult, D.; Majer-Lauterbach, J.; Radujkovic, A.; Blau, I.W.; Bullinger, L.; Müller-Tidow, C.; Dreger, P.; et al. Predicting sinusoidal obstruction syndrome after allogeneic stem cell transplantation with the EASIX biomarker panel. Haematologica 2021, 106, 446–453.
- 121. Gutbier, B.; Jiang, X.; Dietert, K.; Ehrler, C.; Lienau, J.; Van Slyke, P.; Kim, H.; Hoang, V.C.; Maynes, J.T.; Dumont, D.J.; et al. Vasculotide reduces pulmonary hyperpermeability in experimental pneumococcal pneumonia. Crit. Care 2017, 21, 274.
- 122. Latreille, E.; Lee, W.L. Interactions of Influenza and SARS-CoV-2 with the Lung Endothelium: Similarities, Differences, and Implications for Therapy. Viruses 2021, 13, 161.
- 123. Venkat, P.; Yan, T.; Chopp, M.; Zacharek, A.; Ning, R.; Van Slyke, P.; Dumont, D.; Landschoot-Ward, J.; Liang, L.; Chen, J. Angiopoietin-1 Mimetic Peptide Promotes Neuroprotection after Stroke in Type 1 Diabetic Rats. Cell Transplant. 2018, 27, 1744–1752.

- 124. Venkat, P.; Ning, R.; Zacharek, A.; Culmone, L.; Liang, L.; Landschoot-Ward, J.; Chopp, M. Treatment with an Angiopoietin-1 mimetic peptide promotes neurological recovery after stroke in diabetic rats. CNS Neurosci. Ther. 2021, 27, 48–59.
- 125. Trieu, M.; van Meurs, M.; van Leeuwen, A.L.I.; Van Slyke, P.; Hoang, V.; Geeraedts, L.M.G.J.; Boer, C.; van den Brom, C.E. Vasculotide, an Angiopoietin-1 Mimetic, Restores Microcirculatory Perfusion and Microvascular Leakage and Decreases Fluid Resuscitation Requirements in Hemorrhagic Shock. Anesthesiology 2018, 128, 361–374.
- 126. Sanwal, R.; Joshi, K.; Ditmans, M.; Tsai, S.S.H.; Lee, W.L. Ultrasound and Microbubbles for Targeted Drug Delivery to the Lung Endothelium in ARDS: Cellular Mechanisms and Therapeutic Opportunities. Biomedicines 2021, 9, 803.
- 127. Subramaniyam, D.; Virtala, R.; Pawłowski, K.; Clausen, I.G.; Warkentin, S.; Stevens, T.; Janciauskiene, S. TNF-alpha-induced self expression in human lung endothelial cells is inhibited by native and oxidized alpha1-antitrypsin. Int. J. Biochem. Cell Biol. 2008, 40, 258–271.
- 128. Immenschuh, S.; Vijayan, V.; Janciauskiene, S.; Gueler, F. Heme as a Target for Therapeutic Interventions. Front. Pharmacol. 2017, 8, 146.
- 129. Giannoni, L.; Morin, F.; Robin, M.; Peyneau, M.; Schlageter, M.H.; Desmier, D.; Pagliuca, S.; Sutra Del Galy, A.; Sicre de Fontbrune, F.; Xhaard, A.; et al. Human-Derived α1-Antitrypsin is Still Efficacious in Heavily Pretreated Patients with Steroid-Resistant Gastrointestinal Graft-versus-Host Disease. Biol. Blood Marrow Transplant. 2020, 26, 1620–1626.

Retrieved from https://encyclopedia.pub/entry/history/show/46735