# Updates in Anticoagulation Therapy Monitoring

Subjects: Pathology | Hematology Contributor: Hannah L McRae

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anticoagulant anticoagulation warfarin

# 1. Introduction

Since the initial discovery of heparin in 1916, there have been countless clinical and scientific advances in the pharmacophysiology of anticoagulation. This started from the commercial production and first clinical trials involving heparin in the 1930s, to the discovery of coumarin in the 1940s and the subsequent development of warfarin as a rodenticide in 1948. This was then followed by decades-long widespread clinical use of heparin and warfarin and the more recent development of new, targeted oral anticoagulants in the past 10–15 years <sup>[1][2][3]</sup>. Anticoagulation is indicated in a broad range of clinical scenarios, including (but not limited to) the management of venous and/or arterial thromboembolism, treatment of disseminated intravascular coagulation, the flushing of lines such as in hemodialysis, cardiopulmonary bypass, or extracorporeal membrane oxygenation (ECMO). Anticoagulation can also be used prophylactically in patients with atrial fibrillation, artificial valves, and in the post-operative and critical care settings <sup>[4]</sup>.

Anticoagulants are often categorized according to the mechanism of action (<u>Table 1</u>). Heparins are subcategorized as unfractionated heparin (UFH) (given intravenously or subcutaneously) and low molecular weight heparins (LMWH) including enoxaparin (Lovenox<sup>®</sup>/Clexane<sup>®</sup>, Sanofi, Paris, France), dalteparin (Fragmin<sup>®</sup>, Pfizer, New York, NY, USA), tinzaparin (Innohep<sup>®</sup>, LEO Pharma, Ballerup, Denmark) and nadroparin (Fraxiparine<sup>®</sup>/Fraxodi<sup>®</sup>, Aspen Pharmacare, Durban, South Africa), which are administered subcutaneously. Warfarin, acenocoumarol, and phenprocoumon are known as vitamin K antagonists, and have historically been the most ubiquitous oral anticoagulants prescribed for long-term anticoagulation. Factor Xa inhibitors consist of fondaparinux (Atrixa<sup>®</sup>, Aspen Pharmacare, Durban, South Africa)) (a synthetic anticoagulant), which is administered subcutaneously, as well as the direct oral anticoagulants (DOAC) rivaroxaban (Xarelto<sup>®</sup>, Janssen Pharmaceuticals, Beerse, Belgium), apixaban (Eliquis<sup>®</sup>, Bristol-Myers Squibb, New York, NY, USA), edoxaban (Savaysa<sup>®</sup>, Daiichi-Sankyo, Tokyo, Japan), and betrixaban (Bevyxxa<sup>®</sup>, Portola, South San Francisco, CA, USA). The final class of anticoagulants is the direct thrombin inhibitors (DTI), which include dabigatran (Pradaxa<sup>®</sup>, Boehringer Ingelheim, Ingelheim am

Rhein, Germany) (a DOAC), and the parenterally administered bivalirudin (Angiomax<sup>®</sup>, Sandoz, Holzkirchen, Germany) and argatroban (Acova<sup>®</sup>, Texas Biotechnology Corporation, Houston, TX, USA).

Anticoagulation Category	Medication Name(s)	Mechanism of Action	Route(s) of Administration
Vitamin K Antagonists	Warfarin, Acenocoumarol, Phenprocoumon	Inhibition of vitamin K epoxy reductase to decrease the synthesis of vitamin K-dependent coagulation factors	Oral
Heparin (Unfractionated)	Heparin	Inhibition of thrombin and several activated coagulation factors (including Xa) by binding to and enhancing the activity of antithrombin III	Intravenous or Subcutaneous paretneral injection
Heparin (Low Molecular Weight)	Enoxaparin, Dalteparin, Tinxaparin, Nadroparin	Binds to antithrombin III and inhibits thrombin to a much lesser extent than unfractionated heparin; primarily inhibits factor Xa	Subcutaneous parenteral injection
Factor Xa Inhibitors	Fondaparinux *, Rivaroxaban, Apixaban, Edoxaban, Betrixaban	Prevents the cleaving of prothrombin by factor Xa to form thrombin	Fondaparinux- Subcutaneous parenteral injection Rivaroxaban, apixban, edoxaban, betrixaban- Oral
Factor IIa Inhibitors (Direct Thrombin Inhibitors)	Dabigatran, Bivalirudin, Argatroban	Directly binds to and inhibit thrombin	Dabigatran- Oral Bivalirudin- Intravenous Argatroban- Intravenous or Subcutaneous parenteral injection

**Table 1.** Anticoagulants categorized by mechanism of action.

\* Fondaparinux, while technically a synthetic low molecular weight heparin, is considered an indirect factor Xa inhibitor.

In addition to the logistics associated with the different types of administration of anticoagulation, the clinical indication for each anticoagulant varies due to discrepancies in the risk of adverse drug events (particularly thrombotic vs. hemorrhagic risk), therapeutic index, mechanism of drug clearance (i.e., hepatic vs. renal), drug half-life, requirements for therapeutic monitoring, and potential and mechanism of anticoagulant reversal <sup>[4][5]</sup>.

### 2. Vitamin K Antagonists

Vitamin K antagonists (VKA) are oral anticoagulants that inhibit the vitamin K epoxy reductase enzyme, which is required for the conversion of vitamin K to its active form, vitamin KH2. The vitamin K-dependent coagulation factors (II, VII, IX, and X) depend on vitamin KH2 to become synthesized by the liver <sup>[6]</sup>. Warfarin is the most common VKA used clinically in the United States, while others such as acenocoumarol and phenprocoumon are frequently used in other countries. VKA are the most commonly prescribed oral anticoagulants worldwide, though fewer patients are being prescribed VKA now as more Xa- and IIa-inhibiting direct oral anticoagulants have become increasingly prevalent in the past decade <sup>[Z]</sup>. VKA are clinically indicated in the treatment and prophylaxis of venous thromboembolism (VTE) and pulmonary embolism, and in the setting of heart failure, atrial fibrillation, acute coronary syndrome, prosthetic heart valve, stroke, and antiphospholipid syndrome <sup>[B][9]</sup>. Contraindications include bleeding diathesis, thrombocytopenia, central nervous system tumors, major trauma, uncontrolled hypertension, active bleeding, and pregnancy, as VKA cross the placenta and may induce fetal hemorrhage as well as increases the risk for bleeding complications during delivery <sup>[9][10]</sup>.

One of the main advantages to VKA therapy is the body of research and evidence-based practice guidelines that stem from decades of use worldwide. As a result, there is a high degree of clinical familiarity with the drug. In addition, VKA are cheap and easily accessible compared to DOACs; a 2018 study in Britain revealed that DOACs are prescribed to 31% of patients treated for atrial fibrillation, but account for approximately 93% of National Health Service (NHS) expenditure on anticoagulants (referring to prescription costs only) <sup>[11]</sup>. VKA have also been shown to be safer and more efficacious than other oral anticoagulants in patients with certain conditions, such as prosthetic heart valves and recurrent thrombosis in the setting of antiphospholipid syndrome <sup>[12][13]</sup>. VKA are also quickly and easily reversed, which is necessitated by many scenarios from planned surgeries to major trauma and intracranial hemorrhage. Depending on the urgency and extent of international normalized ratio (INR) correction required, reversal can be achieved by VKA discontinuation (or abruption), the administration of oral or IV vitamin K, transfusion of fresh frozen plasma (FFP), and replacement of vitamin K-dependent coagulation factors via infusion of prothrombin complex concentrates.

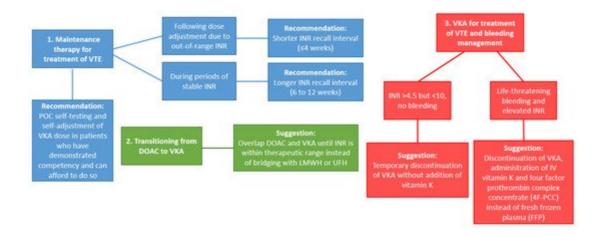
However, there are several adverse effects associated with VKA therapy that make DOAC a better option in some cases, including high rates of serious bleeding complications. According to the results of 33 meta-analyses, the rate of major VKA-related bleeding events is 7.2 per 100 patient-years, and fatal bleeds occur at a rate of 1.3 per 100 patient-years <sup>[14]</sup>. VKA are also shown to be unpredictable and associated with high rates of thromboembolic and bleeding complications in patients with atrial fibrillation; a study of 6454 patients with atrial fibrillation revealed that patients were outside the therapeutic range almost 50% of the time, thus increasing the risk for either thrombosis (below the range) or bleeding (above the range) <sup>[15]</sup>. Furthermore, VKA require frequent monitoring, and diet and co-medications can have considerable implications in patients taking VKA, either enhancing or inhibiting their anticoagulant effect <sup>[9]</sup>.

VKA also have a delayed onset of action as compared to other anticoagulants, typically requiring 24–72 h, and approximately 5–7 days to reach their peak therapeutic effect after initiation. The half-life is approximately 40 h on average (ranging from 20–60 h); however, there is great variability in half-life duration between patients <sup>[8]</sup>. The therapeutic level of VKA is measured by prothrombin time (PT) and INR, which is a standardized ratio developed

by the World Health Organization (WHO) in the 1980s specifically for VKA monitoring, as PT varies greatly between laboratories <sup>[16]</sup>. WHO developed its procedure to mitigate the discrepancy in tissue factor (TF) activity between PT reagents and thus allow for the expression of PT results on a common scale, i.e., INR. The INR is derived from the International Sensitivity Index (ISI), which was developed by WHO and quantifies the reactivity of individual PT reagents and analyzers. In addition, each center has its own geometric mean PT (MNPT), which is the average PT calculated from a least 20 normal donors of both sexes, tested on the same local analyzer and under the same test conditions as the patient PT. The formula for INR is INR = (patient's PT/MNPT)<sup>ISI</sup> <sup>[17][18]</sup>.

In order to properly assess INR after initiation of VKA therapy, the baseline PT, activated partial thromboplastin times (aPTT), and INR values are obtained; this ensures proper calculation of the therapeutic target, and is especially important in patients with naturally prolonged PT and/or aPTT, such as in the case of antiphospholipid syndrome. In addition, it is advisable to assess the patient's liver function to identify potential issues with the metabolism of VKA or baseline hemostatic issues <sup>[9]</sup>. VKA have a narrow therapeutic index (typically an INR of 2.0–3.0, though this may be higher in the setting of artificial heart valves or conditions such as antiphospholipid syndrome), and the dosages required to achieve this range are inconsistent from patient to patient. The correlation between dose and anticoagulant response can be affected by genetic factors as well as environmental variables, such as dietary and nutritional intake, drug interactions, illness and injury, etc., all of which affect the absorption and pharmacokinetics of VKA and vitamin K <sup>[5]</sup>. Depending on fluctuations in INR, clinicians may either adjust the patient's warfarin dosing accordingly, or instruct them to alter their dietary habits.

Careful monitoring via INR is typically recommended at the initiation of VKA therapy, and is usually performed daily in hospitalized patients, and one to three times per week in outpatients until the dose is properly adjusted. Monitoring may be more frequent in patients for whom there is difficulty achieving INR within the therapeutic range, and may eventually decrease to intervals between every two to four weeks once the INR has stabilized for at least one week <sup>[9]</sup>. Traditionally VKA monitoring has required patients to travel to an outpatient laboratory or clinic and undergo a venous blood draw for each INR; however, in the past 10-15 years several point-of-care (POC) devices have been developed and approved for both clinical and home use. This allows patients to be tested either at their regular clinic appointments (thus alleviating the need for extra travel), or at their convenience at home using a personal INR meter, from which the results are uploaded to their electronic medical records for direct supervision by their medical provider. Perhaps unsurprisingly, patient self-testing has been associated with improved quality of life and cost savings as compared to traditional INR monitoring <sup>[19]</sup>. However, recent studies have shown that POC INR devices are associated with decreased precision when the INR is greater than 3.0, and general unreliability once the INR exceeds 4.0 <sup>[20][21]</sup>. These devices are therefore best suited to patients who are compliant with their diet and VKA therapy, and who maintain stable INRs within the therapeutic range and with little variability longterm. As new data emerges, the American Society of Hematology (ASH) guidelines for best practices in VKA monitoring and reversal continue to be updated (Figure 1)  $\frac{[22]}{2}$ .



**Figure 1.** Updates to recommendations for VKA management (Based on ASH guidelines <sup>[22]</sup>). Abbreviations: VTE: venous thromboembolism; POC: point-of-care; VKA: Vitamin K antagonists; INR: international normalized ratio; DOAC: Direct oral anticoagulant; LMWH: low molecular weight heparins; UFH: unfractionated heparin.

## 3. Heparin

Heparin is a glycosaminoglycan containing a pentasaccharide that binds to and enhances the activity of antithrombin III by inducing a conformational change, thereby inhibiting thrombin and several activated coagulation factors (XIIa, IXa, XIa, and Xa) <sup>[23]</sup>. Heparin is not readily absorbed by the gastrointestinal tract and is thus administered either intravenously or subcutaneously. Heparins are comprised of UFH and LMWH. UFH consists of a mixture of polysaccharides with varying molecular weights averaging approximately 15,000 Daltons. LMWH has shorter polysaccharide chains and average molecular weights between 4000–6000 Daltons. LMWH does not inhibit thrombin as readily as UFH; however, LMWH and UFH are thought to inhibit factor Xa to a similar degree <sup>[24]</sup>.

#### 3.1. Unfractionated Heparin

UFH is the most pervasive anticoagulant used in the inpatient population worldwide for multiple reasons, including treatment or prophylaxis of VTE, stroke and transient ischemic attack (TIA), acute coronary syndromes, cardiac surgeries including cardioversion, and in the perioperative and critical care settings <sup>[5]</sup>. It is also used to flush lines to avoid contact factor activation such as in hemodialysis, ECMO, and cardiopulmonary bypass machines. In addition, UFH is considered safe for use in all populations including neonates, children, and pregnant women.

UFH has a short half-life of 30 min when administered as a continuous, intravenous (IV) infusion, and 90 min when administered subcutaneously via parenteral injections. It is easily reversible using protamine sulfate; however, in the absence of protamine, the short half-life allows for reversal by simple discontinuation of UFH administration <sup>[24]</sup>. Nevertheless, there are significant limitations to the use of UFH, as it has a highly variable dose–response relationship and as such requires frequent monitoring to ensure therapeutic levels, is unable to be administered orally, and is associated with complications such as heparin-induced thrombocytopenia (HIT) and increased risk of bleeding events as compared to LMWH <sup>[25]</sup>.

Therapeutic dosing of UFH is typically achieved by the IV administration of an initial bolus followed by weightbased or calculated, fixed-dose heparin dosing via continuous infusion, that can be modified as needed depending on the bleeding risk <sup>[26][27]</sup>. Prophylactic UFH is typically administered in 5000 U subcutaneous parenteral injections, either two or three times per day; while there is conflicting evidence available on which regimen is more effective. Meta-analytic data suggests that 5000 U three times per day is more efficacious than twice per day for VTE prophylaxis, despite the higher bleeding risk <sup>[28]</sup>. After administration, UFH is removed from circulation via a combination of the saturable mechanism, by which heparin binds with high affinity to endothelial cells and is cleared by the reticuloendothelial system and the non-saturable mechanism, i.e., renal excretion <sup>[29]</sup>.

The historical gold standard for UFH monitoring has been the use of serial aPTT that are typically measured frequently (within 2 h of initiation of continuous IV infusion, and every 6 h thereafter). The therapeutic range is approximately 1.5–2.5 times the patient's baseline aPTT <sup>[23][24][30]</sup>. The UFH dose may then be adjusted in relation to the aPTT, and monitoring may become less frequent as the target range is achieved and sustained. While this therapeutic aPTT range is widely recognized and utilized by clinicians, there is only limited evidence supporting this guideline. There are several complicating factors that can make aPTT monitoring difficult and unreliable; aPTT is sensitive to other variables such as coagulation factor consumption in the setting of bleeding or thrombosis, decreased synthesis of coagulation factors in the setting of liver dysfunction or disorders such as hemophilia and von Willebrand disease, and interferences such as the presence of a lupus anticoagulant, which would prolong the baseline aPTT <sup>[31][32]</sup>. In addition, there is known to be wide variability in the sensitivity of aPTT reagents, and individual laboratories are therefore recommended to define their own therapeutic aPTT ranges for safe and reliable heparin monitoring <sup>[33]</sup>.

UFH may also be monitored via anti-factor Xa (anti-Xa) activity, and the question of whether this method is more efficacious than the use of aPTT remains controversial. UFH anti-Xa assays specifically measure the ability of heparin-bound antithrombin III to inhibit factor Xa <sup>[34]</sup>. Studies have shown that the use of anti-Xa is more efficient in achieving the target therapeutic range of UFH as compared to aPTT; however, this has not been shown to have an effect on clinical outcomes <sup>[35][36]</sup>. In addition, while both aPTT and anti-Xa can be performed on the same automated coagulation analyzers, anti-Xa is reportedly difficult to standardize and has lower precision than aPTT tests <sup>[34][37][38]</sup>. Furthermore, many smaller hospitals and laboratories do not offer anti-Xa as it is more specialized and expensive, and in general there is less knowledge among clinicians surrounding the utility and interpretation of anti-Xa results as compared to aPTT. Anti-Xa is also not feasible in patients with recent direct oral factor Xa inhibitor use, as these patients may have residual anti-Xa activity at the start of UFH therapy <sup>[27]</sup>. A new product known as DOAC-Stop<sup>®</sup> (Haematex Research, Hornsby, Australia) has been shown to mitigate the interference of DOACs with anti-Xa and other standard coagulation assays, though despite its potential clinical utility, DOAC-Stop<sup>®</sup> has only been evaluated in clinical trials and is not yet available for commercial use <sup>[39]</sup>.

On the other hand, anti-Xa may mitigate some of the issues surrounding aPTT monitoring in certain patients for which aPTT is unreliable, such as those with lupus anticoagulant. In addition, the aPTT may also be falsely prolonged in the setting of elevated C-reactive protein (CRP), thereby causing discrepancies in aPTT and anti-Xa results in this patient population. Anti-Xa is therefore recommended for monitoring heparin therapy in patients with

elevated CRP <sup>[40]</sup>. Interestingly, a 2012 study of 539 hospitalized adults receiving UFH with a total of 2,321 paired aPTT and anti-Xa values found that there may be some clinical utility in measuring both aPTT and anti-Xa, since patients with disproportionately prolonged aPTT values as compared to anti-Xa had worse clinical outcomes. The study concluded that the concurrent measurement of aPTT and anti-Xa could be useful in stratifying bleeding risk and assist in determining the appropriate dosing regimen <sup>[36]</sup>.

There are some exceptions to the use of aPTT or anti-Xa for UFH monitoring, including interventional cardiology patients, who often receive very high doses of UFH intraoperatively, such as during cardiac catheterization, coronary artery bypass grafting (CABG), or left ventricular implant device (LVAD) implantation, as well as patients on ECMO. The activated clotting time (ACT) is a point-of-care test used for high-dose heparin monitoring and measures the inhibition of the contact and common pathway (factor X-Xa) activation. ACT is the preferred test in these settings due to the short time from sampling to results, size and portability (allowing for measurement at bedside and during transport), user-friendliness and ability to be performed by non-laboratory personnel, no need for a central laboratory, and quick confirmation of UFH reversal with protamine sulfate <sup>[41]</sup>. However, ACT is prone to interference by other anticoagulants, especially DOACs, which are common in patients undergoing interventional cardiac procedures. This can result in largely variable clotting times, thus challenging the standard therapeutic target of 300 s and resulting in the potential for under- or over-dosing of UFH <sup>[42]</sup>. Additional challenges to UFH monitoring include patients with heparin resistance or antithrombin III deficiency, for which both aPTT and anti-Xa (and the addition of antithrombin III concentrates in antithrombin III-deficient patients, if clinically indicated) are typically used <sup>[27]</sup>.

#### 3.2. Low Molecular Weight Heparin

There are multiple commercially available preparations of LMWH worldwide, including enoxaparin, dalteparin, tinzaparin, and nadroparin. Each variant of LMWH is chemically and pharmacologically distinct, including different ratios of factor Xa vs. thrombin inhibition, meaning that the clinical efficacy and safety of each drug varies as well <sup>[27]</sup>. Enoxaparin has the widest range of clinical indications due to the breadth of clinical data regarding safety and efficacy across many patient populations. As such, it is the most commonly marketed and prescribed LMWH <sup>[43]</sup>. However, there are few studies to date that have compared clinical outcomes in patients taking different LMWH products. LMWH is administered via subcutaneous parenteral injection and has similar clinical indications as UFH, including the treatment and prophylaxis of VTE (including during pregnancy), treatment of myocardial infarction and unstable angina, and prevention of clotting in extracorporeal circuits <sup>[44]</sup>. LMWH is also the recommended anticoagulant for the treatment of cancer-associated VTE <sup>[45]</sup>.

Clinicians usually weigh the pros and cons of anticoagulation with LMWH as opposed to UFH in order to determine the best regimen for their patients. LMWH is often preferable over UFH for several reasons. Firstly, it is more readily absorbed and involves less endothelium and protein binding, which results in greater bioavailability. It also has a longer half-life of 4 h, allowing for injection only once or twice daily. LMWH has been shown to have better correlation between dosage and anticoagulant response, which allows for fixed-dose administration and less frequent monitoring, if at all <sup>[24][27]</sup>. In addition, LMWH carries a lower risk of complications such as HIT, bleeding,

and is associated with a lower risk of osteoporosis as compared to UFH <sup>[27][46]</sup>. LMWH injections also lend themselves to both inpatient and outpatient use, though long-term use is often associated with bruising and the deterioration of injection sites, and overall lower patient satisfaction as compared to oral anticoagulants <sup>[44][47]</sup>.

Other limitations of LMWH as compared to UFH include a delayed onset of action (up to 30 min as opposed to instantaneous functionality in the case of intravenous UFH bolus), and the longer half-life makes urgent reversal more difficult <sup>[27]</sup>. Protamine sulfate can be used for reversal in the absence of alternative solutions; however, it is known to be less effective at reversing anti-Xa activity than antithrombin activity <sup>[48]</sup>. Since LMWH is renally cleared, it has a prolonged half-life in patients with renal failure, which is associated with a higher risk of accumulation and subsequent bleeding complications <sup>[49]</sup>.

Anti-Xa is the gold standard for monitoring LMWH therapy, as aPTT is not significantly affected. LMWH is thought to have lower requirements for monitoring in general as compared to UFH, primarily due to fixed-dose administration, the longer half-life, and improved bioavailability, as discussed previously. In addition, studies have shown that the anticoagulant effect and risk of bleeding are not consistently correlated with plasma anti-Xa levels, and that weight-adjusted LMWH dosing has proven to be safe and effective, thus alleviating the need for close monitoring <sup>[50]</sup>. Furthermore, despite the increase in use of DOACs and resultant increase in availability of anti-Xa testing, not all patients or clinics (especially in rural areas) have routine access to a laboratory that offers anti-Xa testing, and increased turnaround time is impractical when making dose adjustments <sup>[27]</sup>. There is also known to be substantial variability between specific anti-Xa assays, which is affected by both the specific reagent and analyzer used, in addition to the aPTT reagent used for correlation of the therapeutic range. In turn, there is concern that this lack of standardization could potentially negatively affect clinical outcomes <sup>[51][52][53]</sup>.

In patients for whom LMWH monitoring is required, anti-Xa levels should be obtained at their peak 4 h postadministration <sup>[54]</sup>. Dose adjustments in patients with renal insufficiency are based on anti-Xa levels; anti-Xa based enoxaparin dose reduction in particular has been shown to reduce the risk of bleeding in these patients <sup>[46][55]</sup>. Some studies have shown that the area under the thrombin generation curve, known as the endogenous thrombin potential (ETP), may be useful in LMWH and UFH monitoring, since it measures the total amount of thrombin formed in vivo as opposed to the limited quantity of thrombin formed in traditional coagulation assays (e.g., aPTT) <sup>[56][57][58][59]</sup>. While this assay may have potential, both for heparin monitoring and for hemostatic evaluation as a whole, it is not currently standardized or validated for clinical use <sup>[60]</sup>.

### 4. Fondaparinux

Fondaparinux is a synthetic anticoagulant derived from a pentasaccharide sequence that functions similarly to LMWH in that it inhibits factor Xa, but not thrombin. It is known as an indirect factor Xa inhibitor because it inhibits factor Xa by way of selectively and reversibly binding to antithrombin III with a higher affinity than UFH and LMWH <sup>[61]</sup>. Fondaparinux is indicated in the treatment and prophylaxis of VTE, is often prescribed in the setting of acute coronary syndrome, and as an alternative to heparin in patients diagnosed with HIT, as it does not interact with platelets or platelet factor 4 <sup>[62]</sup>. It has 100% bioavailability after subcutaneous administration, reaching its peak

concentration in 1.5–2 h after injection <sup>[63]</sup>. The half-life of fondaparinux is approximately 15–17 h, and anticoagulant functionality remains for 2–5 days after injection in patients with normal renal function. It is administered once daily and dosed according to body weight and indication for use (i.e., therapeutic vs. prophylactic dosing). Protamine sulfate is ineffective in the reversal of fondaparinux, and no specific reversal agents are directly approved for this purpose. However, the administration of recombinant activated factor VII has shown to aid in the cessation of bleeding in fondaparinux-related hemorrhage <sup>[64]</sup>.

Fondaparinux does not require monitoring in the majority of cases; however, anti-Xa may be used in certain cases in which levels must be acutely determined, such as in high-risk patients with renal insufficiency, and should be performed approximately three hours after administration <sup>[62]</sup>. Since anti-Xa is not officially approved for fondaparinux monitoring, the therapeutic range is not well-established or standardized, and thus varies between clinical laboratories <sup>[65]</sup>.

#### References

- 1. Barrowcliffe, T.W. History of heparin. In Heparin–A Century of Progress; Lever, R., Mulloy, B., Page, C.P., Eds.; Springer: Berlin/Heidhelberg, Germany, 2012; Volume 207, pp. 3–22.
- 2. Petitou, M.; Casu, B.; Lindahl, U. 1976–1983, A critical period in the history of heparin: The discovery of the antithrombin binding site. Biochimie 2003, 85, 83–89.
- 3. Wardrop, D.; Keeling, D. The story of the discovery of heparin and warfarin. Br. J. Haematol. 2008, 141, 757–763.
- Harter, K.; Levine, M.; Henderson, S.O. Anticoagulation drug therapy: A review. West. J. Emerg. Med. 2015, 16, 11–17.
- 5. Alquwaizani, M.; Buckley, L.; Adams, C.; Fanikos, J. Anticoagulants: A review of the pharmacology, dosing, and complications. Curr. Emerg. Hosp. Med. Rep. 2013, 1, 83–97.
- Ageno, W.; Gallus, A.S.; Wittkowsky, A.; Crowther, M.; Hylek, E.M.; Palareti, G. Oral anticoagulant therapy: Antithrombotic Therapy and Prevention of Thrombosis, 9th ed; American College of Chest Physicians Evidence-Based Clinical Practice Guidelines. Chest 2012, 141, e44S–e88S.
- 7. van Hove, K.H.; Leng, M.G. Trends in anticoagulant prescribing: A review of local policies in english primary care. BMC Health Serv. Res. 2020, 20, 279.
- 8. Patel, S.; Singh, R.; Preuss, C.V.; Patel, N. Warfarin; StatPearls [Internet]; StatPearls Publishing. Available online: (accessed on 27 November 2020).
- 9. Hull, R.D.G.; David, A.; Vazquez, S.R. Warfarin and Other VKAS: Dosing and Adverse Effects. Tirnauer, J.S., Ed.; UpToDate. Available online: (accessed on 4 December 2020).

- 10. Abadi, S.; Einarson, A.; Koren, G. Use of warfarin during pregnancy. Can. Fam. Physician 2002, 48, 695–697.
- 11. Burn, J.; Pirmohamed, M. Direct oral anticoagulants versus warfarin: Is new always better than the old? Open Heart 2018, 5, e000712.
- 12. Gaasch, W.H.; Konkle, B.A. Antithrombotic Therapy for Surgical Prosthetic Heart Valves and Surgical Valve Repair: Indications. Yeon, S.B., Ed.; Available online: (accessed on 4 December 2020).
- 13. Dufrost, V.; Risse, J.; Zuily, S.; Wahl, D. Direct oral anticoagulants use in antiphospholipid syndrome: Are these drugs an effective and safe alternative to warfarin? A systematic review of the literature. Curr. Rheumatol. Rep. 2016, 18, 74.
- 14. Pirmohamed, M. Warfarin: Almost 60 years old and still causing problems. Br. J. Clin. Pharmacol. 2006, 62, 509–511.
- 15. Boulanger, L.; Kim, J.; Friedman, M.; Hauch, O.; Foster, T.; Menzin, J. Patterns of use of antithrombotic therapy and quality of anticoagulation among patients with non-valvular atrial fibrillation in clinical practice. Int. J. Clin. Pr. 2006, 60, 258–264.
- 16. Riley, R.S.; Rowe, D.; Fisher, L.M. Clinical utilization of the international normalized ratio (INR). J. Clin. Lab. Anal. 2000, 14, 101–114.
- 17. Poller, L. International Normalized Ratios (INR): The first 20 years. J. Thromb. Haemost. 2004, 2, 849–860.
- van den Besselaar, A.M.H.P.; Chantarangkul, V.; Angeloni, F.; Binder, N.B.; Byrne, M.; Dauer, R.; Gudmundsdottir, B.R.; Jespersen, J.; Kitchen, S.; Legnani, C.; et al. International collaborative study for the calibration of proposed International Standards for thromboplastin, rabbit, plain, and for thromboplastin, recombinant, human, plain. J. Thromb. Haemost. 2017, 16, 142–149.
- Phibbs, C.S.; Love, S.R.; Jacobson, A.K.; Edson, R.; Su, P.; Uyeda, L.; Matchar, D.B. At-Home versus in-clinic INR monitoring: A cost–utility analysis from the home INR study (THINRS). J. Gen. Intern. Med. 2016, 31, 1061–1067.
- Arline, K.; Rodriguez, C.; Sanchez, K. Reliability of point-of-care international normalized ratio measurements in various patient populations. Point Care: J. Near-Patient Test. Technol. 2020, 19, 12–18.
- 21. Wool, G.D. Benefits and pitfalls of point-of-care coagulation testing for anticoagulation management: An ACLPS critical re-view. Am. J. Clin. Pathol. 2019, 151, 1–17.
- 22. Witt, D.M.; Nieuwlaat, R.; Clark, N.P.; Ansell, J.; Holbrook, A.; Skov, J.; Shehab, N.; Mock, J.; Myers, T.; Dentali, F.; et al. American Society of Hematology 2018 guidelines for management of

venous thromboembolism: Optimal management of anticoagulation therapy. Blood Adv. 2018, 2, 3257–3291.

- 23. Bussey, H.; Francis, J.L. Heparin overview and issues. Pharmacotherapy 2004, 24, 103s–107s.
- 24. Blann, A.D.; Landray, M.J.; Lip, G.Y.H. ABC of antithrombotic therapy: An overview of antithrombotic therapy. Br. Med.Assoc. 2002, 325, 762–765.
- 25. Robertson, L.; Jones, L.E. Fixed dose subcutaneous low molecular weight heparins versus adjusted dose unfractionated heparin for the initial treatment of venous thromboembolism. Cochrane Database Syst Rev 2017, 2, Cd001100.
- 26. Bernardi, E.; Piccioli, A.; Oliboni, G.; Zuin, R.; Girolami, A.; Prandoni, P. Nomograms for the administration of unfractionated heparin in the initial treatment of acute thrombo-embolism—An overview. Thromb. Haemost. 2000, 84, 22–26.
- 27. Hull, R.D.G.; David, A.; Burnett, A.E. Heparin and LMW heparin: Dosing and adverse effects. Tirnauer, J.S., Ed.; UpToDate. Available online: (accessed on 4 November 2019).
- 28. Mahan, C.E.; Pini, M.; Spyropoulos, A.C. Venous thromboembolism prophylaxis with unfractionated heparin in the hos-pitalized medical patient: The case for thrice daily over twice daily dosing. Intern. Emerg. Med. 2010, 5, 299–306.
- 29. Boneu, B.; Caranobe, C.; Sie, P. Pharmacokinetics of heparin and low molecular weight heparin. Baillieres Clin Haematol. 1990, 3, 531–544.
- Appert-Flory, A.; Fischer, F.; Jambou, D.; Buvat, S.; Mahagne, M.-H.; Toulon, P. Monitoring unfractionated heparin treatments. stability of plasma anti-Xa activity up to 4 h in citrated tubes. Blood 2019, 134, 3403.
- 31. Rountree, K.M.; Lopez, P.P. Partial Thromboplastin Time; Updated 21 August 2020; StatPearls Publishing LLC: Treasure Island, FL, USA, 2020.
- 32. Eikelboom, J.W.; Hirsh, J. Monitoring unfractionated heparin with the aPTT: Time for a fresh look. Thromb. Haemost. 2006, 96, 547–552.
- 33. Gouin-Thibaut, I.; Martin-Toutain, I.; Peynaud-Debayle, E.; Marion, S.; Napol, P.; Alhenc-Gelas, M. Monitoring unfractionated heparin with APTT: A French collaborative study comparing sensitivity to heparin of 15 APTT reagents. Thromb. Res. 2012, 129, 666–667.
- Ignjatovic, V.; Summerhayes, R.; Gan, A.; Than, J.; Chan, A.; Cochrane, A.; Bennett, M.; Horton, S.; Shann, F.; Lane, G.; et al. Monitoring unfractionated heparin (UFH) therapy: Which anti factor Xa assay is appropriate? Thromb. Res. 2007, 120, 347–351.
- 35. Coons, J.C.; Iasella, C.J.; Thornberg, M.; Fitzmaurice, M.G.; Goehring, K.; Jablonski, L.; Leader, D.; Meyer, A.; Seo, H.; Benedict, N.J.; et al. Clinical outcomes with unfractionated heparin

monitored by anti-factor Xa vs. activated partial thrombo-plastin time. Am. J. Hematol. 2019, 94, 1015–1019.

- 36. Zehnder, J.; Price, E.; Jin, J. Controversies in heparin monitoring. Am. J. Hematol. 2012, 87, S137–S140.
- Cuker, A.; Ptashkin, B.; Konkle, B.A.; Pipe, S.W.; Whinna, H.C.; Zheng, X.L.; Cines, D.B.; Pollak, E.S. Interlaboratory agreement in the monitoring of unfractionated heparin using the anti-factor Xa-correlated activated partial thromboplastin time. J. Thromb. Haemost. 2009, 7, 80–86.
- Raby, A.; Moffat, K.A.; Flynn, G.; Crowther, M.A.; Cuker, A. Interlaboratory variation in heparin monitoring: Lessons from the quality management program of ontario coagulation surveys. Thromb. Haemost. 2010, 104, 837–844.
- Favresse, J.; Lardinois, B.; Sabor, L.; Devalet, B.; Vandepapeliere, J.; Braibant, M.; Lessire, S.; Chatelain, B.; Jacqmin, H.; Douxfils, J.; et al. Evaluation of the DOAC-Stop® procedure to overcome the effect of DOACs on several thrombophilia screening tests. TH Open: Companion J. Thromb. Haemost. 2018, 2, e202–e209.
- 40. Kostousov, V.; Devaraj, S.; Bruzdoski, K.; Hensch, L.; Hui, S.; Teruya, J. C-reactive proteininduced activated partial thromboplastin time prolongation in heparinized samples is attenuated by elevated factor VIII. Int. J. Lab. Hematol. 2021, 43, 139–142.
- 41. Horton, S.; Augustin, S. Activated clotting time (ACT). In Haemostasis: Methods and Protocols; Monagle, P., Ed.; Humana Press: Totowa, NJ, USA, 2013; pp. 155–167.
- Martin, A.-C.; Kyheng, M.; Foissaud, V.; Duhamel, A.; Marijon, E.; Susen, S.; Godier, A. Activated clotting time monitoring during atrial fibrillation catheter ablation: Does the anticoagulant matter? J. Clin. Med. 2020, 9, 350.
- 43. Merli, G.J.; Groce, J.B. Pharmacological and clinical differences between low-molecular-weight heparins: Implications for pre-scribing practice and therapeutic interchange. Pharmacol. Ther. 2011, 35, 95–105.
- 44. Solari, F.; Varacallo, M. Low molecular weight heparin (LMWH). In StatPearls [Internet]; StatPearls Publishing, 2019. Available online: (accessed on 16 December 2020).
- 45. Kearon, C.; Akl, E.; Comerota, A.J.; Prandoni, P.; Bounameaux, H.; Goldhaber, S.Z.; Nelson, M.E.; Wells, P.S.; Gould, M.K.; Francesco, D.; et al. Antithrombotic Therapy for VTE Disease: Antithrombotic Therapy and Prevention of Thrombosis, 9th ed; American College of Chest Physicians Evidence-Based Clinical Practice Guidelines. Chest 2012, 141, e419S–e496S.
- 46. Lim, W.D.; Francesco, D.; Eikelboom, J.W.; Crowther, M.A. Meta-Analysis: Low-molecular-weight heparin and bleeding in patients with severe renal insufficiency. Ann. Intern. Med. 2006, 144, 673–684.

- 47. Seaman, S.; Nelson, A.; Noble, A. Cancer-associated thrombosis, low-molecular-weight heparin, and the patient experience: A qualitative study. Patient Prefer. Adherence 2014, 8, 453–461.
- van Veen, J.J.; Maclean, R.M.; Hampton, K.K.; Laidlaw, S.; Kitchen, S.; Toth, P.; Makris, M. Protamine reversal of low molecular weight heparin: Clinically effective? Blood Coagul. Fibrinolysis 2011, 22, 565–570.
- 49. Hirsh, J.; Raschke, R. Heparin and Low-Molecular-Weight heparin: The Seventh ACCP Conference on antithrombotic and thrombolytic therapy. Chest 2004, 126, 188S–203S.
- 50. Bounameaux, H.; De Moerloose, P. Is laboratory monitoring of low-molecular-weight heparin therapy necessary? No. J. Thromb. Haemost. 2004, 2, 551–554.
- 51. Toulon, P.; Smahi, M.; De Pooter, N. APTT therapeutic range for monitoring unfractionated heparin therapy. Significant impact of the anti–Xa reagent used for correlation. J. Thromb. Haemost. 2021.
- Hollestelle, M.J.; van der Meer, F.J.; Meijer, P. Quality performance for indirect Xa inhibitor monitoring in patients using international external quality data. Clin. Chem. Lab. Med. 2020, 58, 1921–1930.
- 53. Smahi, M.; De Pooter, N.; Hollestelle, M.J.; Toulon, P. Monitoring unfractionated heparin therapy: Lack of standardization of anti-Xa activity reagents. J. Thromb. Haemost. 2020, 18, 2613–2621.
- 54. Egan, G.; Ensom, M.H.H. Measuring anti-factor Xa activity to monitor low-molecular-weight heparin in obesity: A critical review. Can. J. Hosp. Pharm. 2015, 68, 33–47.
- 55. Crowther, M.; Lim, W. Low molecular weight heparin and bleeding in patients with chronic renal failure. Curr. Opin. Pulm. Med. 2017, 13, 409–413.
- 56. Thomas, O.; Lybeck, E.; Strandberg, K.; Tynngård, N.; Schött, U. Monitoring low molecular weight heparins at therapeutic levels: Dose-Responses of, and correlations and differences between aPTT, anti-factor Xa and thrombin generation assays. PLoS ONE 2015, 10, e0116835.
- 57. Al Dieri, R.; Alban, S.; Béguin, S.; Hemker, H.C. Fixed dosage of low-molecular-weight heparins causes large individual variation in coagulability, only partly correlated to body weight. J. Thromb. Haemost. 2005, 4, 83–89.
- al Dieri, R.; Alban, S.; Beguin, S.; Coenraad Hemker, H. Thrombin generation for the control of heparin treatment, comparison with the activated partial thrombo-plastin time. J. Thromb. Haemost. 2004, 2, 1395–1401.
- 59. Gionis, M.N.; Ioannou, C.V.; Katsamouris, A.N.; Katonis, P.; Balalis, K.; Sfyridaki, K.; Elalamy, I.; Gerotziafas, G.T. The study of the thrombin generation mechanism and the effect of low molecular weight heparin as thromboprophylaxis in patients undergoing total knee and hip replacement. Thromb. Res. 2013, 132, 685–691.

- 60. Duarte, R.C.F.; Ferreira, C.N.; Rios, D.R.A.; Dos Reis, H.J.; Carvalho, M.D.G. Thrombin generation assays for global evaluation of the hemostatic system: Perspectives and limitations. Rev. Bras. Hematol. Hemoter. 2017, 39, 259–265.
- 61. Donat, F.; Duret, J.P.; Santoni, A.; Cariou, R.; Necciari, J.; Magnani, H.; de Greef, R. The pharmacokinetics of fondaparinux sodium in healthy volunteers. Clin. Pharmacokinet. 2002, 41, 1–9.
- 62. Bauer, K.A. Fondaparinux: Dosing and adverse effects; Tirnauer, J.S., Ed.; UpToDate; 2019; Available online: (accessed on 18 December 2020).
- 63. Lassen, M.R.; Bauer, K.A.; Eriksson, B.I.; Turpie, A.G. Postoperative fondaparinux versus preoperative enoxaparin for prevention of venous thromboembolism in elective hip-replacement surgery: A randomised double-blind comparison. Lancet 2002, 359, 1715–1720.
- 64. Bijsterveld, N.R.; Moons, A.H.; Boekholdt, S.M.; van Aken, B.E.; Fennema, H.; Peters, R.J.; Meijers, J.C.; Buller, H.R.; Levi, M. Ability of recombinant factor VIIa to reverse the anticoagulant effect of the pentasaccharide fondapa-rinux in healthy volunteers. Circulation 2002, 106, 2550– 2554.
- Johnson, P.N.; Benefield, E.C.; Bui, P.-Y.N.; Gausman, J.N.; Marlar, R.A.; Gessouroun, M.R. Fondaparinux monitoring: Need for a local Fondaparinux-Calibrated anti–F actor Xa assay. J. Pediatr. Pharmacol. Ther. 2013, 18, 318–319.

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