

Nutrition, Gastrointestinal Peptides, and Endocannabinoids in Childhood Cancer

Subjects: Oncology

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Childhood cancer is a rarely occurring disease. However, it is the second most frequent cause of death among children. It is estimated that each year cancer will be diagnosed in 400,000 children worldwide. The nutritional status at diagnosis is a prognostic indicator and influences the treatment tolerance. Both malnutrition and obesity increase the risk of mortality and complications during treatment. It is necessary to constantly search for new factors that impair the nutritional status. The endocannabinoid system (ECS) is a signaling system whose best-known function is regulating energy balance and food intake, but it also plays a role in pain control, embryogenesis, neurogenesis, learning, and the regulation of lipid and glucose metabolism. Its action is multidirectional, and its role is being discovered in an increasing number of diseases.

Keywords: cancer ; children ; nutritional status ; endocannabinoid system

1. Nutritional Status in Children with Cancer

Nutritional status is the biochemical, structural, and functional state of the body resulting from the level of coverage of energy and nutrient requirements and the action of factors influencing absorption and metabolism ^[1]. In children diagnosed with cancer, the nutritional status is one of the prognostic values influencing treatment tolerance, quality of life, drug metabolism, and overall survival ^{[2][3][4][5]}. Malnutrition and overweight may occur at diagnosis or appear during and after oncological therapy but should not be treated as a normal condition at any stage of treatment. Despite the growing interest in this topic, malnutrition among cancer patients remains a serious problem, ranging between 40–90% in lower-middle-income countries and between 0–30% in high-income countries ^[6]. This value varies depending on the type and stage of cancer, phase of treatment, and assessment method ^[2]. In addition, patients with high-risk treatment protocols are more likely to be malnourished ^{[7][8]}. During oncological treatment, in some types of cancer, weight gain is also observed. According to Iniesta R. et al. ^[9], the prevalence of overnutrition ranged from 8% to 78%. It is believed that the first nutritional status assessment has to be performed at diagnosis and should be repeated regularly during treatment ^[2]. The most noticeable changes in the nutritional status occur in the first months after diagnosis ^{[7][10]}. In a prospective study, Paciarotti et al. ^[11] have shown that after the first three months of treatment, the content of adipose tissue in children with leukemia increased to 130% of the norm, while in children diagnosed with other cancer, it decreased from 78% at diagnosis to 70% of the norm after three months of treatment. Furthermore, weight loss of >5% in the first 3 months of treatment and >10% after 6 months were associated with poorer survival ^[5].

1.1. Nutritional Disorders in Children Diagnosed with Various Types of Cancer

1.1.1. Leukemia

Leukemia is the most common childhood cancer ^[12]. It was showed that obesity and undernutrition are associated with worse survival in children with acute myeloid leukemia (AML) ^[13]. Moreover, a higher BMI at diagnosis was associated with worse event-free survival (EFS), poorer overall survival (OS), and higher mortality in children with AML and those with acute lymphoblastic leukemia (ALL) ^{[14][15][16][17]}. On the other hand, Orgel E. et al. ^[18] showed that BMI at diagnosis wasn't as important as underweight and obesity occurring more than half of the time between induction and maintenance. It was also indicated that obesity during induction increases the risk of persistent minimal residual disease (MRD) ^[19]. However, It does not confirm the relationship between BMI and EFS ^[20], MRD ^[21], and increased risk of recurrence ^{[22][23]}.

It is also worth noting that children with leukemia are at risk of low muscle mass. Suzuki D et al. ^[24] have assessed the content of skeletal muscle in patients with ALL using CT imaging at the L3 level. Skeletal muscle loss was demonstrated in all patients after the induction, while sarcopenia developed in almost 30% of the group ^[24].

1.1.2. Solid Tumors

It is difficult to unequivocally define the prevalence of nutritional disorders in children with solid tumors due to the varied results of individual ones resulting from different methods and times of nutritional assessment, as well as a different selection of the group [6]. It is believed that the risk of malnutrition in children with solid tumors is higher compared to other types of childhood cancers [7][25][26][27][28], although not all confirm this [29][30]. It was indicated that both too high and low BMI is associated with worse OS [31][32], worse response to treatment [33], and a higher risk of toxicity [34] and complications [35]. On the other hand, Sharib JM. et al. [33] do not point to malnutrition as a factor associated with increased treatment toxicity. Tenardi R et al. [36] carried out a retrospective assessment of the nutritional status of children with Ewing sarcoma and osteosarcoma, where a high risk of experiencing extreme body changes was observed [36]. Burke [37] observed that the loss of >10% of body weight was associated with an increased number of days of hospitalization [37]. Lifson L.F. et al. [38] have shown that malnutrition in children with Wilms' tumor reaches 66%, but no statistically significant relationship was found between nutritional status and survival [39][40]. In children with neuroblastoma, malnutrition is noticeable at diagnosis, and BMI decreases after 6 months of treatment, but no relationship has been found between BMI and survival [40]. In a prospective one by Avarnival et al. [41], it has been shown that the BMI of children with solid tumors decreased during the first 6 months of treatment and then gradually increased. It should be emphasized that the risk of malnutrition in children with solid tumors differs depending on the type of nutritional assessment method used. Children with solid tumors have a higher risk of malnutrition, regarding the MUAC, TSFT, and AMC indicators, compared to other methods [42]. This is caused by the tumor mass masking the real body weight, which impairs seemingly correct BMI measurements [42]. That is why it is so important to use measurements employing arm anthropometry, bioimpedance, or, if possible, DXA for these children. Furthermore, Joffe L. et al. [43] demonstrated by using single-slice T12-L1 images from routinely obtained chest CT scans that children with solid tumors lose skeletal muscle and fat in the early stages of therapy.

1.1.3. Central Nervous System (CNS) Tumors

Brain tumors are the second most common cancer in children after leukemia [12]. Tsutsumi et al. [44] have estimated that at diagnosis, 6.7% of children with CNS tumors were malnourished, while 23.3% were overweight. Iniesta R. et al. [7] confirm that patients with brain tumors had the highest risk of being overweight and obese compared to other types of cancer. In a prospective study, Brinksma A. et al. [10] have shown that in the first three months of treatment, most children with brain tumors increased the rate of WFA, BMI, and had a higher content of adipose tissue and lower lean body mass compared to children with solid tumors and hematological neoplasms. Musiol K. et al. [45] observed that in children with brain tumors, BMI was the lowest during the maintenance and was significantly different compared to the control group. After the end of treatment, BMI increased significantly [45].

1.2. Bone Health in Children with Cancer

The effect of treatment on the bone mineral content is also a very important issue because 40% of bone mass is formed in childhood [46]. Children treated for cancer show worse bone formation and higher bone resorption [47]. Added to the causes of bone structure disorders should be anti-cancer drugs like methotrexate, ifosfamide, cyclosporine, doxorubicin, cisplatin, and glucocorticosteroids, as well as radiotherapy, bone marrow transplantation, and decreased physical activity [47].

Children with cancer have a higher risk of developing nutritional disorders than healthy children. Children with solid tumors are believed to be at greater risk of malnutrition than children with leukemia, who are more likely to be overweight and obese. Both malnutrition and obesity have a negative impact on survival, the occurrence of treatment toxicity, and EFS. The nutritional status assessment should be carried out regularly during therapy, and the assessment methods should be adapted to the type of tumor and the child's age. During treatment, it is also necessary to remember and counteract the long-term effects of anti-cancer therapy, e.g., the occurrence of osteopenia and osteoporosis in cancer survivors, because this group has a significantly increased risk of their occurrence.

2. Regulation of Appetite in Children with Cancer

2.1. Appetite Regulation

The regulation of appetite in humans is a complex mechanism influenced by many factors [48]. In the CNS, the hypothalamus is a key area influencing the regulation of appetite [48]. The starvation center is in the lateral hypothalamic area (LHA), and the satiety center is in the ventromedial hypothalamus (VMH) [48][49]. They are influenced by neuropeptides and hormonal signals from tissues and organs [50]. The integration of circulating signals of hunger and satiety takes place in the arcuate nucleus (ARC), within which there are two opposing neuronal systems [50]. The first of them is the orexigenic system, which stimulates appetite through neuropeptide Y (NPY) and agouti-related peptide

(AgRP) [51]. The second one is the anorexigenic system suppressing appetite through a proopiomelanocortin (POMC) and the amphetamine-regulated transcript (CART) [51]. Then, signals are transmitted to the paraventricular hypothalamic nucleus (PVN), where they are integrated and modified [52]. The neurons of PVN send axons that secrete corticotropin-releasing hormone (CRH), thyrotropin-releasing hormone (TRH), and oxytocin (OXT) [52]. The arcuate nucleus also communicates with VMH, which secretes mainly anorexigenic brain-derived neurotrophic factor (BDNF) [52], and LHA, which secretes the appetite-stimulating peptides orexin-A (OxA), orexin-B (OxB), and melanin-concentrating hormone (MCH) [50].

Peripheral appetite regulators include gastrointestinal and adipose tissue hormones reaching the CNS via the bloodstream [49][53] but not all of them can cross the blood-brain barrier [54]. Gut peptides also affect the brain via vagal afferent fibers [50]. Leptin is a peptide produced mainly by white adipose tissue, and its concentration in the body positively correlates with the BMI and the amount of adipose tissue [55]. The action of leptin in the human body is multidirectional, but best known is its participation in the regulation of hunger and satiety, where it stimulates the POMC/CART system and inhibits the secretion of NPY [48][50]. Ghrelin, a gastrointestinal hormone produced in humans mainly in the stomach by type A enteroendocrine cells, has the opposite effect as leptin [56]. Stimulation of the starvation center is the main function of ghrelin, which negatively correlates with the BMI and concentration of leptin and insulin [49][53][57]. The lesser known functions of ghrelin are modulation of taste sensation, glucose metabolism, and gut motility [49]. Ghrelin level positively correlates with the severity of anorexia and cancer cachexia in adult patients [58][59][60]. Another important regulator is insulin, a long-term signal of satiety that can cross the blood-brain barrier [50]. Insulin stimulates leptin synthesis and inhibits NPY/AgRP neurons [49][50]. Glucagon-like peptide 1 (GLP-1) inhibits gastric emptying and reduces appetite, and stimulates the pancreas to secrete insulin [61]. This hormone has an important role in glucose metabolism [61]. Another peptide inhibiting food intake is peptide tyrosine-tyrosine (PYY), secreted in the distal intestine after a meal, especially after a protein-rich one [62]. Cholecystokinin (CCK) is synthesized mainly in the duodenum and jejunum, but also in the CNS [63]. It suppresses appetite and stimulates intestinal motility and secretion of insulin, glucagon, and pancreatic enzymes [64].

Moreover, genetic factors also play a role in regulating the level of intestinal hormones and thus the appetite. This was confirmed by Czogala et al. [65], who assessed the importance of FTO and PLAG1 gene expression in the context of gastrointestinal and adipose tissue hormone levels. The results indicate that the level of FTO and PLAG1 expression positively correlated with the concentration of leptin in the blood serum and negatively with CCK and GLP-1, while the expression and methylation of FTO negatively correlated with the levels of resistin and visfatin [65].

2.2. The Causes of Appetite and Nutritional Status Disorders in Children with Cancer

During cancer treatment, the action of neurohormones, gastrointestinal, and adipose tissue hormones also may change [66]. Only a few have shown gastrointestinal peptide dysfunction in children with cancer, and most have been conducted in children with leukemia. Fayh et al. [67], which showed a wide variation in results. Most of the included ones [67] looked at the concentration of leptin, but in only one research was a higher level of leptin observed in children with cancer [68], while the remaining one was found lower concentration or no difference compared with the control group [67]. Only two were looked at ghrelin concentration, and one of them indicated lower ghrelin levels in children with cancer, which increased in later stages of treatment [68]. As the causes of the varied results, the authors indicate different types of cancers, treatments, and ages of children [67]. Agyrou et al. [69] (2019) presented an overview of research on ghrelin, leptin, and adiponectin levels in children with ALL. They noticed that in most situations, the leptin level was higher and the adiponectin level was lower at diagnosis [69]. Furthermore, Carvalho Gomes CC et al. [70] assessed the levels of appetite-regulating hormones in children with ALL during the induction at three-time points. Statistically significant changes have been observed in the level of ghrelin, which positively correlated with food consumption [70]. The concentration of leptin, insulin, and cortisol did not change significantly during the 28 days [70]. Barbosa-Corte et al. [71] have shown that malnourished children with solid tumors and lymphomas have a lower leptin concentration than well-nourished children. Musial et al. [45] observed no statistically significant differences between leptin levels in children with CNS tumors and the control group and no correlation between leptin concentration and BMI. Statistically, an insignificant lower leptin level at diagnosis was observed in patients with brain tumors compared to the control group (7.04 vs. 16.38 ng/mL) and malnourished children [45]. Changes in gastrointestinal peptides have also been indicated by Skoczniński et al. [72], who observed that the concentration of CCK, ghrelin, and GLP-1 and the expression of their genes were significantly lower before bone transplant compared to 6 months after transplantation. Moreover, the concentrations of peptides in the test group were significantly lower than in the control group of healthy children [72]. The authors indicate that it may be caused by damage to the gastrointestinal mucosa, and the measurement of the concentrations of selected peptides may be a marker of gastrointestinal regeneration [72].

Appetite is also altered by disturbed gastrointestinal tract motility, taste, and smell disturbances, occurring at 45–84% for taste and 5–60% regarding the smell of adult cancer patients [73]. Cancer patients also exhibit increased protein catabolism and lipolysis, as well as inhibition of lipoprotein lipase [73][74]. The deterioration of the body condition leads to the development of cachexia characterized by unintentional weight loss, including muscle and fat tissue, marked weakness, dysfunctional immunity, decreased intestinal peristalsis, and abnormal functioning of the heart and other key organs and systems of the body [75].

Appetite regulation in humans is very complex and involves both CNS centers and peripheral factors. During oncological treatment, the action of hunger and satiety centers is disturbed by the action of proinflammatory cytokines, substances secreted by cancer, and metabolic disorders. In children with cancer, changes in the level of gastrointestinal peptides such as CCK, ghrelin, leptin, and GLP-1 are also observed. However, the results of individual ones are contradictory, and this issue requires further research.

3. The Role of Endocannabinoids System in Childhood Cancer

3.1. Physiology of ECS

ECS is a system of endogenous cannabinoids, receptors, enzymes, and transport proteins, discovered in the 90s [76][77]. It is found in mammals, other vertebrates, and some invertebrates [76][78]. In humans, it is already present in the embryo, while cannabinoid receptors in the brain are detected in the 14th week of fetal life [79]. Moreover, blocking CB1 receptors in mice in the first 24 h of life inhibited the suckling of milk [80]. Added to the best known and first discovered ECS ligands should be included anandamide (AEA) and 2-arachidoyl glycerol (2-AG) [81][82]. Both endocannabinoids are formed “on-demand” [83]. They are lipid derivatives of arachidonic acid (AA) belonging to omega-6 polyunsaturated fatty acids [76]. The precursor of AEA is N-acylphosphatidylethanolamine (NAPE), from which in the brain, kidneys, liver, lungs, spleen, and heart, anandamide and phosphatidic acid are formed using N-acyl phosphatidylethanolamine phospholipase D (NAPE-PLD) [76][78][84]. The 2-arachidonoyl glycerol is formed from diacylglycerol (DAG) using diacylglycerol lipase (DAGL) and phospholipase C [85][86][87][88]. Endocannabinoids have to leave the cell to fulfill their function, and due to their polar nature, the eCB membrane transporter must be involved. The termination of endocannabinoid signaling is intracellular [89]. AEA is mainly degraded by fatty acid amide hydrolase (FAAH) to ethanolamine and arachidonic acid, while 2-AG is hydrolyzed by monoacylglycerol lipase (MAGL) to arachidonic acid and glycerol [90]. It is worth noting that arachidonic acid formed due to hydrolysis is a substrate for the production of prostaglandins [76].

In the ECS, there are two main types of receptors—CB1 and CB2, which are G protein-coupled receptors (GPCRs) [77]. CB1 receptors are located primarily in the brain—most of them in the basal ganglia, cerebellum, hippocampus, and cortex [89]. They are also located in endocrine glands, thyroid and adrenal cells, ovaries, testes, uterus, and placenta, in the gastrointestinal tract, and in adipose tissue, where endocannabinoids activate lipoprotein lipase and fat deposition [91]. In addition, CB1 receptors are also found in the vagus nerve endings [91]. CB2 receptors are located mainly in cells of the immune system—on the surface of B lymphocytes, macrophages, monocytes, and NK cells. Moreover, they are also in the spleen, tonsils, and hematopoietic cells [92], and in the CNS they are located mainly in microglia [69]. Anandamide has a high affinity for the CB1 receptor and low affinity for CB2, while 2-AG can bind to both receptors [77].

3.2. The Role of the ECS in the Regulation of Appetite

The ECS in the human body works in a multidirectional way, and its role is still being investigated [76]. The best-known function of ECS is the regulation of energy balance and food consumption [76]. Other known functions of the ECS include pain control, thermogenesis, sleep cycle regulation, embryogenesis, neurogenesis, learning, and memory, as well as regulation of lipid and glucose metabolism [76][90][92].

In the appetite control process, CB1 receptors in appetite-regulating regions of the hypothalamus are involved, as well as CB1 receptors located in the limbic system, digestive tract, and adipose tissue [90]. It is worth noting that by the presence of these receptors in the limbic system, ECS takes part in the hedonic evaluation of food [49]. The ECS also decreases gastric acid secretion and gastrointestinal motility [93]. For an animal model confirms the role of endocannabinoids in regulating the energy balance. Mice with knockout CB1 receptors had lower body weight, were resistant to hyperphagia [94], and were insensitive to the action of leptin [95]. Activation of the CB1 receptors in OUN leads to increasing motivation for palatable foods and increasing odor sensitivity, which leads to a reduction of satiety feeling and increased food intake (**Figure 1**) [96][97]. ECS also interacts with gastrointestinal peptides. Activation of the ECS leads to ghrelin secretion, which increases appetite (**Figure 1**) [83]. This is a two-way action because the ECS system stimulates the secretion of ghrelin in the digestive system, while ghrelin stimulates the synthesis of 2-AG [97]. Furthermore, in CB1 knockout mice, ghrelin didn't show an anorexigenic effect [98]. It has also been known that leptin levels negatively correlate with endocannabinoid

concentration ^{[91][99]} (**Figure 1**). Activation of CB1 receptors also leads to an increase in insulin secretion, somatostatin, glucagon, and visfatin ^[91]. Furthermore, cholecystikinin reduces the expression of CB1 receptors ^[100].

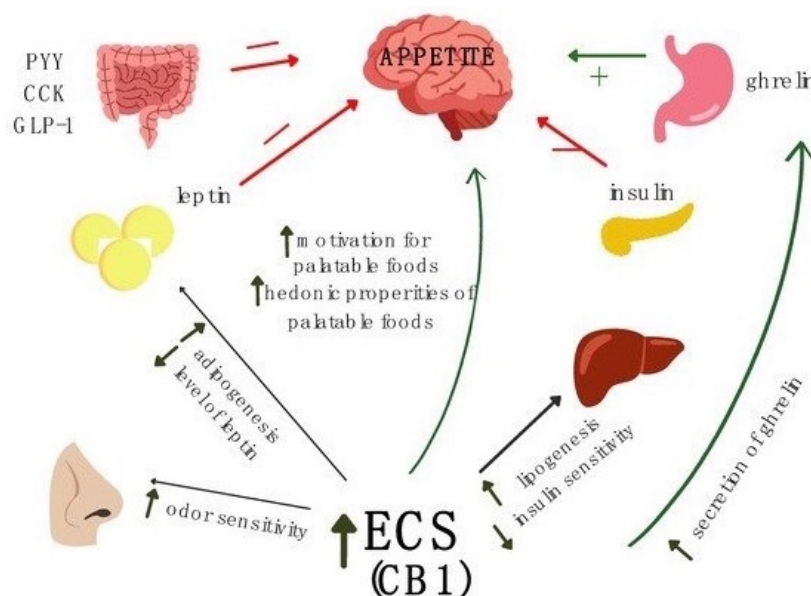


Figure 1. The role of endocannabinoid system (ECS) and gastrointestinal peptides in the regulation of appetite.

A high-fat and high-calorie diet can also modulate the ECS. Tagliamonte et al. ^[101] have shown that overweight and obese people have lower plasma AEA levels after switching from the Western to the Mediterranean diet, possibly due to increased intake of polyunsaturated fatty acids and decreased consumption of saturated fatty acids. In another study, Tagliamonte et al. ^[102] have shown that fat and energy intake can influence the concentration of endocannabinoids, NAE, and NAPE.

3.3. The Role of ECS in Childhood Cancer

The potential role of ECS in the development and course of diseases is still being investigated. It is currently known that in children it plays a role in the pathogenesis of such diseases as immune thrombocytopenia, juvenile idiopathic arthritis, type 2 diabetes, inflammatory bowel disease, celiac disease, obesity, and inflammation of the nervous system ^[103]. There is no data on the role of ECS in oncological diseases in children. Most are carried out among adults, where the anti-cancer properties of cannabinoids have been demonstrated in breast and pancreatic cancer, melanoma, lymphoma, and brain tumors ^[92]. Among children, there are single ones assessing the importance of the cannabinoid system and synthetic cannabinoids. Andradas ^[92] points out that most of the conducted ones concern acute lymphoblastic leukemia, which indicates that cannabinoids destroy cancer cells both in vivo and in vitro and that cannabinoids THC and CBD interact with vincristine, cytarabine, and doxorubicin in vitro ^{[104][105]}. It has also been shown that synthetic cannabinoids inhibit rhabdomyosarcoma growth ^[106] and reduce the viability and invasiveness of neuroblastoma cells ^[107]. Furthermore, synthetic cannabinoids induced cell cycle arrest of osteosarcoma cells ^[108]. In the case of brain tumors, it has been shown that in the group of children with low-grade gliomas, the level of CB1 expression was a predictor of spontaneous involution ^[109].

The role of ECS in children's oncology is still little known. In vitro ones were indicated that the anti-cancer effects of cannabinoids on leukemia, neuroblastoma, rhabdomyosarcoma, and osteosarcoma cells. Furthermore, cannabinoids can enhance the toxic effects of drugs. Another interesting issue is the interaction of endocannabinoids with gastrointestinal peptides. Endocannabinoids correlate positively with ghrelin secretion and negatively with leptin secretion. This topic also requires future research, especially in children with cancer.

References

1. Barr, R.D.; Stevens, M.C.G. The influence of nutrition on clinical outcomes in children with cancer. *Pediatr. Blood Cancer* 2020, 67, e28117.
2. Viani, K.; Trehan, A.; Manzoli, B.; Schoeman, J. Assessment of nutritional status in children with cancer: A narrative review. *Pediatr. Blood Cancer* 2020, 67, e28211.

3. Rogers, P.C. Nutritional status as a prognostic indicator for pediatric malignancies. *J. Clin. Oncol.* 2014, 32, 1293–1294.
4. Brinksma, A.; Huizinga, G.; Sulkers, E.; Kamps, W.; Roodbol, P.; Tissing, W. Malnutrition in childhood cancer patients: A review on its prevalence and possible causes. *Crit. Rev. Oncol. Hematol.* 2012, 83, 249–275.
5. Triarico, S.; Rinninella, E.; Cintoni, M.; Capozza, M.A.; Mastrangelo, S.; Mele, M.C.; Ruggiero, A. Impact of malnutrition on survival and infections among pediatric patients with cancer: A retrospective study. *Eur. Rev. Med. Pharmacol. Sci.* 2019, 23, 1165–1175.
6. Diakatou, V.; Vassilakou, T. Nutritional Status of Pediatric Cancer Patients at Diagnosis and Correlations with Treatment, Clinical Outcome and the Long-Term Growth and Health of Survivors. *Children* 2020, 7, 218.
7. Iniesta, R.R.; Paciarotti, I.; Davidson, I.; McKenzie, J.M.; Brougham, M.F.H.; Wilson, D.C. Nutritional status of children and adolescents with cancer in Scotland: A prospective cohort study. *Clin. Nutr. ESPEN* 2019, 32, 96–106.
8. Pribnow, A.K.; Ortiz, R.; Báez, L.F.; Mendieta, L.; Luna-Fineman, S. Effects of Malnutrition on Treatment-Related Morbidity and Survival of Children with Cancer in Nicaragua. *Pediatr. Blood Cancer* 2017, 64, e26590.
9. Iniesta, R.R.; Paciarotti, I.; Brougham, M.F.; McKenzie, J.M.; Wilson, D.C. Effects of pediatric cancer and its treatment on nutritional status: A systematic review. *Nutr. Rev.* 2015, 73, 276–295.
10. Brinksma, A.; Roodbol, P.F.; Sulkers, E.; Kamps, W.A.; de Bont, E.S.; Boot, A.M.; Burgerhof, J.G.; Tamminga, R.Y.; Tissing, W.J. Changes in nutritional status in childhood cancer patients: A prospective cohort study. *Clin. Nutr.* 2015, 34, 66–73.
11. Paciarotti, I.; McKenzie, J.M.; Davidson, I. Short term effects of childhood cancer and its treatments on nutritional status: A prospective cohort study. *EC Nutr.* 2015, 3, 528–540.
12. World Health Organization. CureAll Framework: WHO Global Initiative for Childhood Cancer: Increasing Access, Advancing Quality, Saving Lives. License: CC BY-NC-SA 3.0 IGO. 2021. Available online: <https://apps.who.int/iris/handle/10665/347370> (accessed on 11 January 2021).
13. Lange, B.J.; Gerbing, R.B.; Feusner, J.; Skolnik, J.; Sacks, N.; Smith, F.O.; Alonzo, T.A. Mortality in overweight and underweight children with acute myeloid leukemia. *JAMA* 2005, 293, 203–211.
14. Orgel, E.; Genkinger, J.M.; Aggarwal, D.; Sung, L.; Nieder, M.; Ladas, E.J. Association of body mass index and survival in pediatric leukemia: A meta-analysis. *Am. J. Clin. Nutr.* 2016, 103, 808–817.
15. Saenz, A.M.; Stapleton, S.; Hernandez, R.G.; Hale, G.A.; Goldenberg, N.A.; Schwartz, S.; Amankwah, E.K. Body Mass Index at Pediatric Leukemia Diagnosis and the Risks of Relapse and Mortality: Findings from a Single Institution and Meta-Analysis. *J. Obes.* 2018, 2018, 7048078.
16. Amankwah, E.K.; Saenz, A.M.; Hale, G.A.; Brown, P.A. Association between body mass index at diagnosis and pediatric leukemia mortality and relapse: A systematic review and meta-analysis. *Leuk. Lymphoma* 2016, 57, 1140–1148.
17. Butturini, A.M.; Dorey, F.J.; Lange, B.J.; Henry, D.W.; Gaynon, P.S.; Fu, C.; Franklin, J.; Siegel, S.E.; Seibel, N.L.; Rogers, P.C.; et al. Obesity and Outcome in Pediatric Acute Lymphoblastic Leukemia. *J. Clin. Oncol.* 2007, 20, 2063–2069.
18. Orgel, E.; Sposto, R.; Malvar, J.; Seibel, N.L.; Ladas, E.; Gaynon, P.S.; Freyer, D.R. Impact on survival and toxicity by duration of weight extremes during treatment for pediatric acute lymphoblastic leukemia: A report from the Children's Oncology Group. *J. Clin. Oncol.* 2014, 1, 1331–1337.
19. Orgel, E.; Tucci, J.; Alhushki, W.; Malvar, J.; Sposto, R.; Fu, C.H.; Freyer, D.R.; Abdel-Azim, H.; Mittelman, S.D. Obesity is associated with residual leukemia following induction therapy for childhood B-precursor acute lymphoblastic leukemia. *Blood* 2014, 124, 3932–3938.
20. Hijiya, N.; Panetta, J.C.; Zhou, Y.; Kyzer, E.P.; Howard, S.C.; Jeha, S.; Razzouk, B.I.; Ribeiro, R.C.; Rubnitz, J.E.; Hudson, M.M.; et al. Body mass index does not influence pharmacokinetics or outcome of treatment in children with acute lymphoblastic leukemia. *Blood* 2006, 108, 3997–4002.
21. Browne, E.K.; Jeha, S.; Cheng, C.; Relling, M.V.; Campana, D.; Pui, C.H.; Inaba, H. The effect of body mass index at diagnosis on clinical outcome in children with newly diagnosed acute lymphoblastic leukemia. *Blood Cancer J.* 2017, 7, e531.
22. Aldhafiri, F.K.; McColl, J.H.; Reilly, J.J. Prognostic significance of being overweight and obese at diagnosis in children with acute lymphoblastic leukemia. *J. Pediatr. Hematol. Oncol.* 2014, 36, 234–236.
23. Jaime-Pérez, J.C.; Turrubiates-Hernández, G.A.; García-Salas, G.; de la Torre-Salinas, A.M.; Áncer-Rodríguez, P.; Villarreal-Martínez, L.; Gómez-Almaguer, D. The Influence of Nutritional Status at Diagnosis of Childhood B-Cell Acute

24. Suzuki, D.; Kobayashi, R.; Sano, H.; Hori, D.; Kobayash, K. Sarcopenia after induction therapy in childhood acute lymphoblastic leukemia: Its clinical significance. *Int. J. Hematol.* 2018, 107, 486–489.
25. Tah, P.C.; Nik Shanita, S.; Pohm, B.K. Nutritional status among pediatric cancer patients: A comparison between hematological malignancies and solid tumors. *J. Spec. Pediatr. Nurs.* 2012, 17, 301–311.
26. Yoruk, M.A.; Durakbasa, C.U.; Timur, C.; Sahin, S.S.; Taskin, E.C. Assessment of Nutritional Status and Malnutrition Risk at Diagnosis and Over a 6-Month Treatment Period in Pediatric Oncology Patients with Hematologic Malignancies and Solid Tumors. *J. Pediatr. Hematol. Oncol.* 2018, 41, e308–e321.
27. Viani, K.; Barr, R.D.; Filho, V.O.; Ladas, E.J. Nutritional status at diagnosis among children with cancer referred to a nutritional service in Brazil. *Hematol. Transfus. Cell Ther.* 2021, 43, 389–395.
28. Garófolo, A.; Lopez, F.A.; Petrilli, A.S. High prevalence of malnutrition among patients with solid non-hematological tumors as found by using skinfold and circumference measurements. *Sao Paulo Med. J.* 2005, 123, 277–281.
29. Dos Lemos, P.S.M.; de Oliveira, F.L.C.; Caran, E.M.M. Nutritional Status of Children and Adolescents at Diagnosis of Hematological and Solid Malignancies. *Rev. Bras. Hematol. Hemoter.* 2014, 36, 420–423.
30. Radhakrishnan, V.; Ganesan, P.; Rajendranath, R.; Ganesan, T.S.; Sagar, T.G. Nutritional Profile of Pediatric Cancer Patients at Cancer Institute, Chennai. *Indian J. Cancer* 2015, 52, 207.
31. Joffe, L.; Dwyer, S.; Glade, B.J.L.; Frazier, A.L.; Ladas, E.J. Nutritional status and clinical outcomes in pediatric patients with solid tumors: A systematic review of the literature. *Semin. Oncol.* 2019, 46, 48–56.
32. Goldstein, G.; Shemesh, E.; Frenkel, T.; Jacobson, J.M.; Toren, A. Abnormal body mass index at diagnosis in patients with Ewing sarcoma is associated with inferior tumor necrosis. *Pediatr. Blood Cancer* 2015, 62, 1892–1896.
33. Sharib, J.M.; Cyrus, J.; Horvai, A.; Gray, H.F.K.; Neuhaus, J.; Matthay, K.K.; Goldsby, R.; Marina, N.; DuBois, S.G. Predictors of acute chemotherapy-associated toxicity in patients with Ewing sarcoma. *Pediatr. Blood Cancer* 2012, 59, 611–616.
34. Altaf, S.; Enders, F.; Jeavons, E.; Krailo, M.; Barkauskas, D.A.; Meyers, P.; Arndt, C. High-BMI at diagnosis is associated with inferior survival in patients with osteosarcoma: A report from the Children's Oncology Group. *Pediatr. Blood Cancer* 2013, 60, 2042–2046.
35. Hingorani, P.; Seidel, K.; Krailo, M.; Mascarenhas, L.; Meyers, P.; Marina, N.; Conrad, E.U.; Hawkins, D.S. Body mass index (BMI) at diagnosis is associated with surgical wound complications in patients with localized osteosarcoma: A report from the Children's Oncology Group. *Pediatr. Blood Cancer* 2011, 57, 939–942.
36. Tenardi, R.D.; Frühwald, M.C.; Jürgens, H.; Herttroijs, D.; Bauer, J. Nutritional status of children and young adults with Ewing sarcoma or osteosarcoma at diagnosis and during multimodality therapy. *Pediatr. Blood Cancer* 2012, 59, 621–626.
37. Burke, M.E.; Lyden, E.R.; Meza, J.L.; Ladas, E.J.; Dasgupta, R.; Wiegner, E.A.; Arndt, C.A. Does body mass index at diagnosis or weight change during therapy predict toxicity or survival in intermediate risk rhabdomyosarcoma? A report from the Children's Oncology Group Soft Tissue Sarcoma Committee. *Pediatr. Blood Cancer* 2013, 60, 748–753.
38. Lifson, L.F.; Hadley, G.P.; Wiles, N.L.; Pillay, K. Nutritional status of children with Wilms' tumour on admission to a South African hospital and its influence on outcome. *Pediatr. Blood Cancer* 2017, 64, e26382.
39. Fernandez, C.V.; Anderson, J.; Breslow, N.E.; Dome, J.S.; Grundy, P.E.; Perlman, E.J.; Green, D.M. Anthropomorphic measurements and event-free survival in patients with favorable histology Wilms tumor: A report from the Children's Oncology Group. *Pediatr. Blood Cancer* 2009, 52, 254–258.
40. Small, A.G.; Thwe, L.M.; Byrne, J.A.; Lau, L.; Chan, A.; Craig, M.E.; Cowell, C.T.; Garnett, S.P. Neuroblastoma, body mass index, and survival: A retrospective analysis. *Medicine* 2015, 94, e713.
41. Aarnivala, H.; Pokka, T.; Soininen, R.; Möttönen, M.; Harila-Saari, A.; Niinimäki, R. Trends in age- and sex-adjusted body mass index and the prevalence of malnutrition in children with cancer over 42 months after diagnosis: A single-center cohort study. *Eur. J. Pediatr.* 2020, 179, 91–98.
42. Tazi, I.; Hidane, Z.; Zafad, S.; Harif, M.; Benchekroun, S.; Ribeiro, R. Nutritional status at diagnosis of children with malignancies in Casablanca. *Pediatr. Blood Cancer* 2008, 51, 495–498.
43. Joffe, L.; Shen, W.; Shadid, G.; Jin, Z.; Ladas, E.J. Skeletal muscle and adipose tissue changes in the first phase of treatment of pediatric solid tumors. *Cancer Med.* 2021, 10, 15–22.
44. Tsutsumi, R.C.; Speridião, P.G.L. Children with brain tumors: A study of nutritional status on hospital admission. *Semin. Ciências Biol. Saúde* 2021, 42, 51.

45. Musiol, K.; Sobol, G.; Mizia-Malarz, A.; Wos, H. Leptin concentration and nutritional status in the course of treatment in children with brain tumours—Preliminary report. *Childs Nerv. Syst.* 2014, 30, 131–136.
46. Barr, R.D.; Ladas, E.J. The role of nutrition in pediatric oncology. *Expert Rev. Anticancer Ther.* 2020, 20, 109–116.
47. Marcucci, G.; Beltrami, G.; Tamburini, A.; Body, J.J.; Confavreux, C.B.; Hadji, P.; Holzer, G.; Kendler, D.; Napoli, N.; Pierroz, D.D.; et al. Bone health in childhood cancer: Review of the literature and recommendations for the management of bone health in childhood cancer survivors. *Ann. Oncol.* 2019, 30, 908–920.
48. Perry, B.; Wang, Y. Appetite regulation and weight control: The role of gut hormones. *Nutr. Diabetes* 2012, 2, e26.
49. Woodward, O.R.M.; Gribble, F.M.; Reimann, F.; Lewis, J.E. Gut peptide regulation of food intake—Evidence for the modulation of hedonic feeding. *J. Physiol.* 2022, 600, 1053–1078.
50. Hariyanto, T.I.; Kurniawan, A. Appetite problem in cancer patients: Pathophysiology, diagnosis, and treatment. *Cancer Treat. Res. Commun.* 2021, 27, 100336.
51. Minor, R.K.; Chang, J.W.; de Cabo, R. Hungry for life: How the arcuate nucleus and neuropeptide Y may play a critical role in mediating the benefits of calorie restriction. *Mol. Cell. Endocrinol.* 2009, 299, 79–88.
52. Rios, M. BDNF and the central control of feeding: Accidental bystander or essential player? *Trends Neurosci.* 2013, 36, 83–90.
53. Austin, J.; Marks, D. Hormonal regulators of appetite. *Int. J. Pediatr. Endocrinol.* 2009, 2009, 141753.
54. Camilleri, M. Peripheral mechanisms in appetite regulation. *Gastroenterology* 2015, 148, 1219–1233.
55. Suzuki, K.; Simpson, K.A.; Minnion, J.S.; Shillito, J.C.; Bloom, S.R. The role of gut hormones and the hypothalamus in appetite regulation. *Endocr. J.* 2010, 57, 359–372.
56. Date, Y.; Kojima, M.; Hosoda, H.; Sawaguchi, A.; Mondal, M.S.; Suganuma, T.; Matsukura, S.; Kangawa, K.; Nakazato, M. Ghrelin, a novel growth hormone-releasing acylated peptide, is synthesized in a distinct endocrine cell type in the gastrointestinal tracts of rats and humans. *Endocrinology* 2000, 141, 4255–4261.
57. Sovetkina, A.; Nadir, R.; Fung, J.N.M.; Nadjarpour, A.; Beddoe, B. The Physiological Role of Ghrelin in the Regulation of Energy and Glucose Homeostasis. *Cureus* 2020, 12, e7941.
58. Davis, M.P.; Dreicer, R.; Walsh, D.; Lagman, R.; LeGrand, S.B. Appetite and cancer-associated anorexia: A review. *J. Clin. Oncol.* 2004, 22, 1510–1517.
59. Wolf, I.; Sadetzki, S.; Kanety, H.; Kundel, Y.; Pariente, C.; Epstein, N.; Oberman, B.; Catane, R.; Kaufman, B.; Shimon, I. Adiponectin, ghrelin, and leptin in cancer cachexia in breast and colon cancer patients. *Cancer* 2006, 106, 966–973.
60. Kerem, M.; Ferahkose, Z.; Yilmaz, U.T.; Pasaoglu, H.; Ofluoglu, E.; Bedirli, A.; Salman, B.; Sahin, T.T.; Akin, M. Adipokines and ghrelin in gastric cancer cachexia. *World J Gastroenterol.* 2008, 14, 3633–3641.
61. Salehi, M.; Purnell, J.Q. The Role of Glucagon-Like Peptide-1 in Energy Homeostasis. *Metab. Syndr. Relat. Disord.* 2019, 17, 183–191.
62. Price, S.L.; Bloom, S.R. Protein PYY and its role in metabolism. *Front. Horm. Res.* 2014, 42, 147–154.
63. Little, T.J.; Horowitz, M.; Feinle-Bisset, C. Role of cholecystokinin in appetite control and body weight regulation. *Obes. Rev.* 2005, 6, 297–306.
64. Rehfeld, J.F. Cholecystokinin-From Local Gut Hormone to Ubiquitous Messenger. *Front. Endocrinol.* 2017, 8, 47.
65. Czogała, W.; Strojny, W.; Schab, M.; Grabowska, A.; Miklusiak, K.; Kowalczyk, W.; Łazarczyk, A.; Tomasik, P.; Skoczeń, S. FTO and PLAG1 Genes Expression and FTO Methylation Predict Changes in Circulating Levels of Adipokines and Gastrointestinal Peptides in Children. *Nutrients* 2021, 13, 3585.
66. Ezeoke, C.C.; Morley, J.E. Pathophysiology of anorexia in the cancer cachexia syndrome. *J. Cachexia Sarcopenia Muscle* 2015, 6, 287–302.
67. Fayh, A.P.T.; de Lima Bezerra, A.D.; Friedman, R. Appetite hormones in children and adolescents with cancer: A systematic review of observational studies. *Nutr. Hosp.* 2018, 35, 201–210.
68. Park, S.H.; Jung, M.H.; Chung, N.G.; Suh, B.K.; Lee, B.C. Serum ghrelin and leptin concentrations in children with cancer: Comparisons with normal children. *Korean J. Pediatr.* 2007, 50, 90511.
69. Argyrou, C.; Hatziagapiou, K.; Theodorakidou, M.; Nikola, O.A.; Vlahopoulos, S.; Lambrou, G.I. The role of adiponectin, LEPTIN, and ghrelin in the progress and prognosis of childhood acute lymphoblastic leukemia. *Leuk. Lymphoma* 2019, 60, 2158–2169.
70. Gomes, C.C.; Silva, C.C.G.D.; Nascimento, P.R.P.D.; Lemos, T.M.A.M.; Marcadenti, A.; Markoski, M.M.; Fayh, A.P.T. Nutritional status and appetite-regulating hormones in early treatment of acute lymphoblastic leukemia among children

and adolescents: A cohort study. *Sao Paulo Med. J.* 2020, 138, 118–125.

71. Barbosa-Cortés, L.; Klunder-Klunder, M.; López-Alarcón, M.; Márquez, H.R.; López-Aguilar, E.; Tapia-Marcial, A. Nutritional status and cytokine concentration during chemotherapy in Mexican children: A longitudinal analysis. *Nutrition* 2019, 57, 46–51.
72. Skoczko, S.; Rej, M.; Kwiecińska, K.; Pietrys, D.; Tomasik, P.J.; Wójcik, M.; Strojny, W.; Dłużniewska, A.; Klimasz, K.; Fijorek, K.; et al. Gastrointestinal peptides in children before and after hematopoietic stem cell transplantation. *BMC Cancer* 2020, 20, 306.
73. Co-Reyes, E.; Li, R.; Huh, W.; Chandra, J. Malnutrition and obesity in pediatric oncology patients: Causes, consequences, and interventions. *Pediatr. Blood Cancer* 2012, 15, 1160–1167.
74. Suzuki, H.; Asakawa, A.; Amitani, H.; Nakamura, N.; Inui, A. Cancer cachexia—Pathophysiology and management. *J. Gastroenterol.* 2013, 48, 574–594.
75. Skipworth, R.J.; Stewart, G.D.; Dejong, C.H.; Preston, T.; Fearon, K.C. Pathophysiology of cancer cachexia: Much more than host-tumour interaction? *Clin. Nutr.* 2007, 26, 667–676.
76. Lu, H.C.; Mackie, K. Review of the Endocannabinoid System. *Biol. Psychiatry Cogn. Neurosci. Neuroimaging* 2021, 6, 607–615.
77. Watkins, B.A. Diet, endocannabinoids, and health. *Nutr. Res.* 2019, 70, 32–39.
78. Iannotti, F.A.; Di Marzo, V. The gut microbiome, endocannabinoids and metabolic disorders. *J. Endocrinol.* 2021, 248, R83–R97.
79. Rutkowska, M.; Jamonit, J. Involvement of the Cannabinoid System in the Regulation of Food Intake. *Adv. Clin. Exp. Med.* 2005, 14, 1011–1017.
80. Frider, E.; Foox, A.; Rosenberg, E.; Faigenboim, M.; Cohen, V.; Barda, L.; Blau, H.; Mechoulam, R. Milk intake and survival in newborn cannabinoid CB1 receptor knockout mice: Evidence for a “CB3” receptor. *Eur. J. Pharmacol.* 2003, 461, 27–34.
81. Devane, W.A.; Hanus, L.; Breuer, A.; Pertwee, R.G.; Stevenson, L.A.; Griffin, G.; Gibson, D.; Mandelbaum, A.; Etinger, A.; Mechoulam, R. Isolation and structure of a brain constituent that binds to the cannabinoid receptor. *Science* 1992, 258, 1946–1949.
82. Mechoulam, R.; Ben-Shabat, S.; Hanus, L.; Ligumsky, M.; Kaminski, N.E.; Schatz, A.R.; Gopher, A.; Almog, S.; Martin, B.R.; Compton, D.R. Identification of an endogenous 2-monoglyceride, present in canine gut, that binds to cannabinoid receptors. *Biochem. Pharmacol.* 1995, 50, 83–90.
83. Horn, H.; Böhme, B.; Dietrich, L.; Koch, M. Endocannabinoids in Body Weight Control. *Pharmaceuticals* 2018, 11, 55.
84. Rahaman, O.; Ganguly, D. Endocannabinoids in immune regulation and immunopathologies. *Immunology* 2021, 164, 242–252.
85. Meccariello, R.; Santoro, A.; D'Angelo, S.; Morrone, R.; Fasano, S.; Viggiano, A.; Pierantoni, R. The Epigenetics of the Endocannabinoid System. *Int. J. Mol. Sci.* 2020, 21, 1113.
86. Zou, S.; Kumar, U. Cannabinoid Receptors and the Endocannabinoid System: Signaling and Function in the Central Nervous System. *Int. J. Mol. Sci.* 2018, 19, 833.
87. Tarragon, E.; Moreno, J.J. Cannabinoids, Chemical Senses, and Regulation of Feeding Behavior. *Chem. Senses* 2019, 44, 73–89.
88. Brunt, T.M.; Bossong, M.G. The neuropharmacology of cannabinoid receptor ligands in central signaling pathways. *Eur. J. Neurosci.* 2022, 55, 909–921.
89. Schulz, P.; Hryhorowicz, S.; Rychter, A.M.; Zawada, A.; Słomski, R.; Dobrowolska, A.; Krela-Kaźmierczak, I. What Role Does the Endocannabinoid System Play in the Pathogenesis of Obesity? *Nutrients* 2021, 13, 373.
90. Blankman, J.L.; Cravatt, B.F. Chemical probes of endocannabinoid metabolism. *Pharmacol. Rev.* 2013, 65, 849–871.
91. Borowska, M.; Czarnywojtek, A.; Sawicka-Gutaj, N.; Woliński, K.; Płazińska, M.T.; Mikołajczak, P.; Ruchała, M. The effects of cannabinoids on the endocrine system. *Endokrynol. Pol.* 2018, 69, 705–719.
92. Andradas, C.; Truong, A.; Byrne, J.; Endersby, R. The Role of Cannabinoids as Anticancer Agents in Pediatric Oncology. *Cancers* 2021, 13, 157.
93. Izzo, A.A.; Sharkey, K.A. Cannabinoids and the gut: New developments and emerging concepts. *Pharmacol. Ther.* 2010, 126, 21–38.
94. Ravinet, T.C.; Delgorge, C.; Menet, C.; Arnone, M.; Soubrié, P. CB1 cannabinoid receptor knockout in mice leads to leanness, resistance to diet-induced obesity and enhanced leptin sensitivity. *Int. J. Obes. Relat. Metab. Disord.* 2004,

95. Cardinal, P.; Bellocchio, L.; Clark, S.; Cannich, A.; Klugmann, M.; Lutz, B.; Marsicano, G.; Cota, D. Hypothalamic CB1 cannabinoid receptors regulate energy balance in mice. *Endocrinology* 2012, 153, 4136–4143.
96. Abdalla, M.M. Central and peripheral control of food intake. *Endocr. Regul.* 2017, 51, 52–70.
97. Gatta-Cherifi, B.; Cota, D. New insights on the role of the endocannabinoid system in the regulation of energy balance. *Int. J. Obes.* 2016, 40, 210–219.
98. Kola, B.; Farkas, I.; Christ-Crain, M.; Wittmann, G.; Lolli, F.; Amin, F.; Harvey-White, J.; Liposits, Z.; Kunos, G.; Grossman, A.B.; et al. The orexigenic effect of ghrelin is mediated through central activation of the endogenous cannabinoid system. *PLoS ONE* 2008, 3, e1797.
99. Di Marzo, V.; Goparaju, S.K.; Wang, L.; Liu, J.; Bátkai, S.; Járαι, Z.; Fezza, F.; Miura, G.I.; Palmiter, R.D.; Sugiura, T.; et al. Leptin-regulated endocannabinoids are involved in maintaining food intake. *Nature* 2001, 410, 822–825.
100. Komorowski, J.; Stepień, H. The role of the endocannabinoid system in the regulation of endocrine function and in the control of energy balance in humans. *Postepy Hig. Med. Dosw.* 2007, 61, 99–105.
101. Tagliamonte, S.; Laiola, M.; Ferracane, R.; Vitale, M.; Gallo, M.A.; Meslier, V.; Pons, N.; Ercolini, D.; Vitaglione, P. Mediterranean diet consumption affects the endocannabinoid system in overweight and obese subjects: Possible links with gut microbiome, insulin resistance and inflammation. *Eur. J. Nutr.* 2021, 60, 3703–3716.
102. Tagliamonte, S.; Gill, C.I.R.; Pourshahidi, L.K.; Slevin, M.M.; Price, R.K.; Ferracane, R.; Lawther, R.; O'Connor, G.; Vitaglione, P. Endocannabinoids, endocannabinoid-like molecules and their precursors in human small intestinal lumen and plasma: Does diet affect them? *Eur. J. Nutr.* 2021, 60, 2203–2215.
103. Argenziano, M.; Tortora, C.; Bellini, G.; Di Paola, A.; Punzo, F.; Rossi, F. The Endocannabinoid System in Pediatric Inflammatory and Immune Diseases. *Int. J. Mol. Sci.* 2019, 20, 5875.
104. Scott, K.A.; Dagleish, A.G.; Liu, W.M. Anticancer effects of phytocannabinoids used with chemotherapy in leukaemia cells can be improved by altering the sequence of their administration. *Int. J. Oncol.* 2017, 51, 369–377.
105. Liu, W.M.; Scott, K.A.; Shamash, J.; Joel, S.; Powles, T.B. Enhancing the in vitro cytotoxic activity of Delta9-tetrahydrocannabinol in leukemic cells through a combinatorial approach. *Leuk. Lymphoma* 2008, 49, 1800–1809.
106. Oesch, S.; Walter, D.; Wachtel, M.; Pretre, K.; Salazar, M.; Guzman, M.; Velasco, G.; Schafer, B.W. Cannabinoid receptor 1 is a potential drug target for treatment of translocation-positive rhabdomyosarcoma. *Mol. Cancer Ther.* 2009, 8, 1838–1845.
107. Fisher, T.; Golan, H.; Schiby, G.; PriChen, S.; Smoum, R.; Moshe, I.; Peshes-Yaloz, N.; Castiel, A.; Waldman, D.; Gallily, R.; et al. In vitro and in vivo efficacy of non-psychoactive cannabidiol in neuroblastoma. *Curr. Oncol.* 2016, 23, S15–S22.
108. Notaro, A.; Sabella, S.; Pellerito, O.; Di Fiore, R.; De Blasio, A.; Vento, R.; Calvaruso, G.; Giuliano, M. Involvement of PAR-4 in cannabinoid-dependent sensitization of osteosarcoma cells to TRAIL-induced apoptosis. *Int. J. Biol. Sci.* 2014, 10, 466–478.
109. Sredni, S.T.; Huang, C.C.; Suzuki, M.; Pundy, T.; Chou, P.; Tomita, T. Spontaneous involution of pediatric low-grade gliomas: High expression of cannabinoid receptor 1 (CNR1) at the time of diagnosis may indicate involvement of the endocannabinoid system. *Childs Nerv. Syst.* 2016, 32, 2061–2067.