

Pathogenesis of Ocular Adnexal Marginal Zone Lymphomas

Subjects: **Hematology**

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Ocular adnexal marginal zone lymphoma (OAMZL) is a distinct type of lymphoma that presents in tissues around the eyeball. The lymphoma develops from mature B lymphocytes that have been triggered by antigens for prolonged times. It seems that the B cells often recognize autoantigens. The lymphoma cells often carry specific chromosomal gains and, in some cases, chromosomal translocations. A main factor in the development of this lymphoma is the constitutive activation of the NF- κ B pathway, which occurs through various types of genetic alterations. Further key pathogenetic mechanisms involve epigenetic changes, indicated by recurrent mutations in epigenetic regulators.

ocular adnexal lymphoma

extranodal marginal zone lymphoma

mucosa-associated tissue

MALT lymphoma

NF- κ B

ocular adnexa

orbit

1. Precursor Lesions

Several precursor lesions potentially developing to Ocular adnexal marginal zone lymphoma (OAMZL) have been described, among them orbital pseudotumors (idiopathic orbital inflammatory disease (IOID)), reactive lymphoid hyperplasia (RLH), and IgG4-related disease ^[1]. Both exogenous antigens and autoantigens can trigger the abovementioned precursor lesions in the ocular adnexa. Precursor lesions as inflammatory non-malignant states have in common that they result in chronic antigenic stimulation, which may lead to activation of the NF- κ B signaling pathway, to chromosomal alterations, and to other genetic and epigenetic alterations. This multistep process can drive lymphoma development ^[2].

OAMZL arising in the context of IgG4-related disease has been repeatedly reported as OAMZL with IgG4-positive cells or infiltrated by IgG4-positive cells ^{[3][4][5][6][7]}. In some cases, the differentiation between the two entities is challenging, since OAMZL exhibits IgG4-positive plasma cells in up to 62% of cases ^[8]. Histopathologically, obliteration of venous vessels is specific. Plasma cells in IgG4-related disease are polytypic. In IgG4-related disease, eosinophilia, high IgE titers, polyclonal hypergammaglobulinemia, and often elevated serum IgG4 levels are observed ^[9]. Infraorbital nerve enlargement is a unique feature on MRI scans ^[5]. Upregulation of activation-induced cytidine deaminase (AID)—the master factor for somatic hypermutation and class-switch recombination of immunoglobulin genes—was observed in IgG4-related ophthalmic disease and OAMZL, whereas AID expression was lower in IgG4-negative OAMZL ^[10]. AID might be a driver for oncogenesis in the development of IgG4-related ophthalmic disease to IgG4-positive OAMZL.

2. Antigen Stimulation

Chronic (auto)antigenic stimulation via chronic inflammation, infection, or autoimmune disease is supposed to be a relevant pathogenic mechanism in the development of primary MALT lymphoma in general ^[11]. A multistep process is supposed to promote survival and growth advantages of stimulated B cells, which may finally give rise to monoclonal B-cell populations. Various mechanisms are described causing this antigenic stimulation.

2.1. Infectious Agents

Chlamydomphila psittaci

Chlamydia are human pathogenic intracellular bacteria that are typically transmitted via infected birds. Mostly, infections are asymptomatic, but they can cause pneumonia, chronic conjunctivitis, pericarditis, and hepatitis ^[12]. *Chlamydomphila psittaci* can induce immune reactions cross-reacting with autoantigens, leading to insufficient elimination of the pathogen and induction of lymphoma development ^{[13][14][15]}.

The prevalence of *C. psittaci* in OAL seems to be region-specific. In most studies involving patients from Italy or Korea, *C. psittaci* was repeatedly detected by PCR and other methods in OAL cases. Other *Chlamydia* species were predominantly observed in China (*C. pneumonia*) and the UK (*C. trachomatis*) ^{[13][16][17]}. In other countries, however—including Japan, the USA, Cuba, the UK, the Netherlands, France, and Germany—no evidence for *C. psittaci* in samples of OAL was observed ^{[18][19][20][21][22][23][24][25][26]}.

(1)Other BacteriaAmong 308 OAL patients analyzed in 11 studies, *Helicobacter pylori* was detected in 23% of the lymphomas ^[27]. However, there is the risk of contamination of OAL biopsy specimens by the rather prevalent *H. pylori* during sampling, so the true incidence may be substantially lower. Notably, the prevalence of OAL patients with *H. pylori*-positive gastric infections is overall no higher than in the general population ^[27], arguing against a significantly increased risk of OAL development in individuals with chronic gastric *H. pylori* infection. A recent study confirmed a lack of association between gastric *H. pylori* infection and OAL incidence ^[28]; none of 18 OAL cases in that study showed *H. pylori* DNA in the lymphoma tissue. Thus, the role of *H. pylori* in the development of OAL is still unresolved.

(2)Viral PathogensIn several studies on a potential viral etiology of OAL, no viruses were detected in the lymphomas ^{[26][29]}. For chronic hepatitis C virus (HCV) infections, the association with marginal-zone lymphomas is especially well known ^{[30][31]}. Although there are clear indications for a role of HCV in the pathogenesis of some types of B-cell lymphoma, its role in OAMZL is less clear ^[32].

For human-immunodeficiency-virus-infected patients, a higher risk of developing marginal-zone lymphomas has been described, but there are no reports on higher incidences of OAMZL ^[33]. Regarding Epstein–Barr virus, cytomegalovirus, and human papilloma virus, there are also no reports on higher infection rates in patients with OAL ^{[29][34]}.

2.2. Autoimmune Diseases

The detailed mechanisms of lymphomagenesis in the context of autoimmunity remain unclear. The occurrence of lymphomas in association with rheumatoid arthritis, Sjögren's syndrome, Hashimoto thyroiditis, and other autoimmune diseases is well described [35][36]. A meta-analysis of 20 studies including patients with the abovementioned autoimmune diseases revealed that lymphomas are more common in these patients than in healthy subjects [37]. The most common lymphoma subtype occurring in patients with autoimmune diseases is marginal-zone lymphoma [38]. Disease activity, the presence of rheumatoid factor, and/or cryoglobulinemia in patients with autoimmune diseases are prognostic factors for lymphoma development, reflecting a continuing immune stimulation [39]. Apoptotic resistance—mediated by high BCL2 expression, activation of NF-κB, and overexpression of B-cell activating factor (BAFF)—is increased in autoimmune diseases [40]. **Figure 1** provides a proposed scenario of OAMZL pathogenesis.

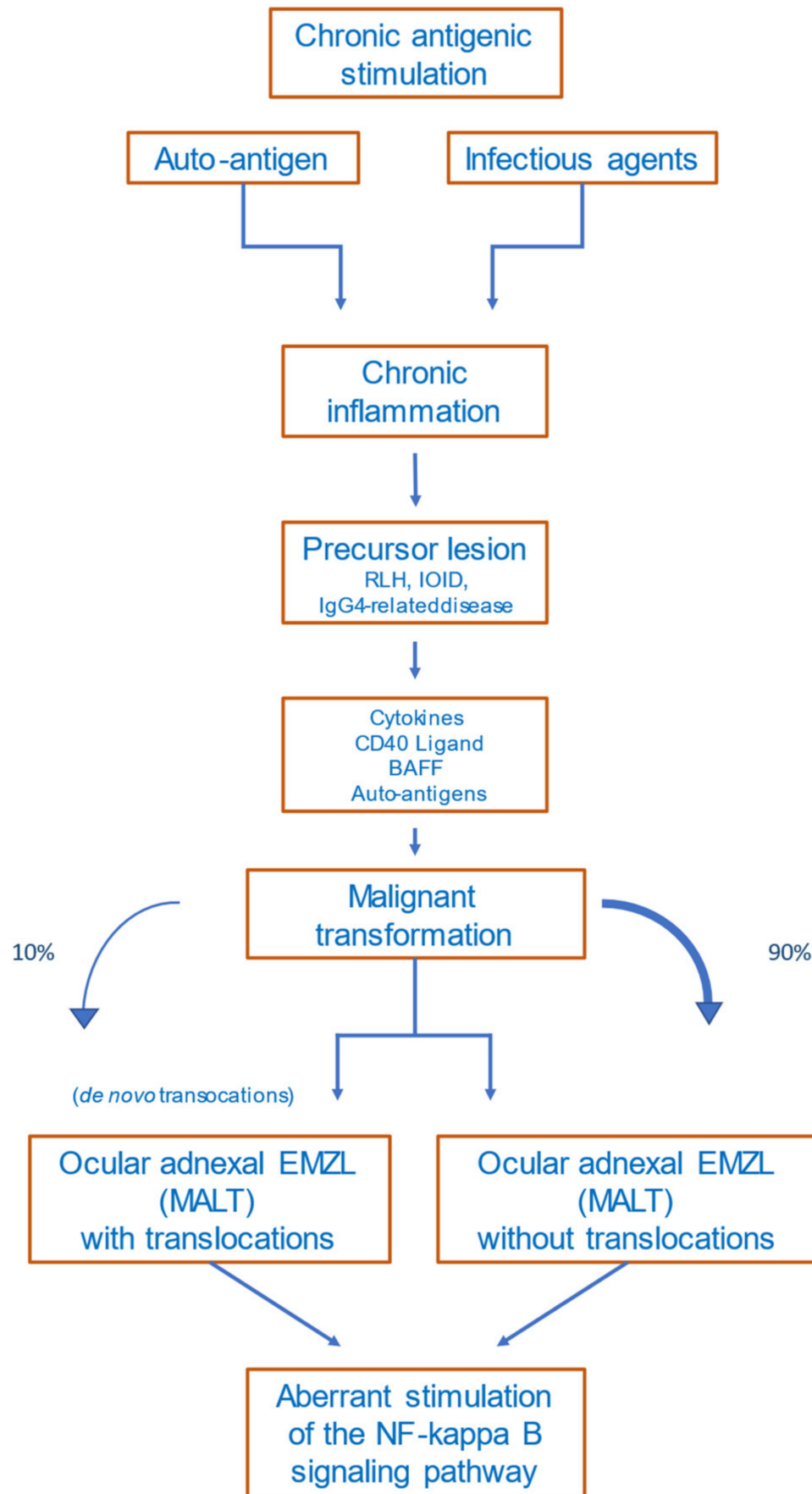


Figure 1. Proposed scheme of OAMZL pathogenesis. RLH: reactive lymphoid hyperplasia; IOID: idiopathic orbital inflammatory disease; BAFF: B-cell activating factor; EMZL: extranodal marginal-zone lymphoma; MALT: mucosa-associated lymphatic tissue.

3. Chromosomal Aberrations

3.1. Translocations

In OAMZL, chromosomal translocations leading to constitutive activation of the NF-κB signaling pathway have been described [41]; these include t(11;18)(q21;q21)/*BIRC3-MALT1*, often accompanied by a trisomy 3 [42], and t(14;18)(q32;q21)/*IGH-MALT1* (Table 1). The t(11;18)(q21;q21) juxtaposes *BIRC3* (previously also known as *API2*) to *MALT1*, resulting in a fusion gene; this translocation has been detected in 10–15% of OAL cases [43][44]. The t(14;18)(q32;q21) translocation brings the *MALT1* gene under control of the *IGH* locus enhancers, causing constitutive expression of *MALT1*, and is present in around 5–10% of OAMZLs. *MALT1* is a protease and an important mediator of canonical NF-κB signaling [45].

Table 1. Genetic alterations of OAMZL.

Chromosomes or Genes Affected	Type of Genetic Alteration	Pathway or Main Function	Approximate Frequency (%)	References
Chromosomal alterations				
Trisomy 3	Chromosomal gain	unclear (FOXP1?)	30–60	[46][47][48]
Trisomy 18	Chromosomal gain	unclear	20–55	[46][47][48]
t(11;18)(q21;q21)	<i>BIRC3-MALT1</i> translocation	NF-κB pathway	10–15	[44][49]
t(14;18)(q32;q21)	<i>IGH-MALT1</i> translocation	NF-κB pathway	5–10	[43][46][50]
t(3;14)(p14.1;q32)	<i>FOXP1-IGH</i> translocation	B-cell development and survival (NF-κB pathway)	5–15	[51][52]
Gene mutations				
<i>TNFAIP3</i>	Deletions, non-synonymous mutations	NF-κB pathway	30–50	[53][54][55][56][57]
<i>MYD88</i>	Non-synonymous mutations (mostly p.L265P)	NF-κB pathway	5–35	[54][57][58][59][60]

Chromosomes or Genes Affected	Type of Genetic Alteration	Pathway or Main Function	Approximate Frequency (%)	References
<i>NOTCH1</i>	Non-synonymous mutations (mostly HD and PEST domains)	NOTCH pathway	2–10	[54][56][61]
<i>NOTCH2</i>	Non-synonymous mutations (mostly TAD and PEST domains)	NOTCH pathway	5–10	[54][62]
<i>KMT2D</i>	Non-synonymous mutations	Epigenetic regulation	5–20	[54][56][57][62]
<i>CREBBP</i>	Non-synonymous mutations	Epigenetic regulation	15	[26][56]
<i>TBL1XR1</i>	Non-synonymous mutations (mostly WD40 domain)	Regulation of nuclear receptor activity (NF-κB and AP1 pathway)	10–20	[26][56][57][61]
<i>JAK3</i>	Non-synonymous mutations	JAK/STAT signaling	5–10	[26][57]
<i>CABIN1</i>	deletions, Non-synonymous mutations	NFAT signaling	30%	[63]
<i>RHOA</i>	deletions, Non-synonymous mutations	Rho signaling	26%	[63]

Non-synonymous mutations: includes damaging point mutations, small insertions/deletions.

The t(3;14)(p14.1;q32)/*FOXP1-IGH* was observed in around 5–15% of OALs, and leads to constitutive expression of FOXP1 [51][52][64]. FOXP1 is a transcription factor that supports B-cell survival, and can cooperate with NF-κB, so that for this translocation event there is also a link to the NF-κB pathway [65].

Further translocations occur in OAMZL with lower prevalence. These include t(1;14)(p22;q34) juxtaposing *BCL10* to the *IGH* locus, or t(5;11) with unknown translocation partners [66]. Notably, translocations are observed in OAMZL at lower frequencies than in MALT lymphomas occurring in other regions, and the various EMZLs show distinct patterns of recurrent chromosomal translocations [64].

3.2. Copy Number Variations

The most frequent copy number variations in OAMZL are trisomy 3 and trisomy 18, in approximately 30–60% and 20–55% of patients, respectively (Table 1) [46][47]. Trisomy 3 is more common in persons above 50 years of age, whereas trisomy 18 is observed mostly in younger, female patients. The cases with trisomy 18 have more lymphoproliferative lesions, less nodularity, and are associated with recurrent disease [67]. The pathogenetic effect of these trisomies in OAMZL is unknown.

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4. Genetic Alterations in Particular Signaling Pathways

4.1. Nuclear Factor Kappa B (NF-κB) Pathway

NF-κB is a transcription factor family that plays a critical role in B-cells' activation, development, and survival [68]. The NF-κB pathway is normally only transiently activated in B cells by binding of various ligands to receptors, including Toll-like receptors, the TNF-α receptor, the BCR, CD40, and others. The intracellular signaling transduction is mediated via a canonical and a non-canonical pathway [64]. A major role of deregulated activation of this pathway in OAMZL is already indicated by the fact that the three most frequent chromosomal translocations of OAMZL cause, or at least contribute to, NF-κB activation, as discussed above. A further major driver for constitutive NF-κB activation in OAMZL is genetic alterations leading to functional changes of TNF-α-inducible protein 3 (TNFAIP3), previously also called A20. Somatic deletions and/or point mutations lead to inactivation of this negative regulator of the classical NF-κB pathway [55]. In OAMZL, destructive *TNFAIP3* mutations and/or deletions were observed in 30–50% of cases [53][56] (Table 1). *TNFAIP3* is the most frequently mutated gene detected so far in OAMZL. It has been reported that *IGH/MALT1* translocations are mutually exclusive with *TNFAIP3* mutations/deletions in OAMZL [55], indicating that these are alternative mechanisms for deregulated NF-κB activation in OAMZL.

To identify potential mutations in further genes of the NF-κB signaling pathway, sequencing analyses were performed by several groups. In 24 OAMZL samples, sequencing of hotspots in the genes *CARD11*, *MYD88*, and *CD79B*, known to be frequently mutated in other B-cell lymphomas, revealed no mutations [69]. In a targeted sequencing approach of genes involved in the NF-κB signaling pathway performed by the researchers' team with 63 patients, mutations in *TNFAIP3* (27% of cases), *MYD88* (19%), and *BCL10* (6%) were observed [54]. Further genes, mutated to lower frequencies, were *TNIP1*, *NFKBIA*, *BIRC3*, *CARD11*, and *CD79B*. Only a few genes encoding components of the non-canonical NF-κB pathway were mutated (*MAP3K14*, *BIRC3*, and *CYLD*), whereas other mutated genes were involved in the canonical pathway [54]. A further study analyzing the frequency of *MYD88* L265P mutations in primary OAL found the gene to be mutated in 36% of patients [58]. In a targeted next-generation sequencing (NGS) approach to OAL including 20 samples, with 17 of them being primary OAL, 25% of cases exhibited mutations in the TIR domain of *MYD88* [59]. Two of the three studies mentioned carefully excluded lymphoplasmacytic lymphoma/Waldenström macroglobulinemia by analyzing paraproteins and plasmacytic differentiation—especially with IgM [54][59], which is necessary when analyzing *MYD88* mutations. Further NGS-based approaches using whole genomes, whole exomes, and targeted sequencing confirmed and extended these findings [26][56][62].

4.2. NOTCH Pathway

The NOTCH signaling pathway is important in cell differentiation; it is active in many cell types regulating cell development, differentiation, and homeostasis, and is involved in many malignant diseases, including lymphomas [70][71][72]. The NOTCH signaling pathway cross-interacts with the NF-κB signaling pathway as its upstream regulator [73][74][75]. Non-synonymous *NOTCH1* or *NOTCH2* mutations were observed in up to 10% of OAMZLs,

with a similar pattern as described for other B-cell lymphomas. There is a clustering of the mutations in the HD and PEST domains of NOTCH1, as well as downstream of the ankyrin repeats in the intracellular domain of NOTCH2 [54][56]. For both genes, these types of mutations cause a gain of function, as the inhibitory C-terminal PEST domains are removed or otherwise inactivated. Copy number gains in the NOTCH target *HES4* may be a further mechanism of enforced NOTCH pathway activity in OAMZL [26][76].

4.3. NFAT Signaling

A recent exome sequencing study provided the first evidence for recurrent alterations in the NFAT signaling pathway in OAMZL. Frequent deletions and destructive mutations were detected in the gene encoding the negative NFAT signaling regulator *CABIN1* (30% of cases with mutations), as well as rarer mutations in NFAT members themselves and other NFAT signaling pathway components.

5. Epigenetic Regulators

Alterations in epigenetic modifiers occur in several types of lymphoma [77]; additionally, in OAMZL, genes encoding epigenetic regulators are mutated. Among these genes are *KMT2D* (approximately 5–20% of cases) and *CREBBP* (ca. 15%). In individual studies, mutations in the epigenetic regulators *KMT2C* and *EP300* have been detected, such that a clear determination of the overall frequency of such alterations needs further investigation [54][56][62]. *KMT2C* and *KMT2D*, belonging to the mixed-lineage leukemia (MLL) family of histone methyltransferases, methylate Lys-4 of histone H3. Mutations leading to inactivation of these genes lead to diminished global H3K4 methylation in follicular and diffuse large B-cell lymphoma [78]. Conditional deletion of *Kmt2d* in different developmental stages of B cells in mice resulted in an increased number of germinal-center B cells and enhanced proliferation. From these results, the authors suggested the *KMT2D* acts as tumor suppressor gene. *KMT2C*, which functions very similarly to *KMT2D*, and can partially replace a loss of *KMT2D*, is therefore also supposed to be tumorigenic in case of a loss [79]. *CREBBP* and *EP300* are related histone and non-histone acetyltransferases, which regulate transcriptional activity in several signaling pathways via chromatin remodeling. In follicular and diffuse large B-cell lymphoma, monoallelic deletions/mutations result in defects of acetylation of the oncoprotein *BCL6* and the tumor suppressor *p53*.

6. Additional Mutated Genes

In addition to the genes in major signaling pathways or involving epigenetic regulators already mentioned, further genes recurrently mutated in OAMZL include *TBL1XR1* [57]. *TBL1XR1* is an essential regulator of transcriptional repression, and contributes to canonical NF-κB activation [80]. This gene can activate the transcription of transcription factors such as NF-κB and JUN [81], and may therefore contribute to the strong NF-κB activity in OAML. *TBL1XR1* is mutated in various tumors and lymphomas, promoting tumor cell survival. In OAMZL, mutations were consistently detected in 10–20% of cases, so *TBL1XR1* is one of the most frequently mutated genes in this type of lymphoma (Table 1). *TBL1XR1* mutations have been linked to a poor prognosis in aggressive lymphomas [82][83]. In a small cohort of patients with OAL of the MALT subtype, *TBL1XR1* mutations were

associated with unique morphometric phenotypes [84]; the cells exhibited significantly lower circularity and solidity as analyzed via computational digital image analysis.

The JAK/STAT signaling pathway is necessary for cytokine signaling and immune regulation, and plays an important role in various types of lymphoma [85]. Activating mutations in *JAK3*, known to cause constitutive activation of the JAK/STAT signaling pathway, were observed in up to 10% of patients with OAMZL [26]. Interestingly, patients with mutant *JAK3* exhibited a shorter progression-free survival [26].

In a recent study, deletions and non-synonymous mutations in the *RHOA* gene were detected in 26% of OAMZL studies [63]; this points to a potential role of altered Rho signaling in OAMZL, but further studies are needed in order to clarify the consequences of the mutations detected in *RHOA*.

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