

Immunotherapy for Cutaneous Malignant Melanoma

Subjects: [Dermatology](#) | [Pharmacology & Pharmacy](#)

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Malignant melanoma (mM) is the leading cause of death among cutaneous malignancies. While its incidence is increasing, the most recent cancer statistics show a small but clear decrease in mortality rate. This trend reflects the introduction of novel and more effective therapeutic regimens, including the two cornerstones of melanoma therapy: immunotherapies and targeted therapies. Unlike chemotherapies or radiation, in which the therapy directly induces cancer cell death, immunotherapies stimulate the patient's immune system to control and eliminate the tumor. Advantages of immunotherapies over traditional cancer treatments include increased durability for long-term control or even cure and more precisely targeted anti-tumor activity that spares healthy tissues, many times with comparable or even reduced overall toxicity. The high immunogenicity and somatic mutation burden of melanoma likely contribute to the success of immunotherapy. Treatments combining immunotherapies with targeted therapies, which disable the carcinogenic products of mutated cancer cells, have further increased treatment efficacy and durability. Toxicity and resistance, however, remain critical challenges to the field. There are three types of immunotherapies currently approved by the US Food and Drug Administration (FDA) for the treatment of advanced melanoma: (1) T-cell stimulating cytokines (i.e. interferon (IFN)- α 2b and interleukin-2 (IL-2)); (2) T-cell exhaustion-mitigating immune checkpoint inhibitors (ICI); and (3) a dendritic cell (DC)-activating oncolytic virus (T-VEC). Still others, such as adoptive cell transfer (ACT), hold strong promise for the future.

metastatic melanoma

targeted therapy

immunotherapy

combined therapies

tumor relapse

tumor resistance

1. Early Immunotherapies

1.1. Interferon- α 2b

Interferon (IFN)- α 2b is a recombinant form of human IFN- α with antiviral and antitumor properties. It was the first immunotherapy approved for melanoma, first as an adjuvant treatment in 1996 and then as first-line therapy in 1998. By binding to IFN receptors 1 and 2, the drug triggers multiple dose- and time-dependent immunostimulatory effects, including upregulation of major histocompatibility complex 1 (MHC1) on tumor cells, enhanced activation of anti-tumor cytotoxic T lymphocytes (CTLs), depression of T regulatory cells (Tregs), enhanced dendritic cell (DC) response, and decreased intercellular adhesion molecule (ICAM) expression ^{[1][2][3]}. In 1996, high-dose (HD) IFN- α 2b became the first adjuvant therapy, approved for use in stage IIB and III melanoma patients following surgical resection. Initial trials demonstrated significantly improved 5-year relapse-free survival (RFS) (37% vs 26%) ^{[4][5]}.

HD IFN- α 2b treatment is limited by high toxicity, with studies reporting dose reductions in 28-55% of patients and toxicity attrition rates of 10-26% (6). HD IFN- α 2b remained the standard adjuvant therapy for high-risk melanoma until ipilimumab approval in 2015

Peginterferon- α 2b (Peg-IFN), which has a longer half-life than IFN- α 2b, was approved for adjuvant use in 2011 after demonstrating significant improvement in 7-year RFS compared to observation (39.1% vs 34.6%) but, like HD IFN- α 2b, could not provide OS benefit [6][7].

Low dose (LD) IFN- α 2b was approved as first-line therapy for stage II melanoma patients in 1998 based on a trial showing improved 5-year RFS (43% vs 51%) and a trend toward improved OS (24% vs 32%) compared to observation [8]. It does not have significant clinical benefit in mM [9].

Today, IFN- α is no longer a first-line agent for most patients; however, it may still have utility as an auxiliary immunostimulatory agent, enhancing the clinical benefits of other immunotherapies.

1.2 High Dose Interleukin-2

Interleukin-2 (IL-2) is a T-cell growth factor that leads to cytokine production and preferential expansion of CD8+ T-cells, NK cells, and Tregs. In 1998, HD intravenous (IV) administration of IL-2 became the first FDA-approved immunotherapy for the treatment of metastatic melanoma (mM) [10][11]. Durable tumor responses have been well documented in a subset of mM patients, with 5-10% of patients achieving complete response and even more achieving increased disease stability [10][11][12]. A recent one is of IL-2-responsive mM patients who exclusively received HD IL-2 for systemic therapy confirms prolonged clinical and survival benefits [13]. As with IFN- α therapy, the use of HD IL-2 treatment is limited by the relatively high incidence of grade 3 and 4 toxicities, which requires the drug to be administered in an intensive inpatient setting [14]. The efficacy of treatment is further limited by the drug's activation of anti-inflammatory Tregs, which limit CD8+ activation and effector functions. Drugs targeting specific subunits of the IL-2 receptor, such as the recombinant IL-2 receptor $\beta\gamma$ -biased agonist NKTR-214 (Bempegaldesleukin), have shown promise in the targeted expansion of anti-tumor T and NK cells with limited expansion of Tregs and dramatically reduced toxicity [15][16][17][18][19][20].

While rarely used as a single or first-line agent today, HD IL-2 remains a second- or third-line option that provides a possible survival benefit to patients who have failed treatment with first-line agents [21]. Many trials combining HD or low-dose IL-2 therapy with additional therapies are ongoing.

2. Immune Checkpoint Inhibitors

2.1. Cytotoxic T lymphocyte-associated antigen 4 (CTLA4) inhibitors

CTLA4 is an immune-inhibitory molecule expressed on the surface of activated T-cells. Together with its immune-activating counterpart, CD28, CTLA4 creates a critical immune checkpoint that must be overcome to achieve a durable immune response [22]. CTLA4 is naturally upregulated in situations of chronic T-cell stimulation to prevent

uncontrolled immune reactions and inappropriate development of autoimmunity. In the TME, however, this system backfires: chronic presentation of tumor antigens to T-cells inhibits the immune system from mounting an anti-tumor immune response and contributes to the immune evasion that allows continued tumor growth [23][24][25][26].

Ipilimumab was approved as the first immune checkpoint inhibitor (ICI) in 2011, the same year that vemurafenib was approved to block BRAF-mediated growth signaling. Ipilimumab is an anti-CTLA4 human IgG antibody. By preventing the interaction of CTLA4 and its ligands, the drug allows T-cells to bypass the inhibitory immune checkpoint and mount a response against tumor antigens. Phase III trials of previously-treated mM patients demonstrated improved OS compared to gp100, a melanoma antigen immunostimulant with limited anti-tumor effects (10.1 vs 6.4 months, $p = 0.0026$) [27]. A metanalysis of pooled data from nearly 2,000 mM patients treated with ipilimumab (both pre-treated and treatment-naïve) reported an increase in the 3-year OS rate to 22% (95% CI [20, 24%]), a dramatic increase from ~5% achieved by previous standard-of-care therapies [28]. Perhaps even more importantly, the OS survival curve plateaued after 3 years, maintaining the ~20% OS rate for the entirety of the 10+ year follow-up [28]. Thus, ipilimumab became both the first therapy to provide an OS benefit in advanced melanoma and the first to demonstrate that long-term durable mM disease control is possible with systemic therapy [29].

While responses to ipilimumab are durable, the response rates are low, ranging from 5-10%. Clinical trials have provided little insight into possible biomarkers of response. Attempts to improve response rates by adding ipilimumab to dacarbazine therapy were somewhat successful (15% vs 10%) and demonstrated a survival benefit over dacarbazine alone (OS 11.2 vs 9.1 months, $p < 0.001$). However, these benefits came at the cost of high toxicity (rate of grade 3/4 AEs: 56.3% vs 27.5%, $p < 0.001$) [30]. Even as a monotherapy, ipilimumab is relatively toxic with immune-related toxicities occurring in 60-80% of patients, 10-26% of which are grade 3/4 reactions [31]. Perhaps unsurprisingly, severe AEs, which are often immune-related AEs (irAE), were found to be associated with improved ORRs [32]. Ipilimumab is still the only approved CTLA4 inhibitor for mM, though ipilimumab monotherapy is not a first-line therapy by ASCO guidelines [33].

2.2. Programmed cell death protein 1 (PD-1) and PD-1 ligand (PD-L1) inhibitors

Like CTLA4, PD-1 is an inhibitory immune checkpoint receptor expressed by activated T cells. When PD-1 binds its receptors, PD-L1 and PD-L2, signaling through the SHP1/2 pathway downregulates the transcription factors necessary for T-cell effector functions, growth, and survival [34]. In healthy tissues, PD-L1 is broadly expressed and upregulated in response to proinflammatory cytokines [35]. Melanoma tumor and TME cells upregulate PD-L1 in response to tumor-infiltrating lymphocytes (TIL), suggesting that PD-L1 expression is used as a mechanism of immune evasion by the cancerous cells [36][37][38].

In 2014, two anti-PD-1 monoclonal antibodies, Pembrolizumab and nivolumab, were approved for treatment-resistant mM after demonstrating superiority over ipilimumab. Early trials of pembrolizumab monotherapy demonstrated improved 6- and 12-month PFS and RR (6-month PFS: pembrolizumab=47% vs ipilimumab=26%; 12-month PFS: P=74-68% vs I=58%; RR= P=33%, I=12%) [39][40]. Two and five-year follow-up studies and real-world findings of pembrolizumab monotherapy confirm its superior OS and durable antitumor immune activity for

both treatment naïve and pre-treated mM patients [41][42][43][44][45]. Similarly, the CheckMate 067 trial first demonstrated that nivolumab monotherapy confers a significantly greater PFS compared to ipilimumab in treatment-naïve mM patients (nivolumab: 6.9 months 95%[4.3, 9.5]; ipilimumab: 2.9 months 95%[2.8, 3.4] [46]. Follow-up data from 2019 then demonstrated superior 5-year OS rates (nivolumab = 44%, ipilimumab = 26%) [47]. Nivolumab has proven to be effective in a range of melanoma tumor subtypes, including both treatment-naïve and pre-treated tumors with either WT or mutant BRAF status [48][49].

The two PD-1 inhibitors differ by epitope binding location and target affinity strength but are equally effective as monotherapies by OS (pembrolizumab = 22.6 mo, nivolumab = 23.9 mo, $p = 0.91$) and time to next-line therapy or death (pembrolizumab = 15.7 mo, nivolumab = 10.8 mo, $p = 0.16$) [50][51]. Both are also relatively well-tolerated with lower rates of grade 3/4 toxicity (14% with pembrolizumab and 4% with nivolumab) than chemotherapy, ipilimumab, and most targeted therapies [52][50]. AEs during nivolumab therapy are associated with improved ORRs [50].

Both PD-1 inhibitors are also effective in the adjuvant setting. A five-year one on adjuvant pembrolizumab demonstrated significant increased RFS, decreased risk of distant metastasis or death (HR 0.60 95%[0.49,0.73]), and sustained treatment effect compared to placebo [51][53]. Interestingly, adjuvant pembrolizumab also proved efficacious in patients with PD-L1-negative and undetermined tumors [39][53]. In pre-treated stage IV melanoma patients with no evidence of residual disease, adjuvant nivolumab alone or in combination with ipilimumab proved similarly effective in increasing RFS compared to placebo [54]. These drugs are the current first-line adjuvant therapy for resected WT melanoma. Patients with resected BRAF-mutant melanomas may choose between pembrolizumab, nivolumab, or dabrafenib-trametinib combination therapy as first-line adjuvant therapy [33].

Optimal utilization of ICIs is hindered by several major challenges, including resistance and poor predictability of patient response. Approximately 30% of melanoma patients have an innate resistance to PD-1 inhibitors and 25% of responders acquire resistance during treatment [55][56][57]. CTLA4 inhibitors face a similar challenge [27]. Mechanisms of resistance likely include specific tumor cell genetics (loss-of-function mutations in *JAK1/2* [58]), differing expression levels of tumor cell surface proteins (for example, MHC I expression [59], alternate ICIs [60], and epigenetic T-cell changes limiting effector function and memory [61]). Efforts to increase durability by combining ICIs with auxiliary agents such as PEG-IFN [62] or hydroxychloroquine [63] have had mixed results, with none providing a clear clinical benefit. Unfortunately, increased toxicity often outweighs any benefit to durability or RR that auxiliary agents provide.

A few studies have identified markers associated with more successful clinical outcomes. For example, independent biomarkers associated with favorable OS of mM patients treated with pembrolizumab include high relative eosinophil count (>1.5%), high relative lymphocyte count (>17.5%), and absence of non-soft tissue or lung metastasis. Patients meeting none of these criteria have a poor prognosis with pembrolizumab [64]. Others have identified that the occurrence of immune-mediated AEs may be associated with better ORR, OS, and PFS with nivolumab and ipilimumab monotherapies but not with pembrolizumab [65]. Another—albeit much smaller ($n = 40$)—it was also found that PD-L1 expression on circulating tumor cells may also be a predictive biomarker for PD-1 inhibitor response, suggesting that liquid biopsy may provide clinically relevant information during treatment

selection [66]. However, subgroup analyses have demonstrated the PD-1 blockade still provides clinical benefits in PD-1 negative tumors [46]. Conflicting evidence on the subject makes using tumor PD-L1 expression as a predictive marker for PD-L1 inhibitor response or overall prognosis for mM controversial [67][68].

2.3. Combination ICI Therapy

Combining CTLA4 and PD-1 blockade is more effective than either class in monotherapy [69][70], yet carries a significantly higher risk of severe toxicity. As monotherapies, nivolumab and ipilimumab have grade 3/4 AE rates of 16-27% and 27%, respectively. When used together, this rate increases to 55-71% [46][54]. Reducing toxicity while maintaining the clinical benefit of combination therapy may be possible with alternative dosing strategies. Regimens of standard-dose pembrolizumab (200 mg) with either 150mg or 50 mg reduced-dose ipilimumab show a meaningful reduction in toxicity (grade 3-5 toxicity rate <26%) without a significant reduction in ORR (PEM200+IPI50: ORR, 55%, and CR, 16%; PEM200+IPI100: ORR, 61%, and CR, 25%). In fact, 12-month PFS and OS rates are actually higher with these regimens (12-mo PFS: 65% for PEM200+IPI50; 82% for PEM200+IPI100; OS: >90% for both) compared to standard dosage and previous alternative dosages (12-mo PFS: 46-53% with standard dosing, 47% and 68% with alternative dosing; 12-mo OS: 73-89%) [71][72][73][74][75]. Larger trials are still necessary.

2.4. Novel immune checkpoint inhibitors

The second generation of PD1 and CTLA4 ICIs are emerging. These new agents include anti-PD1 antibody HX008 [75], anti-PDL1 monoclonal antibody LP002 (NCT04756934), anti-CTLA4 antibody ONC-392 [76]. Lymphocyte activation gene 3 (LAG3) is another T-cell inhibitory checkpoint receptor, the upregulation of which may be a resistance mechanism to PD1 inhibition therapy [77]. Another promising ICI target is TIM-3 (T-cell immunoglobulin and mucin domain 3). TIM-3 blockade restores anti-tumor functions in *ex vivo* of previously exhausted NK and effector T-cells [78] and enhances cancer vaccine-induced antitumor responses in murine melanoma models [79]. A bispecific anti-PD-1 and TIM-3 antibody (RO7121661/RG7769) demonstrated superior anti-tumor TIL activity, IFN- γ secretion, and tumor growth control compared to the monospecific PD-1 antibody in mouse models [80]. The agent has recently entered phase I human trials (NCT03708328).

3. Oncolytic virus therapy

Cancer cells achieve a neoplastic phenotype by genetic and epigenetic mutations. These mutations, however, impair signaling pathways (*RAS*, *WNT*, *PTEN*, *RB1*, *TP53*) that are essential for the intra-cellular antiviral response [81]. Recent advances in genetic modification, such as CRISPR, have allowed researchers to create anti-neoplastic viruses that exploit the vulnerability of mutated cells to viral infection while sparing healthy cells [82].

Talimogene laherparepvec (T-VEC), an oncolytic human herpes simplex virus 1(HSV-1), is the first and only oncolytic virus (OV) approved for metastatic and unresectable melanoma. When T-VEC is injected directly into the tumor site, it promotes the secretion of granulocyte-macrophage colony-stimulating factor (GM-CSF) to activate

DCs and increase tumor antigen presentation to T-cells. In the phase 3 OPTiM trial, 64% of directly injected and 34% of uninjected non-visceral lesions decreased in size by >50%. Complete resolution of lesions occurred in 47% of injected lesions, 22% of non-injected non-visceral lesions, and 9% of non-injected visceral lesions. When compared to recombinant GM-CSF administration, T-VEC demonstrated higher durable RR (16% vs 2.1%, $p = 0.001$), ORR (26% vs 5.7%), and OS (23.3 months, $p = 0.051$). Severe toxicity rates were only 2% [83]. Laboratory evidence shows that T-VEC has increased efficacy in melanomas with INF γ -JAK-STAT pathway mutations [84]. Since dysregulation of INF γ is a common mechanism of resistance to ICI therapy, ongoing trials are investigating T-VEC as a salvage or combination therapy (NCT04330430, NCT04068181).

Systemic administration of OV therapy is also being explored. However, maintaining viral titers capable of generating an anti-tumor response after systemic administration has proved challenging to systemic OV monotherapy [85][86]. Trials are also investigating their role as sensitizing agents or within combination immunotherapies. Systemic OVs may still have a role as priming agents or within combination therapy (NCT04152863).

4. The Future of Melanoma Immunotherapy

Over the past 8 years, immunotherapy has revolutionized the treatment of mM, offering patients more treatment options with higher efficacy and less toxicity. Two-year overall survival rates have risen dramatically from ~10% to ~60% [47]. Further into the identification of melanoma neoantigens and their immunogenic potentials is essential for the advancement of the field. The ability to create individualized therapies specific to each patient's tumor and immune landscape has the potential to revolutionize melanoma therapy. However, significant advances in rapid tumor-cell sequencing and vaccine production must first be achieved. In the shorter term, combination therapy and melanoma vaccines show promise for improving the efficacy, response rates, and durability of current first-line immunotherapies.

4.1 Sequencing and combining therapies

Using combined therapies to treat mM may be the easiest way to achieve longer-lasting disease control, overcome innate resistance, evade adaptive resistance to immunotherapy, and optimize clinical response. There is significant interest in finding the best combinations of the two most effective approved therapy classes—targeted and ICI therapy (NCT02631447, NCT03235245, NCT02902029, NCT02224781). Such may also address the two major roadblocks in the deployment of these therapies: rapid resistance development and modest response rates.

4.2 Neoadjuvant therapy

Neoadjuvant therapy is typically used to reduce tumor burden and allow for less extensive surgeries. Before immunotherapy, neoadjuvant systemic therapy was not standard-of-care for mM treatment, likely because the risks of delaying surgery outweighed the limited benefits these therapies could provide. However, preclinical data suggest that this may not be true for immunotherapy [87], especially for therapies targeting T-cell function and

proliferation. Theoretically, initiating immunotherapy while the major tumor mass is still present may induce a stronger anti-tumor T-cell response. Indeed, a small feasibility one confirmed these results, demonstrating that patients receiving neoadjuvant and then adjuvant treatment had significantly more expansion of tumor-resident T-cell clones than patients who received the same treatment courses exclusively as adjuvant [88]. Neoadjuvant immunotherapy also seems to outperform adjuvant therapy in comparative studies with event-free survival benefit [89]. However, the sample size was small and the toxicity profile of the neoadjuvant arm was disappointing. Larger trials are currently underway to investigate neoadjuvant regimens that preserve efficacy while limiting toxicity (NCT02977052) with promising initial results [90].

4.3 Adoptive cell transfer with tumor-infiltrating lymphocytes

In melanoma, the presence of tumor-infiltrating lymphocytes (TILs) is associated with more favorable OS, RFS, and DSS/MSS [91]. Adoptive cell transfer (ACT) of TILs is the process of expanding autologous lymphocytes *in vitro*, usually aided by IL-2, IL-7, IL-15, and/or IL-21, followed by re-infusion to the patient [92]. This strategy circumvents many limitations of other immunotherapies. For example, *in vitro* TIL culture allows for the selective expansion of lymphocytes with the strongest effector function and the highest tumor-antigen affinity. Using autologous cells from resected tumor specimens avoids issues of rejection and allows each treatment to be uniquely targeted to the patient's specific tumor antigens [93]. Since expansion and activation occur without the suppressive effects of the TME, higher numbers of activated lymphocytes ($>10^{11}$ TILs) can be achieved. This also allows for pretreatment manipulation of the patient's immune system to optimize the efficacy of ACT or other planned immunotherapies without compromising the anti-tumor response. Greater response rates have been achieved when lymphodepletion proceeds and IV IL-2 follows ACT, both of which promote T-cell homeostatic cytokine production [94][95][96].

Since TIL-ACT regimens are not yet standardized, the degree of treatment efficacy reported in clinical trials has varied. Disease progression and overall survival after ACT-TIL are dependent on the expansion of neoepitope-specific CD8+ T-cells [97]. ORRs typically range from 28% - 45% [92]. Five-year follow-up found notable durability and suggests curative potential. Of the 22% CRs, all but one remained disease-free after 3 years, resulting in 100% 3-year and 95% 5-year survival rates [98]. It is especially exciting that these results occurred in challenging mM cases, in which patients had a median of 3 metastatic sites and had all failed first-line treatments, including 20% who had failed ICI therapy.

Overall, patients who receive TIL-ACT after failing ICI treatment have lower ORRs (56% vs 24%) and OS (28.5 vs 11.6 months) than ICI-naïve patients [99]. The same is true for patients with BRAF V600E/K mutations who failed prior targeted therapy (ORR: 21% vs 60% if naïve; OS 9.3 vs 50.7 months) [99]. This is likely because the poor immunogenicity and complex resistance mechanisms that allow tumors to evade ICIs also limit the efficacy of TIL-ACT [92]. However, an ongoing one of TIL-ACT in treatment-resistant mM has demonstrated an 80% disease control rate. Considering the higher toxicity rates and similar response rates of other second-line treatments, such as nivolumab or ipilimumab, TIL-ACT may be the best option for some patients resistant to alternative treatments [100][101].

Limitations to ACT-TIL are similar to those of other immunotherapies. As discussed above, resistance remains a central issue. Similarly, target identification, predictability of immunogenicity, and anti-tumor specificity (sparing healthy tissues) are essential for ACT-TIL success, but solutions remain in the early stages of development. Protocols for TIL expansion, antigen identification, pre-treatment immunodepletion, and post-infusion TIL maintenance (for example, IL-2 dosing) must be optimized for time, cost, efficacy, and safety in order to make this therapy feasible on a larger scale.

4.4 ACT with T-CARs

Another approach to ACT utilizes autologous T-cells modified *ex vivo* with cell-surface chimeric antigen receptors (CAR-T cells). The extracellular component of the CAR is a variable region of a synthetic antibody. It is attached to a T-cell signaling moiety and co-stimulatory domains, which allows MHC-independent T-cell activation [\[102\]](#). CAR-T cells can thus target tumors cells that have downregulated MHCs as an immune-escape mechanism [\[103\]](#). Success with CAR-T ACT for the treatment of hematologic malignancies sparked the investigation into the therapy for solid malignancies. However, success in mM clinical trials has been limited

4.5 Melanoma Vaccines

The five major categories of melanoma vaccines currently in development include (1) melanoma cell-targeted vaccines, (2) dendritic cell (DC) vaccines, (3) peptide-based vaccines, (4) vector-based vaccines, and (5) mRNA or DNA vaccines. Unlike preventative immunizations, cancer vaccines are therapeutic, activating the patient's immune system to incite an anti-tumor response against a known cancer or to prevent disease recurrence in the adjuvant setting.

Whole-cell vaccines use modified melanoma cells to simultaneously expose the immune system to many potential melanoma antigens, circumventing the need to identify the most immunogenic antigens for each tumor [\[104\]](#). DC vaccines are used to directly inject activated or modified DCs into the tumor site to increase anti-tumor T-cell activation. Peptide vaccines supply tumor-specific or tumor-associated antigen (i.e. gp100, MART-1/MelanA, tyrosinase) fragments that can be presented by professional APCs to induce effector T-cell activation. Vector vaccines use recombinant viral vectors to deliver tumor antigen transgenes directly to APCs. Within the APCs, the transgenes are expressed to produce high concentrations of tumor antigens that can be presented on both MHCI and MHCII for enhanced T-cell activation. The simultaneous expression of viral proteins by the delivered vectors boosts the immunogenicity of the vaccine [\[105\]](#). Therapeutic mRNA vaccines have garnered significant excitement after the advancements and efficacy demonstrated in the COVID-19 pandemic response. While still in the early stages, mRNA vaccines may have the potential to induce the targeted expression of nearly any protein. Using an mRNA approach avoids safety concerns associated with DNA and viral vector vaccines. Therapeutic mechanisms under investigation include enhancing the expression of tumor-specific antigens in DCs, mRNA-mediated delivery of specific anti-tumor or anti-ICI antibodies, and programming cancer cells to express suicidal intracellular proteins [\[106\]](#)[\[107\]](#)[\[108\]](#).

4.6 Toll-like receptor agonists

Toll-like receptor (TLR) activation and the resulting pro-inflammatory cytokine release is a critical step in the induction of both the innate and adaptive immune response [\[109\]](#). As poor immunogenicity continues to limit the efficacy of melanoma immunotherapies in some patients, TLRs are a logical ancillary agent that provides pro-inflammatory modulation of the tumor microenvironment

4.7 Predictive markers and personalized medicine

Predicting response rates, toxicity, and durability present a major challenge to current mM immunotherapies. The melanoma and immune oncology communities are investing significant resources to identify predictive biomarkers [\[110\]](#) that would allow treatments to be better optimized for each patient's therapeutic goals. Identification of tumor neoantigens and predictability of immunogenicity poses another issue. There are over 16,200 distinct class I HLA alleles, each with distinct peptide-binding preferences. Predicting which epitopes will likely be presented by each patient's APCs is key to the future of immunotherapies such as ACT, OV, and melanoma vaccines as this interaction ultimately determines the immunogenicity of a given neoantigen. Some recent progress has been made: The HLAthena model can predict endogenous HLA-binding peptides with >75% accuracy [\[111\]](#). The Tumor Neoantigen Selection Alliance (TESLA) developed a bioinformatic-informed model of tumor epitope immunogenicity capable of filtering out 98% of non-immunogenic peptides with a precision of over 0.70 [\[112\]](#). However, no tool currently exists that can accurately predict if a specific neoantigen-HLA combination will be recognized by an individual's TCRs. More informed models will require a larger and more diverse data set. The accessibility and affordability of next-generation molecular and functional diagnostics may one day allow each patient to receive personalized immunotherapy, optimized specifically to their tumor and goals.

References

1. Kirkwood JM, Richards T, Zarour HM, et al. Immunomodulatory effects of high-dose and low-dose interferon α 2b in patients with high-risk resected melanoma. *Cancer*. 2002;95(5):1101-12.
2. Tarhini AA, Gogas H, Kirkwood JM. IFN- α in the treatment of melanoma. *J Immunol*. 2012;189(8):3789-93.
3. Ascierto PA, Napolitano M, Celentano E, et al. Regulatory T cell frequency in patients with melanoma with different disease stage and course, and modulating effects of high-dose interferon- α 2b treatment. *Journal of Translational Medicine*. 2010;8(1):76.
4. Kirkwood JM, Strawderman MH, Ernstoff MS, Smith TJ, Borden EC, Blum RH. Interferon alfa-2b adjuvant therapy of high-risk resected cutaneous melanoma: the Eastern Cooperative Oncology Group Trial EST 1684. *Journal of clinical oncology*. 1996;14(1):7-17.

5. Hauschild A, Gogas H, Tarhini A, et al. Practical guidelines for the management of interferon- α -2b side effects in patients receiving adjuvant treatment for melanoma: Expert opinion. *Cancer: Interdisciplinary International Journal of the American Cancer Society*. 2008;112(5):982-94.
6. Eggermont A, Suci S, Testori A, et al. Long-term results of the randomized phase III trial EORTC 18991 of adjuvant therapy with pegylated interferon alfa-2b versus observation in resected stage III melanoma. *J Clin Oncol*. 2012;30(31):3810-8.
7. Eggermont AM, Suci S, Santinami M, et al. Adjuvant therapy with pegylated interferon alfa-2b versus observation alone in resected stage III melanoma: final results of EORTC 18991, a randomised phase III trial. *The Lancet*. 2008;372(9633):117-26.
8. Inman JL, Russell GB, Savage P, Levine EA. Low-dose adjuvant interferon for stage III malignant melanoma. *Am Surg*. 2003;69(2):127-30.
9. Grob JJ, Dreno B, de la Salmoniere P, et al. Randomised trial of interferon α -2a as adjuvant therapy in resected primary melanoma thicker than 1.5 mm without clinically detectable node metastases. *The Lancet*. 1998;351(9120):1905-10.
10. Atkins MB, Lotze MT, Dutcher JP, et al. High-Dose Recombinant Interleukin 2 Therapy for Patients With Metastatic Melanoma: Analysis of 270 Patients Treated Between 1985 and 1993. *Journal of Clinical Oncology*. 1999;17(7):2105-.
11. Fyfe G, Fisher RI, Rosenberg SA, Sznol M, Parkinson DR, Louie AC. Results of treatment of 255 patients with metastatic renal cell carcinoma who received high-dose recombinant interleukin-2 therapy. *Journal of Clinical Oncology*. 1995;13(3):688-96.
12. Hughes T, Klairmont M, Broucek J, Iodice G, Basu S, Kaufman HL. The prognostic significance of stable disease following high-dose interleukin-2 (IL-2) treatment in patients with metastatic melanoma and renal cell carcinoma. *Cancer Immunology, Immunotherapy*. 2015;64(4):459-65.
13. Clark JI, Curti B, Davis EJ, et al. Long-term progression-free survival of patients with metastatic melanoma or renal cell carcinoma following high-dose interleukin-2. *Journal of Investigative Medicine*. 2021;69(4):888-92.
14. Davar D, Ding F, Saul M, et al. High-dose interleukin-2 (HD IL-2) for advanced melanoma: a single center experience from the University of Pittsburgh Cancer Institute. *Journal for ImmunoTherapy of Cancer*. 2017;5(1):74.
15. Bernatchez C, Haymaker CL, Hurwitz ME, et al. Effect of a novel IL-2 cytokine immune agonist (NKTR-214) on proliferating CD8⁺T cells and PD-1 expression on immune cells in the tumor microenvironment in patients with prior checkpoint therapy. *Journal of Clinical Oncology*. 2017;35(15_suppl):2545-.
16. Khushalani NI, Diab A, Ascierto PA, et al. CA045-001: A phase III, randomized, open label study of bimepegaldesleukin (NKTR-214) plus nivolumab (NIVO) versus NIVO monotherapy in patients

- (pts) with previously untreated, unresectable or metastatic melanoma (MEL). *Journal of Clinical Oncology*. 2019;37(15_suppl):TPS9601-TPS.
17. Diab A, Hurwitz ME, Cho DC, et al. NKTR-214 (CD122-biased agonist) plus nivolumab in patients with advanced solid tumors: Preliminary phase 1/2 results of PIVOT. *J Clin Oncol*. 2018;36(15_suppl):3006.
 18. Parisi G, Saco JD, Salazar FB, et al. Persistence of adoptively transferred T cells with a kinetically engineered IL-2 receptor agonist. *Nature Communications*. 2020;11(1):660.
 19. Levin AM, Bates DL, Ring AM, et al. Exploiting a natural conformational switch to engineer an interleukin-2 'superkine'. *Nature*. 2012;484(7395):529-33.
 20. Sockolosky JT, Trotta E, Parisi G, et al. Selective targeting of engineered T cells using orthogonal IL-2 cytokine-receptor complexes. *Science*. 2018;359(6379):1037-42.
 21. Buchbinder EI, Dutcher JP, Daniels GA, et al. Therapy with high-dose Interleukin-2 (HD IL-2) in metastatic melanoma and renal cell carcinoma following PD1 or PDL1 inhibition. *Journal for ImmunoTherapy of Cancer*. 2019;7(1):49.
 22. Collins AV, Brodie DW, Gilbert RJ, et al. The interaction properties of costimulatory molecules revisited. *Immunity*. 2002;17(2):201-10.
 23. Maker AV, Attia P, Rosenberg SA. Analysis of the cellular mechanism of antitumor responses and autoimmunity in patients treated with CTLA-4 blockade. *The Journal of Immunology*. 2005;175(11):7746-54.
 24. Intlekofer AM, Thompson CB. At the bench: preclinical rationale for CTLA-4 and PD-1 blockade as cancer immunotherapy. *Journal of leukocyte biology*. 2013;94(1):25-39.
 25. Ribas A, Comin-Anduix B, Economou JS, et al. Intratumoral immune cell infiltrates, FoxP3, and indoleamine 2,3-dioxygenase in patients with melanoma undergoing CTLA4 blockade. *Clin Cancer Res*. 2009;15(1):390-9.
 26. Hanson DC, Canniff PC, Primiano MJ, et al. Preclinical *in vitro* characterization of anti-CTLA4 therapeutic antibody CP-675,206. *Cancer Research*. 2004;64(7 Supplement):877-.
 27. Hodi FS, O'Day SJ, McDermott DF, et al. Improved survival with ipilimumab in patients with metastatic melanoma. *New England Journal of Medicine*. 2010;363(8):711-23.
 28. Schadendorf D, Hodi FS, Robert C, et al. Pooled analysis of long-term survival data from phase II and phase III trials of ipilimumab in unresectable or metastatic melanoma. *Journal of clinical oncology*. 2015;33(17):1889.
 29. Eggermont AM, Chiarion-Sileni V, Grob J-J, et al. Adjuvant ipilimumab versus placebo after complete resection of stage III melanoma: long-term follow-up results of the European

- Organisation for Research and Treatment of Cancer 18071 double-blind phase 3 randomised trial. *European Journal of Cancer*. 2019;119:1-10.
30. Robert C, Thomas L, Bondarenko I, et al. Ipilimumab plus dacarbazine for previously untreated metastatic melanoma. *New England Journal of Medicine*. 2011;364(26):2517-26.
 31. Fellner C. Ipilimumab (yervoy) prolongs survival in advanced melanoma: serious side effects and a hefty price tag may limit its use. *Pharmacy and Therapeutics*. 2012;37(9):503.
 32. Weber J, Thompson JA, Hamid O, et al. A randomized, double-blind, placebo-controlled, phase II study comparing the tolerability and efficacy of ipilimumab administered with or without prophylactic budesonide in patients with unresectable stage III or IV melanoma. *Clinical cancer research*. 2009;15(17):5591-8.
 33. Seth R, Messersmith H, Kaur V, et al. Systemic therapy for melanoma: ASCO guideline. *Journal of Clinical Oncology*. 2020;38(33):3947-70.
 34. Sharpe AH, Pauken KE. The diverse functions of the PD1 inhibitory pathway. *Nature Reviews Immunology*. 2018;18(3):153-67.
 35. Merelli B, Massi D, Cattaneo L, Mandalà M. Targeting the PD1/PD-L1 axis in melanoma: Biological rationale, clinical challenges and opportunities. *Critical Reviews in Oncology/Hematology*. 2014;89(1):140-65.
 36. Hino R, Kabashima K, Kato Y, et al. Tumor cell expression of programmed cell death-1 ligand 1 is a prognostic factor for malignant melanoma. *Cancer: Interdisciplinary International Journal of the American Cancer Society*. 2010;116(7):1757-66.
 37. Kleffel S, Posch C, Barthel Steven R, et al. Melanoma Cell-Intrinsic PD-1 Receptor Functions Promote Tumor Growth. *Cell*. 2015;162(6):1242-56.
 38. Taube JM, Anders RA, Young GD, et al. Colocalization of inflammatory response with B7-h1 expression in human melanocytic lesions supports an adaptive resistance mechanism of immune escape. *Sci Transl Med*. 2012;4(127):127ra37-ra37.
 39. Eggermont AM, Blank CU, Mandala M, et al. Adjuvant pembrolizumab versus placebo in resected stage III melanoma. *New England Journal of Medicine*. 2018;378(19):1789-801.
 40. Moser JC, Wei G, Colonna SV, Grossmann KF, Patel S, Hyngstrom JR. Comparative-effectiveness of pembrolizumab vs. nivolumab for patients with metastatic melanoma. *Acta Oncologica*. 2020;59(4):434-7.
 41. Hamid O, Robert C, Daud A, et al. Five-year survival outcomes for patients with advanced melanoma treated with pembrolizumab in KEYNOTE-001. *Annals of Oncology*. 2019;30(4):582-8.
 42. Schachter J, Ribas A, Long GV, et al. Pembrolizumab versus ipilimumab for advanced melanoma: final overall survival results of a multicentre, randomised, open-label phase 3 study (KEYNOTE-

- 006). *The Lancet*. 2017;390(10105):1853-62.
43. Robert C, Ribas A, Schachter J, et al. Pembrolizumab versus ipilimumab in advanced melanoma (KEYNOTE-006): post-hoc 5-year results from an open-label, multicentre, randomised, controlled, phase 3 study. *The Lancet Oncology*. 2019;20(9):1239-51.
44. Cowey CL, Liu FX, Black-Shinn J, et al. Pembrolizumab Utilization and Outcomes for Advanced Melanoma in US Community Oncology Practices. *J Immunother*. 2018;41(2):86-95.
45. Liu FX, Ou W, Diede SJ, Whitman ED. Real-world experience with pembrolizumab in patients with advanced melanoma: A large retrospective observational study. *Medicine (Baltimore)*. 2019;98(30):e16542-e.
46. Larkin J, Chiarion-Sileni V, Gonzalez R, et al. Combined Nivolumab and Ipilimumab or Monotherapy in Untreated Melanoma. *New England Journal of Medicine*. 2015;373(1):23-34.
47. Hodi FS, Chiarion-Sileni V, Gonzalez R, et al. Nivolumab plus ipilimumab or nivolumab alone versus ipilimumab alone in advanced melanoma (CheckMate 067): 4-year outcomes of a multicentre, randomised, phase 3 trial. *The Lancet Oncology*. 2018;19(11):1480-92.
48. Robert C, Long GV, Brady B, et al. Nivolumab in previously untreated melanoma without BRAF mutation. *New England journal of medicine*. 2015;372(4):320-30.
49. Weber JS, D'Angelo SP, Minor D, et al. Nivolumab versus chemotherapy in patients with advanced melanoma who progressed after anti-CTLA-4 treatment (CheckMate 037): a randomised, controlled, open-label, phase 3 trial. *The lancet oncology*. 2015;16(4):375-84.
50. Weber JS, Hodi FS, Wolchok JD, et al. Safety profile of nivolumab monotherapy: a pooled analysis of patients with advanced melanoma. 2017.
51. Robert C, Schachter J, Long GV, et al. Pembrolizumab versus ipilimumab in advanced melanoma. *New England Journal of Medicine*. 2015;372(26):2521-32.
52. Ribas A, Wolchok JD, Robert C, et al. P0116 Updated clinical efficacy of the anti-PD-1 monoclonal antibody pembrolizumab (MK-3475) in 411 patients with melanoma. *European Journal of Cancer*. 2015;51:e24.
53. Eggermont AMM, Blank CU, Mandalà M, et al. Adjuvant pembrolizumab versus placebo in resected stage III melanoma (EORTC 1325-MG/KEYNOTE-054): distant metastasis-free survival results from a double-blind, randomised, controlled, phase 3 trial. *The Lancet Oncology*. 2021;22(5):643-54.
54. Zimmer L, Livingstone E, Hassel JC, et al. Adjuvant nivolumab plus ipilimumab or nivolumab monotherapy versus placebo in patients with resected stage IV melanoma with no evidence of disease (IMMUNED): a randomised, double-blind, placebo-controlled, phase 2 trial. *The Lancet*. 2020;395(10236):1558-68.

55. Ribas A, Hamid O, Daud A, et al. Association of pembrolizumab with tumor response and survival among patients with advanced melanoma. *Jama*. 2016;315(15):1600-9.
56. Jenkins RW, Barbie DA, Flaherty KT. Mechanisms of resistance to immune checkpoint inhibitors. *British journal of cancer*. 2018;118(1):9-16.
57. Sharma P, Hu-Lieskovan S, Wargo JA, Ribas A. Primary, adaptive, and acquired resistance to cancer immunotherapy. *Cell*. 2017;168(4):707-23.
58. Shin DS, Zaretsky JM, Escuin-Ordinas H, et al. Primary resistance to PD-1 blockade mediated by JAK1/2 mutations. *Cancer discovery*. 2017;7(2):188-201.
59. Lee JH, Shklovskaya E, Lim SY, et al. Transcriptional downregulation of MHC class I and melanoma de- differentiation in resistance to PD-1 inhibition. *Nature Communications*. 2020;11(1):1897.
60. Koyama S, Akbay EA, Li YY, et al. Adaptive resistance to therapeutic PD-1 blockade is associated with upregulation of alternative immune checkpoints. *Nature Communications*. 2016;7(1):10501.
61. Spranger S, Bao R, Gajewski TF. Melanoma-intrinsic β -catenin signalling prevents anti-tumour immunity. *Nature*. 2015;523(7559):231-5.
62. Davar D, Wang H, Chauvin J-M, et al. Phase Ib/II Study of Pembrolizumab and Pegylated-Interferon Alfa-2b in Advanced Melanoma. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology*. 2018;36(35):JCO1800632-JCO.
63. Mehnert JM, Mitchell TC, Huang AC, et al. BMM (BRAF Autophagy and MEK Inhibition in Melanoma): A Phase I/II Trial of Dabrafenib, Trametinib, and Hydroxychloroquine in Advanced BRAFV600-mutant Melanoma. *Clinical Cancer Research*. 2022:OF1-OF9.
64. Weide B, Martens A, Hassel JC, et al. Baseline biomarkers for outcome of melanoma patients treated with pembrolizumab. *Clinical Cancer Research*. 2016;22(22):5487-96.
65. Robert C, Hwu W-J, Hamid O, et al. Long-term safety of pembrolizumab monotherapy and relationship with clinical outcome: A landmark analysis in patients with advanced melanoma. *European Journal of Cancer*. 2021;144:182-91.
66. Khatkhat MA, Reid A, Freeman J, et al. PD-L1 Expression on Circulating Tumor Cells May Be Predictive of Response to Pembrolizumab in Advanced Melanoma: Results from a Pilot Study. *Oncologist*. 2020;25(3):e520-e7. Epub 2019/12/05.
67. Gadiot J, Hooijkaas AI, Kaiser AD, van Tinteren H, van Boven H, Blank C. Overall survival and PD-L1 expression in metastasized malignant melanoma. *Cancer*. 2011;117(10):2192-201.
68. Kaunitz GJ, Cottrell TR, Lilo M, et al. Melanoma subtypes demonstrate distinct PD-L1 expression profiles. *Laboratory Investigation*. 2017;97(9):1063-71.

69. Wolchok JD, Kluger H, Callahan MK, et al. Nivolumab plus ipilimumab in advanced melanoma. *N Engl J Med*. 2013;369:122-33.
70. Larkin J, Chiarion-Sileni V, Gonzalez R, et al. Five-year survival with combined nivolumab and ipilimumab in advanced melanoma. *New England Journal of Medicine*. 2019;381(16):1535-46.
71. Long GV, Atkinson V, Cebon JS, et al. Standard-dose pembrolizumab in combination with reduced-dose ipilimumab for patients with advanced melanoma (KEYNOTE-029): an open-label, phase 1b trial. *The Lancet Oncology*. 2017;18(9):1202-10.
72. Long GV, Robert C, Butler MO, et al. Standard-dose Pembrolizumab plus alternate-dose Ipilimumab in advanced melanoma: KEYNOTE-029 cohort 1C, a phase 2 randomized study of two dosing schedules. *Clinical Cancer Research*. 2021;27(19):5280-8.
73. Lebbé C, Meyer N, Mortier L, et al. Evaluation of two dosing regimens for nivolumab in combination with ipilimumab in patients with advanced melanoma: results from the phase IIIb/IV CheckMate 511 trial. *Journal of clinical oncology*. 2019;37(11):867.
74. Carlino MS, Menzies AM, Atkinson V, et al. Long-term follow-up of standard-dose pembrolizumab plus reduced-dose ipilimumab in patients with advanced melanoma: KEYNOTE-029 Part 1B. *Clinical Cancer Research*. 2020;26(19):5086-91.
75. Zhang J, Huang Y, Xi G, Zhang F. HX008: a humanized PD-1 blocking antibody with potent antitumor activity and superior pharmacologic properties. *mAbs*. 2020;12(1):1724751.
76. Zhang Y, Du X, Liu M, et al. Hijacking antibody-induced CTLA-4 lysosomal degradation for safer and more effective cancer immunotherapy. *Cell Research*. 2019;29(8):609-27.
77. Ruffo E, Wu RC, Bruno TC, Workman CJ, Vignali DAA. Lymphocyte-activation gene 3 (LAG3): The next immune checkpoint receptor. *Seminars in Immunology*. 2019;42:101305.
78. da Silva IP, Gallois A, Jimenez-Baranda S, et al. Reversal of NK-Cell Exhaustion in Advanced Melanoma by Tim-3 Blockade. *Cancer Immunology Research*. 2014;2(5):410-22.
79. Baghdadi M, Nagao H, Yoshiyama H, et al. Combined blockade of TIM-3 and TIM-4 augments cancer vaccine efficacy against established melanomas. *Cancer Immunology, Immunotherapy*. 2013;62(4):629-37.
80. Deak LLC, Seeber S, Perro M, et al. RG7769 (PD1-TIM3), a novel heterodimeric avidity-driven T cell specific PD-1/TIM-3 bispecific antibody lacking Fc-mediated effector functions for dual checkpoint inhibition to reactivate dysfunctional T cells. *AACR*; 2020.
81. Pikor LA, Bell JC, Diallo J-S. Oncolytic viruses: exploiting cancer's deal with the devil. *Trends in cancer*. 2015;1(4):266-77.
82. Pol J, Kroemer G, Galluzzi L. First oncolytic virus approved for melanoma immunotherapy. *OncoImmunology*. 2016;5(1):e1115641.

83. Andtbacka RH, Kaufman HL, Collichio F, et al. Talimogene Laherparepvec Improves Durable Response Rate in Patients With Advanced Melanoma. *J Clin Oncol*. 2015;33(25):2780-8. Epub 20150526.
84. Nguyen T-T, Ramsay L, Ahanfeshar-Adams M, et al. Mutations in the IFN γ -JAK-STAT pathway causing resistance to immune checkpoint inhibitors in melanoma increase sensitivity to oncolytic virus treatment. *Clinical Cancer Research*. 2021;27(12):3432-42.
85. Kaufman HL, Bommareddy PK. Two roads for oncolytic immunotherapy development. *J Immunother Cancer*. 2019;7(1):26. Epub 20190201.
86. Garcia M, Moreno R, Gil-Martin M, et al. A phase 1 trial of oncolytic adenovirus ICOVIR-5 administered intravenously to cutaneous and uveal melanoma patients. *Human gene therapy*. 2019;30(3):352-64.
87. Liu J, Blake SJ, Yong MCR, et al. Improved Efficacy of Neoadjuvant Compared to Adjuvant Immunotherapy to Eradicate Metastatic Disease. *Cancer Discovery*. 2016;6(12):1382-99.
88. Blank CU, Rozeman EA, Fanchi LF, et al. Neoadjuvant versus adjuvant ipilimumab plus nivolumab in macroscopic stage III melanoma. *Nature medicine*. 2018;24(11):1655-61.
89. Amaria RN, Menzies AM, Burton EM, et al. Neoadjuvant systemic therapy in melanoma: recommendations of the International Neoadjuvant Melanoma Consortium. *The Lancet Oncology*. 2019;20(7):e378-e89.
90. Rozeman EA, Hoefsmit EP, Reijers ILM, et al. Survival and biomarker analyses from the OpACIN-neo and OpACIN neoadjuvant immunotherapy trials in stage III melanoma. *Nature Medicine*. 2021;27(2):256-63.
91. Fu Q, Chen N, Ge C, et al. Prognostic value of tumor-infiltrating lymphocytes in melanoma: a systematic review and meta-analysis. *OncolImmunology*. 2019;8(7):e1593806.
92. Dafni U, Michielin O, Lluesma SM, et al. Efficacy of adoptive therapy with tumor-infiltrating lymphocytes and recombinant interleukin-2 in advanced cutaneous melanoma: a systematic review and meta-analysis. *Annals of Oncology*. 2019;30(12):1902-13.
93. Rosenberg SA, Restifo NP. Adoptive cell transfer as personalized immunotherapy for human cancer. *Science*. 2015;348(6230):62-8.
94. Ellebaek E, Iversen TZ, Junker N, et al. Adoptive cell therapy with autologous tumor infiltrating lymphocytes and low-dose Interleukin-2 in metastatic melanoma patients. *Journal of translational medicine*. 2012;10(1):1-12.
95. Goff SL, Dudley ME, Citrin DE, et al. Randomized, prospective evaluation comparing intensity of lymphodepletion before adoptive transfer of tumor-infiltrating lymphocytes for patients with metastatic melanoma. *Journal of Clinical Oncology*. 2016;34(20):2389.

96. Forget M-A, Haymaker C, Hess KR, et al. Prospective analysis of adoptive TIL therapy in patients with metastatic melanoma: response, impact of anti-CTLA4, and biomarkers to predict clinical outcome. *Clinical Cancer Research*. 2018;24(18):4416-28.
97. Kristensen NP, Heeke C, Tvingsholm SA, et al. Neoantigen-reactive CD8+ T cells affect clinical outcome of adoptive cell therapy with tumor-infiltrating lymphocytes in melanoma. *The Journal of Clinical Investigation*. 2022;132(2).
98. Rosenberg SA, Yang JC, Sherry RM, et al. Durable complete responses in heavily pretreated patients with metastatic melanoma using T-cell transfer immunotherapy. *Clinical cancer research*. 2011;17(13):4550-7.
99. Seitter SJ, Sherry RM, Yang JC, et al. Impact of Prior Treatment on the Efficacy of Adoptive Transfer of Tumor-Infiltrating Lymphocytes in Patients with Metastatic Melanoma. *Clinical Cancer Research*. 2021;27(19):5289-98.
100. Zimmer L, Apuri S, Eroglu Z, et al. Ipilimumab alone or in combination with nivolumab after progression on anti-PD-1 therapy in advanced melanoma. *European journal of cancer*. 2017;75:47-55.
101. Sarnaik AA, Hamid O, Khushalani NI, et al. Lifileucel, a tumor-infiltrating lymphocyte therapy, in metastatic melanoma. *Journal of Clinical Oncology*. 2021;39(24):2656.
102. Zhang E, Xu H. A new insight in chimeric antigen receptor-engineered T cells for cancer immunotherapy. *Journal of Hematology & Oncology*. 2017;10(1):1.
103. Beatty GL, Gladney WL. Immune escape mechanisms as a guide for cancer immunotherapy. *Clinical cancer research*. 2015;21(4):687-92.
104. Keenan BP, Jaffee EM. Whole cell vaccines--past progress and future strategies. *Semin Oncol*. 2012;39(3):276-86.
105. Larocca C, Schlom J. Viral vector-based therapeutic cancer vaccines. *Cancer J*. 2011;17(5):359-71.
106. Jain R, Frederick JP, Huang EY, et al. MicroRNAs enable mRNA therapeutics to selectively program cancer cells to self-destruct. *Nucleic acid therapeutics*. 2018;28(5):285-96.
107. Van Hoecke L, Verbeke R, Dewitte H, et al. mRNA in cancer immunotherapy: beyond a source of antigen. *Molecular Cancer*. 2021;20(1):48.
108. Bidram M, Zhao Y, Shebardina NG, et al. mRNA-Based Cancer Vaccines: A Therapeutic Strategy for the Treatment of Melanoma Patients. *Vaccines*. 2021;9(10):1060.
109. Bourquin C, Pommier A, Hotz C. Harnessing the immune system to fight cancer with Toll-like receptor and RIG-I-like receptor agonists. *Pharmacological research*. 2020;154:104192.

110. Baker RG, Hoos AX, Adam SJ, et al. The Partnership for Accelerating Cancer Therapies. *Cancer J.* 2018;24(3):111-4.
111. Sarkizova S, Klaeger S, Le PM, et al. A large peptidome dataset improves HLA class I epitope prediction across most of the human population. *Nature Biotechnology.* 2020;38(2):199-209.
112. Wells DK, van Buuren MM, Dang KK, et al. Key parameters of tumor epitope immunogenicity revealed through a consortium approach improve neoantigen prediction. *Cell.* 2020;183(3):818-34. e13.

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