

# Life Cycle of *Puccinia striiformis* f. sp. *tritici*

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Stem rust or black rust, caused by *Puccinia graminis* f. sp. *tritici* Erikss. and E. Henn. (*Pgt*), leaf rust or brown rust, caused by *Puccinia triticina* Erikss. (*Pt*) and the wheat stripe rust or yellow rust, caused by *Puccinia striiformis* Westend. f. sp. *tritici* Erikss. (*Pst*), is a historically crucial economic disease that occurs in almost all wheat-growing regions worldwide.

*Puccinia striiformis* f. sp. *tritici*

alternate hosts

stripe rust of wheat

## 1. Introduction

Wheat is the most cultivated cereal crop, universal staple food, and a host for many pathogens. The most serious threat to wheat crops is a group of rust fungi causing severe yield losses worldwide [1]. Stem rust or black rust, caused by *Puccinia graminis* f. sp. *tritici* Erikss. and E. Henn. (*Pgt*), leaf rust or brown rust, caused by *Puccinia triticina* Erikss. (*Pt*) and the wheat stripe rust or yellow rust, caused by *Puccinia striiformis* Westend. f. sp. *tritici* Erikss. (*Pst*), is a historically crucial economic disease that occurs in almost all wheat-growing regions worldwide [2][3]. The early history of mankind is full of fears and threats to these devastating rust pathogens. Since the discovery of rust pathogens, numerous investigations have been conducted on their life cycles for the management of the diseases caused by these fungi. The tenacity of rust fungi as destructive pathogens throughout the wheat-growing areas in the world is attributed to the special features of the pathogen, for example, the production of a large number of spores, inter and intracontinental wind dissemination, and the ability to change genetically resulting in new races with increased virulence diversity [4]. Generally, the disease occurs in the northern and southern areas of temperate regions. Recently, the wheat stripe rust disease has become more severe in some warmer areas than before [5], endangering global food security [6]. Hot summers and dry seasons are the bottlenecks for the survival of *Pst*. The disease can be controlled by growing resistant cultivars, suitable cultural practices, and the appropriate use of chemical fungicides. Resistant cultivars are the most effective, economical and environmentally friendly approach to combat with the wheat stripe rust pathogen. However, the *Pst* population is highly dynamic and variable, which makes it difficult to develop highly resistant wheat cultivars with durable resistance [6].

In the US Pacific Northwest, barberry is essential for the wheat stem rust but does not play a role for the wheat stripe rust pathogen [7][8][9]. Barberry may serve as an alternate host for *Pst* in the Himalayan region under natural conditions [10][11][12]. In eastern Africa and western Asia, barberry plants have been found, but their association with the wheat stripe rust disease epidemics has not been confirmed [13][14]. To date, the evidence of natural infection of barberry by *Pst* has been observed only in China, but at a low frequency [15][16]. Similarly, *Pst* has not been found on barberry plants in southeastern Sweden, but *Pgt* is common on the alternate host plants in this region [17].

The use of genetic techniques in the past ten years has achieved some advancement in understanding the plant-microbe interaction. The wheat stripe rust pathogen is an obligate, biotrophic parasite, having five distinct spore stages and two hosts to complete their life cycle. A macrocyclic life cycle comprises of uredinial, telial, basidial, pycnial and aecial stages. Like other rust pathogens, *Pst* is also highly specific to their primary host plants, for example, cereal crops and grasses, and the alternate host plants, for example, *Berberis* and *Mahonia* spp. The primary hosts can be the same but generally, the alternate hosts are different for different *Puccinia* spp. Based on their host specificity and morphological characteristics, the *Puccinia* spp. are further divided into formae speciales or varieties. For example, stripe rust on wheat is caused by *P. striiformis* f. sp. *tritici* (*Pst*), on barley by *P. striiformis* f. sp. *hordei* (*Psh*); stem rust on wheat by *P. graminis* f. sp. *tritici* (*Pgt*), and on oat by *P.*

*graminis* f. sp. *avenae* (*Pga*), and on rye by *P. graminis* f. sp. *secalis* (*Pgs*); and crown rust on oat by *P. coronata* var. *avenae* (*Pca*), and on barley by *P. coronata* var. *hordei* (*Pch*) [18].

The genetic diversity of *Pst* in Australia, Europe and North America indicated a clonal population structure of the pathogen [19]. On the other hand, the *Pst* populations of Gansu Province, China, were found to have high genetic diversities and produce abundant telia, indicating possible sexual recombination in this region [20][21]. Jin et al. [13] reported barberry as an alternate host for *P. pseudostriformis* (Syn. *P. striiformis* f. sp. *poae*) under natural conditions in Minnesota in the US and *Pst* under controlled conditions. The possible role of *Berberis* spp. as a sexual host of *Pst* has attained much importance, particularly in the US, China, and Pakistan [7][9][11][15][18]. *Mahonia aquifolium*, under experimental conditions, has also been identified to be susceptible to *Pst* [22].

## 2. Stripe Rust: Outlook

Wheat rusts have created major famines throughout history, causing substantial economic losses [23]. Currently, the most severe rust disease is the wheat stripe rust disease, causing more than 60% yield losses under favorable conditions [1][24][25]. The disease is named as yellow rust or stripe rust due to yellow colored stripes in lines between leaf veins in adult-plants but the urediniospores are in clusters (not in stripes) when the infection is at the seedlings stage [26]. Urediniospores are dikaryotic and produced asexually on the primary host plant. In the case of severe disease epidemics, stripe rust uredinial infection occurs on leaves, spikes, spikelets, glumes, awns and kernels. With the increase in temperature or at the maturity stage of the plant, the production of urediniospores comes to an end. The uredinia start converting into black colored telia containing teliospores [27]. Teliospores may serve as the survival structures which can cause infection on the alternate hosts under favorable environmental conditions or become a dead-end due to severe climatic conditions or incompatibility with the alternate host plants [28]. Teliospores are thick walled and germinate to produce haploid basidiospores [18]. These basidiospores directly penetrate the alternate host epidermal cells and cause infection and produce pycnia on the upper side of leaves and aecia on the lower side of leaves. Under favorable conditions both the pycnial and aecial infections are observed on stems, pedicels and peduncles [12]. The aeciospores infect wheat crops resulting in the formation of urediniospores. In contrast to uredinial re-infection on the same primary host plants or grasses, the aeciospores cannot re-infect the alternate host plants.

The wheat stripe rust pathogen belongs to the genus—*Puccinia*, family—Pucciniaceae, order—Pucciniales, class—Pucciniomycetes, division—Basidiomycota and kingdom—Fungi. *Pst* can undergo long-distance dispersal and it has caused numerous invasions [29][30] associated with austere economical losses [19][31][32][33]. Several cases of incursions of economic importance have been reported for *Pst* but only recently their origin was confirmed [11][30]. In the early 20th century, *Pst* was reported for the first time in South and North America [34][35], most likely spreading from north-western Europe [11][32] and introduced accidentally in Australia from north-western Europe in 1979 through human activity [36]. *Pst* strains detected in South Africa in 1996 were genetically related to populations in the Mediterranean regions and Middle Eastern ones, possibly spread by the wind [11][37]. *Pst* has become important in the context of invasions and recolonizations through the emergence of new races and strains in previously non-colonized areas. For example, since 2000, the emergence of two aggressive strains of *Pst*, PSTS1, and PSTS2, in the geographical expansion of *Pst* epidemics into the southeastern US and western Australia, where the disease was not previously a serious problem [27][38]. Similarly, since 2011, invasive strains of the wheat stripe rust pathogen, Warrior and Kranich, have largely replaced the pre-existing northwestern European *Pst* populations of the pathogen [30][39].

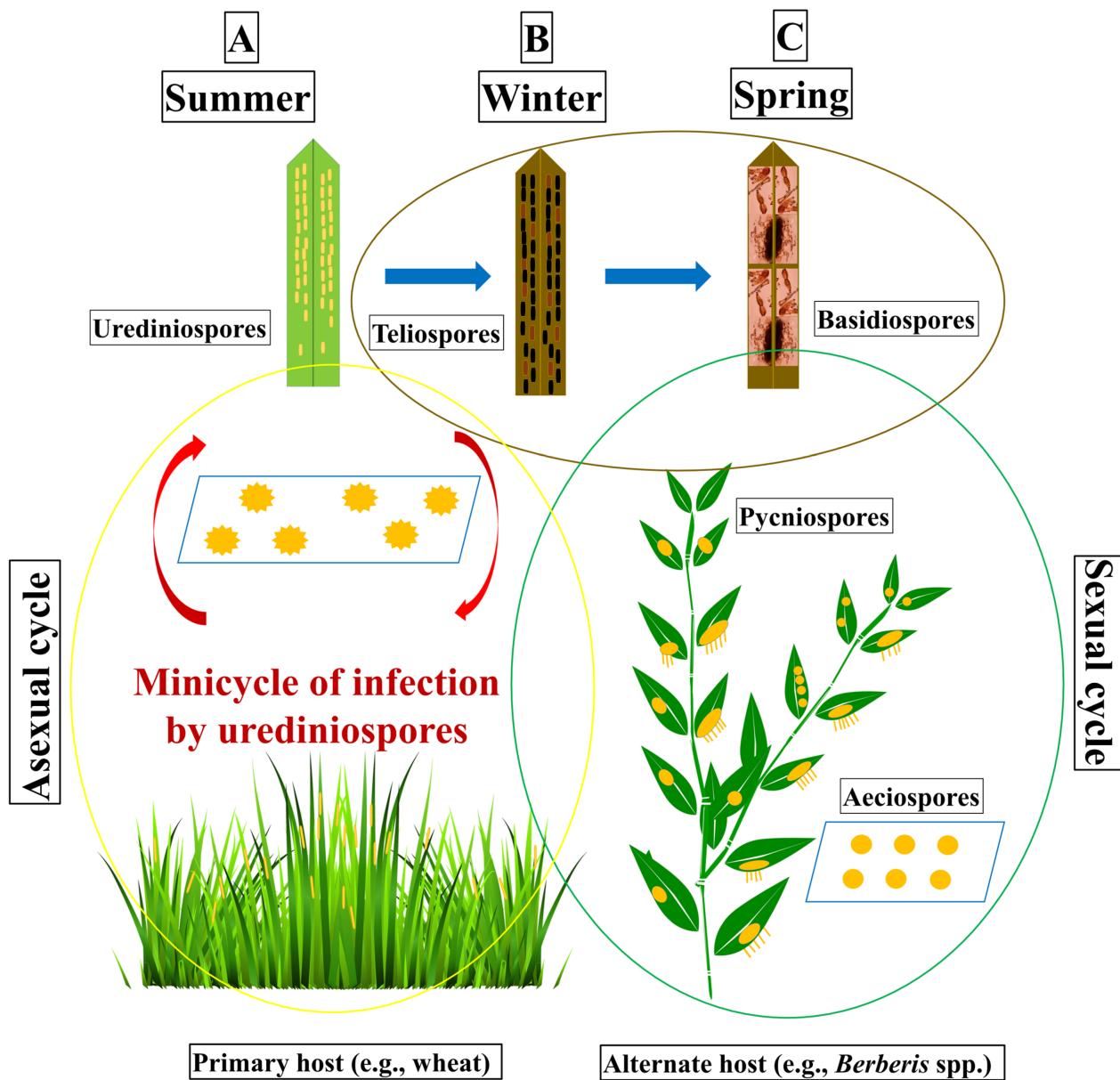
*Pst* has been reported in the areas of the US (Pacific Northwest), eastern Asia (northwestern and southwestern China), Oceania (Australia, New Zealand), southern Asia (India, Pakistan, and Nepal), western Europe (eastern England), the Arabian Peninsula (Yemen) and eastern Africa (Ethiopia, Kenya) [40]. In the last two decades, the emergence of more aggressive races of *Pst* having the ability to cause high epidemic potential even in warmer regions [26] is generally the result of mutation, somatic hybridization and sexual recombination. The potential role of alternate hosts in pathogenic diversity is of much

importance. However, it is still unknown by which mechanisms new races evolve. The high reproduction ability, long distance dissemination, adaptation to different environmental conditions, and several host species make *Pst* a highly diversified pathogen [41]. The threat of new virulent races of this pathogen emphasizes the need to understand the mechanisms involved in the genetic diversity of *Pst* and the role of aecial hosts in sexual reproduction to encounter the possible attacks of *Pst* in the future [26].

### 3. The Life Cycle of *Pst*

The complete life cycle of *Pst* has five spore stages, different from each other, on two phylogenetically distinct host plants, a cereal as the primary host or asexual host, and *Berberis* spp. as the alternate host or sexual host [13]. The dikaryotic (N + N') single-celled urediniospores appear on the primary host through the breaking of epidermal cells, and each uredinia harboring yellow-colored numerous urediniospores. The repeated asexual cycles on the primary host may cause wide-scale epidemics on the cereal hosts [42]. The single uredinia, assembled in lesions, forms typical stripes on the leaves of adult plants and produce urediniospores in 10 to 18 days after infection under optimal conditions. The uredinal lesions expand longitudinally upon the production of new uredinia. With the start of the senescence of infected leaves, *P. striiformis* starts producing telia resulting in the creation of many two-celled oblong-clavate teliospores. These teliospore cells contain a diploid nucleus (N + N') formed by karyogamy. The germinating teliospores produce ellipsoid haploid (N) basidiospores. Basidiospores are uninucleate or binucleate haploid spores produced from a germinating diploid teliospores. Basidiospores cause infection on the alternate host (e.g., barberry), resulting in oblong-shaped pycniospores (N) on the adaxial surface of the leaf, following the formation of dikaryotic (N + N') aecia on the abaxial surface of the leaf. Pycniospores are haploid (N), sexually derived spores (spermatium) formed in a pycnium (spermogonium) of rust fungi. Finally, aeciospores infect the primary host resulting urediniospores into the wheat leaves. The life cycle of *Pst* may take place during two regular growing seasons of the asexual host. *Pst* is an obligate biotrophic fungus which depends on a living host for its development and reproduction [32][42][43].

The sexual phase starts when the two-celled teliospores germinate and produce basidiospores, attached to a sterigmatum. Basidiospores (N) infect barberry leaves, resulting in the formation of pycnia (N), covered with pycnial nectar, formed on the upper side of the leaf. The haploid pycniospores of (-) and haploid hyphae (+) fuse together through plasmogamy and form aeciospores (N + N) on the abaxial side of the barberry leaf [44]. Aeciospores are dikaryotic produced in a cup-shaped aecium of a rust fungus. The asexual infection process in cereal hosts starts via a urediniospore-germ-tube, penetrating a stoma and then differentiating into a substomatal vesicle, resulting in two to three primary infection hyphae, which develop haustorial mother cells. These cells are separated from their respective hyphae by a septum. Haustorial mother cells penetrate the plant cell walls and form haustoria, highly specialized structures, representing the primary interface between host and the pathogen. Haustoria take water and nutrients from the host tissues and also make signaling between hosts and pathogens by producing effector molecules, like avirulence gene products. Young haustoria have a spherical shape, whereas older haustoria appeared more branched which allows the fungus to extend the area of the contact zone into the host and uptake nutrients more efficiently [32]. Basidiospores can only infect alternate hosts (like barberry) but cannot infect primary hosts (like wheat). Basidiospores infect epidermal cells through direct penetration while urediniospores infect through host stromata. *Pst* generally infects common wheat (*Triticum aestivum* L.), cultivated emmer wheat (*T. dicoccum* Schrank), triticale (*Triticosecale*), durum wheat (*T. turgidum* var. *durum* L.) and wild emmer wheat (*T. dicoccoides* Korn); as well as cultivated barley (*Hordeum vulgare* L.) and rye (*Secale cereale* L.). The complete life cycle of *Pst* is shown in **Figure 1**.



**Figure 1.** Schematic diagram of the life cycle of *Puccinia striiformis* f. sp. *tritici* (Pst), divided into three phases from left to right. Phase **A**, usually starts late in the spring and continues into the summer until the harvesting of the wheat crop. It consists of the asexual cycle which takes place on the primary host (e.g., wheat). The infection may take place by aeciospores or urediniospores resulting in the formation of yellow-colored stripes of uredinia on the wheat leaves. Urediniospores re-infect wheat plants in the same field or in the neighboring wheat fields to continue the mini-cycle of somatic reproduction until the conditions become unfavorable. With the increase in temperature or at crop maturity, the urediniospores turn into teliospores. Phase **B**, the teliospores survive and germinate to produce basidiospores. Phase **C**, the basidiospores infect the new leaves of alternate hosts (e.g., barberry) in the spring and produce pycniospores on the adaxial surface of leaves. Pycniospores produce aecial cups on the abaxial surface of the leaves after fertilization. Aecial cups contain aeciospores which can infect the primary hosts.

## References

1. Wellings, C. Global status of stripe rust: A review of historical and current threats. *Euphytica* 2011, 179, 129–141.
2. McIntosh, R.A.; Wellings, C.R.; Park, R.F. *Wheat Rusts: An Atlas of Resistance Genes*; Kluwer Academic Publishers: Dordrecht, The Netherlands; East Melbourne, Australia, 1995; pp. 1–200.
3. Singh, R.P.; William, H.M.; Huerta-Espino, J.; Rosewarne, G. Wheat rust in Asia: Meeting the challenges with old and new technologies. In Proceedings of the 4th International Crop Science Congress, Brisbane, Australia, 26 September–1 October 2004; Available online: file:///D:/1.%20Ph.%20D%20Sajid/Globalization%202050/Taylor%20and%20Francis%20Review%20Revision/141\_singh (accessed on 25 May 2020).
4. Brown, J.K.M.; Hovmöller, M. Aerial dispersal of pathogens on the global and continental scales and its impact on plant disease. *Science* 2002, 297, 537–541.
5. Hovmöller, M.; Walter, S.; Justesen, A.F. Escalating threat of wheat rusts. *Science* 2010, 329, 369.
6. Luo, H.; Wang, X.; Zhan, G.; Wei, G.; Zhou, X.; Zhao, J.; Huang, L.; Kang, Z. Genome-wide analysis of simple sequence repeats and efficient development of polymorphic SSR markers based on whole genome re-sequencing of multiple isolates of the wheat stripe rust fungus. *PLoS ONE* 2015, 10, e0130362.
7. Wang, M.N.; Wan, A.M.; Chen, X.M. Barberry is more important as an alternate host for stem rust than for stripe rust in the U.S. Pacific Northwest. *Plant Dis.* 2015, 99, 1507–1516.
8. Cheng, P.; Chen, X.M. Virulence and molecular analyses support asexual reproduction of *Puccinia striiformis* f. sp. *tritici* in the U.S. Pacific Northwest. *Phytopathology* 2014, 104, 1208–1220.
9. Wang, M.N.; Chen, X.M. Barberry does not function as an alternate host for *Puccinia striiformis* f. sp. *tritici* in the U.S. Pacific Northwest due to Teliospore degradation and barberry phenology. *Plant Dis.* 2015, 99, 1500–1506.
10. Ali, S.; Leconte, M.; Walker, A.-S.; Enjalbert, J.; De Vallavieille-Pope, C. Reduction in the sex ability of worldwide clonal populations of *Puccinia striiformis* f.sp. *tritici*. *Fungal Genet. Biol.* 2010, 47, 828–838.
11. Ali, S.; Leconte, M.; Rahman, H.; Saqib, M.S.; Gladieux, P.; Enjalbert, J.; De Vallavieille-Pope, C. A high virulence and pathotype diversity of *Puccinia striiformis* f. sp. *tritici* at its centre of diversity, the Himalayan region of Pakistan. *Eur. J. Plant Pathol.* 2014, 140, 275–290.
12. Mehmood, S.; Sajid, M.; Zhao, J.; Khan, T.; Zhan, G.; Huang, L.; Kang, Z. Identification of *Berberis* species collected from the Himalayan Region of Pakistan Susceptible to *Puccinia striiformis* f. sp. *tritici*. *Plant Dis.* 2019, 103, 461–467.
13. Jin, Y.; Szabo, L.J.; Carson, M. Century-old mystery of *Puccinia striiformis* life history solved with the identification of *Berberis* as an alternate host. *Phytopathology* 2010, 100, 432–435.
14. Jin, Y. Role of *Berberis* spp. as alternate hosts in generating new races of *Puccinia graminis* and *P. striiformis*. *Euphytica* 2011, 179, 105–108.
15. Zhao, J.; Wang, L.; Wang, Z.; Chen, X.; Zhang, C.; Yao, J.; Zhan, G.; Chen, W.; Huang, L.; Kang, Z.; et al. Identification of eighteen *Berberis* species as alternate hosts of *Puccinia striiformis* f. sp. *tritici* and virulence variation in the pathogen isolates from natural infection of barberry plants in China. *Phytopathology* 2013, 103, 927–934.
16. Wang, Z.; Zhao, J.; Chen, X.; Peng, Y.; Ji, J.; Zhao, S.; Lv, Y.; Huang, L.; Kang, Z. Virulence variations of *Puccinia striiformis* f. sp. *tritici* isolates collected from *Berberis* spp. in China. *Plant Dis.* 2016, 100, 131–138.

17. Berlin, A.; Kyaschenko, J.; Justesen, A.F.; Yuen, J. Rust fungi forming aecia on *Berberis* spp. in Sweden. *Plant Dis.* 2013, 97, 1281–1287.
18. Zhao, J.; Zhao, S.L.; Peng, Y.L.; Qin, J.F.; Huang, L.L.; Kang, Z.S. Investigation on geographic distribution and identification of six *Berberis* spp. serving as alternate host for *P. striiformis* f. sp. *tritici* in Linzhi, Tibet. *Acta Phytopathol. Sin.* 2016, 46, 103–111.
19. Hovmöller, M.; Justesen, A.F.; Brown, J.K.M. Clonality and long-distance migration of *Puccinia striiformis* f. sp. *tritici* in north-west Europe. *Plant Pathol.* 2002, 51, 24–32.
20. Mboup, M.; Leconte, M.; Gautier, A.; Wan, A.; Chen, W.; De Vallavieille-Pope, C.; Enjalbert, J. Evidence of genetic recombination in wheat yellow rust populations of a Chinese oversummering area. *Fungal Genet. Biol.* 2009, 46, 299–307.
21. Duan, X.; Tellier, A.; Wan, A.; Leconte, M.; De Vallavieille-Pope, C.; Enjalbert, J. *Puccinia striiformis* f. sp. *tritici* presents high diversity and recombination in the over-summering zone of Gansu, China. *Mycologia* 2010, 102, 44–53.
22. Wang, M.N.; Chen, X.M. First report of Oregon grape (*Mahonia equifolium*) as an alternate host for the wheat stripe rust pathogen (*Puccinia striiformis* f. sp. *tritici*) under artificial conditions. *Plant Dis.* 2015, 97, 839.
23. Roelfs, A.P.; Singh, R.P.; Saari, E.E. *Rust Diseases of Wheat: Concepts and Methods of Disease Management*; CIMMYT: Mexico City, Mexico, 1992; pp. 1–81.
24. Ellis, J.; Lagudah, E.; Spielmeyer, W.; Dodds, P.N. The past, present and future of breeding rust resistant wheat. *Front. Plant Sci.* 2014, 5, 641.
25. Villaseñor-Espín, O.M.; Huerta-Espino, J.; Gerardo Leyva Mir, S.; Villaseñor-Mir, H.E.; Singh, R.P.; Sergio Sandoval-Islas, J.; Espitia-Rangel, E. Genetics of the yellow rust resistance in adult plantas of wheat cultivars. *Rev. Fitotec. Mex.* 2009, 32, 217–223. Available online: <http://www.revistafitotecniamexicana.org/documentos/32-3/6a.pdf> (accessed on 25 May 2020).
26. Khanfri, S.; Boulif, M.; Lahlali, R. Yellow rust (*Puccinia striiformis*): A serious threat to wheat production worldwide. *Not. Sci. Biol.* 2018, 10, 410–423.
27. Chen, X. Epidemiology and control of stripe rust on wheat. *Can. J. Plant Pathol.* 2005, 27, 314–337.
28. Chen, X. Pathogens which threaten food security: *Puccinia striiformis*, the wheat stripe rust pathogen. *Food Secur.* 2020, 12, 239–251.
29. Zadoks, J.C. Yellow rust on wheat studies in epidemiology and physiologic specialization. *Eur. J. Plant Pathol.* 1961, 67, 69–256.
30. Hovmöller, M.; Walter, S.; Bayles, R.A.; Hubbard, A.; Flath, K.; Sommerfeldt, N.; Leconte, M.; Czembor, P.C.; Rodriguez-Algabe, J.; Thach, T.; et al. Replacement of the European wheat yellow rust population by new races from the centre of diversity in the near-Himalayan region. *Plant Pathol.* 2015, 65, 402–411.
31. Stubbs, R. Stripe Rust. In *Diseases, Distribution, Epidemiology, and Control*; Elsevier: Amsterdam, The Netherlands, 1985; Volume II, pp. 61–101.
32. Hovmöller, M.; Sørensen, C.; Wälter, S.; Justesen, A.F. Diversity of *Puccinia striiformis* on cereals and grasses. *Annu. Rev. Phytopathol.* 2011, 49, 197–217.
33. De Vallavieille-Pope, C.; Ali, S.; Leconte, M.; Enjalbert, J.; Délos, M.; Rouzet, J. Virulence dynamics and regional structuring of *Puccinia striiformis* f. sp. *tritici* in France between 1984 and 2009. *Plant Dis.* 2012, 96, 131–140.

34. Carleton, M.A. A serious new wheat rust in this country. *Science* 1915, 42, 58–59.
35. Rudorf, W.; Job, M. Untersuchungen bezüglich des Spezialisierung von *Puccinia graminis* *tritici*. *Puccinia trilicina* und *Puccinia glumarum* *tritici*, sowie über Resistenz und ihre Vererbung in verschiedenen Kreuzungen. *Pflanzenzüchtung*. 1934, 19, 333–365.
36. Wellings, C.R. *Puccinia striiformis* in Australia: A review of the incursion, evolution, and adaptation of stripe rust in the period 1979–2006. *Aust. J. Agric. Res.* 2007, 58, 567–575.
37. Boshoff, W.H.P.; Pretorius, Z.A.; Van Niekerk, B.D. Establishment, distribution, and pathogenicity of *Puccinia striiformis* f. sp. *tritici* in South Africa. *Plant Dis.* 2002, 86, 485–492.
38. Milus, E.A.; Kristensen, K.; Hovmöller, M. Evidence for increased aggressiveness in a recent widespread strain of *Puccinia striiformis* f. sp. *tritici* causing stripe rust of wheat. *Phytopathology* 2009, 99, 89–94.
39. Hubbard, A.; Lewis, C.M.; Yoshida, K.; Ramírez-Gonzalez, R.; De Vallavieille-Pope, C.; Thomas, J.; Kamoun, S.; Bayles, R.; Uauy, C.; Saunders, D.G.O.; et al. Field pathogenomics reveals the emergence of a diverse wheat yellow rust population. *Genome Biol.* 2015, 16, 23.
40. Waqar, A.; Khattak, S.H.; Begum, S.; Rehman, T.; Rabia, R.; Shehzad, A.; Ajmal, W.; Zia, S.S.; Siddiqi, I.; Ali, G.M.; et al. Stripe rust: A review of the disease, Yr genes and its molecular markers. *Sarhad J. Agric.* 2018, 34, 188–201.
41. Wan, A.; Wang, X.; Kang, Z.; Chen, X. Variability of the stripe rust pathogen. In Stripe Rust; Springer Science and Business Media LLC: Berlin/Heidelberg, Germany, 2017; pp. 35–154.
42. Chen, W.; Wellings, C.; Chen, X.; Kang, Z.; Liu, T. Wheat stripe (yellow) rust caused by *Puccinia striiformis* f. sp. *tritici*. *Mol. Plant Pathol.* 2014, 15, 433–446.
43. Rodriguez-Algaba, J.; Wälter, S.; Sørensen, C.; Hovmöller, M.; Justesen, A.F. Sexual structures and recombination of the wheat rust fungus *Puccinia striiformis* on *Berberis vulgaris*. *Fungal Genet. Biol.* 2014, 70, 77–85.
44. Rodriguez-Algaba, J.; Sørensen, C.; Labouriau, R.; Justesen, A.F.; Hovmöller, M. Genetic diversity within and among aecia of the wheat rust fungus *Puccinia striiformis* on the alternate host *Berberis vulgaris*. *Fungal Biol.* 2017, 121, 541–549.

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