## Prototheca spp. in Bovine Infections

Subjects: Microbiology

Contributor: Simona Nardoni, Francesca Mancianti

Prototheca microalgae, although still considered uncommon etiologic agents, represent an insidious intruder, threatening cattle herd health and determining productive losses. Increasing numbers of clinical cases globally identified would indicate these microalgae as emerging pathogens. They can be isolated from a wide variety of environmental and non-environmental sources, due also to their ability to produce biofilm. This ability to spread and contaminate a huge variety of substrates, as well as the high resistance to elevated temperatures, renders *Prototheca* prevention a very hard task. In addition, early infection signs are subtle and difficult to detect. The poor response to conventional antimycotic drugs represents an additional challenge when facing this infection. Although it would seem unrealistic to completely eradicate the exposure risk of cows to these microalgae, the adoption of proper on-farm protocols and management, with the highest attention to hygiene measures, would be beneficial in reducing the magnitude of this problem. Keeping the attention focused on early diagnosis, together with the development of new, alternative, and effective agents and formulations, would be strongly advised to prevent, treat, and control *Prototheca* infections.

Keywords: microalgae; Prototheca bovis; Prototheca blaschkeae; Prototheca ciferrii; bovine protothecosis; mastitis; cattle

Prototheca spp. (family Chlorellaceae, order Chlorellales, class Trebouxiophyceae) are achlorophyllous microalgae, widely distributed in the environment and repeatedly reported as responsible for human and animal disease. Bovine mastitis represents the most important form of Prototheca infection in animals and consists of clinical or subclinical forms. Dairy-cattle-associated Prototheca species are Prototheca bovis, Prototheca blaschkeae, and Prototheca ciferrii [1]. The ecology of Prototheca is not fully elucidated yet. These organisms can be recovered from animal waste, sludge, sewage, rivers, and fountains, preferring moist areas with high organic contents. Bovine protothecosis is reported to occur worldwide, in the presence of large dairy herds, mostly in tropical and temperate areas [2]. Predisposing factors to protothecosis are reported to be unclean or repeated intramammary infusions, and antibiotic drug treatments in the udder, where Prototheca would act as an opportunistic pathogen favored by antibiotic-induced suppression of the local flora [3][4] [5][6][7][8]. Prototheca spp. can survive a wide range of environmental conditions as well as disinfectants [9][10]. Prototheca spp. can produce biofilm [11][12]. Bovine mammary gland can be infected by P. bovis following teat trauma by mechanical milking, and subsequent contamination of the teat orifice by environmental organic matter [13][14]. Infection of the mammary gland is often subclinical, without any visual sign, and can be revealed by raised somatic cell count only, although the high result is not continuous [6][15].

An environmental control approach would include action in stables, aisles, run as well in milking parlor. The main strategy is devoted to controlling algal amounts in the environment enhancing hygiene measures by using conventional and natural disinfectants, as well as physical tools  $^{[16]}$ . To date, no treatment protocol has been proven fully effective in controlling *Prototheca* spp. infection in dairy cows  $^{[2]}$ .

## References

- 1. Jagielski, T.; Bakuła, Z.; Gawor, J.; Maciszewski, K.; Kusber, W.-H.; Dyląg, M.; Nowakowska, J.; Gromadka, R.; Karnkowska, A. The genus Prototheca (Trebouxiophyceae, Chlorophyta) revisited: Implications from molecular taxonomic studies. Algal Res. 2019, 43, 101639.
- 2. Libisch, B.; Picot, C.; Ceballos-Garzon, A.; Moravkova, M.; Klimesová, M.; Telkes, G.; Chuang, S.-T.; Le Pape, P. Prototheca infections and ecology from a One Health perspective. Microorganisms 2022, 10, 938.
- 3. Pieper, L.; Godkin, A.; Roesler, U.; Polleichtner, A.; Slavic, D.; Leslie, K.E.; Kelton, D.F. Herd Characteristics and cowlevel factors associated with Prototheca mastitis on dairy farms in Ontario, Canada. J. Dairy Sci. 2012, 95, 5635–5644.
- 4. Sobukawa, H.; Watanabe, M.; Kano, R.; Ito, T.; Onozaki, M.; Hasegawa, A.; Kamata, H. In vitro algaecide effect of disinfectants on Prototheca zopfii genotypes 1 and 2. J. Vet. Med. Sci. 2011, 73, 1527–1529.

- 5. Park, H.-S.; Moon, D.C.; Hyun, B.-H.; Lim, S.-K. Short Communication: Occurrence and persistence of Prototheca zopfii in dairy herds of Korea. J. Dairy Sci. 2019, 102, 2539–2543.
- Huilca-Ibarra, M.P.; Vasco-Julio, D.; Ledesma, Y.; Guerrero-Freire, S.; Zurita, J.; Castillejo, P.; Barceló Blasco, F.; Yanez, L.; Changoluisa, D.; Echeverría, G.; et al. High Prevalence of Prototheca bovis infection in dairy cattle with chronic mastitis in Ecuador. Vet. Sci. 2022, 9, 659.
- 7. Branko, S.; Vasilev, D.; Karabasil, N.; Vučurović, I.; Čobanović, N.; Babić, M.; Katić, V. Molecular Identification of Prototheca zopfii genotype 2 mastitis isolates and their influence on the milk somatic cell count. Vet. Arh. 2017, 87, 249–258.
- 8. Watts, J.L. Diagnostic Procedure in Veterinary Bacteriology and Mycology; Academic Press: Cambridge, MA, USA, 1990
- 9. Salerno, T.; Ribeiro, M.G.; Langoni, H.; Siqueira, A.K.; da Costa, E.O.; Melville, P.A.; Bueno, V.F.F.; Yamamura, A.A.M.; Roesler, U.; da Silva, A.V. In vitro algaecide effect of sodium hypochlorite and iodine-based antiseptics on Prototheca zopfii strains isolated from bovine milk. Res. Vet. Sci. 2010, 88, 211–213.
- 10. Lassa, H.; Jagielski, T.; Malinowski, E. Effect of different heat treatments and disinfectants on the survival of Prototheca zopfii. Mycopathologia 2011, 171, 177–182.
- 11. Gonçalves, J.L.; Lee, S.H.I.; de Paula Arruda, E.; Pedroso Galles, D.; Camargo Caetano, V.; Fernandes de Oliveira, C.A.; Fernandes, A.M.; Veiga dos Santos, M. Biofilm-producing ability, and efficiency of sanitizing agents against Prototheca zopfii isolates from bovine subclinical mastitis. J. Dairy Sci. 2015, 98, 3613–3621.
- 12. Tashakkori, N.; Rahmani, H.K.; Khoramian, B. Genotypic and phenotypic diversity of Prototheca spp. recovered from bovine mastitis in terms of antimicrobial resistance and biofilm formation ability. BMC Vet. Res. 2022, 18, 452.
- 13. Pal, M.; Abraha, A.; Rahman, M.T.; Dave, P. Protothecosis: An emerging algal disease of humans and animals. Int. J. Life Sci. Biotechnol. Pharm. Res. 2014, 3, 1–13.
- 14. Da Costa, E.O.; Ribeiro, A.R.; Watanabe, E.T.; Pardo, R.B.; Silva, J.B.; Sanches, R.B. An increased incidence of mastitis caused by Prototheca species and Nocardia species on a farm in São Paulo, Brazil. Vet. Res. Commun. 1996, 20, 237–241.
- 15. Tenhagen, B.A.; Hille, A.; Schmidt, A.; Heuwieser, W. Shedding patterns and somatic cell counts in milk fromquarters chronically infected with Prototheca spp. Dtsch. Tierarztl. Wochenschr. 2005, 112, 44–48.
- 16. Bozzo, G.; Dimuccio, M.M.; Casalino, G.; Ceci, E.; Corrente, M. New approaches for risk assessment and management of bovine protothecosis. Saudi J. Biol. Sci. 2022, 29, 103368.

Retrieved from https://encyclopedia.pub/entry/history/show/110836