

Endocannabinoid System Present in the Glioblastoma Tumour Microenvironment

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Contributor: Mendhi Henna Dasram, Pavesan Naidoo, Roderick B. Walker, Sandile M. Khamanga

The highly aggressive and invasive glioblastoma (GBM) tumour is the most malignant lesion among adult-type diffuse gliomas, representing the most common primary brain tumour in the neuro-oncology practice of adults. With a poor overall prognosis and strong resistance to treatment, this nervous system tumour requires new, innovative treatment. GBM is a polymorphic tumour consisting of an array of stromal cells and various malignant cells contributing to tumour initiation, progression, and treatment response. Cannabinoids possess anti-cancer potencies against glioma cell lines and in animal models. To improve existing treatment, cannabinoids as functionalised ligands on nanocarriers were investigated as potential anti-cancer agents. The GBM tumour microenvironment is a multifaceted system consisting of resident or recruited immune cells, extracellular matrix components, tissue-resident cells, and soluble factors.

Keywords: endocannabinoid system ; glioblastoma tumour ; glioblastoma tumour microenvironment

1. Introduction

The latest version of the Central Nervous System (CNS) tumour classification published by the World Health Organization (WHO) summarises updates from the Consortium to inform molecular and practical approaches to CNS tumour taxonomy work ^[1]. The classification of tumours by the World Health Organization is an important tool for the diagnosis and treatment of brain tumours, including glioblastoma multiforme, the most common and aggressive malignant primary brain tumour in adults ^[2]. GBMs are highly invasive and diffuse tumours characterised by rapid proliferation, angiogenesis, and resistance to therapy ^[3]. Despite significant progress in our understanding of GBM biology and the development of novel therapeutic approaches, the prognosis for GBM patients remains poor ^[4].

One of the main challenges in treating GBM is the highly complex and dynamic nature of the GBM tumour microenvironment, which plays a crucial role in tumour growth, invasion, and resistance to therapy ^[5]. The GBM tumour microenvironment comprises various cell types, including tumour cells, astrocytes, microglia, endothelial cells, and immune cells, as well as extracellular matrix components, growth factors, and cytokines ^{[6][7]}. The interactions between these components create a highly heterogeneous and dynamic environment that facilitates tumour progression and adaptation to therapy ^[8]. Recent studies have highlighted the importance of the macroenvironment and microbiome in GBM pathogenesis and treatment ^{[9][10]}. Tumours release factors that drive the orchestration of an environment in the host that involves the crosstalk between multiple distal compartments at places beyond tumour beds ^[11]. Systemic alterations include changes in the bone marrow's functioning, where myelopoiesis is especially heavily altered in the presence of a tumour ^{[12][13]}. Distal hormonal signals and inflammatory mediators generated through interactions with commensal microorganisms also facilitate the formation of premetastatic niches where disseminated tumour cells call home, lay dormant, and eventually develop into growing metastatic ^{[14][15][16][17]}. Together, these inflammatory, tumour-promoting pro-metastatic networks form a systemic "macroenvironment" in tumour-bearing hosts that influence distant tissues' function and the tumour itself ^[18]. In the current era of personalised medicine, identifying and comprehensively understanding cancer's pathophysiological mechanisms are crucial for tailoring therapies based on grade, histological features, molecular subtypes, aggressiveness, and treatment response.

The endocannabinoid system (ECS) is a widespread neuromodulatory network that plays a role in the development and maturation of nervous systems by modulating network function and neuronal activity ^[19]. G-protein coupled cannabinoid receptors including the canonical receptor subtypes cannabinoid receptor type 1 (CB1-R) and cannabinoid receptor type 2 (CB2-R), endogenous cannabinoids known as endocannabinoids (e.g., anandamide and 2-arachidonoylglycerol), and the proteins that synthesize and degrade endocannabinoids, fatty acid amide hydrolase (FAAH) and monoacylglycerol lipase (MAGL), comprise the endocannabinoid system ^{[20][21]}. In addition to the enzymes involved in the biosynthesis and degradation of endocannabinoids, the other "non-canonical" extended signalling network of the ECS include receptors

GPR55 and PPAR α , inotropic cannabinoid receptors (TRP channels), protein transporters (FABP family), and other fatty acid derivatives [22][23][24][25]. While cannabinoid receptors are present in most tissues, CB1-R is primarily found mainly in the CNS, moderately found in adipose, endocrine, lymphoid, and female tissues and in smaller amounts found in other tissues [26]. The endocannabinoid system has emerged as a potential target for treating GBM [27]. The ECS is a complex signalling system that plays a crucial role in maintaining homeostasis in the body [28]. The ECS involves various physiological processes, including pain modulation, appetite, mood, and immune function [29]. Recent evidence suggests that the ECS is dysregulated in GBM and that its manipulation could represent a promising therapeutic strategy [30][31]. In particular, CB1-R and CB2-R are expressed in GBM cells and the tumour microenvironment, including immune cells and endothelial cells [32][33][34]. Activation of these receptors has been shown to induce antitumour effects in preclinical studies, including inhibition of tumour cell proliferation, migration, invasion, and angiogenesis [35].

Moreover, cannabinoid-induced apoptosis has been reported in GBM cells [36]. Despite the promising preclinical data, the clinical translation of most cannabinoids for the treatment of GBM faces significant challenges, including poor water solubility, limited bioavailability, and poor pharmacokinetics [37]. These limitations have led to the investigation of novel drug delivery systems, including nanocarriers, which have shown promising results in preclinical studies [38]. Nanocarriers are nanoscale drug delivery systems that can encapsulate hydrophobic drugs, such as cannabinoids, and protect them from degradation, enhance their solubility, and increase their bioavailability [39].

2. Cannabinoids as a Promising Adjuvant in the Treatment of GBM

The endocannabinoid system which includes endocannabinoids and the enzymes that synthesise and degrade them, and the transporters and G-protein coupled receptors involved in their signalling have been found in glioblastoma cells [31][40]. The ECS is a homeostatic system that uses lipid-derived signalling molecules to regulate a wide range of physiological functions [41]. Studies have shown high levels of cannabinoid receptors, CB1-R and CB2-R, as well as the transient receptor potential vanilloid 1 receptor expressed on glioblastoma cells, which are regulated by genetic and epigenetic mechanisms [42]. Although the expression levels obtained by immunohistochemistry are heterogeneous and dependent on the age of the patient and the histopathological origin of the brain tumour cells, CB2-R expression has been positively correlated with tumour grade and upregulated in most glioblastomas [43]. According to immunohistochemical analysis, both CB1 and CB2 receptors were detected in around 38% and 54% of glioblastoma endothelial cells, respectively [44]. CB2-R expression levels were found to be higher than CB1 in glioblastoma tissues. These findings suggest that selective CB2-R agonists could potentially serve as crucial targets for the treatment of glioma. The term “cannabinoids” originally described bioactive constituents of the *Cannabis sativa* plant. It is now an umbrella term covering a broad range of compounds subsectioned into the synthetic cannabinoids, the phytocannabinoids, and the endogenous cannabinoids, most of which are ligands which bind to endogenous cannabinoid (e.g., CB1-R and CB2-R) and other G-protein coupled receptors [44][45]. The endogenous cannabinoids are naturally occurring lipid mediators that are synthesised from the membrane phospholipids of cells [46].

Table 1 provides an overview of the main classes of cannabinoids: classical cannabinoids, non-classical cannabinoids, aminoalkylindoles, and eicosanoids [47]. The table summarises the structural characteristics, formulation strategies, and metabolism for each class. This information can be used to understand the unique properties of each class of cannabinoids.

Table 1. Cannabinoids: A Classification Based on Structural Features and Pharmacological Effects.

Classical Cannabinoids
Classical cannabinoids are the most well-known group of cannabinoids, and they are found in the cannabis plant. They have a highly lipophilic structure and poor water solubility due to their characteristic tricyclic terpenophenolic structure [48]. This lipophilicity facilitates easy passage through the lipid bilayers of cell membranes, influencing their absorption and distribution. Classical cannabinoids are extensively metabolised in the liver, primarily by cytochrome P450 enzymes, leading to a variety of metabolites, some of which are active and contribute to its pharmacological effects [49][50]. The high lipophilicity and poor water solubility of classical cannabinoids pose challenges in formulating them for aqueous-based delivery systems [51]. Techniques like nanoemulsions, liposomes, or microencapsulation may be employed to enhance solubility and bioavailability. Examples: THC, CBD, CBN
Non-Classical Cannabinoids
Non-classical cannabinoids, often synthetic cannabinoids that are not found in the cannabis plant, can be designed to have specific physicochemical properties [52]. They may be more potent and selective for cannabinoid receptors than classical cannabinoids [53]. They may be designed to have increased metabolic stability, thereby prolonging their duration of action [54]. However, their synthetic nature might lead to unpredictable metabolism and potential toxic metabolites. Formulation strategies would depend on the specific properties of the compound. Solubility enhancement and targeted delivery systems could be key considerations. Examples: CP 47497, CP 55940

Aminoalkylindoles

Aminoalkylindoles have a simpler, more stable structure compared to classical cannabinoids. The aminoalkylindole chemical class can be subdivided into four groups: naphthoylindoles, phenylacetylindoles, benzoylindoles, and naphthylmethylindoles [59]. This influences their interaction with cannabinoid receptors, making them more selective for cannabinoid receptors [56]. These compounds generally have high lipophilicity and may show significant brain penetration due to their ability to cross the blood–brain barrier efficiently. Similar to classical cannabinoids, addressing solubility and stability issues is critical [57]. There's also a need to consider the potential for rapid onset of action due to efficient CNS penetration. Examples: WIN-55212-2, JWH-018

Eicosanoids

Endocannabinoids, including anandamide and 2-AG, are derived from fatty acids, making them lipophilic structures [58][59]. This allows easier cellular uptake and interaction with cannabinoid receptors [60]. Endocannabinoids are rapidly metabolised in the body, which can limit their therapeutic use unless modifications or delivery systems are employed to stabilise them [61]. Enhancing stability and prolonging the duration of action are primary goals. Techniques might include the use of enzyme inhibitors to prevent rapid degradation or using advanced delivery systems to target specific tissues. Examples: Anandamide, 2-AG

Cannabinoid receptor activation can lead to the modulation of downstream signalling pathways in glioblastoma cells, including the PI3K/Akt/mTOR pathway, the MAPK/ERK pathway, and the c-Jun N-terminal kinase (JNK) pathway [62][63]. The activation of these pathways can have diverse effects on cell proliferation, differentiation, survival, and migration, depending on the specific context and the balance of signalling inputs. In addition to the modulation of signalling pathways, cannabinoids can also regulate gene expression in glioblastoma cells. For example, some cannabinoids, such as THC, have the capacity to influence the expression of the tumour suppressor gene for p53 [64], while inhibiting the expression of genes involved in cell cycle progression and angiogenesis, such as cyclin A and D1 and VEGF [65][66][67][68]. The molecular mechanisms of cannabinoid action in glioblastoma are complex and involve both receptor-dependent and -independent pathways. In addition to the modulation of the ECS and downstream signalling pathways, cannabinoids can also interact with other targets, such as ion channels, other G protein-coupled receptors, and nuclear receptors [69][70][71][72]. An increasing number of preclinical models and clinical studies have investigated the anti-cancer effects of cannabinoids on a variety of cancers [73]. Reports have shown a dysregulation of cannabinoid receptors and endogenous ligands present in the tumour microenvironment of cancerous tumours; however, the 'endocannabinoid's system role suggests both pro-tumourigenic and anti-cancer effects based on the type and site of cancer [74]. Some authors attribute these inconsistencies to an incomplete elucidation of this complex biological system, the bystander effect or the heterogeneity of receptors present in the disease state [42]. An important systematic review that the 2017 National Academy of Sciences committee used to review the health effects of cannabis-focused on gliomas and identified 2260 studies, of which 35 met the inclusion criteria [75][76]. Sixteen of these studies were *in vivo* studies which described the anti-cancer effects of cannabinoids on glioma tumours [31]. Meanwhile, many *in vitro* and preclinical studies in animal models have successfully shown the anti-cancer effects of cannabinoids based on the reduction of tumour growth via the inhibition of tumour cell proliferation and angiogenesis, the tumour microenvironment, induction of tumour cell death, and inhibition of invasion through the genetic or pharmacological modulation of cannabinoid and other receptors [68][77][78][79][80][81]. A study assessing the need for the addition of serum to *in vitro* testing conditions of cannabinoids reaffirmed the importance of mimicking the tumour microenvironment *in vitro* and warned about the high degree to which cannabinoids bind to plastic *in vitro* [82]. This is because the tumour microenvironment is a complex and dynamic environment that can influence the efficacy of cannabinoids. By mimicking the tumour microenvironment *in vitro*, researchers can develop more accurate and predictive models of cannabinoid activity [83]. This can help to prevent clinical failure associated with differences between *in vitro* models and human subjects. A study investigated the *in vitro* and *in vivo* efficacy of cannabidiol (CBD) in neuroblastoma, a nervous system tumour in children [84]. Two cannabinoids, tetrahydrocannabinol (THC) and CBD were experimentally tested to determine the effects of the compounds on invasiveness, programmed cell death, viability, and cell cycle distribution in human neuroblastoma cells *in vitro*. The cannabinoids were also evaluated for their ability to reduce the growth of tumour xenografts *in vivo* in mice. The results showed that both THC and CBD had antitumourigenic activity *in vitro*. However, CBD was more active than THC in reducing the invasiveness, apoptosis, viability, and cell cycle distribution of neuroblastoma cells. *In vivo*, CBD also showed greater efficacy than THC in reducing the growth of tumour xenografts in mice. Further studies are needed to confirm these findings and to evaluate the safety and efficacy of CBD in clinical trials.

The levels of endocannabinoids and expression of their receptors present in the glioblastoma microenvironment are dysregulated in the disease state and this dysregulation is thought to contribute to the growth and progression of GBM tumours [27]. **Figure 1** shows the pathways triggered by cannabinoid receptor interaction, which affect the hallmarks of cancer associated with glioblastoma tumours.

Legend

- ⬆ Pathway activation
- ⬇ Pathway inhibition
- VEGF: vascular endothelial growth factor
- PKC/p38: protein kinase C
- ERK: extracellular signal-regulated kinase
- MMPs: metalloproteinase
- TIMPs: tissue inhibitor of metalloproteinases
- TRPV-2: transient receptor potential cation channel subfamily V member 2
- AML1a: acute myeloid leukemia 1a
- GSCs: glioma stem cells
- ROS: reactive oxygen species
- GSH: glutathione
- P38-MAPK: P38 mitogen-activated protein kinase
- PI3K/Akt: phosphoinositide 3-kinase/ protein kinase B
- Raf/MEK/ERK: rapidly accelerated fibrosarcoma/ mitogen-activated protein kinase kinase/ extracellular signal-regulated kinase
- Akt/mTORC: protein kinase B/ mammalian target of rapamycin complex
- P16ENK4A: protein 16 inhibitor of cyclin-dependent kinase 4A
- E2F1/Cyclin A: E2F transcription factor 1/cyclin A

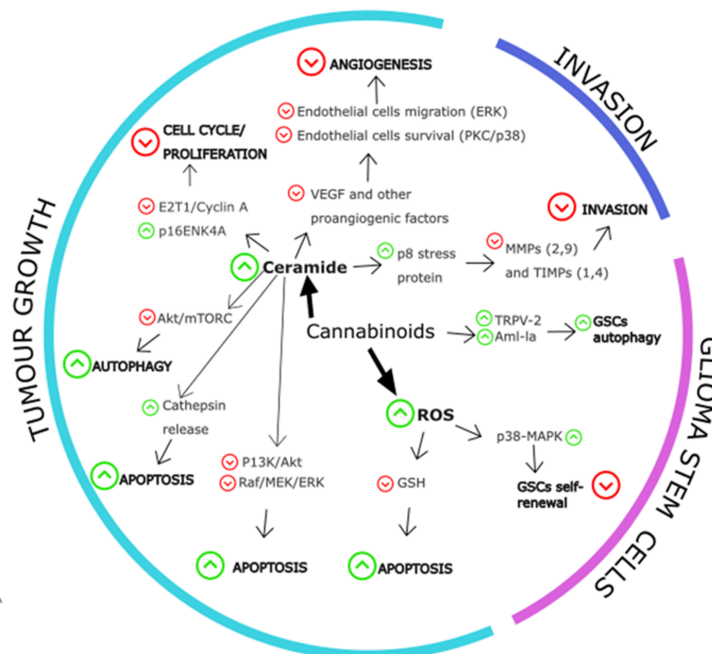


Figure 1. The main molecular mechanisms affected by cannabinoids during the modulation of GBM are depicted where the green arrows represent pathway activation, and the red arrows represent pathway inhibition [85]. Redrawn with permission from Dumitru, C.A., *Cannabinoids in Glioblastoma Therapy: New Applications for Old Drugs*; published by Front Mol Neurosci, 2018. Creative Commons CC BY 4.0. <http://creativecommons.org/licenses/by/4.0/> (accessed on 26 June 2023).

The mechanisms involved in the effect of cannabinoids on GBM tumour growth include cell death-inducing mechanisms, anti-angiogenic mechanisms, and anti-proliferation mechanisms. Cannabinoid-induced cell death is prompted by the activation of the intrinsic apoptosis pathway by cannabinoid-receptor interaction, which results in increased intracellular ceramide, thereby inhibiting pathways PI3K/Akt and Raf1/MEK/ERK [63]. The PI3K/Akt/mTOR pathway is a key signalling pathway that regulates cell proliferation, survival, and metabolism. Activation of CB1 and CB2 receptors by cannabinoids has been shown to inhibit the PI3K/Akt/mTOR pathway in glioblastoma cells, leading to a decrease in cell proliferation and an increase in apoptosis and autophagy [86]. This effect is thought to be mediated by the inhibition of Akt phosphorylation and activation and the downregulation of downstream targets such as mTOR, p70S6K, and 4EBP1 [87].

Cannabinoid-induced apoptosis is also triggered by oxidative stress, as seen when glioma cells treated with CBD caused an increase in reactive oxygen species (ROS) formation [88][89]. The MAPK/ERK pathway is another important signalling pathway that regulates cell proliferation, differentiation, and survival. Activation of CB1 and CB2 receptors by cannabinoid ligands has been shown to modulate the MAPK/ERK pathway in glioblastoma cells, leading to a decrease in angiogenesis and an increase in apoptosis [90][91]. This effect is thought to be mediated by the inhibition of ERK phosphorylation and activation and the downregulation of downstream targets such as c-fos and c-jun.

In a recent study, a standard mix of *cannabis-extracted* active fractions F4 and F5 was found to induce apoptosis and expression of endoplasmic reticulum (ER)-stress-associated genes in glioblastoma cells [92]. The fractions F4 and F5 also inhibited cell migration and invasion, altered cell cytoskeletons, and inhibited colony formation in 2 and 3-dimensional models. The study suggests that combinations of cannabis compounds exert cytotoxic, anti-proliferative, and anti-migratory effects on glioblastoma cells. The JNK pathway is a stress-activated signalling pathway that regulates cell survival and apoptosis [93]. Activation of CB1 and CB2 receptors by cannabinoids has been shown to activate the JNK pathway in glioblastoma cells, leading to increased apoptosis [94]. This effect is thought to be mediated by the activation of JNK phosphorylation and the upregulation of downstream targets such as c-jun. Further research is needed to fully understand the molecular mechanisms of cannabinoid action in glioblastoma, as well as the potential for developing cannabinoid-based therapies for this deadly disease [95].

The physicochemical properties of most traditional cannabinoids, which include high lipophilicity, poor water solubility, and chemical instability, present significant formulation challenges for the development of effective therapies for brain tumours. However, advances in pharmaceutical science and technology are helping to overcome these challenges and to harness the potential of cannabinoids for the treatment of brain tumours. The lipophilic nature of cannabinoids may be beneficial for cannabinoid delivery to the brain but tend to lead to the formation of colloidal aggregates, which artefacts in early drug discovery and proves difficult to achieve suitable solubility and stability in aqueous solutions. However, they possess an attractive composition as nanoparticle formulations for targeted drug delivery. A combination of ligand proteins and

polymers may be used to stabilise the colloidal aggregates, reduce colloid size, and improve longevity in blood circulation [96]. Besides the high hydrophobicity associated with most cannabinoids, including THC, the ability to elicit CB1-R mediated psychoactivity is one of the most noted drawbacks of cannabinoid therapeutic use [97][98]. All sources of evidence investigated in a recent study, including randomised controlled trials, observational studies, and Mendelian Randomisation studies, have consistently indicated the use of cannabis is associated with an increased risk of psychosis and a potentially increased risk of psychiatric symptoms such as mania [99][100]. A systematic review of the safety of cannabinoids for medical use was conducted [101]. There is insufficient data on the safety of cannabinoids, but most studies reported no adverse events (AEs) with acute administration and mild to moderate AEs with chronic administration. The most common AEs reported were drowsiness, fatigue, and dry mouth [102]. An association between cognitive impairment and cannabis has been shown in observational studies and randomized controlled trials, which have also been associated with motor vehicle accidents [103]. While CBD has demonstrated promising efficacy in various clinical trials, it is essential to recognize its intrinsic pharmacological effects, potential adverse drug events, and the possibility of pharmacokinetic and pharmacodynamic drug-drug interactions [104]. Given the increasing prevalence of CBD use among patients with complex medical conditions and treatment regimens, as well as its widespread availability as a consumer product, a comprehensive understanding of CBD's safety profile is paramount [105]. Further research is needed to better understand the safety of cannabinoids for medical use.

References

1. Horbinski, C.; Berger, T.; Packer, R.J.; Wen, P.Y. Clinical Implications of the 2021 Edition of the WHO Classification of Central Nervous System Tumours. *Nat. Rev. Neurol.* 2022, 18, 515–529.
2. Merve, A.; Millner, T.O.; Marino, S. Integrated Phenotype–Genotype Approach in Diagnosis and Classification of Common Central Nervous System Tumours. *Histopathology* 2019, 75, 299–311.
3. Brat, D.J.; Aldape, K.; Colman, H.; Holland, E.C.; Louis, D.N.; Jenkins, R.B.; Kleinschmidt-DeMasters, B.K.; Perry, A.; Reifenberger, G.; Stupp, R.; et al. CIMPACT-NOW Update 3: Recommended Diagnostic Criteria for “Diffuse Astrocytic Glioma, IDH-Wildtype, with Molecular Features of Glioblastoma, WHO Grade IV”. *Acta Neuropathol.* 2018, 136, 805–810.
4. Grech, N.; Dalli, T.; Mizzi, S.; Meilak, L.; Calleja, N.; Zrinzo, A.; Grech, N.; Dalli, T.; Mizzi, S.; Meilak, L.; et al. Rising Incidence of Glioblastoma Multiforme in a Well-Defined Population. *Cureus* 2020, 12, e8195.
5. Perus, L.J.M.; Walsh, L.A. Microenvironmental Heterogeneity in Brain Malignancies. *Front. Immunol.* 2019, 10, 2294.
6. Mukherjee, S.; Pillai, P.P. Current Insights on Extracellular Vesicle-Mediated Glioblastoma Progression: Implications in Drug Resistance and Epithelial-Mesenchymal Transition. *Biochim. Biophys. Acta (BBA)-Gen. Subj.* 2022, 1866, 130065.
7. Abels, E.R.; Maas, S.L.N.; Tai, E.; Ting, D.T.; Broekman, M.L.D.; Breakefield, X.O.; El Khoury, J. GliM&M: Web-Based Tool for Studying Circulating and Infiltrating Monocytes and Macrophages in Glioma. *Sci. Rep.* 2020, 10, 9898.
8. Broekman, M.L.; Maas, S.L.N.; Abels, E.R.; Mempel, T.R.; Krichevsky, A.M.; Breakefield, X.O. Multidimensional Communication in the Microenvirons of Glioblastoma. *Nat. Rev. Neurol.* 2018, 14, 482–495.
9. Allen, B.M.; Hiam, K.J.; Burnett, C.E.; Venida, A.; DeBarge, R.; TenVooren, I.; Marquez, D.M.; Cho, N.W.; Carmi, Y.; Spitzer, M.H. Systemic Dysfunction and Plasticity of the Immune Macroenvironment in Cancer Models. *Nat. Med.* 2020, 26, 1125.
10. Erdman, S.E.; Poutahidis, T. The Microbiome Modulates the Tumor Macroenvironment. *OncolImmunology* 2014, 3, e28271.
11. Sverdlov, E.D. Multidimensional Complexity of Cancer. Simple Solutions Are Needed. *Biochemistry* 2016, 81, 731–738.
12. Del Bianco, P.; Pinton, L.; Magri, S.; Canè, S.; Masetto, E.; Basso, D.; Padovan, M.; Volpin, F.; d'Avella, D.; Lombardi, G.; et al. Myeloid Diagnostic and Prognostic Markers of Immune Suppression in the Blood of Glioma Patients. *Front. Immunol.* 2022, 12, 5672.
13. Ugel, S.; De Sanctis, F.; Mandruzzato, S.; Bronte, V. Tumor-Induced Myeloid Deviation: When Myeloid-Derived Suppressor Cells Meet Tumor-Associated Macrophages. *J. Clin. Investig.* 2015, 125, 3365–3376.
14. Psaila, B.; Lyden, D. The Metastatic Niche: Adapting the Foreign Soil. *Nat. Rev. Cancer* 2009, 9, 285–293.
15. Lah, T.T.; Novak, M.; Breznik, B. Brain Malignancies: Glioblastoma and Brain Metastases. *Semin. Cancer Biol.* 2020, 60, 262–273.

16. Sceneay, J.; Smyth, M.J.; Möller, A. The Pre-Metastatic Niche: Finding Common Ground. *Cancer Metastasis Rev.* 2013, 32, 449–464.
17. Hambardzumyan, D.; Bergers, G. Glioblastoma: Defining Tumor Niches. *Trends Cancer* 2015, 1, 252–265.
18. Gabrilovich, D.I.; Ostrand-Rosenberg, S.; Bronte, V. Coordinated Regulation of Myeloid Cells by Tumours. *Nat. Rev. Immunol.* 2012, 12, 253–268.
19. Lu, H.C.; Mackie, K. Review of the Endocannabinoid System. *Biol. Psychiatry Cogn. Neurosci. Neuroimaging* 2021, 6, 607–615.
20. Ueda, N.; Tsuboi, K.; Uyama, T. Metabolism of Endocannabinoids and Related N-Acylethanolamines: Canonical and Alternative Pathways. *FEBS J.* 2013, 280, 1874–1894.
21. Khan, M.I.; Sobocinska, A.A.; Czarnecka, A.M.; Król, M.; Botta, B.; Szczylik, C. The Therapeutic Aspects of the Endocannabinoid System (ECS) for Cancer and Their Development: From Nature to Laboratory. *Curr. Pharm. Des.* 2016, 22, 1756–1766.
22. Contino, M.; McCormick, P.J. Editorial: The Canonical and Non-Canonical Endocannabinoid System as a Target in Cancer and Acute and Chronic Pain. *Front. Pharmacol.* 2020, 11, 312.
23. Biringer, R.G. Endocannabinoid Signaling Pathways: Beyond CB1R and CB2R. *J. Cell Commun. Signal.* 2021, 15, 335–360.
24. Behl, T.; Makkar, R.; Sehgal, A.; Singh, S.; Makeen, H.A.; Albratty, M.; Alhazmi, H.A.; Meraya, A.M.; Bungau, S. Exploration of Multiverse Activities of Endocannabinoids in Biological Systems. *Int. J. Mol. Sci.* 2022, 23, 5734.
25. Simard, M.; Archambault, A.S.; Lavoie, J.P.C.; Dumais, É.; Di Marzo, V.; Flamand, N. Biosynthesis and Metabolism of Endocannabinoids and Their Congeners from the Monoacylglycerol and N-Acyl-Ethanolamine Families. *Biochem. Pharmacol.* 2022, 205, 115261.
26. Abyadeh, M.; Gupta, V.; Paulo, J.A.; Gupta, V.; Chitranshi, N.; Godinez, A.; Saks, D.; Hasan, M.; Amirkhani, A.; McKay, M.; et al. A Proteomic View of Cellular and Molecular Effects of Cannabis. *Biomolecules* 2021, 11, 1411.
27. Costas-Insua, C.; Guzmán, M. Endocannabinoid Signaling in Glioma. *Glia* 2022, 71, 127–138.
28. Braile, M.; Marcella, S.; Marone, G.; Galdiero, M.R.; Varricchi, G.; Loffredo, S. The Interplay between the Immune and the Endocannabinoid Systems in Cancer. *Cells* 2021, 10, 1282.
29. Matei, D.; Trofin, D.; Iordan, D.A.; Onu, I.; Condurache, I.; Ionite, C.; Buculei, I. The Endocannabinoid System and Physical Exercise. *Int. J. Mol. Sci.* 2023, 24, 1989.
30. Fraguas-Sánchez, A.I.; Martín-Sabroso, C.; Torres-Suárez, A.I. Insights into the Effects of the Endocannabinoid System in Cancer: A Review. *Br. J. Pharmacol.* 2018, 175, 2566–2580.
31. Rocha, F.C.M.; Dos Santos Júnior, J.G.; Stefano, S.C.; Da Silveira, D.X. Systematic Review of the Literature on Clinical and Experimental Trials on the Antitumor Effects of Cannabinoids in Gliomas. *J. Neurooncol.* 2014, 116, 11–24.
32. Wu, X.; Han, L.; Zhang, X.; Li, L.; Jiang, C.; Qiu, Y.; Huang, R.; Xie, B.; Lin, Z.; Ren, J.; et al. Alteration of Endocannabinoid System in Human Gliomas. *J. Neurochem.* 2012, 120, 842–849.
33. De Jesús, M.L.; Hostalot, C.; Garibi, J.M.; Sallés, J.; Meana, J.J.; Callado, L.F. Opposite Changes in Cannabinoid CB1 and CB2 Receptor Expression in Human Gliomas. *Neurochem. Int.* 2010, 56, 829–833.
34. Held-Feindt, J.; Dörner, L.; Sahan, G.; Mehdorn, H.M.; Mentlein, R. Cannabinoid Receptors in Human Astroglial Tumors. *J. Neurochem.* 2006, 98, 886–893.
35. Sredni, S.T.; Huang, C.C.; Suzuki, M.; Pundy, T.; Chou, P.; Tomita, T. Spontaneous Involution of Pediatric Low-Grade Gliomas: High Expression of Cannabinoid Receptor 1 (CNR1) at the Time of Diagnosis May Indicate Involvement of the Endocannabinoid System. *Child's Nerv. Syst.* 2016, 32, 2061–2067.
36. Carracedo, A.; Lorente, M.; Egia, A.; Blázquez, C.; García, S.; Giroux, V.; Malicet, C.; Villuendas, R.; Gironella, M.; González-Feria, L.; et al. The Stress-Regulated Protein P8 Mediates Cannabinoid-Induced Apoptosis of Tumor Cells. *Cancer Cell* 2006, 9, 301–312.
37. Onaivi, E.S.; Singh Chauhan, B.P.; Sharma, V. Challenges of Cannabinoid Delivery: How Can Nanomedicine Help? *Nanomedicine* 2020, 15, 2023–2028.
38. Ngwa, W.; Kumar, R.; Moreau, M.; Dabney, R.; Herman, A. Nanoparticle Drones to Target Lung Cancer with Radiosensitizers and Cannabinoids. *Front. Oncol.* 2017, 7, 208.
39. Deshpande, A.; Patil, T.S. Nanocarrier Technologies for Enhancing the Solubility and Dissolution Rate of Api. In *Medicinal Chemistry with Pharmaceutical Product Development*; Apple Academic Press: Cambridge, MA, USA, 2019; pp. 155–234.

40. Massi, P.; Vaccani, A.; Ceruti, S.; Colombo, A.; Abbracchio, M.P.; Parolaro, D. Antitumor Effects of Cannabidiol, a Nonpsychoactive Cannabinoid, on Human Glioma Cell Lines. *J. Pharmacol. Exp. Ther.* 2004, 308, 838–845.
41. Rakotoarivelo, V.; Sihag, J.; Flamand, N. Role of the Endocannabinoid System in the Adipose Tissue with Focus on Energy Metabolism. *Cells* 2021, 10, 1279.
42. Doherty, G.J.; de Paula, B.H.R. Cannabinoids in Glioblastoma Multiforme—Hype or Hope? *Br. J. Cancer* 2021, 124, 1341–1343.
43. Ellert-Miklaszewska, A.; Grajkowska, W.; Gabrusiewicz, K.; Kaminska, B.; Konarska, L. Distinctive Pattern of Cannabinoid Receptor Type II (CB2) Expression in Adult and Pediatric Brain Tumors. *Brain Res.* 2007, 1137, 161–169.
44. Chakravarti, B.; Ravi, J.; Ganju, R.K. Cannabinoids as Therapeutic Agents in Cancer: Current Status and Future Implications. *Oncotarget* 2014, 5, 5852.
45. Andre, C.M.; Hausman, J.F.; Guerriero, G. Cannabis Sativa: The Plant of the Thousand and One Molecules. *Front. Plant Sci.* 2016, 7, 19.
46. Turcotte, C.; Chouinard, F.; Lefebvre, J.S.; Flamand, N. Regulation of Inflammation by Cannabinoids, the Endocannabinoids 2-Arachidonoyl-Glycerol and Arachidonoyl-Ethanolamide, and Their Metabolites. *J. Leukoc. Biol.* 2015, 97, 1049–1070.
47. Shevyrin, V.A.; Morzherin, Y.Y. Cannabinoids: Structures, Effects, and Classification. *Russ. Chem. Bull.* 2015, 64, 1249–1266.
48. Bow, E.W.; Rimoldi, J.M. The Structure–Function Relationships of Classical Cannabinoids: CB1/CB2 Modulation. *Perspect. Med. Chem.* 2016, 8, 17.
49. Su, M.K.; Seely, K.A.; Moran, J.H.; Hoffman, R.S. Metabolism of Classical Cannabinoids and the Synthetic Cannabinoid JWH-018. *Clin. Pharmacol. Ther.* 2015, 97, 562–564.
50. Bardhi, K.; Coates, S.; Watson, C.J.W.; Lazarus, P. Cannabinoids and Drug Metabolizing Enzymes: Potential for Drug–Drug Interactions and Implications for Drug Safety and Efficacy. *Expert Rev. Clin. Pharmacol.* 2022, 15, 1443–1460.
51. Stella, B.; Baratta, F.; Della Pepa, C.; Arpicco, S.; Gastaldi, D.; Dosio, F. Cannabinoid Formulations and Delivery Systems: Current and Future Options to Treat Pain. *Drugs* 2021, 81, 1513–1557.
52. Bloom, A.S.; Edgemond, W.S.; Moldvan, J.C. Nonclassical and Endogenous Cannabinoids: Effects on the Ordering of Brain Membranes. *Neurochem. Res.* 1997, 22, 563–568.
53. Pop, E. Cannabinoids, Endogenous Ligands and Synthetic Analogs. *Curr. Opin. Chem. Biol.* 1999, 3, 418–425.
54. Zendulka, O.; Dovrtělová, G.; Nosková, K.; Turjap, M.; Šulcová, A.; Hanuš, L.; Juřica, J. Cannabinoids and Cytochrome P450 Interactions. *Curr. Drug Metab.* 2016, 17, 206–226.
55. Shevyrin, V.; Melkozerov, V.; Endres, G.W.; Shafran, Y.; Morzherin, Y. On a New Cannabinoid Classification System: A Sight on the Illegal Market of Novel Psychoactive Substances. *Cannabis Cannabinoid Res.* 2016, 1, 186–194.
56. Shim, J.-Y.; Collantes, E.R.; Welsh, W.J.; Howlett, A.C. Unified Pharmacophoric Model for Cannabinoids and Aminoalkylindoles. In *Molecular Modeling and Prediction of Bioactivity*; Springer: Boston, MA, USA, 2000; pp. 201–206.
57. Mardal, M.; Gracia-Lor, E.; Leibnitz, S.; Castiglioni, S.; Meyer, M.R. Toxicokinetics of New Psychoactive Substances: Plasma Protein Binding, Metabolic Stability, and Human Phase I Metabolism of the Synthetic Cannabinoid WIN 55,212-2 Studied Using in Vitro Tools and LC-HR-MS/MS. *Drug Test. Anal.* 2016, 8, 1039–1048.
58. Burstein, S.H. Eicosanoid Mediation of Cannabinoid Actions. *Bioorg. Med. Chem.* 2019, 27, 2718–2728.
59. Burstein, S.H.; Zurier, R.B. Cannabinoids, Endocannabinoids, and Related Analogs in Inflammation. *AAPS J.* 2009, 11, 109–119.
60. Deutsch, D.G. A Personal Retrospective: Elevating Anandamide (AEA) by Targeting Fatty Acid Amide Hydrolase (FAAH) and the Fatty Acid Binding Proteins (FABPs). *Front. Pharmacol.* 2016, 7, 370.
61. Rouzer, C.A.; Ghebreselasie, K.; Marnett, L.J. Chemical Stability of 2-Arachidonylglycerol under Biological Conditions. *Chem. Phys. Lipids* 2002, 119, 69–82.
62. Laezza, C.; Pagano, C.; Navarra, G.; Pastorino, O.; Proto, M.C.; Fiore, D.; Piscopo, C.; Gazzo, P.; Bifulco, M. The Endocannabinoid System: A Target for Cancer Treatment. *Int. J. Mol. Sci.* 2020, 21, 747.
63. Ellert-Miklaszewska, A.; Ciechomska, I.A.; Kaminska, B. Cannabinoid Signaling in Glioma Cells. In *Advances in Experimental Medicine and Biology*; Springer: Dordrecht, The Netherlands, 2013.
64. Downer, E.J.; Gowran, A.; Murphy, Á.C.; Campbell, V.A. The Tumour Suppressor Protein, P53, Is Involved in the Activation of the Apoptotic Cascade by Δ^9 -Tetrahydrocannabinol in Cultured Cortical Neurons. *Eur. J. Pharmacol.* 2007, 564, 57–65.

65. Sarfaraz, S.; Afaq, F.; Adhami, V.M.; Malik, A.; Mukhtar, H. Cannabinoid Receptor Agonist-Induced Apoptosis of Human Prostate Cancer Cells LNCaP Proceeds through Sustained Activation of ERK1/2 Leading to G 1 Cell Cycle Arrest. *J. Biol. Chem.* 2006, 281, 39480–39491.
66. Galanti, G.; Fisher, T.; Kventsel, I.; Shoham, J.; Gallily, R.; Mechoulam, R.; Lavie, G.; Amariglio, N.; Rechavi, G.; Toren, A. Δ^9 -Tetrahydrocannabinol Inhibits Cell Cycle Progression by Downregulation of E2F1 in Human Glioblastoma Multiforme Cells. *Acta. Oncol.* 2008, 47, 1062–1070.
67. Irrera, N.; D'ascola, A.; Pallio, G.; Bitto, A.; Mannino, F.; Arcoraci, V.; Rottura, M.; Ieni, A.; Minutoli, L.; Metro, D.; et al. β -Caryophyllene Inhibits Cell Proliferation through a Direct Modulation of CB2 Receptors in Glioblastoma Cells. *Cancers* 2020, 12, 1038.
68. Blázquez, C.; González-Feria, L.; Álvarez, L.; Haro, A.; Casanova, M.L.; Guzmán, M. Cannabinoids Inhibit the Vascular Endothelial Growth Factor Pathway in Gliomas. *Cancer Res.* 2004, 64, 5617–5623.
69. O'Reilly, E.M.; Cosgrave, J.M.; Gallagher, W.M.; Perry, A.S. Plant-Derived Cannabinoids as Anticancer Agents. *Trends Cancer* 2022, 8, 350–357.
70. Hinz, B.; Ramer, R. Cannabinoids as Anticancer Drugs: Current Status of Preclinical Research. *Br. J. Cancer* 2022, 127, 1–13.
71. Kolbe, M.R.; Hohmann, T.; Hohmann, U.; Ghadban, C.; Mackie, K.; Zöller, C.; Prell, J.; Illert, J.; Strauss, C.; Dehghani, F. THC Reduces Ki67-Immunoreactive Cells Derived from Human Primary Glioblastoma in a GPR55-Dependent Manner. *Cancers* 2021, 13, 1064.
72. Oesch, S.; Gertsch, J. Cannabinoid Receptor Ligands as Potential Anticancer Agents—High Hopes for New Therapies? *J. Pharm. Pharmacol.* 2010, 61, 839–853.
73. Mangal, N.; Erridge, S.; Habib, N.; Sadanandam, A.; Reebye, V.; Sodergren, M.H. Cannabinoids in the Landscape of Cancer. *J. Cancer Res. Clin. Oncol.* 2021, 147, 2507–2534.
74. Cherkasova, V.; Wang, B.; Gerasymchuk, M.; Fiselier, A.; Kovalchuk, O.; Kovalchuk, I. Use of Cannabis and Cannabinoids for Treatment of Cancer. *Cancers* 2022, 14, 5142.
75. Worster, B.; Hajjar, E.R.; Handley, N. Cannabis Use in Patients with Cancer: A Clinical Review. *JCO Oncol. Pract.* 2022, 18, 743–749.
76. Klimkiewicz, A.; Jasinska, A. The Health Effects of Cannabis and Cannabinoids: The Current State of Evidence and Recommendations for Research. *Psychiatria* 2017, 15, 88–92.
77. Sánchez, C.; Galve-Roperh, I.; Canova, C.; Brachet, P.; Guzmán, M. Δ^9 -Tetrahydrocannabinol Induces Apoptosis in C6 Glioma Cells. *FEBS Lett.* 1998, 436, 6–10.
78. Allister, S.D.; Chan, C.; Taft, R.J.; Luu, T.; Abood, M.E.; Moore, D.H.; Aldape, K.; Yount, G. Cannabinoids Selectively Inhibit Proliferation and Induce Death of Cultured Human Glioblastoma Multiforme Cells. *J. Neurooncol.* 2005, 74, 31–40.
79. End, D.W.; Thoursen, K.; Dewey, W.L.; Carchman, R.A. A Comparative Study of the Disposition of Tetrahydrocannabinol in Neuroblastoma and Glioma Cells in Tissue Culture: Relation to Cellular Impairment. *Mol. Pharmacol.* 1977, 13, 864–871.
80. Velasco, G.; Galve-Roperh, I.; Sánchez, C.; Blázquez, C.; Guzmán, M. Hypothesis: Cannabinoid Therapy for the Treatment of Gliomas? *Neuropharmacology* 2004, 47, 315–323.
81. Bifulco, M.; Laezza, C.; Gazzero, P.; Pentimalli, F. Endocannabinoids as Emerging Suppressors of Angiogenesis and Tumor Invasion (Review). *Oncol. Rep.* 2007, 17, 813–816.
82. Carkaci-Salli, N.; Raup-Konsavage, W.M.; Karelia, D.; Sun, D.; Jiang, C.; Lu, J.; Vrana, K.E. Cannabinoids as Potential Cancer Therapeutics: The Concentration Conundrum. *Cannabis Cannabinoid Res.* 2023.
83. Jo, Y.; Choi, N.; Kim, K.; Koo, H.J.; Choi, J.; Kim, H.N. Chemoresistance of Cancer Cells: Requirements of Tumor Microenvironment-Mimicking in Vitro Models in Anti-Cancer Drug Development. *Theranostics* 2018, 8, 5259–5275.
84. Fisher, T.; Golan, H.; Schiby, G.; Prichen, S.; Smoum, R.; Moshe, I.; Peshes-Yaloz, N.; Castiel, A.; Waldman, D.; Gallily, R.; et al. In Vitro and in Vivo Efficacy of Non-Psychoactive Cannabidiol in Neuroblastoma. *Curr. Oncol.* 2016, 23, S15–S22.
85. Dumitru, C.A.; Sandalcioğlu, I.E.; Karsak, M. Cannabinoids in Glioblastoma Therapy: New Applications for Old Drugs. *Front. Mol. Neurosci.* 2018, 11, 159.
86. Salazar, M.; Carracedo, A.; Salanueva, Í.J.; Hernández-Tiedra, S.; Lorente, M.; Egia, A.; Vázquez, P.; Blázquez, C.; Torres, S.; García, S.; et al. Cannabinoid Action Induces Autophagy-Mediated Cell Death through Stimulation of ER Stress in Human Glioma Cells. *J. Clin. Investig.* 2009, 119, 1359–1372.

87. Ciechomska, I.A.; Gabrusiewicz, K.; Szczepankiewicz, A.A.; Kaminska, B. Endoplasmic Reticulum Stress Triggers Autophagy in Malignant Glioma Cells Undergoing Cyclosporine A-Induced Cell Death. *Oncogene* 2012, 32, 1518–1529.
88. Massi, P.; Valenti, M.; Solinas, M.; Parolaro, D. Molecular Mechanisms Involved in the Antitumor Activity of Cannabinoids on Gliomas: Role for Oxidative Stress. *Cancers* 2010, 2, 1013–1026.
89. Wang, K.; Wang, Q.; Li, Q.; Zhang, Z.; Gao, J.; Fan, C.; Sun, B.; Ni, Q. Cannabinoid WIN 55,212-2 Inhibits Human Glioma Cell Growth by Triggering ROS-Mediated Signal Pathways. *BioMed Res. Int.* 2021, 2021, 6612592.
90. Blázquez, C.; Casanova, M.L.; Planas, A.; Gómez del Pulgar, T.; Villanueva, C.; Fernández-Aceñero, M.J.; Aragonés, J.; Huffman, J.W.; Jorcano, J.L.; Guzmán, M. Inhibition of Tumor Angiogenesis by Cannabinoids. *FASEB J.* 2003, 17, 529–531.
91. Widmer, M.; Hanemann, C.O.; Zajicek, J. High Concentrations of Cannabinoids Activate Apoptosis in Human U373MG Glioma Cells. *J. Neurosci. Res.* 2008, 86, 3212–3220.
92. Peeri, H.; Shalev, N.; Vinayaka, A.C.; Nizar, R.; Kazimirsky, G.; Namdar, D.; Anil, S.M.; Belausov, E.; Brodie, C.; Koltai, H. Specific Compositions of Cannabis Sativa Compounds Have Cytotoxic Activity and Inhibit Motility and Colony Formation of Human Glioblastoma Cells In Vitro. *Cancers* 2021, 13, 1720.
93. De los Reyes Corrales, T.; Losada-Pérez, M.; Casas-Tintó, S. JNK Pathway in CNS Pathologies. *Int. J. Mol. Sci.* 2021, 22, 3883.
94. Fonseca, B.M.; Teixeira, N.A.; Correia-da-Silva, G. Cannabinoids as Modulators of Cell Death: Clinical Applications and Future Directions; Springer: Cham, Switzerland, 2017; pp. 63–88.
95. Rodriguez-Almaraz, J.E.; Butowski, N. Therapeutic and Supportive Effects of Cannabinoids in Patients with Brain Tumors (CBD Oil and Cannabis). *Curr. Treat. Options Oncol.* 2023, 24, 30–44.
96. Ganesh, A.N.; McLaughlin, C.K.; Duan, D.; Shoichet, B.K.; Shoichet, M.S. A New Spin on Antibody-Drug Conjugates: Trastuzumab-Fulvestrant Colloidal Drug Aggregates Target HER2-Positive Cells. *ACS Appl. Mater. Interfaces* 2017, 9, 12195–12202.
97. Guzmán, M.; Duarte, M.J.; Blázquez, C.; Ravina, J.; Rosa, M.C.; Galve-Roperh, I.; Sánchez, C.; Velasco, G.; González-Feria, L. A Pilot Clinical Study of Δ^9 -Tetrahydrocannabinol in Patients with Recurrent Glioblastoma Multiforme. *Br. J. Cancer* 2006, 95, 197–203.
98. Johnson, E.C.; Hatoum, A.S.; Deak, J.D.; Polimanti, R.; Murray, R.M.; Edenberg, H.J.; Gelernter, J.; Di Forti, M.; Agrawal, A. The Relationship between Cannabis and Schizophrenia: A Genetically Informed Perspective. *Addiction* 2021, 116, 3227–3234.
99. Marijuana and Madness; Castle, D.; Murray, R.M.; D'Souza, D.C. (Eds.) Cambridge University Press: Cambridge, UK, 2011.
100. Hindley, G.; Beck, K.; Borgan, F.; Ginestet, C.E.; McCutcheon, R.; Kleinloog, D.; Ganesh, S.; Radhakrishnan, R.; D'Souza, D.C.; Howes, O.D. Psychiatric Symptoms Caused by Cannabis Constituents: A Systematic Review and Meta-Analysis. *Lancet Psychiatry* 2020, 7, 344–353.
101. Larsen, C.; Shahinas, J. Dosage, Efficacy and Safety of Cannabidiol Administration in Adults: A Systematic Review of Human Trials. *J. Clin. Med. Res.* 2020, 12, 129–141.
102. Sawtelle, L.; Holle, L.M. Use of Cannabis and Cannabinoids in Patients with Cancer. *Ann. Pharmacother.* 2021, 55, 870–890.
103. Hostiuc, S.; Moldoveanu, A.; Negoii, I.; Drima, E. The Association of Unfavorable Traffic Events and Cannabis Usage: A Meta-Analysis. *Front. Pharmacol.* 2018, 9, 99.
104. Brown, J.D.; Winterstein, A.G. Potential Adverse Drug Events and Drug–Drug Interactions with Medical and Consumer Cannabidiol (CBD) Use. *J. Clin. Med.* 2019, 8, 989.
105. Buchtova, T.; Lukac, D.; Skrott, Z.; Chroma, K.; Bartek, J.; Mistrik, M. Drug–Drug Interactions of Cannabidiol with Standard-of-Care Chemotherapeutics. *Int. J. Mol. Sci.* 2023, 24, 2885.