

Biosensors for the Detection of L-Amino Acids

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The field of biosensors is filled with reports and designs of various sensors, with the vast majority focusing on glucose sensing. However, in addition to glucose, there are many other important analytes that are worth investigating as well. In particular, L-amino acids appear as important diagnostic markers for a number of conditions. The need to determine L-amino acids from clinical samples has risen. More clinical data appear to demonstrate that abnormal concentrations of L-amino acids are related to various clinical conditions such as inherited metabolic disorders, dyslipidemia, type 2 diabetes, muscle damage, etc.

amino acids

detection

biosensors

diagnostics

clinical condition

disease

1. Typical Concentrations of Total L-Amino Acids in Human Blood

Research articles indicate that human blood contains various L-amino acids. In an early article by Stein and Moore, human blood plasma was investigated and twenty-eight compounds were found to be ninhydrin-positive and identifiable as L-amino acids ^[1]. Quantitative analysis revealed that the L-amino acids present in human blood at the highest concentrations are glutamine, alanine, valine, proline, and lysine. The average concentration of total L-amino acids was determined to be around 37 mg per 100 mL of plasma (or ~3.3 mmol/L). However, the described study investigated a small sample size; in total, plasma from only five individuals was investigated. A more recent study by Schmidt et al. investigated the typical levels of L-amino acids in the blood of various groups based on diet preferences, i.e., meat-eaters, fish-eaters, vegetarians, and vegans. Significant sample sizes (~100) were reached ^[2]. The results for particular L-amino acids were different between the groups; however, the concentrations of total L-amino acids were similar in the range of 4.1–5.2 mmol/L.

2. Overview of L-Amino Acids in Clinical Conditions

The importance of L-amino acids for clinical diagnostics is not sufficiently clear and is often overlooked. Several factors are contributing to this issue, such as the relationship of L-amino acids to many biochemical processes, and, in turn, to many various conditions from metabolic disorders to cancers, giving them lower specificity. Additionally, to this date, the measurement methods and/or devices for L-amino acids are quite expensive and usually they are not routinely applied for clinical sample analysis. The most common clinical conditions related to L-amino acids are discussed in detail and summarized in **Figure 1**.

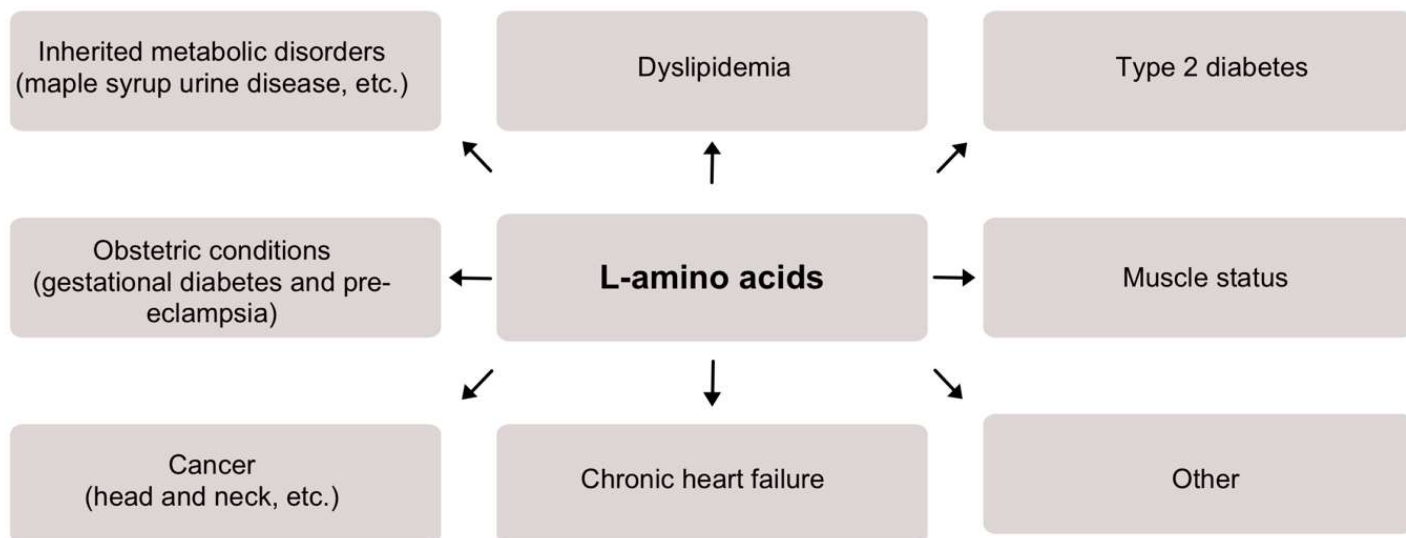


Figure 1. A scheme demonstrating the significance of L-amino acids detection and/or monitoring for various medical conditions.

2.1. Inherited Metabolic Disorders

Inherited metabolic disorders, such as maple syrup urine disease, are closely related to serum L-amino acids both for diagnostic and therapeutic purposes [3][4]. Maple syrup urine disease has been shown to result in an increase in branched-chain L-amino acids (BCAAs) in plasma [5]. Long-term treatment requires restriction of dietary BCAAs and supplementation with a BCAA-free mixture. Frequent monitoring of the plasma concentration of L-amino acids could help identify deficiencies in essential L-amino acids secondary to a protein-restricted diet, and thus guide treatment.

2.2. Dyslipidemia

BCAA, tyrosine, tryptophan, glutamate, and L-homocysteine concentrations are reported to be considerably lower in healthy diet individuals [6]. In one study, forty healthy men and women with a body mass index of 25.6 ± 0.6 were overfed by 1250 kCal per day for 28 days and, as a consequence, their serum BCAAs increased from $397 \pm 10 \mu\text{mol/L}$ to $428 \pm 9 \mu\text{mol/L}$. This highlights the role of food overconsumption in increased BCAA concentrations [7]. Moreover, BCAAs and total L-amino acid levels are increased in obese individuals. Higher concentrations of all BCAAs, phenylalanine, tyrosine, and alanine are associated with a higher fat mass index, and some plasma-free L-amino acids are correlated with parameters used to measure central obesity (e.g., waist circumference, waist-to-hip ratio, BMI) [8][9]. Interestingly, medical studies report that serum amino acid profiles and increased levels of leucine, arginine, valine, proline, phenylalanine, isoleucine, and lysine could predict the development of hypertriglyceridemia later in life in healthy individuals even before they become obese [10]. For example, serum L-amino acid concentrations of 1125 individuals were analyzed with the follow-up data available after 7 years for comparison and a relationship was found between L-amino acids and an increased risk of developing hypertriglyceridemia [11]. In summary, measurements of L-amino acid concentrations can serve as early biomarkers to identify people at high risk of developing dyslipidemia and diseases related to dyslipidemia.

2.3. Type 2 Diabetes

Increased levels of L-amino acids have been reported to be an early manifestation of insulin resistance; i.e., changes in blood concentrations of particular L-amino acids (e.g., sulfur amino acids, tyrosine, phenylalanine) are apparent with obesity and insulin resistance, often before the onset of clinical diagnosis [12][13]. Furthermore, BCAAs and aromatic L-amino acids predict the risk of developing type 2 diabetes. For example, during a study conducted by Wang et al., 2422 normoglycemic individuals were profiled for metabolites. After 12 years, 201 individuals were diagnosed with type 2 diabetes. A significant correlation between the disease and several amino acids, i.e., isoleucine, leucine, valine, tyrosine, and phenylalanine, was established. The risk of future diabetes was concluded to be at least fourfold greater in those with high plasma amino acid concentrations in both the discovery and replication samples [14]. Similar results were obtained from the Southall And Brent REvisited (SABRE) study. Asian-American individuals have been reported to have an increased risk of diabetes compared to Europeans that is not related to obesity or established traditional metabolic measures [15]. In total, 643 Asian and 801 European non-diabetic males were tested for baseline levels of nine L-amino acids in serum. Serum concentrations of isoleucine, phenylalanine, tyrosine, and alanine were significantly higher in the Asian men, while glycemia was similar in the two groups. Diabetes developed in 35% of the Asian and 14% of the European males. Strong associations between L-amino acids and diabetes were also observed in other studies [16][17][18]. In summary, L-amino acids could serve as a predictor of early-onset type 2 diabetes in healthy individuals.

2.4. Obstetric Conditions

There are data showing that L-amino acids are related to some obstetric conditions, such as gestational diabetes and pre-eclampsia. Significant differences in serum levels of particular amino acids (e.g., L-arginine, L-glycine, and 3-hydroxy-isovaleryl carnitine) are observed between females who develop gestational diabetes and controls [19]. According to that article, these differences exist already in the first trimester of pregnancy and may be useful for early screening. Additionally, increased concentrations of L-amino acids were shown to be more prevalent in women with pre-eclampsia [20]. In the reviewed article, maternal and cord blood L-amino acid levels were reported to be significantly higher in women with pre-eclampsia compared to pregnant women without this condition. Most likely, pre-eclampsia is associated with increased placental L-amino acid transport or reduced utilization of fetal L-amino acids.

2.5. Muscle Status

A strong relationship between L-amino acids and muscle status has been reported. For example, L-amino acid imbalance could explain fatigue after long-term exercise [21]. During overtraining, brain tryptophan uptake and 5-hydroxytryptamine synthesis are increased. In a 1993 study, the serum L-amino acid profile was determined in nine athletes before and after competing in the Colmar ultratriathlon. Serum concentrations of 25 L-amino acids (22 proteinogenic L-amino acids and citrulline, α -Aminobutyric acid, and taurine) decreased by 18% with a decrease in 18 individual L-amino acids by 9–56% and an increase in tyrosine, phenylalanine, methionine, cystine, and free tryptophan. In addition, a decrease in body mass (approximately 3.3 kg) and an increase in plasma volume

(approximately 7.6%) were observed. The decrease in intracellular water may be seen as a protein catabolic signal. Interestingly, it was also reported that L-amino acid supplementation could be used to prevent muscle damage. For example, during prolonged exercise, L-branched-chain amino acid (BCAA) supplementation decreases the serum concentration of intramuscular enzymes (creatine kinase and lactate dehydrogenase) and thus may reduce muscle damage [22]. L-amino acids were also reported to be important for patients who have diseases that induce muscle mass decrease. It was reported that the supplementation of BCAAs may reduce the muscle damage for patients with rheumatic diseases. It was demonstrated that plasma BCAAs, aspartic acid, and glutamate concentrations correlate positively with the rate of improvement in biceps femoris muscle atrophy, suggesting that these amino acids are associated with the BCAA-induced increase in muscle mass [23]. It was also demonstrated that, for patients treated in intensive care units, the supplementation of L-amino acids can protect skeletal muscle mass and function; for severely ill patients, a higher provision of protein and L-amino acids has been associated with a lower mortality [24]. Another recent report also indicated that significant amounts of L-amino acids are lost during continuous renal replacement therapy and may indicate the need for clinical intervention for clinical care patients [25]. It was demonstrated that the amount of L-amino acids lost during continuous renal replacement therapy is increased not only by the set-up specifics, but also by individual considerations of the patients (e.g., by a higher systemic concentration of amino acids and by a higher fat-free mass index (mostly muscles)). To summarize, L-amino acids are strongly associated with body muscle status, and they could show muscle damage and help to prevent it.

2.6. Cancer

L-amino acids could be an important parameter for cancer monitoring, diagnosis, and prognosis. It was reported that L-amino acids could be predictors of cancer cachexia and sarcopenia [26]. Cancer cachexia and sarcopenia cause ongoing muscle loss and a higher serum essential/total L-amino acid ratio is associated with sarcopenia in patients with advanced gastrointestinal cancers. It is reported that the levels of L-amino acids are also associated with the stage of disease and prognosis in patients with head and neck cancer [27]. Increased levels of alpha-aminobutyric acid, aminoadipic acid, L-histidine, L-proline, and L-tryptophan are associated with a reduced risk of advanced stage head and neck cancer. Increased levels of beta-alanine, beta-aminobutyric acid, ethanolamine, glycine, isoleucine, 4-hydroxyproline, and phenylalanine are associated with an increased risk of advanced stage head and neck cancer. Increased levels of alpha-aminobutyric acid are associated with increased overall survival, while increased levels of arginine, ethanolamine, glycine, histidine, isoleucine, 4-hydroxyproline, leucine, lysine, 3-methylhistidine, phenylalanine, and serine are associated with decreased overall survival. Other papers also report a relationship between cancers and L-amino acids. For example, in a recent review by Kang, the importance of L-amino acid restriction for cancer therapy was highlighted [28]. It was stated that L-amino acid restriction, in particular L-leucine, could be a simple metabolic intervention for cancer therapy.

2.7. Chronic Heart Failure

Plasma L-amino acids can predict chronic heart failure. It was demonstrated that 17 L-amino acids and two concentration ratios between specific L-amino acids were significantly different in a heart failure group compared

with those in the control [29]. The heart failure group had five specific L-amino acids, i.e., L-monoethanolamine, L-methionine, L-tyrosine, L-methylhistidine, and L-histidine, in correlation with cardiac function indicators such as B-type natriuretic peptide, left ventricle ejection fraction, and others. Moreover, in another study, it was shown that concentrations of essential L-amino acids and L-BCAAs are significantly lower for patients with severe chronic heart failure, which is associated with low nutritional status along with the loss of skeletal muscle [30]. To summarize, research papers indicate that chronic heart failure is related to changes in L-amino acid concentrations.

2.8. Other

A relationship between L-amino acids and other conditions also were reported. Several papers have highlighted the importance of L-amino acids, especially L-BCAAs, for the prediction of septic shock resolution and survival [31][32]. A significant number of papers report that L-amino acids play important roles in brain functioning and could be related to various mental and neurological conditions, such as autism spectrum disorders, cerebral palsy, and Parkinson's disease [33][34][35][36]. Papers have been published indicating a relationship between L-amino acids and liver cirrhosis [37][38] and chronic kidney disease [39]. Some papers also reported that L-amino acids may be related to a patient's frailty and longevity in general [40][41]. For example, Hamed et al. reported that L-amino acids could be important in age-related diseases and that protein and/or L-amino acid restriction could be related with reduced occurrence in cancer, diabetes, and overall mortality [41]. Another paper also indicated that L-amino acids are highly important in aging [42].

3. Biosensors for Measurement of Total L-Amino Acid Concentration

3.1. General Overview of Enzymatic Biosensors

Advantages and disadvantages of all generations of enzymatic electrodes are summarized in **Table 1**.

Table 1. Key advantages and disadvantages of enzymatic biosensors and their generations.

Generation	Key Advantages	Key Disadvantages
First (electrons are being transferred via native mediator, e.g., H ₂ O ₂)	<p>Simple to design</p> <p>Native mediator is constantly being produced by the enzyme, thus cannot be depleted</p> <p>The sensing layer can be separated from the electrode surface</p>	<p>Depended on the concentration of the dissolved oxygen in a solution</p> <p>Lack of sensitivity</p>

Generation	Key Advantages	Key Disadvantages
	The sensing layer can be made using excessive amount of enzyme to thus increase stability	High operational potential (e.g., +0.6 V vs. Ag/AgCl)
Second (electrons are being transferred via synthetic mediator, e.g., tetrathiafulvalene)	<p>Usually do not depend on the concentration of the dissolved oxygen in a solution</p> <p>Have better sensitivities in comparison to first generation</p> <p>The sensing layer can be made using excessive amount of enzyme to thus increase stability</p> <p>Low operational potential (e.g., +0.2 V vs. Ag/AgCl)</p>	<p>Complex design</p> <p>Synthetic mediator needs to be constantly added to the solution</p> <p>Desorption of mediator from the surface can occur</p>
Third (electrons are being transferred directly)	<p>Usually do not depend on the concentration of the dissolved oxygen in a solution</p> <p>Highest sensitivities in comparison to the first and second generations</p> <p>Lowest operational potentials (e.g., 0 V vs. Ag/AgCl)</p> <p>No mediator (native or synthetic) is needed for the electrode operation</p>	<p>Most complex design</p> <p>Can be implemented for a limited number of selective enzymes</p> <p>Low operational stability in comparison to other generations</p>

3.2. Overview of Biosensors for L-Amino Acid Detection

Table 2 summarizes most significant biosensors for L-amino acid detection.

Table 2. A comparative table of most significant biosensors for L-amino acid detection (majority for total L-amino acids, some for specific).

Biosensor	Enzyme, Nanomaterials Used	Analysis Method	Analytical Parameters (as Reported)	Real Sample
Nanjo and Guilbault [43]	Enzyme: L-amino acid oxidase (<i>Crotalus adamanteus</i>) Nanomaterials: N/A	Chronoamperometry	Sensitivity: N/A LOD: 10^{-6} – 10^{-5} M Linear range: N/A Stability: stable for at least 4 months	N/A
Varadi et al. [44]	Enzyme: L-amino acid oxidase (<i>Crotalus adamanteus</i>), D-amino acid oxidase (porcine kidney) Nanomaterials: N/A	Flow-through amperometry	Sensitivity: N/A LOD: N/A Linear range: 0.1–3 mM Stability: 900–1000 measurements	Brewing process samples (ginger and brown beer)
Sarkar et al. [45]	Enzyme: L-amino acid oxidase (<i>Crotalus adamanteus</i>), D-amino acid oxidase (porcine kidney) Nanomaterials: N/A	Chronoamperometry	Sensitivity: N/A LOD: 0.15–0.47 mM Linear range: 0.47–2.5 mM Stability: 40% activity loss after 56 days	Milk, fruit juice, urine
Lata and Pundir [46]	Enzyme: L-amino acid oxidase (goat kidney) Nanomaterials: MWCNT	Linear square voltammetry	Sensitivity: N/A LOD: 0.5 μ M	Fruit juices, alcoholic beverages

Biosensor	Enzyme, Nanomaterials Used	Analysis Method	Analytical Parameters (as Reported)	Real Sample	
			Linear range: 0.5 μM –100 mM Stability: 70% left after 140 days		211, N.; Key, fish- : J. Clin.
Dominguez et al. [47]	Enzyme: L-amino acid oxidase (<i>Gratelia Adamate</i>)*, D-amino acid oxidase (porcine kidney) Nanomaterials: N/A	Chronoamperometry	Sensitivity: 10–480 $\mu\text{A M}^{-1}$ LOD: 1.1–160 μM Linear range: 10^{-6} – 10^{-4} M Stability: around 10 days with no need to regenerate the electrode surface	Grapes	dman, ; :W.; 2017,
Odewunmi et al. [48]	Enzyme: enzyme-free Nanomaterials: AgO nanoparticles	Chronoamperometry	Sensitivity: 4230 $\mu\text{A mM}^{-1} \text{cm}^{-2}$ LOD: 0.42 μM Linear range: 60–500 μM Stability: N/A	Human serum	J.H.; sented ed Trial. C. Food nched- –1080. fat and A 2020,
Garcia-Carmona et al. [49]	Enzyme: enzyme-free Nanomaterials: MWCNT	Differential pulse voltammograms	Sensitivity: 0.012 $\mu\text{A } \mu\text{M}^{-1}$ LOD: 8 μM Linear range: 25–750 μM	Human serum	Eur. J. mino tudy in

Girls. J. Clin. Endocrinol. Metab. 2016, 101, 2047–2055.

Biosensor	Enzyme, Nanomaterials Used	Analysis Method	Analytical Parameters (as Reported)	Real Sample	Reference
			Stability: N/A		ig, T.; and the 14, 37,
			Sensitivity: N/A		/, P.; sulin
Hua et al. [50]	Enzyme: enzyme-free Nanomaterials: tetrahedral copper metal-organic frameworks	Linear sweep voltammetry	LOD: 25 nM Linear range: N/A Stability: no significant change after 6-month storage	Human blood	the fox, betes.
Miškinis et al. [51]	Enzyme: L-amino acid oxidase (<i>Crotalus adamanteus</i>) Nanomaterials: gold nanoparticles	Chronocoulometry	Sensitivity: 0.73 $\mu\text{C}/\mu\text{M}$ LOD: 5.5 μM Linear range: 5.5–100 μM Stability: 50% of the initial activity after 10 days of storage	Human serum and blood	nd, I.F.; al and in -979. mon, chain n in type 07. s, M.; mics.

18. Floegel, A.; Stefan, N.; Yu, Z.; Mühlenbruch, K.; Drogan, D.; Joost, H.-G.; Fritsche, A.; Häring, H.-U.; De Angelis, M.H.; Peters, A.; et al. Identification of Serum Metabolites Associated with Risk of Type 2 Diabetes Using a Targeted Metabonomic Approach. *Diabetes* 2013, 62, 639–648.
19. Nevalainen, J.; Sairanen, M.; Appelblom, H.; Gissler, M.; Timonen, S.; Ryyänen, M. First-Trimester Maternal Serum Amino Acids and Acylcarnitines Are Significant Predictors of Gestational Diabetes. *Rev. Diabet. Stud.* 2016, 13, 236–245.
20. Evans, R.W.; Powers, R.W.; Ness, R.B.; Cropcho, L.J.; Daftary, A.R.; Harger, G.F.; Vergona, R.; Finegold, D.N. Maternal and fetal amino acid concentrations and fetal outcomes during pre-eclampsia. *Reproduction* 2003, 125, 785–790.
21. Lehmann, M.; Huonker, M.; Dimeo, F.; Heinz, N.; Gastmann, U.; Treis, N.; Steinacker, J.M.; Keul, J.; Kajewski, R.; Häussinger, D. Serum Amino Acid Concentrations in Nine Athletes Before and After the 1993 Colmar Ultra Triathlon. *Int. J. Sports Med.* 1995, 16, 155–159.

22. Coombes, J.S.; McNaughton, L.R. Effects of branched-chain amino acid supplementation on serum creatine kinase and lactate dehydrogenase after prolonged exercise. *J. Sports Med. Phys. Fit.* 2000, 40, 240–246.
23. Yoshikawa, N.; Yamamoto, M.; Kuribara-Souta, A.; Uehara, M.; Yamazaki, H.; Tanaka, H. Amino Acid Profile in 18 Patients with Rheumatic Diseases Treated with Glucocorticoids and BCAAs. *J. Nutr. Sci. Vitaminol.* 2021, 67, 180–188.
24. Pavlickova Aimova, P.; Hronek, M.; Zadak, Z. The Importance and Dosage of Amino Acids in Nutritional Support of Various Pathological Conditions in ICU Patients; Biomedical Papers of the Medical Faculty of the Palacky University: Olomouc, Czech Republic, 2014; Volume 158, pp. 346–355.
25. Vicka, V.; Vickiene, A.; Miskinyte, S.; Bartuseviciene, I.; Lisauskiene, I.; Serpytis, M.; Ringaitiene, D.; Sipylaite, J. Role of Fat-Free Mass Index on Amino Acid Loss during CRRT in Critically Ill Patients. *Medicina* 2023, 59, 389.
26. Kitagawa, M.; Haji, S.; Amagai, T. High Serum Essential Amino Acids as a Predictor of Skeletal Muscle Depletion in Patients with Cachexia and Advanced Gastrointestinal Cancers. *Nutr. Clin. Pract.* 2017, 32, 645–651.
27. Cadoni, G.; Giraldi, L.; Chiarla, C.; Gervasoni, J.; Persichilli, S.; Primiano, A.; Settimi, S.; Galli, J.; Paludetti, G.; Arzani, D.; et al. Prognostic Role of Serum Amino Acids in Head and Neck Cancer. *Dis. Markers* 2020, 2020, 2291759.
28. Kang, J.-S. Dietary restriction of amino acids for Cancer therapy. *Nutr. Metab.* 2020, 17, 20.
29. Hakuno, D.; Hamba, Y.; Toya, T.; Adachi, T. Plasma Amino Acid Profiling Identifies Specific Amino Acid Associations with Cardiovascular Function in Patients with Systolic Heart Failure. *PLoS ONE* 2015, 10, e0117325.
30. Tsuji, S.; Koyama, S.; Taniguchi, R.; Fujiwara, T.; Fujiwara, H.; Sato, Y. Nutritional status of outpatients with chronic stable heart failure based on serum amino acid concentration. *J. Cardiol.* 2018, 72, 458–465.
31. Puskarich, M.A.; McHugh, C.; Flott, T.L.; Karnovsky, A.; Jones, A.E.; Stringer, K.A. Serum Levels of Branched Chain Amino Acids Predict Duration of Cardiovascular Organ Failure in Septic Shock. *Shock* 2021, 56, 65–72.
32. Su, L.; Li, H.; Xie, A.; Liu, D.; Rao, W.; Lan, L.; Li, X.; Li, F.; Xiao, K.; Wang, H.; et al. Dynamic Changes in Amino Acid Concentration Profiles in Patients with Sepsis. *PLoS ONE* 2015, 10, e0121933.
33. Mizuno, K.; Tanaka, M.; Nozaki, S.; Yamaguti, K.; Mizuma, H.; Sasabe, T.; Sugino, T.; Shirai, T.; Kataoka, Y.; Kajimoto, Y.; et al. Mental fatigue-induced decrease in levels of several plasma amino acids. *J. Neural Transm.* 2007, 114, 555–561.

34. Yu, X.; Qian-Qian, L.; Cong, Y.; Xiao-Bing, Z.; Hong-Zhu, D. Reduction of essential amino acid levels and sex-specific alterations in serum amino acid concentration profiles in children with autism spectrum disorder. *Psychiatry Res.* 2021, 297, 113675.
35. Tinkov, A.A.; Skalnaya, M.G.; Skalny, A.V. Serum trace element and amino acid profile in children with cerebral palsy. *J. Trace Elem. Med. Biol.* 2021, 64, 126685.
36. Figura, M.; Kuśmierska, K.; Bucior, E.; Szlufik, S.; Kozirowski, D.; Jamrozik, Z.; Janik, P. Serum amino acid profile in patients with Parkinson's disease. *PLoS ONE* 2018, 13, e0191670.
37. Ooi, P.H.; Gilmour, S.M.; Yap, J.; Mager, D.R. Effects of branched chain amino acid supplementation on patient care outcomes in adults and children with liver cirrhosis: A systematic review. *Clin. Nutr. ESPEN* 2018, 28, 41–51.
38. Muto, Y.; Sato, S.; Watanabe, A.; Moriwaki, H.; Suzuki, K.; Kato, A.; Kato, M.; Nakamura, T.; Higuchi, K.; Nishiguchi, S.; et al. Overweight and obesity increase the risk for liver cancer in patients with liver cirrhosis and long-term oral supplementation with branched-chain amino acid granules inhibits liver carcinogenesis in heavier patients with liver cirrhosis. *Hepatol. Res.* 2006, 35, 204–214.
39. Durantou, F.; Lundin, U.; Gayraud, N.; Mischak, H.; Aparicio, M.; Mourad, G.; Daurès, J.-P.; Weinberger, K.M.; Argilés, A. Plasma and Urinary Amino Acid Metabolomic Profiling in Patients with Different Levels of Kidney Function. *Clin. J. Am. Soc. Nephrol.* 2014, 9, 37–45.
40. Zhang, Y.; Zhou, Q.; Yang, R.; Hu, C.; Huang, Z.; Zheng, C.; Liang, Q.; Gong, R.; Zhu, X.; Gong, H.; et al. Serum branched-chain amino acids are associated with leukocyte telomere length and frailty based on residents from Guangxi longevity county. *Sci. Rep.* 2020, 10, 10252.
41. Mirzaei, H.; Suarez, J.A.; Longo, V.D. Protein and amino acid restriction, aging and disease: From yeast to humans. *Trends Endocrinol. Metab.* 2014, 25, 558–566.
42. Dato, S.; Hoxha, E.; Crocco, P.; Iannone, F.; Passarino, G.; Rose, G. Amino acids and amino acid sensing: Implication for aging and diseases. *Biogerontology* 2019, 20, 17–31.
43. Nanjo, M.; Guilbault, G. Enzyme electrode for L-amino acids and glucose. *Anal. Chim. Acta* 1974, 73, 367–373.
44. Varadi, M.; Adanyi, N.; Szabo, E.E.; Trummer, N. Determination of the ratio of D- and L-amino acids in brewing by an immobilised amino acid oxidase enzyme reactor coupled to amperometric detection. *Biosens. Bioelectron.* 1999, 14, 335–340.
45. Sarkar, P.; Tothill, I.E.; Setford, S.J.; Turner, A.P.F. Screen-printed amperometric biosensors for the rapid measurement of L- and D-amino acids. *Analyst* 1999, 124, 865–870.
46. Lata, S.; Pundir, C.S. L-amino acid biosensor based on L-amino acid oxidase immobilized onto NiHCNFe/c-MWCNT/PPy/GC electrode. *Int. J. Biol. Macromol.* 2013, 54, 250–257.

47. Domínguez, R.; Serra, B.; Reviejo, A.J.; Pingarrón, J.M. Chiral Analysis of Amino Acids Using Electrochemical Composite Bienzyme Biosensors. *Anal. Biochem.* 2001, 298, 275–282.
48. Odewunmi, N.A.; Kawde, A.-N.; Ibrahim, M. In-situ single-step electrochemical AgO modified graphite pencil electrode for trace determination of DL-methionine in human serum sample. *Sens. Actuators B Chem.* 2019, 281, 765–773.
49. García-Carmona, L.; Moreno-Guzmán, M.; Sierra, T.; González, M.C.; Escarpa, A. Filtered carbon nanotubes-based electrodes for rapid sensing and monitoring of L-tyrosine in plasma and whole blood samples. *Sens. Actuators B Chem.* 2018, 259, 762–767.
50. Hua, Y.; Lv, X.; Cai, Y.; Liu, H.; Li, S.; Wan, Y.; Wang, H. Highly selective and reproducible electroanalysis for histidine in blood with turn-on responses at a potential approaching zero using tetrahedral copper metal organic frameworks. *Chem. Commun.* 2019, 55, 1271–1274.
51. Miškinis, J.; Ramonas, E.; Gurevičienė, V.; Razumienė, J.; Dagys, M.; Ratautas, D. Capacitance-Based Biosensor for the Measurement of Total Loss of L-Amino Acids in Human Serum during Hemodialysis. *ACS Sens.* 2022, 7, 3352–3359.

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