

Vincristine-Induced Pediatric Peripheral Neuropathy

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Vincristine-induced peripheral neurotoxicity (VIPN) is a very common side effect of vincristine chemotherapy among pediatric patients with cancer.

vincristine

chemotherapy-induced peripheral neuropathy

pediatric cancer

molecular mechanisms

risk factors

prevention

treatment

1. Introduction

Vinka alkaloid (VAs) drugs (vincristine, vinblastine, vinorelbine, vindesine and vinflunine) are a class of microtubule-targeting agents that interfere with the continuous mitotic divisions and cell growth of cancer cells [\[1\]](#). Vincristine is one of the most used VAs in pediatric patients with cancer and it has been incorporated in several poly-chemotherapy regimens for acute lymphoblastic leukemia (ALL), lymphomas, neuroblastoma, sarcomas and central nervous system tumors. However, neurotoxicity is a severe and dose-limiting side effect of vincristine and it may produce delay or discontinuation of the treatment. Vincristine may cause peripheral, progressive (almost distally to proximally) and symmetric nerve damage, due to microtubule structure disruption, inflammatory processes and axonal dysfunction [\[2\]](#).

2. Pharmacokinetics and Pharmacodynamics of Vincristine

Vincristine acts as an inhibitor of the treadmilling process, by the link to the tubulins, which prevents the formation of the microtubules and consequently of the mitotic spindle. In this way, cell division is blocked and the cell dies [\[3\]](#). In particular, vincristine and the other VAs interact with tubulin dimers by binding to approximately sixteen–seventeen different specific binding sites, known as the vinca domain [\[4\]](#). Once the binding site is reached, VAs have a concentration-dependent action: at low concentrations, VAs prevent microtubules from elongating, by binding to the + end and preventing GTP from binding to β -tubulin [\[5\]](#); at high concentrations, they promote the depolarization of microtubules, as explained in [Figure 1](#).

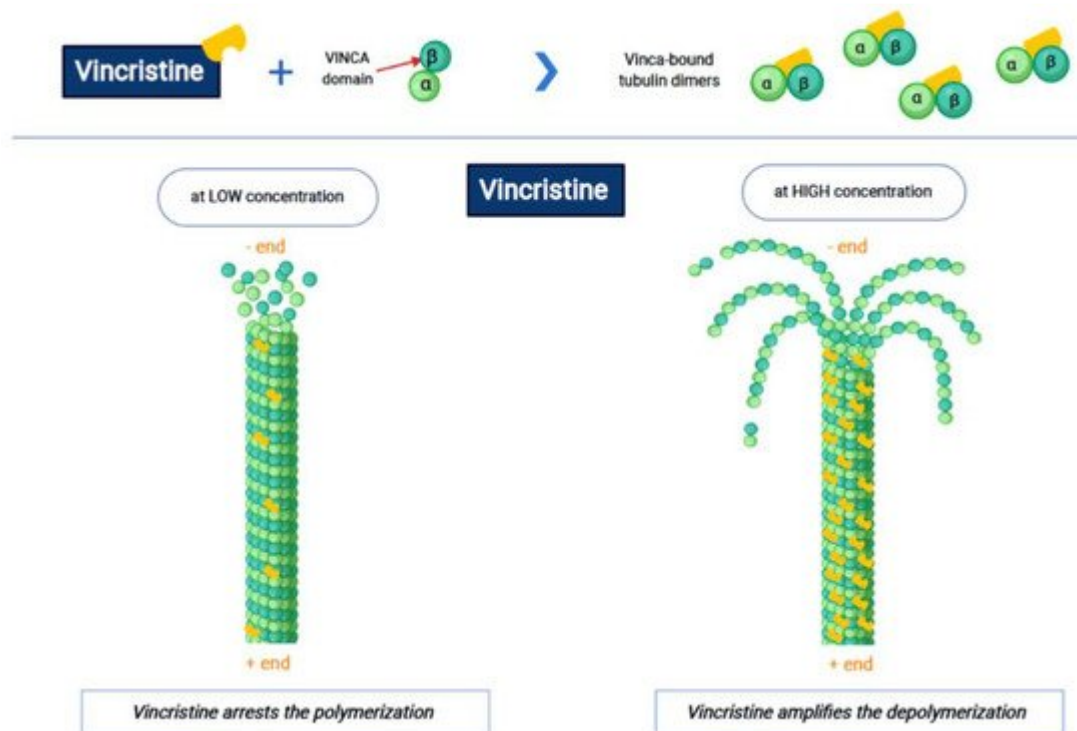


Figure 1. The first part of the figure shows the interaction between vincristine and the tubulin dimers. The second part shows the vincristine concentration-dependent action: the arrest of polymerization at low dose and amplification of depolymerization at a high dose.

Once the formation of the mitotic spindle is blocked, the cell goes into apoptosis with or without p53 activation [6]. The VAs damage does not only concern the mitosis, but also all the other processes that involve the microtubules, such as the inhibition of axon transport, secretion processes, structure disorders and impairment of platelet functions [7]. Vincristine has poor oral bioavailability and is administered intravenously as vincristine sulfate, which is a vesicant. After intravenous administration, vincristine rapidly distributes extensively into most body tissues, with poor penetration across the blood–brain barrier (BBB) and into the central nervous system (CNS). However, it is very neurotoxic and fatal if administered intrathecally, because it produces quickly serious leptomeningitis and ventriculitis [8]. Vincristine metabolism is performed in the liver by the cytochrome p450 CYP3A enzyme system, particularly by CYP3A4 and CYP3A5. Vincristine has a half-life of 85 h and it is eliminated primarily via the biliary route and excreted in the feces; consequently, great attention should be paid in the presence of hyperbilirubinemia [1]. The kidney eliminates a very small amount of the drug [9]. Vincristine has little myelosuppressive effects and is usually given even to leukopenic and thrombocytopenic patients.

3. Pathogenesis of VIPN

The pathogenesis of VIPN is strictly connected to the mechanisms through which vincristine carries out its antineoplastic function. As previously mentioned, vincristine acts primarily by altering the normal assembly and disassembly function of microtubules, with consequent mitosis block and cell death. In addition to this, all other activities that involve microtubules are inevitably compromised. Inside the neurons, microtubules are not involved

in the constitution of the mitotic spindle, since these cells are not in active replication. Microtubules are abundant in neurons, because they make up the skeleton of axons and dendrites, giving them their specialized morphologies [1][10]. In addition to its direct structural and functional damage of nerve cells, vincristine also induces the expression of integrins (immune markers) on the surface of endothelial cells, allowing macrophages to express the CX3CR receptor for the adhesion to the endothelium and the migration into the nervous tissue. This process causes the production of reactive oxygen species (ROS), which act as a chemical mediator for immune-neuronal communication by activating transient receptor potential TRPA1 channels (functionally expressed by the axons of sensory neurons) and evoking pain [11]. Three mechanisms of neurological damage influence and amplify each other, strongly conditioned by patient-related risk factors and treatment-related risk factors, as we explain below. [Figure 2](#) synthesizes the VIPN pathogenesis.

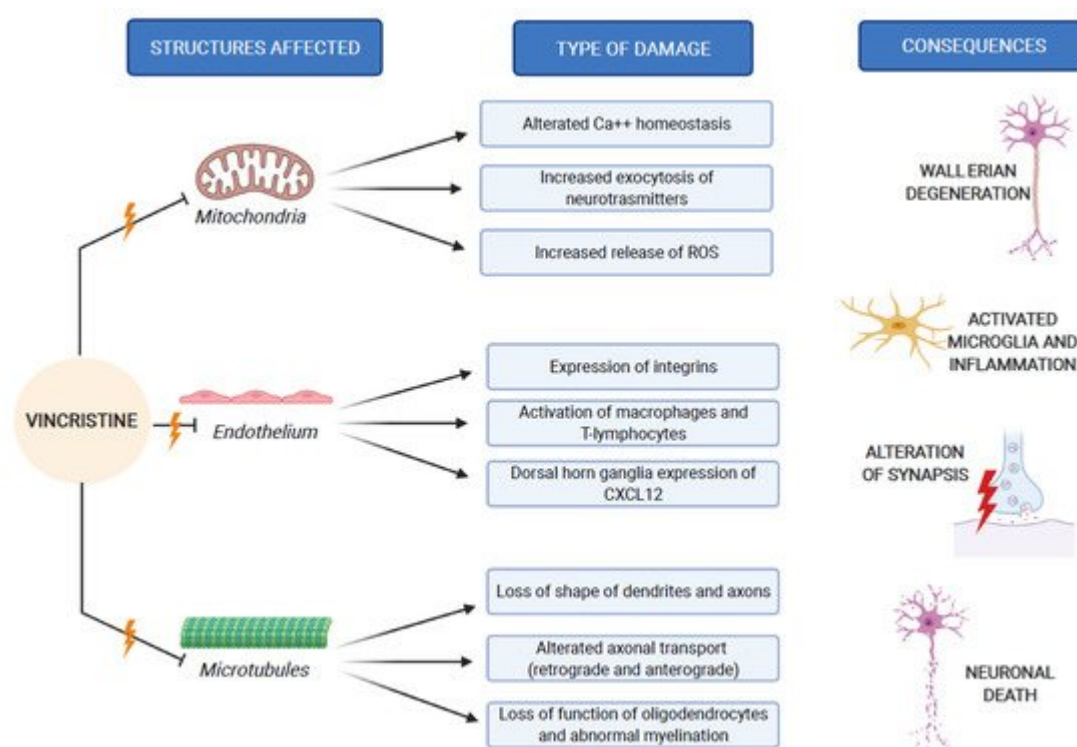


Figure 2. VIPN pathogenesis, which involves mitochondria, endothelium and microtubules of nerve cells.

4. Patient-Related Risk Factors for VIPN

Age, sex and ethnicity represent the three patient-related factors that may influence the occurrence of VIPN. Younger children may be at greater risk of VIPN. Of the 58 drafts evaluated by van de Velde et al., no clear evidence was identified regarding the influence of gender on the incidence of VIPN, while ethnicity seemed to play a significant role [12]. Ethnicity can influence the functionality of the cytochrome p450 3A (CYP3A) family, which plays a main role in the metabolism of vincristine. As evidenced by Madsen et al., other gene polymorphisms may influence the occurrence of VIPN [13]. Among these, CEP72 anomalies seem to facilitate the occurrence of VIPN, as described in several studies, although exhaustive data are not yet available [14][15].

5. Treatment-Related Risk Factors for VIPN

Prolonged treatments and higher single doses of vincristine seem to be related to increased occurrence and severity of VIPN in adult patients, providing validation for a maximum vincristine dose of 2 mg [13][16]. The studies about the association between dose and VIPN in pediatric patients are lacking conclusive results and greater vincristine doses are not exactly associated with peripheral neuropathy in pediatric patients. However, the recommended dose is 0.05–0.065 mg/kg in infants and 1.5 mg/sqm in children, with a maximum of 2 mg/dose and a minimum of a week interval between each dose [12][17]. [Table 1](#) summarizes the treatment-related risk factors for VIPN in children.

Table 1. Treatment-related risk factors for VIPN in children.

Risk Factor	Mechanism	Reference
Dose	Higher dose of vincristine could facilitate VIPN	[18][16]
Administration schedule	Dosing closely may increase the risk of VIPN	[19]
Method of administration	Prolonged administration could reduce the risk of VIPN compared to IV bolus administration	[18]
Concomitant use of other drugs	Azoles, aprepitant and fosaprepitant inhibit CYP3A4, increasing the risk of VIPN	[20][21][22][23][24][25][26][27]

6. Strategies for Prevention and Treatment of VIPN

[Table 2](#) summarizes the principal drugs studied in clinical trials for the prophylaxis and treatment of VIPN in pediatric oncologic patients.

Table 2. Principal drugs investigated for prophylaxis and treatment of VIPN in children.

Drug	Dosage	Patients Age (in Years)	Outcome	Reference
Gabapentin	5–10 mg/kg/day (max 50–70 mg/kg/day)	1–18	No evidence of superiority over opioids for reducing or preventing VIPN pain	[28]
Pyridoxine	150 mg/sqm/day	2–13	Complete resolution of symptoms of VIPN	[29]
Pyridostigmine	3 mg/kg/day	2–13	Complete resolution of symptoms of VIPN	[29]
Pyridoxine and Pyridostigmine	150 mg/sqm/day and 3 mg/kg/day	2–10	Significantly improvement of symptoms	[30]

Drug	Dosage	Patients Age (in Years)	Outcome	Reference
Glutamic acid	250 mg daily for BSA * < 1 sqm, 500 mg daily for BSA ≥ 1 sqm		Prevention of VIPN only in patients aged 13 years or more	[31]
	1.5 g daily (on the day before or on the day of the first dose of VCR)	3–18	Reduced occurrence of VIPN	[32]
Glutamine	6 g/sqm twice daily for 21 days	5–21	Improvement in sensory function and QoL **	[33]

* BSA: body surface area. ** QoL: quality of life.

7. Summary

VIPN is a common side effect of vincristine treatment in pediatric oncologic patients and its pathogenesis seems to be multifactorial, related to patient-related risk factors (age, race, ethnicity and genetic polymorphisms) and treatment-related risk factors (dose, time of infusion and drug–drug interactions). The recognition of risk factors would allow clinicians the prompt identification of patients at higher risk, helping clinicians to manage them in the most appropriate and personalized way. Several molecules (such as gabapentin, pyridoxine and pyridostigmine, glutamic acid and glutamine) have been investigated for the prophylaxis and/or treatment of VIPN in pediatric oncologic patients. Furthermore, there is a lack of consensus about guidelines for the management of VIPN in this setting of patients, and future studies are required for evaluating novel preventive and therapeutic approaches.

References

1. Martino, E.; Casamassima, G.; Castiglione, S.; Cellupica, E.; Pantalone, S.; Papagni, F.; Rui, M.; Siciliano, A.M.; Collina, S. Vinca alkaloids and analogues as anti-cancer agents: Looking back, peering ahead. *Bioorg. Med. Chem. Lett.* 2018, 28, 2816–2826.
2. Carlson, K.; Ocean, A.J. Peripheral neuropathy with microtubule-targeting agents: Occurrence and management approach. *Clin. Breast Cancer* 2011, 11, 73–81.
3. Liu, Y.M.; Chen, H.L.; Lee, H.Y.; Liou, J.P. Tubulin inhibitors: A patent review. *Expert Opin. Ther. Patents* 2014, 24, 69–88.
4. Cormier, A.; Knossow, M.; Wang, C.; Gigant, B. The binding of vinca domain agents to tubulin: Structural and biochemical studies. *Methods Cell Biol.* 2010, 95, 373–390.

5. Pellegrini, F.; Budman, D.R. Review: Tubulin function, actions of antitubulin drugs, and new drug development. *Cancer Investig.* 2005, 23, 264–273.
6. Lobert, S.; Fahy, J.; Hill, B.T.; Duflos, A.; Etievant, C.; Correia, J.J. Vinca alkaloid-induced tubulin spiral formation correlates with cytotoxicity in the leukemic L1210 cell line. *Biochemistry* 2000, 39, 12053–12062.
7. Beck, W.T.; Cass, C.E.; Houghton, P.J. Microtubule-targeting anticancer drugs derived from plants and microbes: Vinca alkaloids, taxanes and epothiolones. In *Holland-Frei Cancer Medicine*, 5th ed.; Bast, R.C., Kufe, D.W., Pollock, R.E., Eds.; BC Decker Inc.: New York, NY, USA, 2003; pp. 680–698.
8. Triarico, S.; Maurizi, P.; Mastrangelo, S.; Attinà, G.; Capozza, M.A.; Ruggiero, A. Improving the brain delivery of chemotherapeutic drugs in childhood brain tumors. *Cancers* 2019, 11, 824.
9. Ruggiero, A.; Ferrara, P.; Attinà, G.; Rizzo, D.; Riccardi, R. Renal toxicity and chemotherapy in children with cancer. *Br. J. Clin. Pharmacol.* 2017, 83, 2605–2614.
10. Baas, P.W.; Rao, A.N.; Matamoros, A.J.; Leo, L. Stability properties of neuronal microtubules. *Cytoskeleton* 2016, 73, 442–460.
11. Old, E.A.; Nadkarni, S.; Grist, J.; Gentry, C.; Bevan, S.; Kim, K.W.; Mogg, A.J.; Perretti, M.; Malcangio, M. Monocytes expressing CX3CR1 orchestrate the development of vincristine-induced pain. *J. Clin. Investig.* 2014, 124, 2023–2036.
12. Van de Velde, M.E.; Kaspers, G.L.; Abbink, F.C.H.; Wilhelm, A.J.; Ket, J.C.F.; van den Berg, M.H. Vincristine-induced peripheral neuropathy in children with cancer: A systematic review. *Crit. Rev. Oncol. Hematol.* 2017, 114, 114–130.
13. Madsen, M.L.; Due, H.; Ejlskjær, N.; Jensen, P.; Madsen, J.; Dybkær, K. Aspects of vincristine-induced neuropathy in hematologic malignancies: A systematic review. *Cancer Chemother. Pharmacol.* 2019, 84, 471–485.
14. Stock, W.; Diouf, B.; Crews, K.R.; Pei, D.; Cheng, C.; Laumann, K.; Mandrekar, S.J.; Luger, S.; Advani, A.; Stone, R.M.; et al. An inherited genetic variant in CEP72 promoter predisposes to vincristine-induced peripheral neuropathy in adults with acute lymphoblastic leukemia. *Clin. Pharmacol. Ther.* 2017, 101, 391–395.
15. Gutierrez-Camino, A.; Martin-Guerrero, I.; Lopez-Lopez, E.; Echebarria-Barona, A.; Zabalza, I.; Ruiz, I.; Guerra-Merino, I.; Garcia-Orad, A. Lack of association of the CEP72 RS924607 TT genotype with vincristine-related peripheral neuropathy during the early phase of pediatric acute lymphoblastic leukemia treatment in a Spanish population. *Pharmacogenet. Genom.* 2016, 26, 100–102.
16. Kanbayashi, Y.; Hosokawa, T.; Okamoto, K.; Konishi, H.; Otsuji, E.; Yoshikawa, T.; Takagi, T.; Taniwaki, M. Statistical identification of predictors for peripheral neuropathy associated with

administration of bortezomib, taxanes, oxaliplatin or vincristine using ordered logistic regression analysis. *Anticancer Drugs* 2010, 21, 877–881.

17. Lavoie Smith, E.M.; Li, L.; Chiang, C.; Thomas, K.; Hutchinson, R.J.; Wells, E.M.; Ho, R.H.; Skiles, J.; Chakraborty, A.; Bridges, C.M.; et al. Patterns and severity of vincristine-induced peripheral neuropathy in children with acute lymphoblastic leukemia. *J. Peripher. Nerv. Syst.* 2015, 20, 37–46.
18. Flatters, S.J.L.; Bennett, G.J. Studies of peripheral sensory nerves in paclitaxel-induced painful peripheral neuropathy: Evidence for mitochondrial dysfunction. *Pain* 2006, 122, 247–257.
19. Stadelmann, C.; Timmler, S.; Barrantes-Freer, A.; Simons, M. Myelin in the central nervous system: Structure, function, and pathology. *Physiol. Rev.* 2019, 99, 1381–1431.
20. Nikanjam, M.; Sun, A.; Albers, M.; Mangalindin, K.; Song, E.; Vempaty, H.; Sam, D.; Capparelli, E.V. Vincristine-associated neuropathy with antifungal usage: A Kaiser Northern California experience. *J. Pediatr. Hematol. Oncol.* 2018, 40, 273–277.
21. Ruggiero, A.; Arena, R.; Battista, A.; Rizzo, D.; Attinà, G.; Riccardi, R. Azole interactions with multidrug therapy in pediatric oncology. *Eur. J. Clin. Pharmacol.* 2013, 69, 1–10.
22. Moriyama, B.; Henning, S.A.; Leung, J.; Falade-Nwulia, O.; Jarosinski, P.; Penzak, S.R.; Walsh, T.J. Adverse interactions between antifungal azoles and vincristine: Review and analysis of cases. *Mycoses* 2012, 55, 290–297.
23. Thackray, J.; Spatz, K.; Steinherz, P.G. Vincristine toxicity with co-administration of fluconazole: Long-term concerns. *Pediatr. Blood Cancer* 2017, 64, 12.
24. Bellmann, R.; Smuszkiewicz, P. Pharmacokinetics of antifungal drugs: Practical implications for optimized treatment of patients. *Infection* 2017, 45, 737–779.
25. Van Schie, R.M.; Brüggemann, R.J.; Hoogerbrugge, P.M.; te Loo, D.M. Effect of azole antifungal therapy on vincristine toxicity in childhood acute lymphoblastic leukaemia. *J. Antimicrob. Chemother.* 2011, 66, 1853–1856.
26. Pana, Z.D.; Roilides, E. Risk of azole-enhanced vincristine neurotoxicity in pediatric patients with hematological malignancies: Old problem–new dilemma. *Pediatr. Blood Cancer* 2011, 57, 30–35.
27. Ruggiero, A.; Rizzo, D.; Catalano, M.; Coccia, P.; Triarico, S.; Attinà, G. Acute chemotherapy-induced nausea and vomiting in children with cancer: Still waiting for a common consensus on treatment. *J. Int. Med. Res.* 2018, 46, 2149–2156.
28. Anghelescu, D.L.; Tesney, J.M.; Jeha, S.; Wright, B.B.; Trujillo, L.; Sandlund, J.T.; Pauley, J.; Cheng, C.; Pei, D.; Pui, C.H. Prospective randomized trial of interventions for vincristine-related neuropathic pain. *Pediatr. Blood Cancer* 2020, 67, 28539.

29. Akbayram, S.; Akgun, C.; Doğan, M.; Sayin, R.; Caksen, H.; Oner, A.F. Use of pyridoxine and pyridostigmine in children with vincristine-induced neuropathy. *Indian J. Pediatr.* 2010, 77, 681–683.
30. Aydin Köker, S.; Gözmen, S.; Demirağ, B.; Ünalp, A.; Karapinar, T.H.; Oymak, Y.; Gürbüz, G.; Öner, E.İ.; Vergin, R.C. Effect of pyridoxine plus pyridostigmine treatment on vincristine-induced peripheral neuropathy in pediatric patients with acute lymphoblastic leukemia: A single-center experience. *Neurol. Sci.* 2021.
31. Bradfield, S.M.; Sandler, E.; Geller, T.; Tamura, R.N.; Krischer, J.P. Glutamic acid not beneficial for the prevention of vincristine neurotoxicity in children with cancer. *Pediatr. Blood Cancer* 2015, 62, 1004–1010.
32. Mokhtar, G.M.; Shaaban, S.Y.; Elbarbary, N.S.; Fayed, W.A. A trial to assess the efficacy of glutamic acid in prevention of vincristine-induced neurotoxicity in pediatric malignancies: A pilot study. *J. Pediatr. Hematol. Oncol.* 2010, 32, 594–600.
33. Sands, S.; Ladas, E.J.; Kelly, K.M.; Weiner, M.; Lin, M.; Ndao, D.H.; Dave, A.; Vahdat, L.T.; Bender, J.G. Glutamine for the treatment of vincristine-induced neuropathy in children and adolescents with cancer. *Support Care Cancer* 2017, 25, 701–708.

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