Anti-Diabetic Mechanisms of Glycyrrhizic acid and Its Derivatives

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Type 2 diabetes mellitus (T2DM) is a chronic metabolic disease, which is characterized by hyperglycemia, chronic insulin resistance, progressive decline in β -cell function, and defect in insulin secretion. It has become one of the leading causes of death worldwide. There is no cure for T2DM, but it can be treated, and blood glucose levels can be controlled. It has been reported that diabetic patients may suffer from the adverse effects of conventional medicine. Therefore, alternative therapy, such as traditional Chinese medicine (TCM), can be used to manage and treat diabetes. Glycyrrhizic acid (GL) and its derivatives are suggested to be promising candidates for the treatment of T2DM and its complications. It is the principal bioactive constituent in licorice, one type of TCM.

glycyrrhizic acid glycyrrhetinic acid ammonium glycyrrhizinate diammonium glycyrrhizinate

type 2 diabetes mellitus

1. GL and Its Derivatives

Glycyrrhizic acid (GL) is an amphiphilic compound, its hydrophilic part is represented by the glucuronic acid residues, and its hydrophobic part is the glycyrrhetinic acid (GA) residue ^[1]. GA is not only a hydrolytic product of GL, but it is also one of the main bioactive components of licorice with two isoforms, 18α -GA and 18β -GA ^{[2][3]}. GL and its aglycon, GA, have promising therapeutic effects, which have attracted the researchers to develop them into clinical medicine, including anti-cancer, anti-inflammatory, anti-viral, anti-diabetic, and hepatoprotective effects ^{[4][5]}. However, studies found that the oral administration of GL is poorly absorbed from the gut, but it is slowly converted into 18β -GA under intestinal bacterial hydrolysis, and the hydrolysis product 18β -GA is absorbed in the gut ^{[§][2]]}. Therefore, due to the poor bioavailability of GL, it has been designed to conjugate with various metals or ammonium to form water-soluble salts for improving its bioavailability, including ammonium glycyrrhizinate (DG), and dipotassium glycyrrhizinate ^{[8][9][10]}. The structures of GL, GA, AG, and DG are shown in **Figure 1**.



Figure 1. Chemical structures of glycyrrhizic acid (GL) and its derivatives. They include two isomers of glycyrrhizic acid, namely, (**a**) 18α-glycyrrhizic acid (18α-GL) and (**b**) 18β-glycyrrhizic acid (18β-GL); two isomers of glycyrrhetinic acid (GA), namely, (**c**) 18α-glycyrrhetinic acid (18α-GA) and (**d**) 18β-glycyrrhetinic acid (18β-GA); (**e**) ammonium glycyrrhizinate (AG); and (**f**) diammonium glycyrrhizinate (DG).

2. Anti-Diabetic Mechanisms of GL and Its Derivatives in T2DM

Type 2 diabetes mellitus (T2DM) is mainly characterized by hyperglycemia, which eventually results in chronic insulin resistance in insulin-sensitive tissues and defective insulin secretion by pancreatic β -cells ^[11]. The antidiabetic effects of GL and its derivatives have been confirmed in vitro and in vivo. The mechanisms include improving glucose tolerance ^[12] and insulin sensitivity ^[13], regulating glucose homeostasis ^[13] and lipid metabolism ^[14], and enhancing insulin secretion ^[15]. A schematic diagram shows the mechanisms of GL and its derivatives in improving insulin resistance in the liver and adipose tissue (**Figure 2**).



Liver / Adipose tissue

Figure 2. Schematic diagram showing the mechanisms of glycyrrhizic acid (GL) and its derivatives in improving insulin resistance in the liver and adipose tissue. GL and its derivatives act on the insulin receptor to regulate gluconeogenesis and enhance glycogen synthesis via PI3K/Akt signaling pathway, and increase glucose uptake via GLUT4, thus regulating glucose homeostasis. On the other hand, they increase lipolysis through PI3K/Akt/HSL pathway and reduce fatty acid synthesis through downregulating SREBP-1c/FAS/SCD1 pathway, thereby regulating lipid metabolism. Therefore, GL and its derivatives improve insulin resistance through improving glucose homeostasis and lipid metabolism. FAS: Fatty acid synthetase; G6Pase: Glucose-6-phosphatase; GLUT4: Glucose transporter 4; GSK-3 β : Glycogen synthase kinase-3 β ; HSL: Hormone-sensitive lipase; IGF-1: Insulin-like growth factor 1; IL-6: Interleukin 6; IRS-1: Insulin receptor substrate 1; IRS-2: Insulin receptor substrate 2; PEPCK: Phosphoenolpyruvate carboxykinase; PI3K: Phosphoinositide 3-kinase; PTP1B: Protein tyrosine phosphatase 1B; SCD1: Stearoyl CoA desaturase 1; SREBP-1c: Sterol regulatory element-binding protein 1c; TNF- α : Tumor necrosis factor- α .

2.1. Insulin Resistance

Insulin is a hormone produced by the pancreas that controls glucose levels in the blood and helps in storing glucose in the insulin-sensitive organs, including skeletal muscle, fat, and the liver, where it is used to provide

energy. The inability of responding to insulin in insulin-sensitive metabolic tissues, known as insulin resistance, is one of the main pathophysiology of T2DM, thus the tissues cannot uptake glucose from the blood easily. As a result, the pancreas produces more insulin to help in reducing blood glucose levels ^{[16][17]}. At a later stage, the pancreatic β-cells cannot compensate for the high demand of insulin; therefore, this eventually leads to β-cell dysfunction and defect in insulin secretion, and hyperglycemia occurs ^[18]. Studies have shown that GL, GA, and DG could improve insulin sensitivity in diabetic rodents ^{[12][13][14][19][20][21][22][23][24][25][26][27][28][29][30].}

There are several indicators of insulin resistance, including homeostasis model assessment of insulin resistance (HOMA-IR), homeostatic model assessment of β -cells (HOMA- β), and quantitative insulin sensitivity check index (QUICKI) ^[31]. HOMA was developed by Matthews et al., and it is used to quantify insulin resistance and β -cell function from fasting blood glucose and insulin levels ^[32], while QUICKI is another similar mathematical transformation from fasting blood glucose and insulin levels ^[33]. A study found that GL reduced HOMA-IR index ^[13] ^[14][22][25][26][27][28], and increased HOMA- β and QUICKI indices ^[25] in diabetic rodents, indicating that GL could improve insulin sensitivity. Moreover, impaired fasting blood glucose levels is another indicator of insulin resistance and is characterized by a notable increase in hepatic insulin resistance ^[34]. GL and 18 β -GA were demonstrated to reduce fasting blood glucose levels in diabetic rodents ^[12][13][14][21][22][25][27][28][29]. In addition, high fasting serum insulin levels is another sign of insulin resistance ^[35], and GL was found to decrease fasting serum insulin levels in diabetic rodents ^[12][13][14][21][22][25][27][28][29].

The insulin sensitivity of metabolic tissues plays a vital role in reducing blood glucose levels, and these tissues mainly include liver, adipose tissue, and skeletal muscle ^[36]. White adipose tissue (WAT) is an endocrine organ, which stores energy as triacylglycerols, and secretes hormones to regulate glucose homeostasis and adipokines to regulate inflammation. It was suggested that hypertrophy in WAT is associated with insulin resistance ^[37]. High-fat diet (HFD) was shown to increase visceral adipocyte size and induce adipose tissue inflammation in HFD-induced obese rats. In contrast, GL could decrease the size of adipocytes, thereby reducing body and WAT weights ^[19]. Moreover, it also reduced the adipocyte size and area in the subcutaneous WAT of rats ^{[27][30]}. Similarly, GL and DG could reduce the weights of WAT in HFD-induced obese rodents ^{[19][23]}. Furthermore, peroxisome proliferator-activated receptor- γ (PPAR- γ) is a nuclear receptor, which is highly expressed in adipose tissue ^[38]. The activation of PPAR- γ has been shown to upregulate the expressions of genes that are related to insulin signaling, thus enhancing insulin sensitivity ^[39]. Studies found that the GL treatment improved insulin sensitivity via upregulating the expression of PPAR- γ in the WAT and skeletal muscle ^{[13][14][21]}. On the other hand, the liver is responsible for glucose uptake and lipid synthesis and metabolism ^[40]. HFD increased liver weight, and GL reduced the liver weight and hepatic steatosis in HFD-induced diabetic mice ^[12].

Insulin receptor (IR) is a transmembrane receptor, which can be activated by insulin and insulin-like growth factor (IGF). IR and insulin receptor substrate (IRS) play a vital role in regulating glucose homeostasis in the pancreas and insulin-sensitive tissues ^[16]. IRS acts as a secondary messenger to transmit signals from insulin to downstream intracellular pathways, thus enhancing insulin sensitivity and the transcription of insulin-related genes ^[41]. GL was found to increase IR mRNA expression and its phosphorylation to enhance insulin sensitivity ^{[19][20]}. In addition, another study found that GL enhanced insulin sensitivity through the phosphorylation of IRS-1 and IRS-2

in HFD-induced diabetic mice ^[12]. Moreover, protein tyrosine phosphatase 1B (PTP1B), the main enzyme involved in IR desensitization, regulates insulin and leptin levels ^[42]. It is a negative regulator of insulin signaling pathway, and its inhibitors have become an attractive strategy to treat T2DM and obesity ^[43]. Interestingly, 18 α -GA and 18 β -GA were identified as competitive PTP1B inhibitors ^{[44][45]}. Two other derivatives of GA, namely, indole- and Nphenylpyrazole-GA, also showed potent non-competitive inhibition of PTP1B ^[46].

Insulin-like growth factor-1 (IGF-1) is a growth hormone that can enhance glucose uptake and reduce hepatic glucose production, thereby improving insulin sensitivity ^[47]. Insulin and IGF-1 regulate many signaling pathways, including Ras/mitogen-activated protein kinase (MAPK), and phosphoinositide 3-kinase (PI3K)/Akt pathway ^[16]. The activation of Ras is involved in the development of T2DM, and the inactivation of Ras genes improved insulin sensitivity ^[48]. In addition, PI3K/Akt pathway was identified as one of the vital pathways that are associated with insulin resistance ^[49]. GA was shown to improve glucose uptake and reverse insulin resistance by targeting Ras proteins and activating PI3K/Akt pathway, respectively ^[24]. Moreover, studies found that advanced glycation end products (AGE)-receptor for AGE (RAGE) axis activated oxidative stress and pro-inflammatory pathways, which contributed to insulin resistance ^{[50][51]}. GL was found to improve insulin resistance and downregulate RAGE expression spontaneously, suggesting that GL might improve insulin sensitivity via suppressing AGE-RAGE axis ^[22].

Glucagon-like peptide-1 (GLP-1), a 30-amino acid peptide hormone, is secreted by gut enteroendocrine cells, which improve insulin resistance in adipocytes ^[52]. It is currently used as a T2DM therapeutic. In addition, the inhibition of intestinal farnesoid X-activated receptor (FXR) or FXR deficiency was shown to promote GLP-1 secretion ^[53], and DG was also found to improve insulin resistance and glucose tolerance through downregulation of FXR ^[23].

Studies have shown that inflammation is associated with insulin resistance, and the most vital pro-inflammatory mediators include tumor necrosis factor- α (TNF- α) and interleukin 6 (IL-6). The inhibition of pro-inflammatory mediators was suggested to be one of the strategies for preventing the development of insulin resistance and the pathogenesis of diabetes ^{[54][55][56][57][58]}. GA was shown to reduce fasting plasma TNF- α and IL-6 levels to enhance insulin-responsive pathways ^[24].

2.2. Glucose Tolerance and Homeostasis

Glucose tolerance is another indicator of diabetes and defined as the ability of the body to dispose glucose. Impaired glucose tolerance is often occurred in T2DM, and the progression from normal to impaired glucose tolerance is caused by insulin resistance ^[59]. Studies have shown that GL could improve glucose tolerance in diabetic rodents ^{[12][13][29]}.

Glucose homeostasis can be affected by many factors, including insulin, glucagon, concentration of free fatty acids (FFA), and nutritional factors ^[60]. Gluconeogenesis is the process of glucose production, and gluconeogenic enzymes play a vital role in this process, including phosphoenolpyruvate carboxykinase (PEPCK), fructose-1,6-

bisphosphatase (FBPase), and glucose-6-phosphatase (G6Pase) ^[61]. Glycogenesis is the process of glycogen synthesis, in which glucose is converted and stored as glycogen in the liver, which is activated by insulin in response to high blood glucose levels. Pyruvate dehydrogenase (PDase), an essential enzyme in glycogen synthesis, is also a substrate of glycogen synthase kinase 3β (GSK- 3β), and GSK- 3β induces pyruvate dehydrogenase E1 α (PDH-E1 α) phosphorylation in response to insulin to promote glycogen synthesis ^[62]. 11 β hydroxysteroid dehydrogenase (11β-HSD) is an enzyme involved in the pathogenesis of metabolic syndrome, which is characterized by hyperglycemia, dyslipidemia, diabetes, and arterial hypertension, and has two isoforms, 11β-HSD1 and 11β-HSD2 ^[63]. 11β-HSD1 is highly expressed in insulin-sensitive tissues, including liver, WAT, and skeletal muscle ^[64]. The inhibition of 11β-HSD1 ameliorated hyperglycemia and improved insulin sensitivity in diabetic mice [65]. In addition, PEPCK is a critical enzyme in gluconeogenesis, and the improved activity of PEPCK leads to elevated glucose output and aggravation of diabetes, whereas the defects of PEPCK result in lethal hypoglycemia ^[66]. Hexose-6-phosphate dehydrogenase (H6PDH), an enzyme that is localized in the endoplasmic reticulum lumen, converts glucose-6-phosphate (G6P) and nicotinamide adenine dinucleotide phosphate (NADP) to produce nicotinamide adenine dinucleotide phosphate hydrogen (NADPH). It provides a high [NADPH]/[NADP⁺] ratio for 11β-HSD1 amplification of intracellular active glucocorticoid production [67]. Moreover, increased 11β-HSD1-dependent active glucocorticoid production is associated with insulin resistance, T2DM, and its cardiovascular complications [68].

Studies found that GL could reduce PEPCK and G6Pase mRNA expressions to suppress gluconeogenesis, and upregulate PDase and GSK-3 β mRNA expressions to increase glycogen synthesis in the liver ^{[12][19][28]}. Similarly, GL reduced PEPCK activities in the liver and kidney of normal rats ^[27], and alleviated the over-activities of PEPCK and G6Pase in the liver and kidney in rats with metabolic syndrome ^[14]. In addition, GL could reduce H6PDH activities in the liver, kidney, WAT, and muscle ^{[27][28]}. Moreover, GL could decrease the activities of 11 β -HSD1 and 11 β -HSD2 in the liver, kidney, WAT, and muscle to reduce blood glucose levels ^{[26][27][28]}. Furthermore, two isoforms of GA have different inhibition activities on 11 β -HSD; 18 α -GA selectively inhibits 11 β -HSD1, but not 11 β -HSD2, while 18 β -GA favorably inhibits 11 β -HSD2 ^[69].

Glucose transporter type 4 (GLUT4) is an insulin-regulated glucose transporter, which is involved in rapid glucose uptake in various kinds of cells to regulate glucose homeostasis, and its downregulation is closely associated with the development of glucose tolerance ^{[70][71][72]}. GA was found to promote GLUT4 expression by targeting Ras protein to regulate MAPK pathway ^[24]. Similarly, GL elevated GLUT4 expression in the skeletal muscle of HFD-induced diabetic rats to improve glucose homeostasis ^[13]. Moreover, GL and GA were shown to enhance insulin-stimulated glucose uptake in 3T3L-1 adipocytes ^[15].

Due to the poor bioavailability of GL, it has been formulated as nanoparticles. GL-loaded nanoparticles improved lipid profile and lowered fasting blood glucose levels in nicotinamide plus streptozotocin (STZ)-induced T2DM rats. Interestingly, the dosages used in GL-loaded nanoparticles were only a quarter of the dosages of the pure GL form ^{[73][74]}. Moreover, a combination of GL-loaded nanoparticles and thymoquinone-loaded nanocapsules was applied, displaying better anti-diabetic activities than when administered separately in nicotinamide and STZ-induced T2DM rats, including decreased blood glucose levels and improved lipid profile ^[74]. Glycosylated hemoglobin A1c is

another biochemical parameter that is used to estimate the severity of diabetes, and its high level is an indicator of poor control of blood glucose levels ^[75]. Moreover, GL-loaded nanoparticles decreased the level of glycosylated hemoglobin A1c ^{[73][74]}. Furthermore, GA, as an efficient liver-specific ligand, was made to be a liver-targeted drug delivery carrier and conjugated with chitosan lactate and poly(ethylene glycol) to form nanoparticles containing siRNA-CREB regulated transcription coactivator 2 (CRTC2). These nanoparticles were shown to be accumulated in the liver after 2 h of treatment, reduce blood glucose levels, and inhibit hepatic gluconeogenesis in rats ^{[76][77]}.

2.3. Lipid Metabolism

The liver is responsible for the regulation of lipid metabolism, uptake of FFA, and export of fat ^[78]. In T2DM, lipid metabolism is altered, and fat uptake and delivery are imbalanced, which is linked with the over-production of very low-density lipoprotein (VLDL) ^{[78][79]}. Many studies found that HFD or high-fat-high-sucrose diet increased serum and hepatic triglyceride (TG), total cholesterol, low-density lipoprotein (LDL) cholesterol, and FFA levels, and decreased high-density lipoprotein (HDL) cholesterol levels in rodents ^{[12][14][19][80][81]}. In contrast, GL could improve serum lipid parameters, including decreases in serum and hepatic FFA, TG, total cholesterol, LDL cholesterol, and VLDL levels, but an increase in HDL cholesterol levels in diabetic rodents ^{[12][14][19][30][73][74][80][81]}. Similarly, GL-loaded nanoparticles could also decrease total cholesterol, TG, LDL, and VLDL levels, and increase HDL levels in diabetic rats ^[73]. Meanwhile, GL could also increase lipid uptake in cardiac and skeletal muscle ^[14].

Lipoprotein lipase (LPL) is a multi-functional glycoprotein enzyme, which plays a vital role in lipid parameters, such as the levels of TG, LDL cholesterol, HDL cholesterol, and VLDL, and its activity is upregulated by apolipoprotein C-II and downregulated by apolipoprotein C-III ^{[82][83]}. Increased LPL activity can be used as an indicator of enhanced lipid uptake into tissues ^[84]. GL was shown to upregulate LPL expression in the kidney, heart, muscle, and WAT ^{[14][21][30][80]}. In addition, other studies showed that GL promoted TG metabolism by inducing LPL activity, which upregulated hepatic apolipoprotein C-III expression and suppressed apolipoprotein C-III expression in HFD-induced diabetic mice ^[12].

Excess FFA and lipid droplets lead to intramuscular accumulation of lipid intermediates, which results in the inhibition of insulin signal transduction to impair insulin sensitivity ^[85]. GL was shown to decrease lipid deposition in the liver, kidney, heart, and muscle in HFD-induced diabetic rodents ^{[12][80]}. Interestingly, a study showed that GL treatment improved insulin sensitivity in HFD-induced obese rats, which might be through decreased lipid deposition ^[80]. Moreover, hepatic lipogenesis is regulated by a transcription factor, sterol regulatory element-binding transcription factor 1c (SREBP-1c), which is activated by insulin and regulates the genes that are involved in fatty acid and TG synthesis, including fatty acid synthetase (FAS) and stearoyl CoA desaturase (SCD) 1 ^[86]. GL was shown to improve lipid metabolism mainly through reducing hepatic lipogenesis and improving fatty acid metabolism, via downregulation of SREBP-1c, FAS, and SCD1 expressions, and upregulation of peroxisome proliferator-activated receptor- α (PPAR- α), carnitine palmitoyl transferase 1 α (CPT1 α), and acyl-coenzyme A dehydrogenase (ACADS), respectively ^[12]. GL and GA were also found to reduce the early stage of adipogenesis and lipid accumulation in 3T3-L1 adipocytes through downregulating CCAAT/enhancer-binding protein (C/EBP)- β and C/EBP- δ , PPAR- γ , and SCD expressions ^{[15][87]}. Similarly, 18 β -GA was found to inhibit lipid accumulation,

suppress adipogenic differentiation via inhibiting Akt phosphorylation and downregulating PPAR-γ and C/EBP-α expressions, and stimulate lipolysis via upregulation of adipose TG lipase (ATGL), hormone-sensitive lipase (HSL), and perilipin (Plin-1) expressions in 3T3-L1 adipocytes ^[88]. However, Yamamoto et al. showed that GL did not stimulate lipolysis in 3T3-L1 adipocytes ^[87].

2.4. Insulin Secretion and Protection of Pancreatic β-Cells

Insulin is a hormone, which is produced by pancreatic β -cells in the islets of Langerhans, and it promotes glucose uptake from the blood into tissues. Insulin secretion is regulated by pancreatic β -cells in response to the increase in blood glucose levels ^[89]. In T2DM, chronic insulin resistance and a progressive decline in β -cell function result in β -cell dysfunction and defect in insulin secretion ^[90]. IRS-2 plays a vital role in the preservation of β -cell mass ^[91], while pancreas duodenum homeobox-1 (PDX-1) plays a role in β -cell survival and function ^[92]. In addition, glucokinase (GLK) acts as a glucose sensor to regulate insulin secretion ^[93]. GA was shown to enhance glucose-stimulated insulin secretion in isolated mouse islets, and upregulate IRS-2, PDX-1, and GLK expressions to improve β -cell viability ^[15]. A schematic diagram of GA-mediated glucose-stimulated insulin secretion in pancreatic β -cells is shown in **Figure 3**.



Figure 3. The mechanisms of glycyrrhetinic acid (GA)-mediated glucose-stimulated insulin secretion in pancreatic β-cells. GA upregulates IRS-2, PDX-1, and GLK expressions to protect β-cells and increase insulin secretion. GLUT2: Glucose transporter 2; GLK: Glucokinase; IRS-2: Insulin receptor substrate 2; PDX-1: Pancreas duodenum homeobox-1.

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