Drug Resistance in Nontuberculous Mycobacteria

Subjects: Microbiology Contributor: Herman Spaink

The genus *Mycobacterium* comprises a multitude of species known to cause serious disease in humans, including *Mycobacterium tuberculosis* and *M. leprae*, the responsible agents for tuberculosis and leprosy, respectively. In addition, there is a worldwide spike in the number of infections caused by a mixed group of species such as the *M. avium*, *M. abscessus* and *M. ulcerans* complexes, collectively called nontuberculous mycobacteria (NTMs). The situation is forecasted to worsen because, like tuberculosis, NTMs either naturally possess or are developing high resistance against conventional antibiotics.

Keywords: nontuberculous mycobacteria ; drug resistance mechanisms ; antimicrobial testing ; drug discovery

1. The Rise of Nontuberculous Mycobacteria

Mycobacteria are a large group of non-motile, rod-shaped bacteria that tend to grow mold-like pellicles on liquid culture media. Out of the 150 species known to this genus, nearly 25 are known to cause disease in humans. The most well-known mycobacteria species are the *M. tuberculosis* and *M. leprae* complexes, with an estimated prevalence rate of 130 (the year 2020) and 2 (the year 2018) cases per 100,000 population, respectively ^{[1][2]}, while all others are collectively called nontuberculous mycobacteria (NTMs) ^[3]. Despite that NTMs are less widespread pathogens for humans than *M. tuberculosis*, they have proven to be an emerging threat to the immunocompromised population ^[4], with an estimated 4.1–14.1 cases per 100,000 population worldwide (2013) ^[5]. NTMs are ubiquitous and can survive in a wide range of environmental conditions, and their infections are difficult to diagnose ^[6]. The most common NTM-related pathologies are pulmonary infections (pulmonary nontuberculous mycobacterial disease) caused by strains from the *M. avium* complex and *M. abscessus* ^{[6][Z]}, but NTMs can also cause skin and soft tissue infections (e.g., *M. marinum* infection and Buruli ulcer caused by *M. ulcerans*), lymphadenitis in immunocompromised children, and even invasive disseminated disease eventually leading to death.

According to Runyon, NTMs can be classified based on the growth rate and pigment formation (Table 1) ^[B]. Types I, II, and III strains are classified as slow-growers because they take seven or more days of growth for forming visible colonies on a subculture plate ^[9]. They are differentiated on their ability to produce pigments only on exposure to light (type I or photochromogens) or also in the dark (type II or scotochromogens), or not being strongly pigmented (type III or non-photochromogens) ^[10]. Type IV strains are regarded as rapid-growers as they take less than seven days to form visible colonies on a subculture plate ^[10]. Generally, slow-growing mycobacteria are much more prevalent than fast-growing ones ^[11] and present higher ratios of drug resistance (with the fast-growing *M. abscessus* being a notable exception) ^[12]. It has been suggested that all mycobacteria evolved from a common ancestral rapid growing mycobacterial strain ^{[13][14]} ^[15].

Table 1. Summary of the nontuberculous mycobacteria (NTMs) mentioned in this review, their classification according to Runyon, and their reported pathogenesis in humans.

Runyon Classification	NTM Species	Pathogenesis in Humans	
Photochromogens Runyon type I	<i>M. kansasii</i> ^{[<u>16]</u>,}	Pulmonary infections	
	M. simiae ^[17]	Skin infections	
		Disseminated infections	
	M. marinum ^{[<u>18]</u>}	Skin and soft tissue infections	
		Disseminated infections	
Scotochromogens Runyon type II	M. gordonae ^[19]	Pulmonary infections	
		Skin infections	
		Disseminated infections	
	M. scrofulaceum ^[20]	Cervical lymphadenitis among children	
		Pulmonary infections	
		Disseminated infections	
Non- photochromogens Runyon type III	<i>M. avium</i> complex (<i>M. avium</i> and <i>M.</i> intracellulare) ^[21]	Pulmonary MAC infections	
		Disseminated infections (mostly in AIDS patients)	
		MAC associated lymphadenitis (in young kids and people with normal immune systems)	
	M. malmoense ^[22]	Pulmonary infections	
		Disseminated infections	
	M. ulcerans ^[18]	Skin diseases (Buruli ulcers)	
Rapid growing Runyon type IV	M. abscessus ^[23]	Pulmonary infections	
		Skin and Soft tissue disease	
		Central nervous system infections	
		Disseminated infections	
	M. chelonae ^[24]	Skin and soft tissue infections	
		Pulmonary infections	
		Disseminated infections	
	M. smegmatis	Widely regarded as nonpathogenic	

Recently, there has been a considerable increase in the number of reported NTM related diseases, including respiratory infections caused by various strains from the *M. avium* complex, *M. kansasii* and *M. abscessus* ^[25]. This is partly because of the awareness of the symptoms caused by these infections and improvements in detection techniques, but also because of an increase in the number of susceptible individuals and that NTM can form biofilms in common household

and hospital sources of infection (such as showerheads, faucets, water distribution systems, plumbing systems, etc.) ^[26] ^[27]. The situation is worrying because, just like tuberculosis, these bacteria have developed high resistance against conventional antibiotics ^[28]. However, these pathogens are still considered opportunistic since they require a combination of constant exposure as well as host susceptibility to infection, and these infections have mainly remained limited to patients with pre-existing lung diseases ^{[25][29]}.

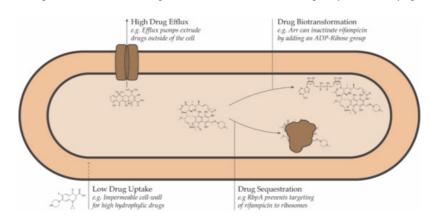
The major NTM that is infecting such individuals suffering from chronic diseases like cystic fibrosis is *M. abscessus*, which is a rapidly growing, intrinsically multidrug-resistant species ^[30]. These infections are often impossible to treat despite prolonged antibiotic therapy, and the therapy may even be contraindicated with lung transplantation, leaving no effective options for treatment ^[31]. While NTM infections were earlier thought to be independently acquired by susceptible individuals, the recent consensus is that such infections are frequently transmitted indirectly from an infected to a healthy individual, for instance, via contaminated hospital equipment ^[32]. Some opportunistic infectious NTM species tend to cluster in specific geographical distributions, and there may be a genetic basis for the susceptibility to their infection in particular patients ^{[11][33][34]}. Finally, relapse and reinfection is a major problem with some NTM infections, like the ones caused by *M. avium* complex ^[35], although it is less so for other species like *M. kansasii* ^[36].

Currently, the treatment for almost all NTM infections is based on macrolide-based antibiotics, such as clarithromycin or azithromycin. For NTM infections caused by the slow-growing group, the regime also includes ethambutol and rifampicin ^[37], while for fast-growers, it includes an aminoglycoside and either cefoxitin, imipenem or tigecycline ^[38]. These treatments are largely empirical, derived from years of clinical practice, can last for as long as 18 months, are costly, and are often associated with drug-related toxicities and side-effects ^[39]. Cure rates range from 80–90% with *M. malmoense* infections to just 30–50% with *M. abscessus* infections ^[40]. Thus, the discovery of new and more efficient therapies against NTMs is an important topic of research. However, a major bottleneck is the low susceptibility of mycobacteria to most antibiotics, including the ones used against tuberculosis ^[41]. A better understanding of the underlying mechanisms behind this drug resistance by improving the available models to study their infection could significantly help in accelerating the drug discovery process.

2. Mechanisms of Drug Resistance in Nontuberculous Mycobacteria

Drug Resistance can be either intrinsic (natural) or acquired $\frac{[42]}{2}$. Intrinsic resistance describes a situation where an organism possesses a set of special features that allows it to tolerate a particular drug or survive in an otherwise hostile chemical environment $\frac{[42]}{2}$. Mechanisms by which NTMs are intrinsically resistant to antibiotics include their thick, impermeable cell walls or their presence in biofilms and granulomas, which effectively decrease drug uptake, as well as the expression of proteins that specifically target clinically used antibacterial compounds.

On the other hand, acquired resistance refers to the case where a resistant strain emerges from a population that was previously drug-sensitive $^{[42]}$. These events are usually related to the prolonged antibiotic treatments required to cure NTM infections. The acquired resistance is particularly severe for NTMs that only have a single copy of genes encoding common target proteins such as ribosomes, thus increasing the risk of acquiring protective mutations with single-drug treatments $^{[4][43]}$. Here, we will focus on the mechanisms of mycobacterial physiology that make them naturally resistant to antimicrobial treatments since Nasiri et al. recently reviewed the mutations that may cause resistance to certain antibiotics in NTM $^{[44]}$.



Conceptually, resistance to antimicrobial drugs can be a result of one or more of the following mechanisms: decreased drug uptake, increased drug efflux, increased drug metabolism, or reduced drug sequestration (<u>Figure 1</u>) ^[45].



3. Models for Drug Discovery against NTM

There is a wide range of techniques that can be employed in the development of new potential antimicrobial therapies against NTMs. These techniques can be classified depending on the tools employ between in silico, in vitro, or in vivo. In general terms, in silico techniques are useful to generate new leads and narrow the search of potential candidates based on prior information at the start of a study or to optimize compounds based on specific targets via virtual simulations. These leads can then be tested for efficacy using standardized in vitro analysis, which allows the determination of their potential antimycobacterial activity. Finally, in vivo animal models can be used to recreate infection environments and are therefore interesting for preclinical evaluation of potential compounds. We summarized the main attributes for each category in <u>Table 2</u>. A recent review by Rampacci et al. explains in detail the different techniques, assays, and preclinical models against NTMs that have been developed, with an emphasis on the newer models ^[46]. We direct the reader to that review for an in-depth description of these methodologies and their read-outs. Here, we will give a brief outline of the most common techniques implemented in the lab and how they complement each other to create an integrated pipeline for drug discovery.

	In Silico	In Vitro	In Vivo
Methods employed	Structure–activity relations, Molecular simulations, Comparative genomics	Antimicrobial effect tests on cultured cells	Tests on live infected animals
Main insights	Molecular basis for drug action	Molecular and cellular effect of drug action	Whole-organism level of drug action
Advantages	High throughput, low-cost, no need for actual chemical synthesis of compounds or bacterial growth	Relatively simple systems and lower cost and time involvement, easy to handle, scalable	Closer to the actual physiological environment
Limitations	Requires prior information and complicated models to simulate molecular events such as docking and drug- target interactions	Needs a high level of standardization and careful experimentation for reproducibility, may not reproduce clinical situations	Requires careful model selection, large organism response is less predictable, ethical considerations, high economical costs
Best-fit stage in drug discovery	Primary (for narrowing the search of potential candidates) or secondary (for optimizing compounds to species-specific targets)	Secondary (for screening initial targets and efficacy determination)	Tertiary (for preclinical evaluation)

Table 2. Summary of the current methods available for the discovery of new antimicrobial therapies in NTMs.

4. Considerations for the Design of Therapies against NTMs

4.1. Optimization of Known Compounds Relevant for Combatting NTM

When designing new possible therapies against NTMs, it is important to consider the mechanisms that confer drug resistance that we reviewed above. For example, the exceptionally high hydrophobicity of mycobacterial cell walls has an important bearing on drug design: the more lipophilic molecules generally show higher permeability and hence are more active. This means that a possible route to developing anti-NTM antibiotics is to synthesize hydrophobic derivatives of existing antibiotics [47]. For instance, ciprofloxacin, when modified by the addition of hydrophobic alkyl substituents, showed higher activity against *M. avium* ^[48]. Similarly, for *M. leprae*, the efficacy of fluoroquinolones improved by incorporating the hydrophobic cyclopropyl groups ^[49].

Compounds that have been identified to be active against other diseases may directly be screened by in vitro bacterial assays, and MIC values may be determined to check the efficiency of the drug ^[50]. However, for *M. abscessus*, the hit rates among drug libraries that are active against neglected diseases like ascariasis, Buruli ulcer, Chagas disease, and malaria is just 1% ^[51], highlighting the great difficulty in finding new drugs for NTMs. A way forward could be to screen the compound libraries active against tuberculosis for their effect against NTM because of the structural similarity and homology of their drug targets ^[52]. In a recently conducted study, 129 compounds known to be active against *M. tuberculosis* were tested against *M. abscessus* and *M. avium*, and their rates were higher than for drugs that are not active against tuberculosis ^[53]. Rifabutin, an antibiotic used for the treatment of tuberculosis, has recently been found to be active against *M. abscessus* ^[50]. Notwithstanding these positive outcomes, most existing drugs specific to tuberculosis are usually ineffective against NTM ^{[53][54]}.

4.2. Synergies and Combination Therapies

In the development of a novel treatment, it should be considered that most of the successful anti-NTM drug therapies involve synergistic effects of two drugs: one antibiotic to disrupt the permeability of the outer membrane in order to ensure entry of the drug into the cell, and the another disrupting at least one vital cellular processes (such as DNA, RNA, or protein and outer membrane synthesis) for inhibiting cell growth ^[55]. For example, the performance of hydrophobic drugs that have intracellular targets can be improved by using them in conjunction with compounds that specifically target cell wall homeostasis. Such synergistic effects have been observed between ethambutol plus rifampicin in *M. avium* ^[56] or vancomycin plus clarithromycin in *M. abscessus* ^[57]. Similar effects can be seen using adjuvants that inhibit specific efflux pumps ^{[58][59]} or that increase the expression of the enzymes required for the biotransformation of a prodrug, thus boosting antibiotic effectivity ^[60].

Moreover, the synergistic effects cannot only improve drug efficacy but also reduce the chances that treatments lead to drug resistance ^{[61][62]}. For instance, the combination of β -lactam antibiotics and β -lactamase inhibitors has shown to be a promising strategy against *M. avium* infections ^[63]. Further, synergies can also be achieved by using a combination of two or more drugs that have the same cellular target. This approach has been validated for *M. abscessus* complex, wherein a dual β -lactam drug regimen proved to be much more effective than a single-drug regimen, with or without β -lactamase inhibitors ^{[64][65]}. In such regimens, each of the β -lactams preferentially targets a different enzyme that is involved in cell wall synthesis, thereby ensuring that "overlapping" effects are minimized, and combinedly, all biochemical pathways are exhaustively targeted ^[65]. Typically, mutations that cause the development of resistance mechanisms can have subsequent "spill-over" effects: the same mechanism may confer resistance to the entire class of drugs that target the same biochemical pathway (cross-resistance), or it can lead to increased vulnerability to other drugs that target a different pathway (collateral sensitivity) ^[66]. Exploiting collateral sensitivity by either combinatorial or cyclical treatment regimens involving multiple drugs can be an effective strategy, as recently demonstrated with drugs directed against *M. marinum* ^[67].

4.3. Host-Directed Therapies

A different approach towards finding more effective treatments against NTM infections is host-directed therapies (HDT), in which specific immune pathways of the host are modulated in such a way that it leads to a better clinical outcome. That is, HDTs aims to empower the host to clear the infection instead of directly targeting bacteria. The ways in which HDTs can help against NTMs are strengthening innate immunity against mycobacterial infections, preventing the growth of the bacilli by inhibiting the essential host-related growth-factors, restoring the immune response suppressed due to the infection, or reducing tissue damage due to hyperinflammation ^{[68][69]}.

HDTs offer several unique advantages over conventional therapies. First, chances of drug-related resistance are considerably reduced because it is difficult for the bacteria to develop completely new mechanisms of interacting with the host quickly and while being under the same hostile immune selection ^[70]. They also offer the possibility of making conventional drugs more effective against already resistant strains by neutralizing pathogen defenses ^[71]. The synergistic role of HDT adjuvants with anti-mycobacterials was demonstrated in a study in which picolinic acid (PA) was shown to potentiate fluoroquinolones against bacteria from the *M. avium* complex ^[72]. Fluoroquinolones are otherwise only very weakly effective against *M. avium* infections. This was attributed to two factors- upregulation of the immune system by PA and chelation of Fe ions by PA, which deprive the bacteria of the essential ions needed for growth ^[73]. In a previous study, PA was shown to inhibit *M. avium* growth inside mouse macrophages by inducing apoptosis- causing morphological changes ^[74].

In addition, some compounds show both host-directed and bacterial-directed actions. Clofazimine, a commonly used drug in *M. leprae* infections, is a good example of a drug that simultaneously affects the host as well as the bacteria. Upon infecting the body, *M. leprae* creates a safe microenvironment for itself inside the macrophages of the host by increasing the accumulation and retarding the breakdown of macrophage lipids ^[75]. It was shown that clofazimine not only helps reverse these two processes but also activates immune reactions in *M. leprae* infected host cells. Hence, effectively, it not just prevents the growth of the bacteria but also actively helps to eliminate it ^[76]. Another example of an antibiotic with a strong effect on the host inflammatory system is minocycline that modulates the endocannabinoid signaling pathway and, in this way, might have HDT potential ^[77].

Finally, considering the mechanisms of defense of the host system can lead to more effective ways of delivering drugs to the target, adding value to existing treatments. An example would be precision-targeting the drug by loading them into the host cells that act as carriers. This approach was demonstrated by loading dendritic cells to deliver amikacin inside alveolar granulomas and thus enhancing the killing of residing mycobacteria ^[78].

However, a major challenge in the development of effective HDTs is that different patients may not have the same immune status, which may depend on factors like the stage of the disease, health of the individual, pre-existing conditions, and genetic makeup $\frac{71}{2}$. This can be a hindrance towards a universal HDT and may require an approach for personalized medicine, much like cancer immunotherapy $\frac{79}{2}$.

References

- 1. Global Tuberculosis Report 2020. Available online: https://www.who.int/teams/global-tuberculosis-programme/tbreports (accessed on 23 January 2021).
- 2. Leprosy. Available online: https://www.who.int/news-room/fact-sheets/detail/leprosy (accessed on 23 January 2021).
- Yates, V.M. Mycobacterial Infections. In Rook's Textbook of Dermatology; Wiley-Blackwell: Oxford, UK, 2010; pp. 1–41. ISBN 9781405161695.
- Griffith, D.E.; Aksamit, T.; Brown-Elliott, B.A.; Catanzaro, A.; Daley, C.; Gordin, F.; Holland, S.M.; Horsburgh, R.; Huitt, G.; Iademarco, M.F.; et al. An official ATS/IDSA Statement: Diagnosis, treatment, and prevention of nontuberculous mycobacterial diseases. Am. J. Respir. Crit. Care Med. 2007, 175, 367–416.
- Kendall, B.; Winthrop, K. Update on the epidemiology of pulmonary nontuberculous mycobacterial infections. Semin. Respir. Crit. Care Med. 2013, 34, 087–094.
- Ratnatunga, C.N.; Lutzky, V.P.; Kupz, A.; Doolan, D.L.; Reid, D.W.; Field, M.; Bell, S.C.; Thomson, R.M.; Miles, J.J. The rise of non-tuberculosis mycobacterial lung disease. Front. Immunol. 2020, 11.
- 7. Gopalaswamy, R.; Shanmugam, S.; Mondal, R.; Subbian, S. Of tuberculosis and non-tuberculous mycobacterial infections–a comparative analysis of epidemiology, diagnosis and treatment. J. Biomed. Sci. 2020, 27, 74.
- 8. Runyon, E.H. Anonymous mycobacteria in pulmonary disease. Med. Clin. N. Am. 1959.
- Kim, C.-J.; Kim, N.-H.; Song, K.-H.; Choe, P.G.; Kim, E.S.; Park, S.W.; Kim, H.-B.; Kim, N.-J.; Kim, E.-C.; Park, W.B.; et al. Differentiating rapid- and slow-growing mycobacteria by difference in time to growth detection in liquid media. Diagn. Microbiol. Infect. Dis. 2013, 75, 73–76.
- 10. Porvaznik, I.; Solovič, I.; Mokrý, J. Non-tuberculous mycobacteria: Classification, diagnostics, and therapy. Adv. Exp. Med. Biol. 2017.
- 11. Johnson, M.M.; Odell, J.A. Nontuberculous mycobacterial pulmonary infections. J. Thorac. Dis. 2014, 6, 210.
- 12. Nessar, R.; Cambau, E.; Reyrat, J.M.; Murray, A.; Gicquel, B. Mycobacterium abscessus: A new antibiotic nightmare. J. Antimicrob. Chemother. 2012.
- 13. Devulder, G.; de Montclos, M.P.; Flandrois, J.P. A multigene approach to phylogenetic analysis using the genus Mycobacterium as a model. Int. J. Syst. Evol. Microbiol. 2005, 55, 293–302.
- 14. Gupta, R.S.; Lo, B.; Son, J. Phylogenomics and comparative genomic studies robustly support division of the genus mycobacterium into an emended genus mycobacterium and four novel genera. Front. Microbiol. 2018, 9.
- 15. Wee, W.Y.; Dutta, A.; Choo, S.W. Comparative genome analyses of mycobacteria give better insights into their evolution. PLoS ONE 2017, 12, e0172831.
- Jagielski, T.; Borówka, P.; Bakuła, Z.; Lach, J.; Marciniak, B.; Brzostek, A.; Dziadek, J.; Dziurzyński, M.; Pennings, L.; van Ingen, J.; et al. Genomic insights into the mycobacterium kansasii complex: An update. Front. Microbiol. 2020, 10, 2918.

- Hamieh, A.; Tayyar, R.; Tabaja, H.; Zein, S.E.L.; Bou Khalil, P.; Kara, N.; Kanafani, Z.A.; Kanj, N.; Bou Akl, I.; Araj, G.; et al. Emergence of Mycobacterium simiae: A retrospective study from a tertiary care center in Lebanon. PLoS ONE 2018, 13, e0195390.
- Franco-Paredes, C.; Marcos, L.A.; Henao-Martínez, A.F.; Rodríguez-Morales, A.J.; Villamil-Gómez, W.E.; Gotuzzo, E.; Bonifaz, A. Cutaneous Mycobacterial infections. Clin. Microbiol. Rev. 2018, 32.
- 19. Douglas, J.G.; Calder, M.A.; Choo-Kang, Y.F.J.; Leitch, A.G. Mycobacterium gordonae: A new pathogen? Thorax 1986, 41, 152–153.
- Suzuki, S.; Morino, E.; Ishii, M.; Namkoong, H.; Yagi, K.; Asakura, T.; Asami, T.; Fujiwara, H.; Uwamino, Y.; Nishimura, T.; et al. Clinical characteristics of pulmonary Mycobacterium scrofulaceum disease in 2001-2011: A case series and literature review. J. Infect. Chemother. 2016, 22, 611–616.
- Han, X.Y.; Tarrand, J.J.; Infante, R.; Jacobson, K.L.; Truong, M. Clinical significance and epidemiologic analyses of Mycobacterium avium and Mycobacterium intracellulare among patients without AIDS. J. Clin. Microbiol. 2005, 43, 4407–4412.
- Doig, C.; Muckersie, L.; Watt, B.; Forbes, K.J. Molecular epidemiology of Mycobacterium malmoense infections in Scotland. J. Clin. Microbiol. 2002, 40, 1103–1105.
- 23. To, K.; Cao, R.; Yegiazaryan, A.; Owens, J.; Venketaraman, V. General Overview of nontuberculous mycobacteria opportunistic pathogens: Mycobacterium avium and Mycobacterium abscessus. J. Clin. Med. 2020, 9.
- 24. Jones, R.S.; Shier, K.L.; Master, R.N.; Bao, J.R.; Clark, R.B. Current significance of the Mycobacterium chelonaeabscessus group. Diagn. Microbiol. Infect. Dis. 2019, 94, 248–254.
- Máiz Carro, L.; Barbero Herranz, E.; Nieto Royo, R. Respiratory infections due to nontuberculous mycobacterias. Med. Clin. 2018, 150, 191–197.
- 26. Nishiuchi, Y.; Iwamoto, T.; Maruyama, F. Infection sources of a common non-tuberculous mycobacterial pathogen, Mycobacterium avium complex. Front. Med. 2017, 4, 27.
- Shah, N.M.; Davidson, J.A.; Anderson, L.F.; Lalor, M.K.; Kim, J.; Thomas, H.L.; Lipman, M.; Abubakar, I. Pulmonary Mycobacterium avium-intracellulare is the main driver of the rise in non-tuberculous mycobacteria incidence in England, Wales and Northern Ireland, 2007–2012. BMC Infect. Dis. 2016.
- Johansen, M.D.; Herrmann, J.L.; Kremer, L. Non-tuberculous mycobacteria and the rise of Mycobacterium abscessus. Nat. Rev. Microbiol. 2020, 18, 392–407.
- Fleshner, M.; Olivier, K.N.; Shaw, P.A.; Adjemian, J.; Strollo, S.; Claypool, R.J.; Folio, L.; Zelazny, A.; Holland, S.M.; Prevots, D.R.; et al. Mortality among patients with pulmonary non-tuberculous mycobacteria disease. Int. J. Tuberc. Lung Dis. 2016.
- 30. Jarand, J.; Levin, A.; Zhang, L.; Huitt, G.; Mitchell, J.D.; Daley, C.L. Clinical and microbiologic outcomes in patients receiving treatment for Mycobacterium abscessus pulmonary disease. Clin. Infect. Dis. 2011.
- 31. Taylor, J.L.; Palmer, S.M. Mycobacterium abscessus chest wall and pulmonary infection in a cystic fibrosis lung transplant recipient. J. Hear. Lung Transplant. 2006.
- Bryant, J.M.; Grogono, D.M.; Rodriguez-Rincon, D.; Everall, I.; Brown, K.P.; Moreno, P.; Verma, D.; Hill, E.; Drijkoningen, J.; Gilligan, P.; et al. Emergence and spread of a humantransmissible multidrug-resistant nontuberculous mycobacterium. Science 2016.
- 33. Hermansen, T.S.; Ravn, P.; Svensson, E.; Lillebaek, T. Nontuberculous mycobacteria in Denmark, incidence and clinical importance during the last quarter-century. Sci. Rep. 2017.
- Baldwin, S.L.; Larsenid, S.E.; Ordway, D.; Cassell, G.; Coler, R.N. The complexities and challenges of preventing and treating nontuberculous mycobacterial diseases. PLoS Negl. Trop. Dis. 2019, 13, e0007083.
- 35. Faverio, P.; Stainer, A.; Bonaiti, G.; Zucchetti, S.C.; Simonetta, E.; Lapadula, G.; Marruchella, A.; Gori, A.; Blasi, F.; Codecasa, L.; et al. Characterizing non-tuberculous mycobacteria infection in bronchiectasis. Int. J. Mol. Sci. 2016.
- 36. Santin, M.; Dorca, J.; Alcaide, F.; Gonzalez, L.; Casas, S.; Lopez, M.; Guerra, M.R. Long-term relapses after 12-month treatment for Mycobacterium kansasii lung disease. Eur. Respir. J. 2009.
- 37. Sim, Y.S.; Park, H.Y.; Jeon, K.; Suh, G.Y.; Kwon, O.J.; Koh, W.-J. Standardized combination antibiotic treatment of Mycobacterium avium complex lung disease. Yonsei Med. J. 2010, 51, 888.
- Wallace, R.J.; Dukart, G.; Brown-Elliott, B.A.; Griffith, D.E.; Scerpella, E.G.; Marshall, B. Clinical experience in 52 patients with tigecycline-containing regimens for salvage treatment of Mycobacterium abscessus and Mycobacterium chelonae infections. J. Antimicrob. Chemother. 2014, 69, 1945–1953.

- Diagnosis and treatment of disease caused by nontuberculous mycobacteria. This official statement of the American Thoracic Society was approved by the Board of Directors, March 1997. Medical Section of the American Lung Association. Am. J. Respir. Crit. Care Med. 1997, 156, S1–S25.
- 40. Mirsaeidi, M.; Farshidpour, M.; Allen, M.B.; Ebrahimi, G.; Falkinham, J.O. Highlight on advances in nontuberculous mycobacterial disease in North America. Biomed Res. Int. 2014, 2014, 919474.
- 41. Wu, M.-L.; Aziz, D.B.; Dartois, V.; Dick, T. NTM drug discovery: Status, gaps and the way forward. Drug Discov. Today 2018, 23, 1502–1519.
- 42. Munita, J.M.; Arias, C.A.; Unit, A.R.; Santiago, A. De HHS Public access mechanisms of antibiotic resistance. HHS Public Access 2016, 4, 1–37.
- 43. Moon, S.M.; Park, H.Y.; Kim, S.-Y.; Jhun, B.W.; Lee, H.; Jeon, K.; Kim, D.H.; Huh, H.J.; Ki, C.-S.; Lee, N.Y.; et al. Clinical characteristics, treatment outcomes, and resistance mutations associated with macrolide-resistant Mycobacterium avium complex lung disease. Antimicrob. Agents Chemother. 2016, 60, 6758–6765.
- 44. Nasiri, M.J.; Haeili, M.; Ghazi, M.; Goudarzi, H.; Pormohammad, A.; Imani Fooladi, A.A.; Feizabadi, M.M. New insights in to the intrinsic and acquired drug resistance mechanisms in mycobacteria. Front. Microbiol. 2017, 8.
- 45. Hayes, J.D.; Wolf, C.R. Molecular mechanisms of drug resistance. Biochem. J. 1990, 272, 281–295.
- 46. Rampacci, E.; Stefanetti, V.; Passamonti, F.; Henao-Tamayo, M. Preclinical models of nontuberculous mycobacteria infection for early drug discovery and vaccine research. Pathogens 2020, 9, 641.
- 47. Rastogi, N.; Frehel, C.; Ryter, A.; Ohayon, H.; Lesourd, M.; David, H.L. Multiple drug resistance in Mycobacterium avium: Is the wall architecture responsible for exclusion of antimicrobial agents? Antimicrob. Agents Chemother. 1981, 20, 666–677.
- 48. Haemers, A.; Leysen, D.C.; Bollaert, W.; Zhang, M.Q.; Pattyn, S.R. Influence of N substitution on antimycobacterial activity of ciprofloxacin. Antimicrob. Agents Chemother. 1990, 34, 496–497.
- 49. Franzblau, S.G.; White, K.E. Comparative in vitro activities of 20 fluoroquinolones against Mycobacterium leprae. Antimicrob. Agents Chemother. 1990, 34, 229–231.
- Chopra, S.; Matsuyama, K.; Hutson, C.; Madrid, P. Identification of antimicrobial activity among FDA-approved drugs for combating Mycobacterium abscessus and Mycobacterium chelonae. J. Antimicrob. Chemother. 2011, 66, 1533– 1536.
- 51. Jeong, J.; Kim, G.; Moon, C.; Kim, H.J.; Kim, T.H.; Jang, J. Pathogen box screening for hit identification against Mycobacterium abscessus. PLoS ONE 2018, 13, e0195595.
- 52. Low, J.L.; Wu, M.-L.; Aziz, D.B.; Laleu, B.; Dick, T. Screening of TB actives for activity against nontuberculous mycobacteria delivers high hit rates. Front. Microbiol. 2017, 8.
- Cowman, S.; Burns, K.; Benson, S.; Wilson, R.; Loebinger, M.R. The antimicrobial susceptibility of non-tuberculous mycobacteria. J. Infect. 2016, 72, 324–331.
- 54. McGuffin, S.; Mullen, S.; Early, J.; Parish, T. 1341. Development of a series of high-throughput screens to identify leads for nontuberculous mycobacteria drug design. Open Forum Infect. Dis. 2019, 6, S485.
- 55. Falkinham, J.O. Challenges of NTM drug development. Front. Microbiol. 2018, 9.
- Zimmer, B.L.; DeYoung, D.R.; Roberts, G.D. In vitro synergistic activity of ethambutol, isoniazid, kanamycin, rifampin, and streptomycin against Mycobacterium avium-intracellulare complex. Antimicrob. Agents Chemother. 1982, 22, 148– 150.
- 57. Mukherjee, D.; Wu, M.-L.; Teo, J.W.P.; Dick, T. Vancomycin and clarithromycin show synergy against Mycobacterium abscessus in vitro. Antimicrob. Agents Chemother. 2017, 61.
- 58. Machado, D.; Cannalire, R.; Santos Costa, S.; Manfroni, G.; Tabarrini, O.; Cecchetti, V.; Couto, I.; Viveiros, M.; Sabatini, S. Boosting Effect of 2-phenylquinoline efflux inhibitors in combination with macrolides against Mycobacterium smegmatis and Mycobacterium avium. ACS Infect. Dis. 2016, 1, 593–603.
- 59. Felicetti, T.; Machado, D.; Cannalire, R.; Astolfi, A.; Massari, S.; Tabarrini, O.; Manfroni, G.; Barreca, M.L.; Cecchetti, V.; Viveiros, M.; et al. Modifications on C6 and C7 positions of 3-phenylquinolone efflux pump inhibitors led to potent and safe antimycobacterial treatment adjuvants. ACS Infect. Dis. 2019, 5, 982–1000.
- Blondiaux, N.; Moune, M.; Desroses, M.; Frita, R.; Flipo, M.; Mathys, V.; Soetaert, K.; Kiass, M.; Delorme, V.; Djaout, K.; et al. Reversion of antibiotic resistance in Mycobacterium tuberculosis by spiroisoxazoline SMARt-420. Science 2017, 355, 1206–1211.
- 61. Zheng, W.; Sun, W.; Simeonov, A. Drug repurposing screens and synergistic drug-combinations for infectious diseases. Br. J. Pharmacol. 2018, 175, 181–191.

- 62. Worthington, R.J.; Melander, C. Combination approaches to combat multidrug-resistant bacteria. Trends Biotechnol. 2013, 31, 177–184.
- 63. Lefebvre, A.-L.; Dubée, V.; Cortes, M.; Dorchêne, D.; Arthur, M.; Mainardi, J.-L. Bactericidal and intracellular activity of β-lactams against Mycobacterium abscessus. J. Antimicrob. Chemother. 2016, 71, 1556–1563.
- 64. Pandey, R.; Chen, L.; Manca, C.; Jenkins, S.; Glaser, L.; Vinnard, C.; Stone, G.; Lee, J.; Mathema, B.; Nuermberger, E.L.; et al. Dual β-lactam combinations highly active against Mycobacterium abscessus complex in vitro. MBio 2019, 10.
- 65. Story-Roller, E.; Maggioncalda, E.C.; Lamichhane, G. Select β-lactam combinations exhibit synergy against Mycobacterium abscessus in vitro. Antimicrob. Agents Chemother. 2019, 63.
- 66. Hetényi, C.; van der Spoel, D. Blind docking of drug-sized compounds to proteins with up to a thousand residues. FEBS Lett. 2006, 580, 1447–1450.
- 67. Johnson, E.O.; Office, E.; Kawate, T.; Orzechowski, M.; Hung, D.T. Discovery of a novel synergistic antimycobacterial combination targeting EfpA using large-scale chemical-genetics. bioRxiv 2019.
- 68. Wallis, R.S.; Hafner, R. Advancing host-directed therapy for tuberculosis. Nat. Rev. Immunol. 2015, 15, 255–263.
- 69. Tobin, D.M. Host-Directed therapies for tuberculosis. Cold Spring Harb. Perspect. Med. 2015, 5, a021196.
- 70. Schwegmann, A.; Brombacher, F. Host-Directed drug targeting of factors hijacked by pathogens. Sci. Signal. 2008, 1, re8.
- Hawn, T.R.; Shah, J.A.; Kalman, D. New tricks for old dogs: Countering antibiotic resistance in tuberculosis with hostdirected therapeutics. Immunol. Rev. 2015, 264, 344–362.
- 72. Cai, S.; Sato, K.; Shimizu, T.; Yamabe, S.; Hiraki, M.; Sano, C.; Tomioka, H. Antimicrobial activity of picolinic acid against extracellular and intracellular Mycobacterium avium complex and its combined activity with clarithromycin, rifampicin and fluoroquinolones. J. Antimicrob. Chemother. 2006, 57, 85–93.
- 73. Gomes, M.S.; Dom, G.; Pedrosa, J.; Boelaert, J.R.; Appelberg, R. Effects of iron deprivation on Mycobacterium avium growth. Tuber. Lung Dis. 1999, 79, 321–328.
- 74. Pais, T.F.; Appelberg, R. Macrophage control of mycobacterial growth induced by picolinic acid is dependent on host cell apoptosis. J. Immunol. 2000, 164, 389–397.
- 75. Tanigawa, K.; Suzuki, K.; Nakamura, K.; Akama, T.; Kawashima, A.; Wu, H.; Hayashi, M.; Takahashi, S.-I.; Ikuyama, S.; Ito, T.; et al. Expression of adipose differentiation-related protein (ADRP) and perilipin in macrophages infected with Mycobacterium leprae. FEMS Microbiol. Lett. 2008, 289, 72–79.
- Degang, Y.; Akama, T.; Hara, T.; Tanigawa, K.; Ishido, Y.; Gidoh, M.; Makino, M.; Ishii, N.; Suzuki, K. Clofazimine modulates the expression of lipid metabolism proteins in Mycobacterium leprae-infected macrophages. PLoS Negl. Trop. Dis. 2012, 6, e1936.
- Tang, J.; Chen, Q.; Guo, J.; Yang, L.; Tao, Y.; Li, L.; Miao, H.; Feng, H.; Chen, Z.; Zhu, G. Minocycline attenuates neonatal germinal-matrix-hemorrhage-induced neuroinflammation and brain edema by activating cannabinoid receptor
 Mol. Neurobiol. 2016, 53, 1935–1948.
- Montes-Worboys, A.; Brown, S.; Regev, D.; Bellew, B.F.; Mohammed, K.A.; Faruqi, I.; Sharma, P.; Moudgil, B.; Antony, V.B. Targeted delivery of amikacin into granuloma. Am. J. Respir. Crit. Care Med. 2010, 182, 1546–1553.
- Rao, M.; Ippolito, G.; Mfinanga, S.; Ntoumi, F.; Yeboah-Manu, D.; Vilaplana, C.; Zumla, A.; Maeurer, M. Improving treatment outcomes for MDR-TB—Novel host-directed therapies and personalised medicine of the future. Int. J. Infect. Dis. 2019, 80, S62–S67.

Retrieved from https://encyclopedia.pub/entry/history/show/17985