

Oxidative Stress in Human Pathology

Subjects: Pathology

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Accumulating evidence shows that oxidative stress plays an essential role in the pathogenesis and progression of many diseases. The imbalance between the production of reactive oxygen species (ROS) and the antioxidant systems has been extensively studied in pulmonary, neurodegenerative and cardiovascular disorders; however, its contribution is still debated in gastrointestinal disorders. Evidence suggests that oxidative stress affects gastrointestinal motility in obesity, and post-infectious disorders by favoring the smooth muscle phenotypic switch toward a synthetic phenotype. Here is to gain insight into the role played by oxidative stress in gas-trointestinal pathologies (GIT), and the involvement of ROS in the signaling underlying the mus-cular alterations of the gastrointestinal tract (GIT).

Keywords: oxidative stress ; gastrointestinal diseases ; gastrointestinal muscle inflammation ; antioxidants

1. Introduction

Oxidative stress in living organisms results from the imbalance between the production of reactive oxygen species (ROS) and the ability to neutralize them. The disparity between excessive reactive molecules and weak endogenous defense leads to damage to cell structures and molecules such as lipids, proteins, and DNA, ultimately contributing to the pathogenesis of a wide range of diseases. ROS, when available in appropriate low amounts, act as signal transduction molecules driving cell activities and also provide cell protection ^[1]. On the other hand, if generated in excess, as in inflammation, ROS can trigger the production of additional highly reactive species ^[2]. Crucially is the oxidative modification of key enzymes or regulatory sites, whose redox modification triggers cell signaling alteration and programmed cell death. Oxidative stress and inflammation are closely linked. Oxidative stress can cause inflammation and this, in turn, induces oxidative stress generating a vicious circle ^{[3][4]} that results in cell damage, which promotes a pro-inflammatory environment ^[5].

Literature data confirm the key role of oxidative stress in etiology of numerous and different diseases (Figure 1), including metabolic syndrome ^[6], atherosclerosis ^[7], cardiovascular disease ^{[8][9]}, cancer ^{[10][11]}, neurodegenerative disorders ^{[12][13]}, diabetes ^[14], infertility ^[15], renal diseases ^[16], and gastrointestinal and hepatic diseases ^[17].

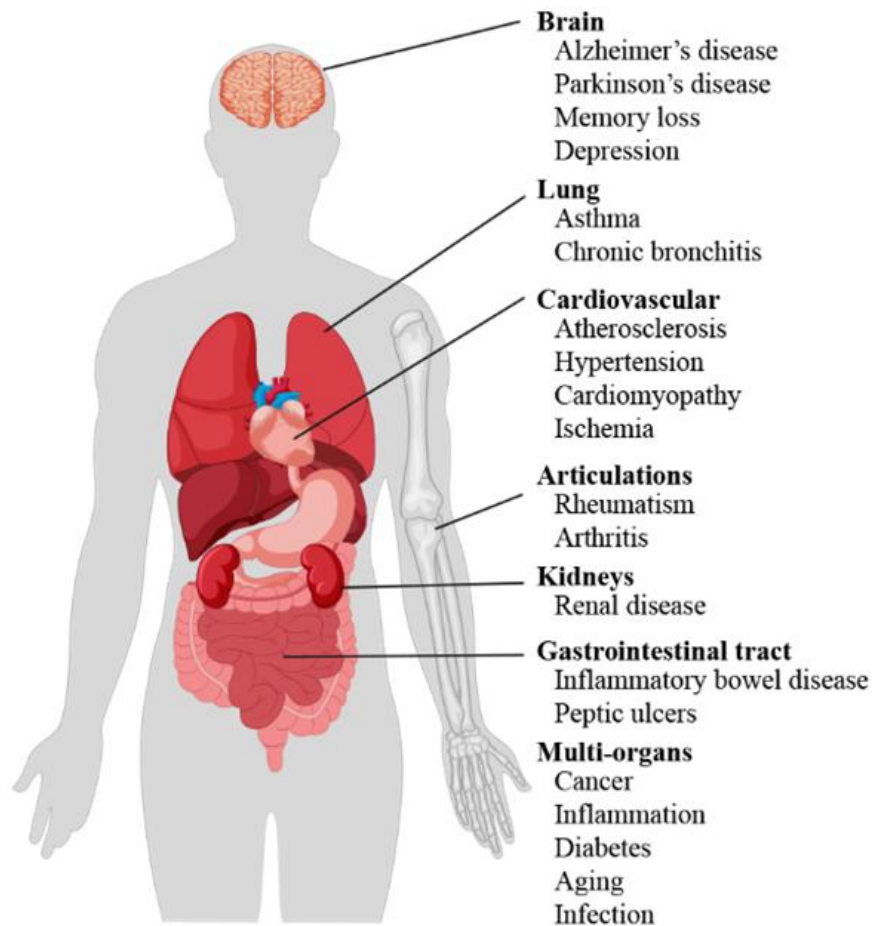


Figure 1. Scheme of oxidative stress-induced diseases in humans.

Being involved in the absorption of nutrients and in the immune response, the gastrointestinal tract (GIT) plays a key role also in the production of ROS. Several evidences highlight how the pathogenesis of various GIT diseases, including colorectal and gastric cancers ^{[18][19][20]}, inflammatory bowel disease (IBD) ^{[21][22]}, and peptic ulcers ^[23], is due, at least in part, to oxidative stress.

The GIT tissue is structured into four layers: the mucosa (epithelium, lamina propria, and muscular mucosae), the submucosa, the muscularis propria (inner circular muscle layer, intermuscular space, and outer longitudinal muscle layer), and the serosa.

The intestinal epithelia are exposed continuously to a wide variety of potentially harmful substances and act as a selective barrier between the tissues and luminal environment of the GIT. There are several stressors, which induce the generation of free radicals and result in oxidative stress and GIT inflammatory responses involving the epithelium and immune/inflammatory cells ^[24]. Although there is enough information on the role played by oxidative stress in the damage of intestinal mucosa, little is known about the involvement of the surrounding muscle layers.

Knowledge of the biochemical mechanisms underlying the alterations induced by oxidative stress at the GIT level, as well as of the physiological responses of the different GIT layers to such stress, is mandatory to better understand either pathogenesis of GIT diseases or to develop new and more effective therapeutic strategies.

This review summarizes the current understanding of the role of oxidative stress in GIT pathophysiology, also discussing the specific molecular mechanisms involved, focusing particular attention on the implication of the muscular layers of the GIT.

2. Oxidative Stress

Oxidative stress occurs when, in tissues and organs, the formation of highly reactive molecules e.g., ROS, reactive nitrogen species (RNS), and reactive sulfur species (RSS), overcome the endogenous antioxidant defense system capacities, leading to cellular damage and dysfunctions that result in a wide range of diseases. The reactive species are constantly generated within cells at low concentrations as a result of normal metabolic processes. They can also results from the exposure to external factors like radiation (X-rays and UV), ozone, air pollutants, cigarette smoke, bacteria, viruses, drugs, etc. ^[25], or as the outcome of an acute or chronic cellular stress. The reactive species can be free radicals

and non-radical oxidants. The free radicals are unstable because of unpaired electrons presence in their outer electron orbit. Since free radicals are highly unstable and reactive, tend to neutralize themselves by reacting with other molecules causing their oxidation [26]. Therefore, by reacting with important biological molecules, including DNA, lipids and proteins, they can cause damage on various levels [27]. Proteins, being among the main components of the cells, represent major targets for free radicals [28]. Free radicals can induce some protein modifications, i.e., unfolding or alteration of protein structure, most of which, fortunately, are essentially harmless events [29]. While the reversible oxidative changes are involved in the regulation of protein activity, irreversible protein changes can lead to their inactivation with consequent lasting harmful cellular effects [29].

The intracellular sources of chemical reactive species are mainly mitochondria, endoplasmic reticulum, lysosomes, peroxisomes, cytosol, and plasma membrane [30] (Figure 2). ROS derive from the chemical reduction of molecular oxygen and, among the main ones, we find: the free radicals, such as superoxide anion radical ($O_2^{\cdot-}$), hydroxyl radical ($\cdot OH$), as well as non-radical oxidant, such as hydrogen peroxide (H_2O_2) and hypochlorous acid ($HClO$) [31]. Among the RNS, the major players are peroxynitrite radical ($ONOO^{\cdot-}$), ozone, and nitric oxide ($\cdot NO$) [32]. The new identified RSS include thiol radical (RS^{\cdot}), and RSS both formed by the reaction between ROS and thiols. Similarly, RSS include radical species, such as (RSR^{\cdot}), glutathionyl radical ($GSSG^{\cdot}$), and non-radicals ones, such as reactive sulfane species (RSR), reactive sulfur substances (SO_2 , SO_3), etc. [33][34]. In particular, RSS are able to trigger both oxidation and reduction reactions with particular tropism for sulfur-containing molecules, such as peptides and proteins [33][34].

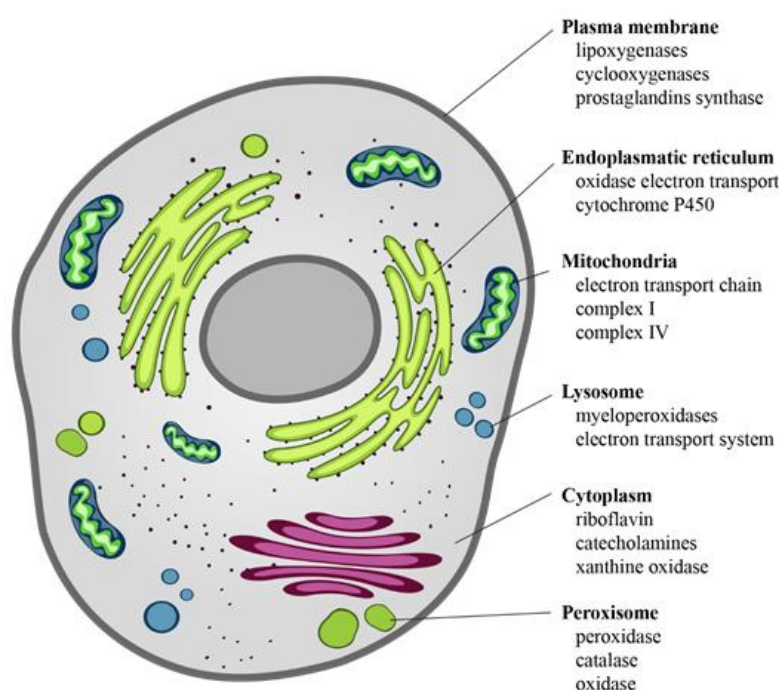


Figure 2. Cellular sources of ROS. ROS are the “by-products” of electron transfer reactions. The major source of ROS is the mitochondrial electron transport chain, followed by the NADPH oxidases present on either side of the plasma membrane. In the smooth endoplasmic reticulum, we find cytochrome P-450 and b5 families, which are responsible for a series of reactions to detoxify fat-soluble drugs and harmful metabolites. Peroxisomes, through their oxidases, are a significant source of total cellular H_2O_2 production. Moreover, they are responsible for dismutation of H_2O_2 to H_2O and O_2 , and of fatty acids β -oxidation. Other enzymes, present free in the cytoplasm, such as xanthine oxidase, aldehyde oxidase, flavoprotein dehydrogenase, and tryptophan dioxygenase can produce ROS during catalytic cycling.

The most important sites of ROS production are the enzymes of the mitochondrial electron transport respiratory chain. Other enzymes catalyze chemical reactions contributing to the ROS formation, among them the homologs of nicotinamide adenine dinucleotide phosphate (NADPH) oxidase, phospholipase A2 (PLA2), uncoupled nitric oxide synthase (NOS) as well as cyclooxygenases (COX), xanthine oxidase (XO), lipoxygenases (LOXs), glucose oxidase, and myeloperoxidase (MPO) [35][36]. NADPH oxidase, an enzyme present in the plasma membrane, was initially discovered in the phagosomes of macrophages, neutrophils, and monocytes. There are six homologs of NADPH oxidase, NOX1, NOX3-5, and dual oxidase (DUOX) 1 and 2, with several intracellular localizations [37]. Literature data showed that NOX1 and DUOX2 have significant roles in *Helicobacter pylori*-induced gastric inflammation, inflammatory bowel disease (IBD), and tumors [38]. XO is present in the cytoplasm and also on the outer surface of the plasma membrane; it is mainly expressed in the liver and small intestinal mucosa within the gastrointestinal tract [39]. LOXs are non-heme iron enzymes that can generate ROS catalyzing oxidation of arachidonic acid (AA). MPO is a heme-enzyme localized in lysosomes of neutrophils, macrophages, and monocytes. Several data demonstrate that MPO activity is increased in inflamed mucosa in ulcerative

colitis and also in *H. pylori*-infected subjects, playing a role in the development of *H. pylori*-induced atrophic gastritis. The chronic oxidative stress related to ulcerative colitis and *H. pylori*-infection could also lead to cancer, often associated with these diseases [40][41]. NOS is a heme-containing monooxygenase that generates NO. There are three different isoforms of NOS: neuronal NOS (nNOS), endothelial NOS (eNOS), and endotoxin or cytotoxin-inducible NOS (iNOS) [42]. In GIT, NOS expression and activity are very important because the generation of NO maintains normal functions of mucosa and plays a cytoprotective role. Indeed, NO regulates blood flow, epithelial secretion, and barrier function of gastric mucosal [43] and represents one of the main enteric neurotransmitters mediating GI muscle relaxation [43][44]. However, NO can also have deleterious effects, and iNOS expression was found increased in chronic ulcerative colitis and peptic ulcer patients. COX enzyme releases AA from the membrane phospholipids and catalyzes AA conversion to prostanooids. COX has two isoforms, COX-1 and COX-2, both of which are expressed in normal human gastric mucosa. COX-1 is constitutively expressed, while COX-2 is induced by inflammation and tumorigenesis [45]. COX-2 has also been reported to have cytoprotective functions in human colon and gastric cancer cells where it was induced during high osmotic stress [46]. Therefore, reactive oxygen species, including oxygen free radicals, are generated by the activity of several types of oxidases. Initially, O_2 is reduced by the addition of electrons, thereby producing $O_2^{\cdot-}$ that can react with other endogenous molecules to generate secondary oxidizing molecules, such as $ONOO^{\cdot}$. Thereafter, the reduction of $O_2^{\cdot-}$ leads to the by-product H_2O_2 that is characterized by a long life span and relative stability. The latter is enzymatically converted into water and O_2 , or possibly into different metabolites, thus extinguishing the radical cascade.

Both $O_2^{\cdot-}$ and H_2O_2 are also important signaling molecules, particularly in vascular smooth muscle cells where they can trigger specific biochemical pathways that regulate the defense mechanisms following exposure to oxidative stress. At the center of these pathways are for example mitogen-activated protein kinases, and tyrosine kinases, and transcription factors [47]. Particularly transcription factors, such as activator protein-1 (AP-1), NF- κ B, and/or NF-E2-related factor (NRF2) have been reported to also participate in redox-modulated cell signaling [48][49].

3. Antioxidants

If the body's antioxidant defense system fails to neutralize the excess free radicals, the imbalance between oxidants and the defense system can lead to pathological conditions, including cancer, cardiovascular disease, neurodegenerative disorders, atherosclerosis, and others. Halliwell and Gutteridge defined antioxidants as "*any substance that delays, prevents or removes oxidative damage to a target molecule*" [50]. All living organisms are endowed with endogenous antioxidant defenses capable of contrasting and removing reactive chemical species. However, these defenses are insufficient to totally remove reactive species and completely prevent oxidative damage to cells, tissues, and organs. The endogenous antioxidants can act at various levels: blocking the formation of radicals, neutralizing them by oxidizing themselves, or delaying the oxidation reactions of other molecules. Moreover, some antioxidants, acting as metal chelators, transform metal pro-oxidants into more stable chemical forms.

The antioxidants were be classified by Gutteridge and Halliwell into three categories: primary, secondary, and tertiary antioxidants, on the bases of their mechanism of action [51]. Primary antioxidants inhibit oxidant formation; secondary antioxidants function as scavengers of ROS, and tertiary antioxidants repair the oxidized molecules. Currently, antioxidants are substantially classified as enzymatic or non-enzymatic (Figure 3).

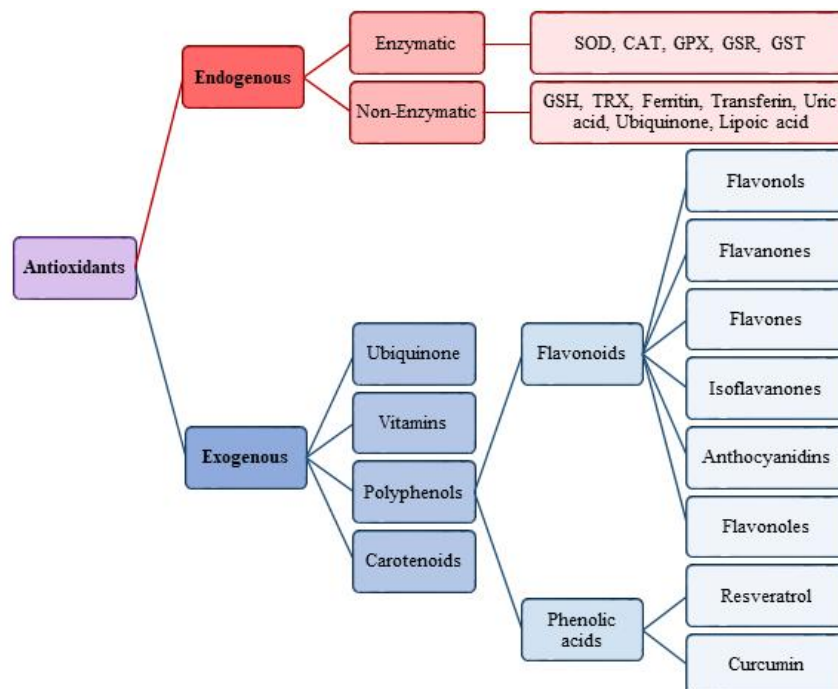


Figure 3. Scheme of endogenous and exogenous antioxidants. SOD, Superoxide dismutase; CAT, Catalase; GPX, Glutathione peroxidase; GSR, Glutathione reductase; GST, Glutathione transferase.

3.1. Enzymatic Antioxidants

Among the enzymatic antioxidants that contribute to the defense against the reactive species, we find catalase (CAT), superoxide dismutase (SOD), glutathione peroxidase (GPX), and glutathione reductase (GSR). Enzymatic antioxidants have both primary and secondary defense functions and represent an endogenous antioxidant system. Glutathione peroxidase, SOD, and catalase are the primary defense that prevents the formation or neutralize reactive species [52][53]. In particular, SOD and catalase provide major antioxidant defenses against ROS.

SOD catalyzes the dismutation of O_2^- into O_2 and H_2O_2 . In humans beings are present three isoforms of SOD [54]: cytosolic copper and zinc-containing enzyme (Cu-Zn-SOD), present in the mitochondrial inter-membranous space; manganese-requiring mitochondrial enzyme (Mn-SOD), present in the mitochondrial matrix; and extracellular Cu-Zn containing SOD (EC-SOD) [55]. H_2O_2 not scavenged by GPX located at the level of the mitochondrial matrix crosses the mitochondrial membrane towards the cytosol, where it can be scavenged either by cytosolic Cu-Zn-SOD or CAT [56]. Increased levels of all three SOD isoforms are present in intestinal tissues from IBD patients, particularly in the epithelium [57], and in patients with ulcer healing [58]. Increased expression of Mn-SOD is associated with colorectal cancer, and it was also found increased in normal mucosa of gastric adenocarcinoma as well as in squamous cell oesophageal carcinoma tissues [59]. Moreover, a gastrointestinal mucosal injury could be prevented by the presence of SOD [60].

CAT, present mainly in peroxisomes, dismutates H_2O_2 to H_2O and O_2 [61]. In humans, CAT has been found virtually in all organs although it is produced largely in liver, kidney, and erythrocytes. Lower catalase activity was observed in colorectal cancer [62], gastric adenocarcinoma, in *H. pylori*-infected stomach [62], and in Crohn's disease [63].

GPX converts glutathione (GSH) into its oxidized form (GSSG), reduces H_2O_2 to H_2O , and lipid hydroperoxides (ROOH) to the corresponding stable alcohols. The GPX reaction is paired to glutathione reductase (GSR), which maintains reduced glutathione (GSH) levels. GSR, GPX, and glutathione S-transferases (GST), form the glutathione system that in the GIT mucosa acts as an antioxidative barrier. This enzyme, generating GSH, is important for the protection of cell membranes, red blood cells, and hemoglobin to oxidative stress [64]. GPX is found in the mitochondria, cytoplasm, and extracellular space [65], and protects cells from the harmful consequences of peroxide decomposition. In humans, there are eight isotypes of GPX. While GPX1 is ubiquitous, GPX2 is specific for the gastrointestinal tract and protects the gut against the absorption of dietary hydroperoxides [66]. Moreover, GPX2 defends the gastrointestinal tract against ROS derived from gut inflammation associated with commensal bacteria [67]. Importantly, glucose-6-phosphate dehydrogenase, while not directly neutralizing the radicals, can be considered an antioxidant enzyme. This oxidoreductase maintains the level of NADPH, thus helping to keep glutathione in its reduced state (GSH) [68] and creating a reducing environment.

Thioredoxin reductase (TrxR) together with thioredoxin (Trx) forms the thioredoxin system. There are three TrxR isoforms: TrxR1 found in the cytoplasm, TrxR2 in mitochondria, and TrxR3 present only in specialized tissues. TrxR, by transferring reducing equivalents from NADPH to thioredoxin, keeps it in its reduced form [69]. It has been shown that a compensatory

upregulation of TrxR mRNA in gastrointestinal cancer was induced by oxidative stress provoked by bile acids [70].

3.2. Non-enzymatic Antioxidants

Among the endogenous non-enzymatic antioxidants, there are glutathione and Trx. Glutathione is ubiquitously expressed mostly in its reduced form, GSH. Glutathione is a strong antioxidant, certainly one of the most important among those that the body can produce. Relevant is its action against both free radicals and molecules such as hydrogen peroxide, nitrites, nitrates, benzoates, and others. An important element for its functioning is NADPH. In fact, this molecule, a derivative of vitamin PP (nicotinic acid), functions as a redox cofactor of the enzyme GSR, which reduced glutathione (GSH) from oxidized glutathione (or GSSG) through electrons transferred from NADPH to GSSG.

Trx contains two free sulfhydryl groups of two cysteine residues. It is involved in the biosynthesis of deoxynucleotides, since it reduces the oxidized ribonucleotide reductase by yielding their hydrogens to the two oxidized sulfhydryl groups of the ribonucleotide reductase. Trx is present in the cytoplasm, membranes, and mitochondria but also in the extracellular space [71]. It showed a cytoprotective action in various inflammatory conditions, and was found to regulate the activity of redox-sensitive transcription factors, which are part of the antioxidant defence system.

Ubiquinone, also known as CoQ, is a lipophilic molecule existing in three different redox states: fully oxidized, partially reduced (ubisemiquinone), and fully reduced (ubiquinol) [72]. It is found in the plasma membrane and in several intracellular membrane including mitochondrial ones where it plays a key role in energy production and ROS generation. In its fully reduced form, CoQ is a potential antioxidant. Experimental studies have shown a protective role of ubiquinone against protein carbonylation and oxidative damage to DNA [73][74]. Furthermore, it is also been shown that ubiquinone can prevent peroxidative damage to membrane phospholipids [75] and regenerate other powerful antioxidants, such as α -tocopherol and ascorbate, by recycling them back to their reduced active forms, thus increasing resources cellular antioxidants [76]. These properties make ubiquinone suitable as a food supplement to improve cellular bioenergetics and to counteract some age-related diseases.

Activator protein-1 (AP-1), nuclear factor kappa B (NF- κ B), and nuclear factor (erythroid-derived 2)-like 2 related factor (NRF2) are three transcription factors that have been reported to be involved in redox-modulated signaling pathways. Indeed, oxidative stress up-regulates NF- κ B activity, and AP-1 and NRF2 activation depends on the environmental and/or intracellular redox state. Under normal conditions, NRF2 is found blocked in the cytosol by its inhibitor, KEAP1. Oxidative modification of KEAP1 and NRF2 phosphorylation result in the release of NRF2 from KEAP1 [77] and its translocation into the nucleus, where it binds with antioxidant response elements involved in activation of gene expression, thereby protecting cells from free radical damage. Therefore, NRF2, through its interaction with antioxidant response element (ARE), is able to modulate the expression of defensive genes coding detoxifying enzymes and antioxidant proteins [78].

3.3. Exogenous Antioxidants

Besides the endogenous enzymatic and non-enzymatic antioxidant defenses, other antioxidants are also utilized by the body, which must necessarily be introduced through diet and for this reason are defined exogenous. In addition to endogenous antioxidants, exogenous ones act through different mechanisms and in different cellular compartments. They are mainly free radical scavengers: they neutralize free radicals, repair oxidized membranes, and decrease reactive oxygen species production [79]. Among the exogenous antioxidants, we find: vitamins (A, C, E, and K), enzyme cofactors (Q10), nitrogen compounds (uric acid), minerals (zinc, Zn and selenium, Se), and polyphenols (flavonoids and phenolic acid) [80]. Metals such as manganese, zinc, copper, iron, and selenium up-regulate the catalytic activity of antioxidant enzymes [81]. It has been indicated that an inadequate dietary intake of these trace minerals may compromise the effectiveness of antioxidant defense mechanisms [82].

Exogenous antioxidants have generated a growing interest in preventing or reducing oxidative stress. In fact, many epidemiological researches have highlighted how the use of foods containing antioxidants and scavengers has a potential protective effect against the disorders caused by oxidative stress [83][84][85][86][87]. By increasing the body's natural antioxidant defenses, or by supplementing with dietary antioxidants, various chronic diseases can be prevented or their progression can be slowed down. Natural antioxidants such as flavonoids, tannins, and polyphenols act by donating electrons to intermediate radicals and play a role in the inhibition of lipid peroxidation. For example, vitamin E, particularly its active form α -tocopherol, protects cells from lipid peroxidation and helps in the prevention of chronic diseases associated with oxidative stress [88][89].

The antioxidant phytochemicals contained in vegetables and fruits are considered a benefits to the health. Indeed, several studies demonstrated that they have antioxidant abilities both in vitro and in vivo [90][91]. Moreover, literature data highlighted that antioxidant phytochemicals can also have anti-inflammatory action [92]. In fact, natural compounds such

as curcumin, resveratrol, and anthocyanins could reduce inflammation via inhibition of prostaglandin production, NF- κ B activity, and specific oxidative enzymes, as well as by increasing anti-inflammatory cytokine (e.g., IL-10) or decreasing pro-inflammatory cytokine (i.e., IL-1 β) production [93][94].

References

1. Janssen-Heininger, Y.M.; Mossman, B.T.; Heintz, N.H.; Forman, H.J.; Kalyanaraman, B.; Finkel, T.; Stamler, J.S.; Rhee, S.G.; van der Vliet, A. Redox-based regulation of signal transduction: Principles, pitfalls, and promises. *Free Radic. Biol. Med.* 2008, 45, 1–17.
2. Vaziri, N.D. Causal link between oxidative stress, inflammation, and hypertension. *Iran. J. Kidney Dis.* 2008, 2, 1–10.
3. Petrie, J.R.; Guzik, T.J.; Touyz, R.M. Diabetes, Hypertension, and Cardiovascular Disease: Clinical Insights and Vascular Mechanisms. *Can. J. Cardiol.* 2018, 34, 575–584.
4. Ishibashi, T. Molecular hydrogen: New antioxidant and anti-inflammatory therapy for rheumatoid arthritis and related diseases. *Curr. Pharm. Des.* 2013, 19, 6375–6381.
5. Ng, C.Y.; Kamisah, Y.; Faizah, O.; Jaarin, K. The role of repeatedly heated soybean oil in the development of hypertension in rats: Association with vascular inflammation. *Int. J. Exp. Pathol.* 2012, 93, 377–387.
6. Vona, R.; Gambardella, L.; Cittadini, C.; Straface, E.; Pietraforte, D. Biomarkers of Oxidative Stress in Metabolic Syndrome and Associated Diseases. *Oxid. Med. Cell Longev.* 2019, 2019, 8267234.
7. Violi, F.; Loffredo, L.; Carnevale, R.; Pignatelli, P.; Pastori, D. Atherothrombosis and Oxidative Stress: Mechanisms and Management in Elderly. *Antioxid. Redox Signal.* 2017, 27, 1083–1124.
8. Cai, H.; Harrison, D.G. Endothelial dysfunction in cardiovascular diseases: The role of oxidant stress. *Circ. Res.* 2000, 87, 840–844.
9. Majunova, M.; Dovinova, I.; Barancik, M.; Chan, J.Y. Redox signaling in pathophysiology of hypertension. *J. Biomed. Sci.* 2013, 20, 69.
10. Klaunig, J.E. Oxidative Stress and Cancer. *Curr. Pharm. Des.* 2018, 24, 4771–4778.
11. Reuter, S.; Gupta, S.C.; Chaturvedi, M.M.; Aggarwal, B.B. Oxidative stress, inflammation, and cancer: How are they linked? *Free Radic. Biol. Med.* 2010, 49, 1603–1616.
12. Islam, M.T. Oxidative stress and mitochondrial dysfunction-linked neurodegenerative disorders. *Neurol. Res.* 2017, 39, 73–82.
13. Singh, A.; Kukreti, R.; Saso, L.; Kukreti, S. Oxidative Stress: A Key Modulator in Neurodegenerative Diseases. *Molecules* 2019, 24, 1583.
14. Wang, J.; Wang, H. Oxidative Stress in Pancreatic Beta Cell Regeneration. *Oxid. Med. Cell Longev.* 2017, 2017, 1930261.
15. Noh, S.; Go, A.; Kim, D.B.; Park, M.; Jeon, H.W.; Kim, B. Role of Antioxidant Natural Products in Management of Infertility: A Review of Their Medicinal Potential. *Antioxidants (Basel)* 2020, 9, 957.
16. Ratliff, B.B.; Abdulmahdi, W.; Pawar, R.; Wolin, M.S. Oxidant Mechanisms in Renal Injury and Disease. *Antioxid. Redox Signal.* 2016, 25, 119–146.
17. Puentes-Pardo, J.D.; Moreno-SanJuan, S.; Carazo, A.; Leon, J. Heme Oxygenase-1 in Gastrointestinal Tract Health and Disease. *Antioxidants (Basel)* 2020, 9, 1214.
18. Kekec, Y.; Paydas, S.; Tuli, A.; Zorludemir, S.; Sakman, G.; Seydaoglu, G. Antioxidant enzyme levels in cases with gastrointestinal cancer. *Eur. J. Intern. Med.* 2009, 20, 403–406.
19. Inokuma, T.; Haraguchi, M.; Fujita, F.; Tajima, Y.; Kanematsu, T. Oxidative stress and tumor progression in colorectal cancer. *Hepatogastroenterology* 2009, 56, 343–347.
20. Zhang, L.; Li, L.; Gao, G.; Wei, G.; Zheng, Y.; Wang, C.; Gao, N.; Zhao, Y.; Deng, J.; Chen, H.; et al. Elevation of GPRC5A expression in colorectal cancer promotes tumor progression through VNN-1 induced oxidative stress. *Int. J. Cancer* 2017, 140, 2734–2747.
21. Grisham, M.B. Oxidants and free radicals in inflammatory bowel disease. *Lancet* 1994, 344, 859–861.
22. Pavlick, K.P.; Laroux, F.S.; Fuseler, J.; Wolf, R.E.; Gray, L.; Hoffman, J.; Grisham, M.B. Role of reactive metabolites of oxygen and nitrogen in inflammatory bowel disease. *Free Radic. Biol. Med.* 2002, 33, 311–322.

23. Peng, Y.C.; Hsu, C.L.; Tung, C.F.; Chou, W.K.; Huang, L.R.; Hung, D.Z.; Hu, W.H.; Yang, D.Y. Chemiluminescence assay of mucosal reactive oxygen species in gastric cancer, ulcer and antral mucosa. *Hepatogastroenterology* 2008, 55, 770–773.
24. Bhattacharyya, A.; Chattopadhyay, R.; Mitra, S.; Crowe, S.E. Oxidative stress: An essential factor in the pathogenesis of gastrointestinal mucosal diseases. *Physiol. Rev.* 2014, 94, 329–354.
25. Pham-Huy, L.A.; He, H.; Pham-Huy, C. Free radicals, antioxidants in disease and health. *Int. J. Biomed. Sci.* 2008, 4, 89–96.
26. Cheeseman, K.H.; Slater, T.F. An introduction to free radical biochemistry. *Br. Med. Bull.* 1993, 49, 481–493.
27. Lobo, V.; Patil, A.; Phatak, A.; Chandra, N. Free radicals, antioxidants and functional foods: Impact on human health. *Pharm. Rev.* 2010, 4, 118–126.
28. Davies, M.J.; Fu, S.; Wang, H.; Dean, R.T. Stable markers of oxidant damage to proteins and their application in the study of human disease. *Free Radic. Biol. Med.* 1999, 27, 1151–1163.
29. Dalle-Donne, I.; Scaloni, A.; Giustarini, D.; Cavarra, E.; Tell, G.; Lungarella, G.; Colombo, R.; Rossi, R.; Milzani, A. Proteins as biomarkers of oxidative/nitrosative stress in diseases: The contribution of redox proteomics. *Mass Spectrom. Rev.* 2005, 24, 55–99.
30. Balaban, R.S.; Nemoto, S.; Finkel, T. Mitochondria, oxidants, and aging. *Cell* 2005, 120, 483–495.
31. Schieber, M.; Chandel, N.S. ROS function in redox signaling and oxidative stress. *Curr. Biol.* 2014, 24, R453–R462.
32. Ray, P.D.; Huang, B.W.; Tsuji, Y. Reactive oxygen species (ROS) homeostasis and redox regulation in cellular signaling. *Cell Signal.* 2012, 24, 981–990.
33. Giles, G.I.; Nasim, M.J.; Ali, W.; Jacob, C. The Reactive Sulfur Species Concept: 15 Years On. *Antioxidants (Basel)* 2017, 6, 38.
34. Ali, S.S.; Ahsan, H.; Zia, M.K.; Siddiqui, T.; Khan, F.H. Understanding oxidants and antioxidants: Classical team with new players. *J. Food Biochem.* 2020, 44, e13145.
35. Munzel, T.; Gori, T.; Bruno, R.M.; Taddei, S. Is oxidative stress a therapeutic target in cardiovascular disease? *Eur. Heart J.* 2010, 31, 2741–2748.
36. Swindle, E.J.; Metcalfe, D.D. The role of reactive oxygen species and nitric oxide in mast cell-dependent inflammatory processes. *Immunol. Rev.* 2007, 217, 186–205.
37. Bedard, K.; Krause, K.H. The NOX family of ROS-generating NADPH oxidases: Physiology and pathophysiology. *Physiol. Rev.* 2007, 87, 245–313.
38. O'Neill, S.; Brault, J.; Stasia, M.-J.; Knaus, U.G. Genetic disorders coupled to ROS deficiency. *Redox Biol.* 2015, 6, 135–156.
39. van der Vliet, A.; Tuinstra, T.J.; Bast, A. Modulation of oxidative stress in the gastrointestinal tract and effect on rat intestinal motility. *Biochem. Pharm.* 1989, 38, 2807–2818.
40. Zhu, H.; Yang, L.; Zhou, B.; Yu, R.; Tang, N.; Wang, B. Myeloperoxidase G-463A polymorphism and the risk of gastric cancer: A case-control study. *Carcinogenesis* 2006, 27, 2491–2496.
41. Ullman, T.A.; Itzkowitz, S.H. Intestinal inflammation and cancer. *Gastroenterology* 2011, 140, 1807–1816.
42. Peek, R.M., Jr.; Fiske, C.; Wilson, K.T. Role of innate immunity in *Helicobacter pylori*-induced gastric malignancy. *Physiol. Rev.* 2010, 90, 831–858.
43. Barrachina, M.D.; Panes, J.; Esplugues, J.V. Role of nitric oxide in gastrointestinal inflammatory and ulcerative diseases: Perspective for drugs development. *Curr. Pharm. Des.* 2001, 7, 31–48.
44. Guerra, D.D.; Bok, R.; Vyas, V.; Orlicky, D.J.; Lorca, R.A.; Hurt, K.J. Akt phosphorylation of neuronal nitric oxide synthase regulates gastrointestinal motility in mouse ileum. *Proc. Natl. Acad. Sci. USA* 2019, 116, 17541–17546.
45. Marnett, L.J. The COXIB experience: A look in the rearview mirror. *Annu. Rev. Pharm. Toxicol.* 2009, 49, 265–290.
46. Mahkonen, A.; Putaala, H.; Mustonen, H.; Rautonen, N.; Puolakkainen, P. *Lactobacillus acidophilus* 74-2 and butyrate induce cyclooxygenase (COX)-1 expression in gastric cancer cells. *Immunopharmacol. Immunotoxicol.* 2008, 30, 503–518.
47. Lassegue, B.; Sorescu, D.; Szocs, K.; Yin, Q.; Akers, M.; Zhang, Y.; Grant, S.L.; Lambeth, J.D.; Griendling, K.K. Novel gp91(phox) homologues in vascular smooth muscle cells: nox1 mediates angiotensin II-induced superoxide formation and redox-sensitive signaling pathways. *Circ. Res.* 2001, 88, 888–894.

48. Brigelius-Flohe, R.; Flohe, L. Basic principles and emerging concepts in the redox control of transcription factors. *Antioxid. Redox Signal.* 2011, 15, 2335–2381.
49. Surh, Y.J.; Kundu, J.K.; Li, M.H.; Na, H.K.; Cha, Y.N. Role of Nrf2-mediated heme oxygenase-1 upregulation in adaptive survival response to nitrosative stress. *Arch. Pharm. Res.* 2009, 32, 1163–1176.
50. Halliwell, B. How to characterize an antioxidant: An update. *Biochem. Soc. Symp.* 1995, 61, 73–101.
51. Halliwell, B.; Gutteridge, J.M. The definition and measurement of antioxidants in biological systems. *Free Radic. Biol. Med.* 1995, 18, 125–126.
52. Halliwell, B.; Aeschbach, R.; Loliger, J.; Aruoma, O.I. The characterization of antioxidants. *Food Chem. Toxicol.* 1995, 33, 601–617.
53. Rahman, K. Studies on free radicals, antioxidants, and co-factors. *Clin. Interv. Aging* 2007, 2, 219–236.
54. Nozik-Grayck, E.; Suliman, H.B.; Piantadosi, C.A. Extracellular superoxide dismutase. *Int. J. Biochem. Cell Biol.* 2005, 37, 2466–2471.
55. Okado-Matsumoto, A.; Fridovich, I. Subcellular distribution of superoxide dismutases (SOD) in rat liver: Cu,Zn-SOD in mitochondria. *J. Biol. Chem.* 2001, 276, 38388–38393.
56. Poyton, R.O.; Ball, K.A.; Castello, P.R. Mitochondrial generation of free radicals and hypoxic signaling. *Trends Endocrinol. Metab.* 2009, 20, 332–340.
57. Kruidenier, L.; Kuiper, I.; van Duijn, W.; Marklund, S.L.; van Hogezand, R.A.; Lamers, C.B.; Verspaget, H.W. Differential mucosal expression of three superoxide dismutase isoforms in inflammatory bowel disease. *J. Pathol.* 2003, 201, 7–16.
58. Naito, Y.; Yoshikawa, T.; Ando, T.; Kishi, A.; Ueda, S.; Oyamada, H.; Kondo, M. Changes in superoxide dismutase activity in the gastric mucosa of peptic ulcer patients. *J. Clin. Gastroenterol.* 1992, 14 (Suppl. 1), S131–S134.
59. Janssen, A.M.; Bosman, C.B.; van Duijn, W.; Oostendorp-van de Ruit, M.M.; Kubben, F.J.; Griffioen, G.; Lamers, C.B.; van Krieken, J.H.; van de Velde, C.J.; Verspaget, H.W. Superoxide dismutases in gastric and esophageal cancer and their prognostic impact in gastric cancer. *Clin. Cancer Res.* 2000, 6, 3183–3192.
60. Klinowski, E.; Broide, E.; Varsano, R.; Eshchar, J.; Scapa, E. Superoxide dismutase activity in duodenal ulcer patients. *Eur. J. Gastroenterol. Hepatol.* 1996, 8, 1151–1155.
61. Schrader, M.; Fahimi, H.D. Peroxisomes and oxidative stress. *Biochim. Biophys. Acta* 2006, 1763, 1755–1766.
62. Chang, D.; Hu, Z.L.; Zhang, L.; Zhao, Y.S.; Meng, Q.H.; Guan, Q.B.; Zhou, J.; Pan, H.Z. Association of catalase genotype with oxidative stress in the predication of colorectal cancer: Modification by epidemiological factors. *Biomed. Environ. Sci.* 2012, 25, 156–162.
63. Iborra, M.; Moret, I.; Rausell, F.; Bastida, G.; Aguas, M.; Cerrillo, E.; Nos, P.; Beltran, B. Role of oxidative stress and antioxidant enzymes in Crohn's disease. *Biochem. Soc. Trans.* 2011, 39, 1102–1106.
64. Chang, J.C.; van der Hoeven, L.H.; Haddox, C.H. Glutathione reductase in the red blood cells. *Ann. Clin. Lab. Sci.* 1978, 8, 23–29.
65. Toppo, S.; Vanin, S.; Bosello, V.; Tosatto, S.C. Evolutionary and structural insights into the multifaceted glutathione peroxidase (Gpx) superfamily. *Antioxid. Redox Signal.* 2008, 10, 1501–1514.
66. Wingler, K.; Muller, C.; Schmehl, K.; Florian, S.; Brigelius-Flohe, R. Gastrointestinal glutathione peroxidase prevents transport of lipid hydroperoxides in CaCo-2 cells. *Gastroenterology* 2000, 119, 420–430.
67. Chu, F.F.; Esworthy, R.S.; Doroshov, J.H. Role of Se-dependent glutathione peroxidases in gastrointestinal inflammation and cancer. *Free Radic. Biol. Med.* 2004, 36, 1481–1495.
68. Thomas, D.; Cherest, H.; Surdin-Kerjan, Y. Identification of the structural gene for glucose-6-phosphate dehydrogenase in yeast. Inactivation leads to a nutritional requirement for organic sulfur. *EMBO J.* 1991, 10, 547–553.
69. Arner, E.S. Focus on mammalian thioredoxin reductases--important selenoproteins with versatile functions. *Biochim. Biophys. Acta* 2009, 1790, 495–526.
70. Lechner, S.; Muller-Ladner, U.; Schlottmann, K.; Jung, B.; McClelland, M.; Ruschoff, J.; Welsh, J.; Scholmerich, J.; Kullmann, F. Bile acids mimic oxidative stress induced upregulation of thioredoxin reductase in colon cancer cell lines. *Carcinogenesis* 2002, 23, 1281–1288.
71. Koharyova, M.; Kolarova, M. Oxidative stress and thioredoxin system. *Gen. Physiol. Biophys.* 2008, 27, 71–84.
72. Wang, Y.; Hekimi, S. Understanding Ubiquinone. *Trends Cell Biol.* 2016, 26, 367–378.
73. Forsmark-Andree, P.; Dallner, G.; Ernster, L. Endogenous ubiquinol prevents protein modification accompanying lipid peroxidation in beef heart submitochondrial particles. *Free Radic. Biol. Med.* 1995, 19, 749–757.

74. Forsmark-Andree, P.; Ernster, L. Evidence for a protective effect of endogenous ubiquinol against oxidative damage to mitochondrial protein and DNA during lipid peroxidation. *Mol. Asp. Med.* 1994, 15, s73–s81.
75. Sarmiento, A.; Diaz-Castro, J.; Pulido-Moran, M.; Kajarabille, N.; Guisado, R.; Ochoa, J.J. Coenzyme Q10 Supplementation and Exercise in Healthy Humans: A Systematic Review. *Curr. Drug Metab.* 2016, 17, 345–358.
76. Bentinger, M.; Brismar, K.; Dallner, G. The antioxidant role of coenzyme Q. *Mitochondrion* 2007, 7, S41–S50.
77. Zhang, D.D.; Hannink, M. Distinct cysteine residues in Keap1 are required for Keap1-dependent ubiquitination of Nrf2 and for stabilization of Nrf2 by chemopreventive agents and oxidative stress. *Mol. Cell Biol.* 2003, 23, 8137–8151.
78. Mann, G.E.; Niehueser-Saran, J.; Watson, A.; Gao, L.; Ishii, T.; de Winter, P.; Siow, R.C. Nrf2/ARE regulated antioxidant gene expression in endothelial and smooth muscle cells in oxidative stress: Implications for atherosclerosis and preeclampsia. *Sheng Li Xue Bao* 2007, 59, 117–127.
79. Berger, M.M. Can oxidative damage be treated nutritionally? *Clin. Nutr.* 2005, 24, 172–183.
80. Sies, H. Total antioxidant capacity: Appraisal of a concept. *J. Nutr.* 2007, 137, 1493–1495.
81. Vertuani, S.; Angusti, A.; Manfredini, S. The antioxidants and pro-antioxidants network: An overview. *Curr. Pharm. Des.* 2004, 10, 1677–1694.
82. Gale, C.R. Dietary antioxidants and dementia. *Int. Psychogeriatr.* 2001, 13, 259–262.
83. Block, G.; Patterson, B.; Subar, A. Fruit, vegetables, and cancer prevention: A review of the epidemiological evidence. *Nutr. Cancer* 1992, 18, 1–29.
84. Yamada, T.; Hayasaka, S.; Shibata, Y.; Ojima, T.; Saegusa, T.; Gotoh, T.; Ishikawa, S.; Nakamura, Y.; Kayaba, K.; Jichi Medical School Cohort Study. Frequency of citrus fruit intake is associated with the incidence of cardiovascular diseases: The Jichi Medical School cohort study. *J. Epidemiol.* 2011, 21, 169–175.
85. Mursu, J.; Virtanen, J.K.; Tuomainen, T.P.; Nurmi, T.; Voutilainen, S. Intake of fruit, berries, and vegetables and risk of type 2 diabetes in Finnish men: The Kuopio Ischaemic Heart Disease Risk Factor Study. *Am. J. Clin. Nutr.* 2014, 99, 328–333.
86. Kruk, J. Association between vegetable, fruit and carbohydrate intake and breast cancer risk in relation to physical activity. *Asian Pac. J. Cancer Prev.* 2014, 15, 4429–4436.
87. Kyro, C.; Skeie, G.; Loft, S.; Landberg, R.; Christensen, J.; Lund, E.; Nilsson, L.M.; Palmqvist, R.; Tjønneland, A.; Olsen, A. Intake of whole grains from different cereal and food sources and incidence of colorectal cancer in the Scandinavian HELGA cohort. *Cancer Causes Control.* 2013, 24, 1363–1374.
88. Azlina, M.F.N.; Qodriyah, M.S.; Kamisah, Y. Tocopherol and Tocotrienol: Therapeutic Potential in Animal Models of Stress. *Curr. Drug Targets* 2018, 19, 1456–1462.
89. Reiter, E.; Jiang, Q.; Christen, S. Anti-inflammatory properties of alpha- and gamma-tocopherol. *Mol. Asp. Med.* 2007, 28, 668–691.
90. Singh, M.; Suman, S.; Shukla, Y. New Enlightenment of Skin Cancer Chemoprevention through Phytochemicals: In Vitro and In Vivo Studies and the Underlying Mechanisms. *Biomed. Res. Int.* 2014, 2014, 243452.
91. Koren, E.; Kohen, R.; Ginsburg, I. Polyphenols enhance total oxidant-scavenging capacities of human blood by binding to red blood cells. *Exp. Biol. Med. (Maywood)* 2010, 235, 689–699.
92. Zhang, Y.J.; Gan, R.Y.; Li, S.; Zhou, Y.; Li, A.N.; Xu, D.P.; Li, H.B. Antioxidant Phytochemicals for the Prevention and Treatment of Chronic Diseases. *Molecules* 2015, 20, 21138–21156.
93. Hutchins-Wolfbrandt, A.; Mistry, A.M. Dietary turmeric potentially reduces the risk of cancer. *Asian Pac. J. Cancer Prev.* 2011, 12, 3169–3173.
94. Dominguez-Avila, J.A.; Villa-Rodriguez, J.A.; Montiel-Herrera, M.; Pacheco-Ordaz, R.; Roopchand, D.E.; Venema, K.; Gonzalez-Aguilar, G.A. Phenolic Compounds Promote Diversity of Gut Microbiota and Maintain Colonic Health. *Dig. Dis. Sci.* 2020, 1–20. doi: 10.1007/s10620-020-06676-7.