

Fungi-Derived β -Glucan on Tumor Progression

Subjects: Oncology

Contributor: Vaclav Vetvicka

β -Glucans have been studied in animal species, from earthworms to humans. They form a heterogenous group of glucose polymers found in fungi, plants, bacteria, and seaweed. β -Glucans have slowly emerged as an important target for the recognition of pathogens.

Keywords: β -glucan ; mushrooms ; cancer ; immune ; health

1. Introduction

To use natural products as a possible remedy is weaved into the history of mankind. People have been appealing to nature to cure various diseases since ancient times. The first documented history of plant preparation and medicinal use is a record on Sumerian clay tablets from the period of 4000 BCE. There is also an approximately 5000-year-old written Indian document about medicinal effects of mushrooms. Indian Ayurveda and traditional Chinese medicine can serve as examples of healing trends which have been developed through empirical experience. A physician of Marcus Aurelius emphatically pioneered the treatment of diseases by using the specific diets of Galen of Pergamum. Several centuries later, J. Lind ^[1] used 200-year-old Dutch knowledge of the benefits of citrus fruits to the health of the sailors on long voyages, and conducted one of the first large-scale clinical medical trials. Japanese legend indicates that monkeys without cancer or any other disease fed on the mushroom *Lentinula edodes*. Medicinal plants are commonly used for therapy of various diseases and represent a significant part of old folk remedies. The healing properties of mushrooms have been known for hundreds of years. The number of mushroom types is estimated to be 140,000—of which only about 10% are known.

In the last 40 years, there has been an ever-increasing interest in the evaluation and use of natural products to reduce the risk of numerous diseases or to treat them directly. Bioactive polysaccharides, often isolated from various mushrooms, functioning as biological response modifiers, have quickly become the most studied natural immunomodulator. β -Glucans are glucose polymers present in the cell wall of yeast, fungi, and mushrooms ^[2]. β -Glucans possess broad immunomodulatory properties, including activation of innate immune functions such as the oxidative burst and adaptive immunity (Figure 1) ^{[3][4][5]}. Research on zymosan beginning in the 1940s was followed by the investigation of β -glucans in the 1960s and 1970s, when scientists established the significant influence of β -glucans on the immune system in relation to cancer treatment, anti-infection immunity, restoration of damaged bone, and the activation of innate-immunity cells (macrophages, dendritic cells, granulocytes, and natural killer (NK) cells) ^[6]. This activation triggers responses of adaptive-immunity cells, such as CD4⁺ or CD8⁺ T cells and B cells, resulting in the inhibition of tumor growth and metastasis ^[7]. Nonetheless, information on the mechanism of antimetastatic action of β -glucans remains limited.

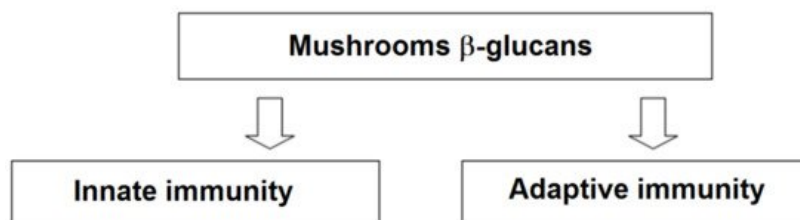


Figure 1. Mushroom-derived β -glucans affect all branches of the immune system.

A variety of polysaccharides from an array of sources can stimulate the immune system. β -Glucans are among these biological response modifiers, and their biological activities are well established (for review, see Vetvicka 2013 ^[8]). Some confusion is found among the more than 30,000 research papers, which is most probably caused by differences in source, primary structure, branching, and molecular weight ^{[9][10][11]}. The most pronounced effects were found in cancer ^{[7][12]} and

infection [13] models, but improvements in wound healing [14], inflammation [15], stress [16], immunotoxicity [17], genoprotection [18], viral infection [19], and allergy [20][21] were also found. For a review of the various biological effects of β -glucan, see Vannucci et al. (2013) [22]. The role of the various cell types is summarized in Figure 2.

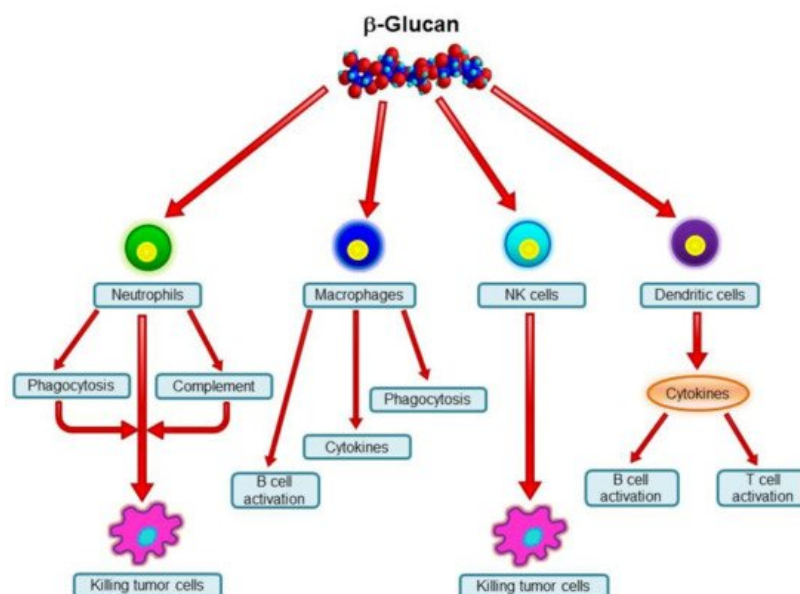


Figure 2. Major effects of β -glucans on immune cells.

In addition, β -glucans have been found to be active in every species tested, in earthworms [23], bees [24], shrimps [25], fish [26], chicken [27], mice, rats [28], rabbits, guinea pigs [29], sheep, pigs [30], cattle [31], and humans. These wide-ranging activities make β -glucan probably the only natural immunomodulator able to activate immune reactions across all species. The fact that β -glucans can easily penetrate the gut wall helps their recognition by other immunocytes and subsequent spreading throughout the body (Figure 3).

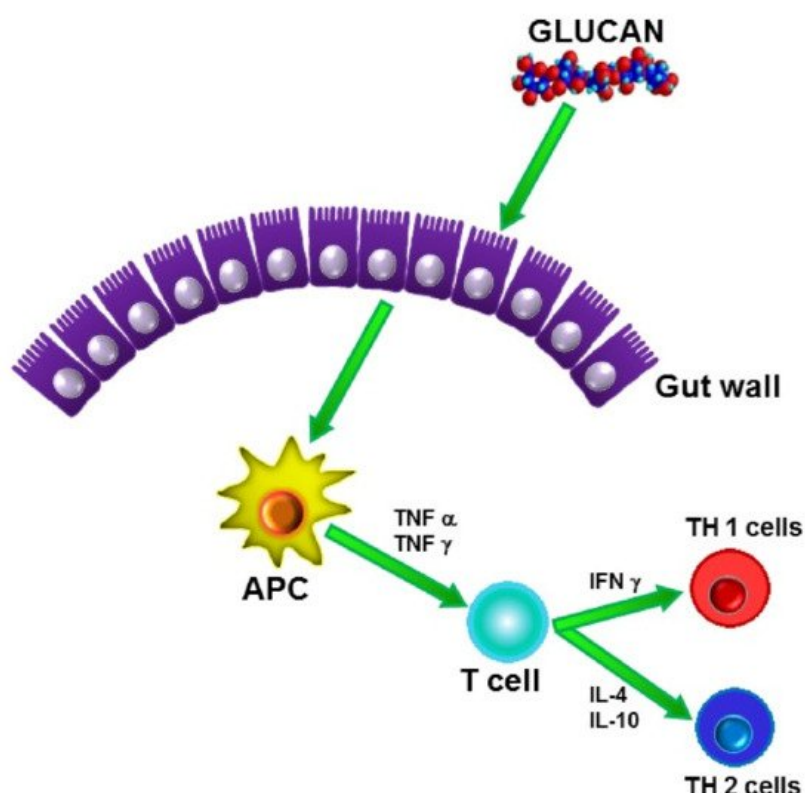


Figure 3. Transport of β -glucan through gut wall.

The fungal kingdom is significant. It not only provides food directly to humans but has also been a source of important drugs; mushroom compounds show promise in cancer immunotherapy [32]. β -Glucans are a group of polysaccharides belonging to the class known as biological response modifiers. They have potential therapeutic use for atherosclerosis, inflammatory disease, type 2 diabetes mellitus, and cancer [33][34][35]. Some hold promise as a new tool for vaccine development [36]. β -Glucans can serve as cellular structural polysaccharides or polysaccharides secreted on the surface of cells (in mushrooms or fungi) [37].

There are many types and sources of β -glucans; some have been studied more thoroughly as possible therapeutic compounds, but many need further research. β -(1 \rightarrow 3)-Glucans trigger different immune responses, and these polysaccharides have been shown to be effective immunostimulatory agents [38][39][40][41][42]. The mechanism behind the beneficial effect of β -glucans and other macrophage stimulators administered via different routes has not yet been clearly identified.

Mushrooms have been used in health care for treating simple and common diseases, like skin diseases and pandemic diseases such as AIDS. There are many investigated beneficial effects of β -glucans, including antioxidant, anti-inflammatory, anti-hypercholesterolemic, anti-viral and anti-cancer [43]. They are rich in carbohydrates, like β - and α -glucans, chitin, hemicellulose, mannans, xylans, and galactans, which make them the right choice for prebiotics. Not all medicinal effects of mushrooms, particularly the effects in cancer treatment, can be precisely explained by the action of one single molecule [32].

Hypercholesterolemia is a major risk factor of atherosclerosis and cardiovascular diseases [44]. Because statin use can be associated with muscle problems and other adverse effects, nonadherence and discontinuation of statin therapy are common and often lead to inadequate control of plasma cholesterol levels, increasing cardiovascular risk. In such situations, the hypocholesterolemic effects of β -glucans and other polysaccharides are attractive.

Various hypolipidemic agents are currently in use, including proprotein convertase subtilisin/kexin type 9 (PCSK9) inhibitors, apolipoprotein B-100 antisense oligonucleotides, cholesteryl ester transfer protein (CETP) inhibitors, and microsomal triglyceride transfer protein (MTTP) inhibitors, as well as yeast polysaccharides (β -glucans and mannans) and compounds derived from natural sources (nutraceuticals), such as glucomannans, plant sterols, berberine, and red yeast rice.

In diabetes mellitus, increased β -glucan intake correlates with improved glycemic control, which is associated with slower progression of this disease. There are differences in the effects on glycemic control and insulin sensitivity between oat (whole and bran) extract and β -glucan extract (taken orally) [45].

In cancer, dietary β -glucans can be used as soluble fiber with potential health-promoting effects. These compounds are believed to act on gut peptides, which are important signaling molecules in the regulation of energy and glucose homeostasis [46]. Fungotherapy for targeted treatment of cancer without harmful effects on healthy tissues is actively being developed, utilizing the active ingredients and mechanisms of action of mushrooms [47]. For the subsequent creation of drugs, the need for further studies on the mechanisms of antitumor/antiviral action of the components of medicinal mushrooms is great—for those growing under natural conditions and for those cultivated by solid-phase and submerged methods under laboratory and factory conditions [48].

2. β -Glucans Isolated from Different Sources

During 50 decades of research, countless types of β -glucan have been isolated, characterized, and tested. In the scientific literature, you can find several hundred different biological components all under the name β -glucan. There are numerous sources for glucan, from yeast to mushrooms to grain. β -Glucans can be relatively easily isolated from yeast, and the popularity of *Saccharomyces cerevisiae* is based on its availability and low cost. The main raison d'être of β -glucan is to form integral parts of the cell wall. Different physicochemical parameters (solubility, primary structure, molecular weight, branching, polymer charge) play a role in determining whether the polysaccharide modulates immune reactions. Some conclusions can be made. Branched or linear 1,4- β -glucans have very limited activity, if any. β -Glucans with 1,6 configurations usually have limited activity. The highest stimulation of defense reactions has been achieved with β -glucans that have a 1,3 configuration with additional branching at the position 0–6 of the 1,3-linked d-glucose residues. Among all β -glucans, those with a 1,3 configuration are best characterized in the literature. Excellent reviews of the relationship between structure and functional activity have been published [9][49]. For detailed insight into the chemistry, structure, and synthesis of β -glucans, see William et al. (2013) [50]. However, it is important to be mindful that no conclusive agreement on which composition or structure results in the most active glucan has yet been reached. Most likely because the precise structural confirmation of β -glucan in solution, usually due to free and independent rotation of glycosidic bonds between individual residues, is unknown. From the computer models we can assume a left- or right-handed triple helix configuration in water [42]. In fungal glucans, some chemical modifications, such as carboxymethylation, result in improvements of biological activity, particularly in antioxidant activity [51].

The choice of source is more historical than science-based. Investigation of β -glucans began in the 60s of the last century. Two lines can be traced in β -glucans history, based on different starting points, but slowly converging. The first one took place mainly in the USA and Europe, the second one in Asia, specifically Japan. Research on β -glucans in the Euro-

American milieu was based on knowledge of immunomodulatory effects of zymosan [52]. When zymosan was examined in detail, β -glucan was identified as a primary effective component. It was subsequently isolated, and the immunological effects were investigated [50][53]. In Asian medicine, consuming different medicinal mushrooms has a long tradition. Initial investigations conducted by Goro Chihara, who isolated lentinan from the shiitake mushroom (*Lentinus edodes*, now *Lentinula edodes* [54]), have led to approval of β -glucan as an official drug. For a summary of mushroom β -glucan action as immunomodulators of cancer immunotherapy, see Ayeka (2018) [32].

3. Mechanisms of Action

Modern chemotherapy of various tumors by cytostatic agents has shown solid efficacy against most cancers; however, further improvement of this approach is limited by adverse effects of antitumor therapy, namely by low selectivity of antitumor drugs, which is related to their toxicity (adverse effects and complications). In this regard, β -glucan has many biological activities and functions such as stimulation of the immune system and anti-inflammatory, antimicrobial, anti-infective, antiviral, antitumor, antioxidant, anticoagulant, cholesterol-lowering, radioprotective, and wound-healing properties [55][56]. This approach, especially chemically modified water-soluble polysaccharides in combination with antitumor drugs, seems appealing for further use in clinical oncology.

β -Glucans are recognized by numerous membrane receptors (Figure 4), which often share common characteristics. The most important β -glucan receptors are Dectin-1, CR3 receptor, and Toll-like receptors. Dectin-1 receptors react to the β -glucan binding by phosphorylation of the tyrosine-based activating motif [57]. The signaling events involve activation of Syk and NF- κ B pathways [58]. For detailed information of molecular interactions of β -glucan with Dectin-1, see Legentil, Paris, Ballet, Trouvelot, Daire, Vetvicka, and Ferrieres (2015) [42].

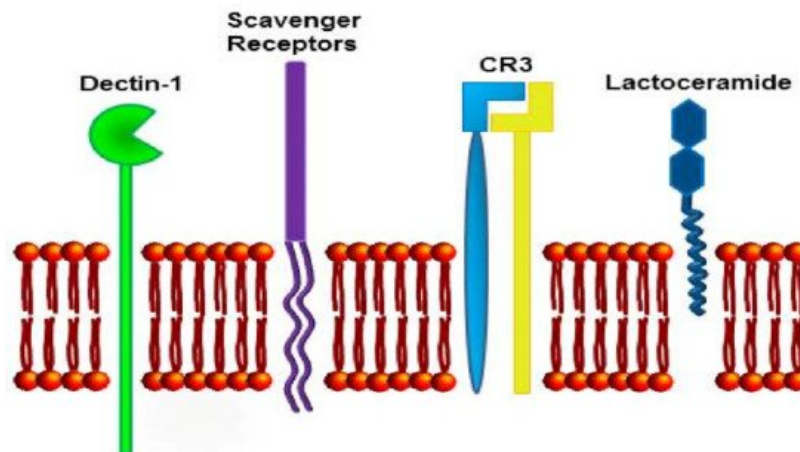


Figure 4. Major β -glucans-binding receptors.

Another important β -glucan receptor is CR3, known also as Mac-1, $\alpha_M \beta_2$ -integrin, or CD11b/CD18. This receptor is a one-membrane glycoprotein made up of two noncovalently linked α and β subunits known as CD11b or α_M and CD18 or β_2 . With respect to β -glucan binding, the key finding was that β -glucan bound to the lectin domain of CR3, and it primed the receptor in response to tumors that bore iC3b and were normally resistant to cellular cytotoxicity. Many human tumors generate an immune response that results in the deposition of antitumor antibodies, leading to the discovery of the significant therapeutic efficacy of combining β -glucan with antitumor monoclonal antibodies. This significant synergy has been demonstrated in a variety of murine tumors [59], as well as in human carcinoma xenograft models [60][61]. Another experimental proof of concept was reached when β -glucan-mediated therapeutic efficacy was restored by passive immunization with either natural antibodies in SCID mice or antitumor monoclonal antibodies in mice with low titers of natural antitumor antibodies. For more details on the mechanisms of effects on cancer suppression, see Li et al. (2010) [62]. However, the full explanation of how β -glucan binds to its receptor/s, and which receptor is more important for its biological effect, is still missing.

Another mechanism is the effect on myeloid-derived suppressor cells. β -glucan was found to stimulate the apoptosis of polymorphonuclear macrophages and dendritic cells and to regulate the differentiation of monocytic macrophages and dendritic cells [63].

An interesting suggestion was presented recently by Geller et al. (2019) [64]. These authors investigated the complex mechanisms at play within the tumor microenvironment and emphasized the need for the development of strategies that target immune cells within the tumor microenvironment. One such intervention can be β -glucan, a natural compound with an immunostimulatory and immunomodulatory potential and therapeutic anticancer effects. β -glucan can modulate the

tumor microenvironment both by bridging innate and adaptive immunity and by switching the phenotype of immunosuppressive cells to an immunostimulatory one. A new role for β -glucan in cancer therapy has been suggested because of an evolving understanding that β -glucan participates in a phenomenon called trained immunity, where innate-immunity cells take on memory phenotypes. This new concept suggests that β -glucan may play an essential part in the prevention and suppression of the growth of various tumors; these effects are important in cancer therapy [64].

Anti-inflammatory activities of some medicinal mushrooms in gut inflammation were shown by Ishimoto et al. (2018) [65]. Those authors performed autodigestion of *Ganoderma lingzhi* and found an enhanced release of hypotensive peptides and an immunomodulatory β -1,3-glucan. Gut inflammation was assessed by measuring the lengths of the intestines and colon, and sepsis was evaluated by means of the survival of the animals.

A main feature of β -glucans is their capacity to function as biological response modifiers, exerting regulatory effects on inflammation and shaping the effector functions of different innate and adaptive immunity cell populations. The potential to interfere with processes involved in the development or control of cancer makes β -glucans interesting candidates as adjuvants in antitumor therapies, as well as in cancer prevention strategies [66]. The recognition by innate immunity cells occurs via ligation of specific PRR, such as Toll-like and C-type lectin-like receptors. Among the latter, Dectin-1 is the best characterized receptor, reported to bind β -glucan from different sources, and is expressed on the surface of monocytes, macrophages, neutrophils, and DC and T lymphocytes [66]. Other receptors, including lactosylceramide receptor, mannose receptor, and complement and scavenger receptors were reported to directly bind β -glucan or to cooperate with Dectin-1 for its recognition; β -glucan was shown to stimulate NK cell cytotoxic activity through direct binding to the NKp30 activating receptor. In vitro, β -glucans can enhance the functional activity of monocytes/macrophages and DC, and activate antimicrobial activity of mononuclear cells and neutrophils.

Mushroom β -glucan may immunomodulate the tumor-associated macrophages in such tumors, like Lewis lung carcinoma [6]. Authors have shown the efficacious effect of mushroom polysaccharides for ameliorating the immune suppression in the tumor microenvironment. Increased M1 phenotype of tumor-associated macrophages and attenuated M2 phenotype of tumor-associated macrophages could be achieved by ingesting mushroom polysaccharides.

The structure of *Grifola fondosa* polysaccharide was identified to be a β -d-(1,3)-linked glucan backbone with a single β -d-(1,6)-linked glucopyranosyl residue branched at C-6 on every third residue. This polysaccharide could interact with poly(A) moiety of a designed antisense oligonucleotide targeting the primary transcript of proinflammatory cytokine TNF α (TNF α -A60). This *Grifola fondosa* polysaccharide-based complex could incorporate TNF α -A60 into the macrophage cells via Dectin-1 receptor and attenuate lipopolysaccharide-induced secretion of TNF α . It was concluded that GFPS could be applied to deliver therapeutic oligonucleotides for the treatment of diseases such as inflammation and cancers [67]. So, β -glucan from *Grifola frondosa* effectively delivers therapeutic oligonucleotide into cells via Dectin-1 receptor and attenuates TNF α gene expression.

In addition to direct stimulation of various cell types involved in the fight against cancer, β -glucan was also found to ameliorate the negative side effects of chemotherapy, including immunosuppression. Recent observation demonstrated that β -glucan-based alleviation of cyclophosphamide-induced severe immunosuppression is caused by regulation of gut microbiota [68].

Another possible mechanism is the inhibition of transformation induced by oncogenes. This was demonstrated with glucans from *Ganoderma lucidum* and *Tricholoma lobayence* using cell transformation caused by *ras* oncogene. Both glucans successfully inhibited cell transformation [69]. The fact that this inhibitory effect required the presence of normal cells cannot be presently explained.

Numerous molecular targets of mushroom-derived β -glucan involve NF- κ B inhibitors, protein kinase inhibitors, modulators of G1/S and G2/M checkpoints, inhibitors of MAPK protein kinase signaling pathways, cyclooxygenase inhibitors, and DNA polymerase inhibitors (for review, see Zaidman, Yassin, Mahajna, and Wasser (2005) [70]). In cases of lentinan, recent observation suggested that the breast cancer progression inhibition is modulated via the Nur77/HIF-1 α axis [71]. The authors speculate that HIFs might be a potential target in breast cancer treatment.

Some mushroom β -glucans can have positive effects on cancer development manifested indirectly. Inflammatory bowel diseases have a clear connection with the development of colorectal cancer; colitis-associated colorectal cancers form over 5% of all colorectal cancers [72], with some studies reporting as high as 43% [73]. β -glucan administration does not only attenuate inflammation and other symptoms of colitis [74], but prevents carcinogenesis via inhibition of P450 1A2 expression [75]. Similarly, oral supplementation with *Pleurotus*-derived β -glucan suppressed expression of proliferation-associated marker proliferating cell nuclear antigen, meaning that β -glucan administration will suppress abnormal proliferative activity of pre- and neoplastic cells, and subsequently suppress the development of cancer [76].

An unknown mechanism was found in a study of β -glucan from *Sparassis crispa*. Using colon cancer as a model, this β -glucan exhibited direct toxicity against various human colon cancer cell lines by destroying membrane integrity, whereas normal colon cells were resistant [77]. β -glucan direct toxicity is extremely rare, so currently there is no explanation of these effects.

Even though most of the effects of β -glucan are manifested via binding to some of the numerous specific receptors, few studies describing direct effects of β -glucan on cells expressing none of these receptors exist. A glucose-based polysaccharide isolated from *Ganoderma lucidum* was found to have direct effects on lung cancer cells, probably via inhibition of phosphorylation of several signaling molecules [78]. A β -glucan from *Pleurotus djamor* used in high doses inhibited and killed ovarian carcinoma cells PA1 [79]. Surprisingly, soluble β -glucan isolated from the same mushroom had no direct cytotoxic effects. Proteoglycan from *Grifola frondosa* modulated expression of some cancer-related genes in both canine and human tumor cells [80]. Our laboratory found somewhat similar results using synthetic glucan-based oligosaccharides [81]. Some direct effects on lung cancer cells were also described for β -glucan from *Antrodia cinnamomea*, with the supposed mechanism being regulation of the TGF β /AKT/GSK3 β axis [82]. β -glucan from *Lentinus edodes* can in some cases inhibit proliferation of breast cancer cells when used simultaneously with hypoxic conditions [71]. The problem of all these studies, however, is the fact that none of them were ever independently repeated. In addition, they all differ in type of β -glucan and in experimental conditions. Therefore, the possible direct effects of β -glucan remain a side note.

4. The Antitumor Effect of β -Glucan in Humans

Mushroom-derived β -glucan-rich polysaccharides are known for their immunomodulatory and antitumor properties. The polysaccharide fraction, mainly β -glucans, is responsible for the immunomodulatory effects. Fungal β -glucans have been proven to activate leukocytes, and this action depends on structural characteristics of β -glucans [83]. Additionally, these compounds may be employed as drug carriers. β -Glucan research is now focused on human studies: clinical trials and epidemiological assessment of the efficacy and safety of mushroom-derived β -glucans for cancer treatment and prevention [84]. Fungal β -glucans can be used as adjuvants for treating cancer patients [85][86]. In general, β -glucans are a promising option for cancer prevention and treatment, especially for cervical cancer. The therapeutic potential of β -glucan alone or as an adjuvant therapy in cervical cancer has been documented; moreover, some authors highlighted β -glucans as drug carriers for preventive and therapeutic use [87]. Glucans and specific proteins are responsible for most of the biological effects of mushrooms, particularly in terms of immunomodulatory and antitumor activities [85]. Human studies represent a minority of the available data, as exemplified by placebo-controlled trials.

β -1,3-d-Glucans and β -1,6-d-glucans (polysaccharides from higher fungi) increase the number of Th1 lymphocytes, which help to prevent allergic reactions. Some β -glucans, like pleuran from oyster mushrooms (*Pleurotus* spp.) or lentinan from shiitake mushrooms (*Lentinula edodes*), have a marked anticarcinogenic activity. In addition to having an immunostimulatory effect, β -glucans may participate in the physiological processes related to the metabolism of fats in the human body. Their therapeutic application causes a decrease in the total cholesterol content of the blood and may contribute to reductions in body weight [88].

Vetvicka and Vetvickova [17][36] compared five different β -glucans, isolated from algae, yeast, bacteria, oats, and a mushroom, by studying their promotion of the phagocytosis of blood cells and the secretion of IL-2, and their suppression of melanoma and breast and lung cancers. In addition, those authors evaluated the impact of β -glucan supplementation on two experimental models of infection. It was concluded that most of the tested β -glucans stimulated phagocytosis and IL-2 secretion, reduced cancer growth, and ameliorated the effects of the experimental infections. Some clinical trials of fungal β -glucans as an adjuvant therapy for cancer started to appear in the literature in the 1980s [86].

Dietary β -glucans are soluble fiber with possible health-promoting effects. Gut peptides serve as important signals for the regulation of energy and glucose homeostasis. This article reviews the effects of different foods enriched in β -glucan on immune responses, inflammation, gut hormones, and cancer. Gut hormones are influenced by the consumption of β -glucan-enriched foods, and in humans, the levels of such peptides as ghrelin and glucagonlike peptides 1 and 2 influence serum glucose concentration, as well as innate and adaptive immunity. Cancer cell progression is also regulated by obesity and glucose dyshomeostasis, which are affected by β -glucan consumption with food, in turn regulating gut hormones [46].

β -glucan alters inflammation via immunostimulatory patterns [89][90]. This phenomenon may be related to a possible effect on gut hormone control (Huang, 2015), although there are opposite opinions [2].

Macrophages perform an important function in all phases of host defense in innate and adaptive immunity through the secretion of cytokines (IL-1, IL-6, IL-8, IL-12, and TNF- α) and inflammatory mediators like nitric oxide (NO) and hydrogen peroxide (H₂O₂) [46]. Moreover, as reported recently, β -glucan (a Dectin-1 ligand) promotes macrophage M1 polarization via the NF- κ B/autophagy pathway. Additionally, Dectin-1 small interfering RNA (siRNA), autophagy inducer rapamycin, and NF- κ B inhibitor SN50 reverse the impact of β -glucan on the autophagy level and macrophage M1 polarization, suggesting that Dectin-1 and NF- κ B act upstream of autophagy [91].

According to the data of Wang, Wu, Chen, Liu, and Chen (2015) [6], obtained in an experimental Lewis lung carcinoma tumor model, oral treatment with *Ganoderma lucidum* or *Antrodia camphorata* polysaccharides significantly reduced the TGF β release into blood serum. Simultaneously, it was shown that oral mushroom β -glucan treatment significantly increased IFN γ mRNA expression but significantly reduced COX2 mRNA expression in the lungs. In addition, IL-12 and IFN γ mRNA expression was significantly increased, but IL-6, IL-10, COX2, and TGF β mRNA expression levels were substantially lowered following oral treatment with mushroom polysaccharides. This highlights a strong beneficial effect of mushroom polysaccharides against immune suppression in the tumor microenvironment [59]. An enhanced M1 phenotype of tumor-associated macrophages (TAM) and their attenuated M2 phenotype could be achieved by ingesting mushroom polysaccharides.

Recently, researchers isolated and compared α - and β -glucans from shiitake mushrooms (*L. edodes*) with different biological activities [92][93]. A polysaccharide-enriched extract obtained from *L. edodes* was subjected to several purification steps to separate three d-glucans containing β -(1,6), β -(1,3), α -(1,6), and α -(1,3) linkages, with subsequent characterization by nuclear magnetic resonance spectroscopy, gas chromatography coupled with mass spectrometry, infrared spectroscopy, size exclusion chromatography, and other methods.

The anticancer effect of β -glucans may be related to their control of inflammation via immunostimulatory patterns [46]; on the other hand, it might be associated with a possible influence on the control of gut hormones [89]. When β -glucans are employed as immunostimulatory agents or adjuvant therapeutics, several receptors have been reported to recognize β -glucans, including Dectin-1, complement receptor 3 (CR3), CD5, and lactosylceramide [94] (Figure 5). Bose et al. [3][4] investigated the participation of various receptors (Dectin-1 and CR3, involved in the oxidative burst) in response to different physical forms of β -glucans in human monocytes.

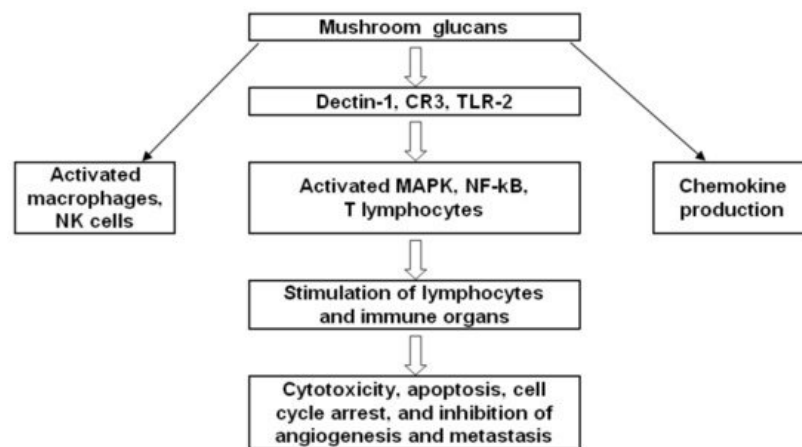


Figure 5. A probable immunomodulatory mechanism of the action of mushroom glucans.

Furthermore, β -glucans can induce the proliferation of human peripheral blood mononuclear cells and maturation of monocyte-derived dendritic cells via cytokine production [95]. Therefore, it seems that β -glucans can stimulate a broad immune response including phagocytosis and proinflammatory events, which may lead to the elimination of infectious agents (e.g., *Staphylococcus aureus*, *Escherichia coli*, *Candida albicans*, *Pneumocystis carinii*, *Listeria monocytogenes*, *Leishmania donovani*, and an influenza virus). Several studies have confirmed in animal models that systemic treatment with β -glucans enhances the migration of neutrophils into a site of inflammation and improves their antimicrobial function [96].

Mushroom compounds modulate the immune system, thereby helping it to fight tumors and other diseases. These compounds can strengthen the immune system by stimulating lymphocytes, NK cells, and macrophages; by enhancing cytokine production; by inhibiting the proliferation of cancer cells; by promoting apoptosis; and by blocking angiogenesis, in addition to being cytotoxic to cancer cells. These compounds encounter intestinal cells, the frontline of the intestinal

immune system, and interact with antigens, thereby taking part in an intestinal immune response and inducing an inflammatory response if necessary. [Figure 6](#) shows the modulation of the immune cells in the tumor microenvironment by β -glucan.

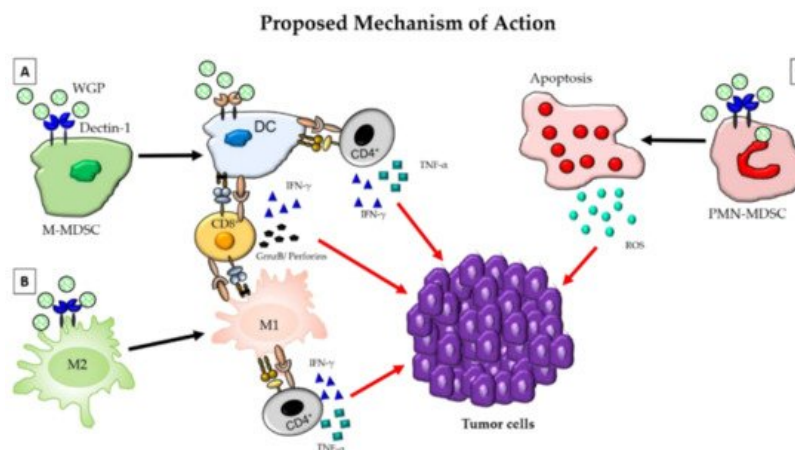


Figure 6. The modulation of the immune cells in the tumor microenvironment by β -glucan. β -glucan binds to the Dectin-1 receptors expressed on cells of the myeloid lineage and is then phagocytosed. In (A), β -glucan can be seen binding to Dectin-1 on an M-MDSC. Binding to the M-MDSC will cause the M-MDSC to switch from a suppressive phenotype to a DC phenotype that can act as an APC. This dendritic cell (DC) will then activate CD4⁺ and CD8⁺ T-cells, where CD4⁺ T-cells will secrete pro-inflammatory cytokines, such as TNF α and IFN- γ , and CD8⁺ T-cells will secrete Granzyme B, perforins, and IFN- γ . The secretion of these pro-inflammatory cytokines by CD4⁺ and CD8⁺ T-cells will lead to the destruction of tumor cells. β -Glucan induces the polarization of suppressive M2 macrophages (B) into inflammatory M1 macrophages. M1 macrophages will then activate Th1 type T-cells, leading to damage to the tumor cells through the secretion of pro-inflammatory cytokines by CD4⁺ and CD8⁺ T-cells. Finally, in (C), β -glucan will bind to the Dectin-1 receptor on polymorphonuclear (PMN)-MDSCs and cause apoptosis of the cell. As the cell undergoes apoptosis, it will produce ROS that will ultimately target the tumor cells, leading to tumor cell death. Overall, these mechanisms together convert a suppressive tumor microenvironment (TME) to an inflammatory TME that has a greater potential to induce the killing of tumors. From Geller, Shrestha, and Yan (2019) [64].

Studies of glucans in human cancers include clinical trials and epidemiological data related to the efficacy and safety of mushroom-derived β -glucans in cancer treatment and prevention [84]. Recently, Medicinal Mushrooms Physician Data Query (PDQ): Health Professional Version [97] presented a cancer information summary for health professionals, providing comprehensive, peer-reviewed, evidence-based information about the use of medicinal mushrooms in the treatment of patients with cancer. According to recent research, β -glucans have a variety of potential therapeutic properties as well as metabolic and beneficial effects on the gastrointestinal tract, and hold promise for further clinical application [98]. Therapeutic effects of fungal β -glucans on human colon cancer were documented [99]; the β -glucans decreased the size of xenografted colon cancer tumors via the stimulation of the immune system and direct cytotoxicity. This type of polysaccharide can also exert synergistic effects with chemotherapeutic agents and other drugs (immune stimulators). An innovative strategy is to utilize β -glucans to deliver nanoparticles containing chemotherapeutic agents to the site of colon cancer, thus improving therapeutic efficacy. Lentinan is a component of the second most cultivated and most popular edible mushroom in the world, known as “xianggu” in China and “shiitake” in Japan. There are 9474 reported cases of lentinan-associated cancer treatments, including lung cancer (3469 cases), gastric cancer, and other malignant tumors [55]. A comprehensive review of mushroom β -glucans in cancer therapy suggested stimulation of immune system as the basic mechanism of action [84]. Fungal β -glucans are promising as an adjuvant therapy for cancer [86], as well as modulating the immune system weakened by radiotherapy and chemotherapy in cancer treatment [32].

References

1. Lind, J. A Treatise of the Scurvy: In Three Parts, Containing an Inquiry into the Nature, Causes, and Cure, of That Disease, Together with a Critical and Chronological View of What Has Been Published on the Subject; Sands, Murray, and Cochran: Edinburgh, UK, 1753.
2. Jayachandran, M.; Chen, J.; Chung, S.S.M.; Xu, B. A critical review on the impacts of beta-glucans on gut microbiota and human health. *J. Nutr. Biochem.* 2018, 61, 101–110.
3. Bose, N.; Chan, A.S.; Guerrero, F.; Maristany, C.M.; Qiu, X.; Walsh, R.M.; Ertelt, K.E.; Jonas, A.B.; Gorden, K.B.; Dudney, C.M.; et al. Binding of soluble yeast beta-glucan to human neutrophils and monocytes is complement-

dependent. *Front. Immunol.* 2013, 4, 230.

4. Bose, N.; Wurst, L.R.; Chan, A.S.; Dudney, C.M.; LeRoux, M.L.; Danielson, M.E.; Will, P.M.; Nodland, S.E.; Patchen, M.L.; Dalle Lucca, J.J.; et al. Differential regulation of oxidative burst by distinct beta-glucan-binding receptors and signaling pathways in human peripheral blood mononuclear cells. *Glycobiology* 2014, 24, 379–391.
5. Vetvicka, V.; Vetvickova, J. B-glucan improves condition of chronic fatigue in mice by stimulation of immunity. *Open Biochem. J.* 2020, 14, 1–8.
6. Wang, W.J.; Wu, Y.S.; Chen, S.; Liu, C.F.; Chen, S.N. Mushroom beta-glucan may immunomodulate the tumor-associated macrophages in the Lewis lung carcinoma. *BioMed Res. Int.* 2015, 2015, 604385.
7. Yoon, T.J.; Koppula, S.; Lee, K.H. The effects of beta-glucans on cancer metastasis. *Anticancer Agents Med. Chem.* 2013, 13, 699–708.
8. Vetvicka, V. [Beta]-glucans as Natural Biological Response Modifiers; Nova Science Publishers, Inc.: New York, NY, USA, 2013.
9. Bohn, J.A.; BeMiller, J.N. (1→3)- β -D-Glucans as biological response modifiers: A review of structure-functional activity relationships. *Carbohydr. Polym.* 1995, 28, 3–14.
10. Leung, P.H.; Zhang, Q.X.; Wu, J.Y. Mycelium cultivation, chemical composition and antitumor activity of a *Tolyposcladium* sp. fungus isolated from wild *Cordyceps sinensis*. *J. Appl. Microbiol.* 2006, 101, 275–283.
11. Zhang, L.; Li, X.; Xu, X.; Zeng, F. Correlation between antitumor activity, molecular weight, and conformation of lentinan. *Carbohydr. Res.* 2005, 340, 1515–1521.
12. Yan, J.; Vetvicka, V.; Xia, Y.; Coxon, A.; Carroll, M.C.; Mayadas, T.N.; Ross, G.D. Beta-glucan, a “specific” biologic response modifier that uses antibodies to target tumors for cytotoxic recognition by leukocyte complement receptor type 3 (CD11b/CD18). *J. Immunol.* 1999, 163, 3045–3052.
13. Novak, M.; Vetvicka, V. Glucans as biological response modifiers. *Endocr. Metab. Immune Disord. Drug Targets* 2009, 9, 67–75.
14. Majtan, J.; Jesenak, M. beta-glucans: Multi-functional modulator of wound healing. *Molecules* 2018, 23, 806.
15. Vetvicka, V.; Vetvickova, J. Combination therapy with glucan and coenzyme Q10 in murine experimental autoimmune disease and cancer. *Anticancer Res.* 2018, 38, 3291–3297.
16. Bashir, K.M.I.; Choi, J.S. Clinical and physiological perspectives of beta-glucans: The past, present, and future. *Int. J. Mol. Sci.* 2017, 18, 1906.
17. Vetvicka, V.; Vetvickova, J. Glucans and cancer: Comparison of commercially available beta-glucans—Part IV. *Anticancer Res.* 2018, 38, 1327–1333.
18. Boulaka, A.; Christodoulou, P.; Vlassopoulou, M.; Koutrotsios, G.; Bekiaris, G.; Zervakis, G.I.; Mitsou, E.K.; Saxami, G.; Kyriacou, A.; Zervou, M.; et al. Genoprotective properties and metabolites of beta-glucan-rich edible mushrooms following their in vitro fermentation by human faecal microbiota. *Molecules* 2020, 25, 3554.
19. Muthusamy, G.; Joardar, S.N.; Samanta, I.; Isore, D.P.; Roy, B.; Maiti, T.K. Dietary administered purified β -glucan of edible mushroom (*Pleurotus florida*) provides immunostimulation and protection in broiler experimentally challenged with virulent Newcastle disease virus. *J. Basic Appl. Zool.* 2020, 81, 55.
20. Jesenak, M.; Banovcin, P.; Rennerova, Z.; Majtan, J. beta-Glucans in the treatment and prevention of allergic diseases. *Allergol. Immunopathol.* 2014, 42, 149–156.
21. Kirmaz, C.; Bayrak, P.; Yilmaz, O.; Yuksel, H. Effects of glucan treatment on the Th1/Th2 balance in patients with allergic rhinitis: A double-blind placebo-controlled study. *Eur. Cytokine Netw.* 2005, 16, 128–134.
22. Vannucci, L.; Krizan, J.; Sima, P.; Stakheev, D.; Caja, F.; Rajsiglova, L.; Horak, V.; Saieh, M. Immunostimulatory properties and antitumor activities of glucans (Review). *Int. J. Oncol.* 2013, 43, 357–364.
23. Beschin, A.; Bilej, M.; Hanssens, F.; Raymakers, J.; Van Dyck, E.; Revets, H.; Brys, L.; Gomez, J.; De Baetselier, P.; Timmermans, M. Identification and cloning of a glucan- and lipopolysaccharide-binding protein from *Eisenia foetida* earthworm involved in the activation of prophenoloxidase cascade. *J. Biol. Chem.* 1998, 273, 24948–24954.
24. Mazzei, M.; Fronte, B.; Sagona, S.; Carrozza, M.L.; Forzan, M.; Pizzurro, F.; Bibbiani, C.; Miragliotta, V.; Abramo, F.; Millanta, F.; et al. Effect of 1,3-1,6 beta-glucan on natural and experimental deformed wing virus infection in newly emerged honeybees (*Apis mellifera ligustica*). *PLoS ONE* 2016, 11, e0166297.
25. Duvic, B.; Soderhall, K. Purification and characterization of a beta-1,3-glucan binding protein from plasma of the crayfish *Pacifastacus leniusculus*. *J. Biol. Chem.* 1990, 265, 9327–9332.
26. Vetvicka, V.; Vannucci, L.; Sima, P. The effects of beta-glucan on fish immunity. *N. Am. J. Med. Sci.* 2013, 5, 580–588.

27. Jacob, J.P.; Pescatore, A.J. Barley beta-glucan in poultry diets. *Ann. Transl. Med.* 2014, 2, 20.
28. Feletti, F.; De Bernardi di Valserra, M.; Contos, S.; Mattaboni, P.; Germogli, R. Chronic toxicity study on a new glucan extracted from *Candida albicans* in rats. *Arzneimittelforschung* 1992, 42, 1363–1367.
29. Ferencik, M.; Kotulova, D.; Masler, L.; Bergendi, L.; Sandula, J.; Stefanovic, J. Modulatory effect of glucans on the functional and biochemical activities of guinea-pig macrophages. *Methods Find. Exp. Clin. Pharmacol.* 1986, 8, 163–166.
30. Benkova, M.; Boroskova, Z.; Soltys, J. [Immunostimulatory effects of certain substances in experimental ascaridiasis in pigs]. *Vet. Med.* 1992, 36, 717–724.
31. Buddle, B.M.; Pulford, H.D.; Ralston, M. Protective effect of glucan against experimentally induced staphylococcal mastitis in ewes. *Vet. Microbiol.* 1988, 16, 67–76.
32. Ayeka, P.A. Potential of mushroom compounds as immunomodulators in cancer immunotherapy: A review. *Evid. Based Complement. Altern. Med.* 2018, 2018, 7271509.
33. Driscoll, M.; Hansen, R.; Ding, C.; Cramer, D.E.; Yan, J. Therapeutic potential of various beta-glucan sources in conjunction with anti-tumor monoclonal antibody in cancer therapy. *Cancer Biol. Ther.* 2009, 8, 218–225.
34. Novak, M.; Vetvicka, V. Beta-glucans, history, and the present: Immunomodulatory aspects and mechanisms of action. *J. Immunotoxicol.* 2008, 5, 47–57.
35. Vetvicka, V.; Vannucci, L.; Sima, P.; Richter, J. Beta glucan: Supplement or drug? From laboratory to clinical trials. *Molecules* 2019, 24, 1251.
36. Vetvicka, V.; Vetvickova, J. Anti-infectious and anti-tumor activities of beta-glucans. *Anticancer Res.* 2020, 40, 3139–3145.
37. Blagodatski, A.; Yatsunskaya, M.; Mikhailova, V.; Tiasto, V.; Kagansky, A.; Katanaev, V.L. Medicinal mushrooms as an attractive new source of natural compounds for future cancer therapy. *Oncotarget* 2018, 9, 29259–29274.
38. Cyprik, W.; Ohman, T.; Eskelinen, E.L.; Matikainen, S.; Nyman, T.A. Quantitative proteomics of extracellular vesicles released from human monocyte-derived macrophages upon beta-glucan stimulation. *J. Proteome Res.* 2014, 13, 2468–2477.
39. Jozefowski, S.; Yang, Z.; Marcinkiewicz, J.; Kobzik, L. Scavenger receptors and beta-glucan receptors participate in the recognition of yeasts by murine macrophages. *Inflamm. Res.* 2012, 61, 113–126.
40. Kogan, G.; Pajtinka, M.; Babincova, M.; Miadokova, E.; Rauko, P.; Slamenova, D.; Korolenko, T.A. Yeast cell wall polysaccharides as antioxidants and antimutagens: Can they fight cancer? *Neoplasma* 2008, 55, 387–393.
41. Kogan, G.; Sandula, J.; Korolenko, T.A.; Falameeva, O.V.; Poteryaeva, O.N.; Zhanaeva, S.Y.; Levina, O.A.; Filatova, T.G.; Kaledin, V.I. Increased efficiency of Lewis lung carcinoma chemotherapy with a macrophage stimulator--yeast carboxymethyl glucan. *Int. Immunopharmacol.* 2002, 2, 775–781.
42. Legentil, L.; Paris, F.; Ballet, C.; Trouvelot, S.; Daire, X.; Vetvicka, V.; Ferrieres, V. Molecular interactions of beta-(1-->3)-glucans with their receptors. *Molecules* 2015, 20, 9745–9766.
43. Teplyakova, T.V.; Ilyicheva, T.N.; Andreeva, I.; Solovyanova, N. The activity of components of true tinder mushroom, *Chaga Inonotus obliquus* (Fr.) Pil. against viruses, bacteria and fungi (Abstract). In Proceedings of the 10th International Medicinal Mushroom Conference, Nantong, China, 19–22 September 2019; p. 117.
44. Johnston, T.P.; Korolenko, T.A.; Pirro, M.; Sahebkar, A. Preventing cardiovascular heart disease: Promising nutraceutical and non-nutraceutical treatments for cholesterol management. *Pharmacol. Res.* 2017, 120, 219–225.
45. He, L.X.; Zhao, J.; Huang, Y.S.; Li, Y. The difference between oats and beta-glucan extract intake in the management of HbA1c, fasting glucose and insulin sensitivity: A meta-analysis of randomized controlled trials. *Food Funct.* 2016, 7, 1413–1428.
46. Baldassano, S.; Accardi, G.; Vasto, S. Beta-glucans and cancer: The influence of inflammation and gut peptide. *Eur. J. Med. Chem.* 2017, 142, 486–492.
47. Shnyreva, A.V.; Shnyreva, A.A.; Espinoza, C.; Padron, J.M.; Trigos, A. Antiproliferative activity and cytotoxicity of some medicinal wood-destroying mushrooms from Russia. *Int. J. Med. Mushrooms* 2018, 20, 1–11.
48. Teplyakova, T.V.; Ilyicheva, T.N.; Markovich, N.A. Prospects for the development of anti-influenza drugs based on medicinal mushrooms (review). *Appl. Biochem. Microbiol.* 2020, 56, 489–496.
49. Šandula, J.; Kogan, G.; Kačuráková, M.; Machová, E. Microbial (1–3)-β-d-glucans, their preparation, physico-chemical characterization and immunomodulatory activity. *Carbohydr. Polym.* 1999, 38, 247–253.

50. William, D.L.; Lowman, D.W.; Reale, M.; Ensley, H.E. Insights into the physicochemical characterization, chemistry, structure and synthesis of (1-3,1-6)- β -glucans. In *Biology and Chemistry of Beta Glucan: Beta-Glucan, Structure, Chemistry and Specific Application*; Vetvicka, V., Novak, M., Eds.; Bentham Science Publishers: Sharjah, United Arab Emirates, 2013; Volume 2.
51. Theis, T.V.; Queiroz Santos, V.A.; Appelt, P.; Barbosa-Dekker, A.M.; Vetvicka, V.; Dekker, R.F.H.; Cunha, M.A.A. Fungal exocellular (1-6)-beta-d-glucan: Carboxymethylation, characterization, and antioxidant activity. *Int. J. Mol. Sci.* 2019, 20, 2337.
52. Pillemer, L.; Ecker, E.E. Anticomplementary factor in fresh yeast. *J. Biol. Chem.* 1941, 137, 139–142.
53. Di Luzio, N.R.; Riggi, S.J. The effects of laminarin, sulfated glucan and oligosaccharides of glucan on reticuloendothelial activity. *J. Reticuloendothel. Soc.* 1970, 8, 465–473.
54. Chihara, G.; Maeda, Y.; Hamuro, J.; Sasaki, T.; Fukuoka, F. Inhibition of mouse sarcoma 180 by polysaccharides from *Lentinus edodes* (Berk.) Sing. *Nature* 1969, 222, 687–688.
55. Zhang, M.; Zhang, Y.; Zhang, L.; Tian, Q. Mushroom polysaccharide lentinan for treating different types of cancers: A review of 12 years clinical studies in China. *Prog. Mol. Biol. Transl. Sci.* 2019, 163, 297–328.
56. Yuan, H.; Lan, P.; He, Y.; Li, C.; Ma, X. Effect of the modifications on the physicochemical and biological properties of beta-glucan—a critical review. *Molecules* 2019, 25, 57.
57. Gantner, B.N.; Simmons, R.M.; Canavera, S.J.; Akira, S.; Underhill, D.M. Collaborative induction of inflammatory responses by dectin-1 and Toll-like receptor 2. *J. Exp. Med.* 2003, 197, 1107–1117.
58. Fang, J.; Wang, Y.; Lv, X.; Shen, X.; Ni, X.; Ding, K. Structure of a beta-glucan from *Grifola frondosa* and its antitumor effect by activating Dectin-1/Syk/NF- κ B signaling. *Glycoconj. J.* 2012, 29, 365–377.
59. Hong, F.; Yan, J.; Baran, J.T.; Allendorf, D.J.; Hansen, R.D.; Ostroff, G.R.; Xing, P.X.; Cheung, N.K.; Ross, G.D. Mechanism by which orally administered beta-1,3-glucans enhance the tumoricidal activity of antitumor monoclonal antibodies in murine tumor models. *J. Immunol.* 2004, 173, 797–806.
60. Cheung, N.K.; Modak, S.; Vickers, A.; Knuckles, B. Orally administered beta-glucans enhance anti-tumor effects of monoclonal antibodies. *Cancer Immunol. Immunother.* 2002, 51, 557–564.
61. Salvador, C.; Li, B.; Hansen, R.; Cramer, D.E.; Kong, M.; Yan, J. Yeast-derived beta-glucan augments the therapeutic efficacy mediated by anti-vascular endothelial growth factor monoclonal antibody in human carcinoma xenograft models. *Clin. Cancer Res.* 2008, 14, 1239–1247.
62. Li, B.; Cai, Y.; Qi, C.; Hansen, R.; Ding, C.; Mitchell, T.C.; Yan, J. Orally administered particulate beta-glucan modulates tumor-capturing dendritic cells and improves antitumor T-cell responses in cancer. *Clin. Cancer Res.* 2010, 16, 5153–5164.
63. Albeituni, S.H.; Ding, C.; Liu, M.; Hu, X.; Luo, F.; Kloecker, G.; Bousamra, M., II; Zhang, H.G.; Yan, J. Yeast-derived particulate beta-glucan treatment subverts the suppression of myeloid-derived suppressor cells (MDSC) by inducing polymorphonuclear MDSC apoptosis and monocytic MDSC differentiation to APC in cancer. *J. Immunol.* 2016, 196, 2167–2180.
64. Geller, A.; Shrestha, R.; Yan, J. Yeast-derived beta-glucan in cancer: Novel uses of a traditional therapeutic. *Int. J. Mol. Sci.* 2019, 20, 3618.
65. Ishimoto, Y.; Ishibashi, K.I.; Yamanaka, D.; Adachi, Y.; Ito, H.; Igami, K.; Miyazaki, T.; Ohno, N. Protection against gut inflammation and sepsis in mice by the autodigested product of the Lingzhi medicinal mushroom, *Ganoderma lingzhi* (agaricomycetes). *Int. J. Med. Mushrooms* 2018, 20, 809–823.
66. Del Corno, M.; Gessani, S.; Conti, L. Shaping the innate immune response by dietary glucans: Any role in the control of cancer? *Cancers* 2020, 12, 155.
67. Cui, H.; Zhu, X.; Huo, Z.; Liao, B.; Huang, J.; Wang, Z.; Song, C.; Hu, X.; Fang, J. A beta-glucan from *Grifola frondosa* effectively delivers therapeutic oligonucleotide into cells via dectin-1 receptor and attenuates TNF α gene expression. *Int. J. Biol. Macromol.* 2020, 149, 801–808.
68. Vetvicka, V.; Vetvickova, J. Combination of glucan, resveratrol and vitamin C demonstrates strong anti-tumor potential. *Anticancer Res.* 2012, 32, 81–87.
69. Hsiao, W.L.; Li, Y.Q.; Lee, T.L.; Li, N.; You, M.M.; Chang, S.T. Medicinal mushroom extracts inhibit ras-induced cell transformation and the inhibitory effect requires the presence of normal cells. *Carcinogenesis* 2004, 25, 1177–1183.
70. Zaidman, B.Z.; Yassin, M.; Mahajna, J.; Wasser, S.P. Medicinal mushroom modulators of molecular targets as cancer therapeutics. *Appl. Microbiol. Biotechnol.* 2005, 67, 453–468.

71. Zhang, X.; Li, T.; Liu, S.; Xu, Y.; Meng, M.; Li, X.; Lin, Z.; Wu, Q.; Xue, Y.; Pan, Y.; et al. beta-glucan from *Lentinus edodes* inhibits breast cancer progression via the Nur77/HIF-1 α axis. *Biosci. Rep.* 2020, 40, BSR20201006.
72. Peyrin-Biroulet, L.; Lepage, C.; Jooste, V.; Gueant, J.L.; Faivre, J.; Bouvier, A.M. Colorectal cancer in inflammatory bowel diseases: A population-based study (1976–2008). *Inflamm. Bowel Dis.* 2012, 18, 2247–2251.
73. Triantafyllidis, J.K.; Nasioulas, G.; Kosmidis, P.A. Colorectal cancer and inflammatory bowel disease: Epidemiology, risk factors, mechanisms of carcinogenesis and prevention strategies. *Anticancer Res.* 2009, 29, 2727–2737.
74. Schwartz, B.; Hadar, Y. Possible mechanisms of action of mushroom-derived glucans on inflammatory bowel disease and associated cancer. *Ann. Transl. Med.* 2014, 2, 19.
75. Okamoto, T.; Kodoi, R.; Nonaka, Y.; Fukuda, I.; Hashimoto, T.; Kanazawa, K.; Mizuno, M.; Ashida, H. Lentinan from shiitake mushroom (*Lentinus edodes*) suppresses expression of cytochrome P450 1A subfamily in the mouse liver. *Biofactors* 2004, 21, 407–409.
76. Lavi, I.; Nimri, L.; Levinson, D.; Peri, I.; Hadar, Y.; Schwartz, B. Glucans from the edible mushroom *Pleurotus pulmonarius* inhibit colitis-associated colon carcinogenesis in mice. *J. Gastroenterol.* 2012, 47, 504–518.
77. Nowacka-Jechalke, N.; Nowak, R.; Lemieszek, M.K.; Rzeski, W.; Gawlik-Dziki, U.; Szpakowska, N.; Kaczynski, Z. Promising potential of crude polysaccharides from *Sparassis crispa* against colon cancer: An in vitro study. *Nutrients* 2021, 13, 161.
78. Hsu, W.H.; Qiu, W.L.; Tsao, S.M.; Tseng, A.J.; Lu, M.K.; Hua, W.J.; Cheng, H.C.; Hsu, H.Y.; Lin, T.Y. Effects of WSG, a polysaccharide from *Ganoderma lucidum*, on suppressing cell growth and mobility of lung cancer. *Int. J. Biol. Macromol.* 2020, 165, 1604–1613.
79. Maity, G.N.; Maity, P.; Choudhuri, I.; Bhattacharyya, N.; Acharya, K.; Dalai, S.; Mondal, S. Structural studies of a water insoluble beta-glucan from *Pleurotus djamor* and its cytotoxic effect against PA1, ovarian carcinoma cells. *Carbohydr. Polym.* 2019, 222, 114990.
80. Alonso, E.N.; Ferronato, M.J.; Gandini, N.A.; Fermento, M.E.; Obiol, D.J.; Lopez Romero, A.; Arevalo, J.; Villegas, M.E.; Facchinetti, M.M.; Curino, A.C. Antitumoral effects of D-Fraction from *Grifola Frondosa* (Maitake) mushroom in breast cancer. *Nutr. Cancer* 2017, 69, 29–43.
81. Vetricka, V.; Saraswat-Ohri, S.; Vashishta, A.; Descroix, K.; Jamois, F.; Yvin, J.C.; Ferrieres, V. New 4-deoxy-(1 \rightarrow 3)-beta-D-glucan-based oligosaccharides and their immunostimulating potential. *Carbohydr. Res.* 2011, 346, 2213–2221.
82. Lin, T.Y.; Tseng, A.J.; Qiu, W.L.; Chao, C.H.; Lu, M.K. A sulfated glucan from *Antrodia cinnamomea* reduces Slug expression through regulation of TGF β /AKT/GSK3 β axis in lung cancer. *Carbohydr. Polym.* 2019, 210, 175–184.
83. Volman, J.J.; Helsper, J.P.; Wei, S.; Baars, J.J.; van Griensven, L.J.; Sonnenberg, A.S.; Mensink, R.P.; Plat, J. Effects of mushroom-derived beta-glucan-rich polysaccharide extracts on nitric oxide production by bone marrow-derived macrophages and nuclear factor-kappaB transactivation in Caco-2 reporter cells: Can effects be explained by structure? *Mol. Nutr. Food Res.* 2010, 54, 268–276.
84. Aleem, E. beta-Glucans and their applications in cancer therapy: Focus on human studies. *Anticancer Agents Med. Chem.* 2013, 13, 709–719.
85. Motta, F.; Gershwin, M.E.; Selmi, C. Mushrooms and immunity. *J. Autoimmun.* 2021, 117, 102576.
86. Steimbach, L.; Borgmann, A.V.; Gomar, G.G.; Hoffmann, L.V.; Rutckeviski, R.; de Andrade, D.P.; Smiderle, F.R. Fungal beta-glucans as adjuvants for treating cancer patients—A systematic review of clinical trials. *Clin. Nutr.* 2020, in press.
87. Chaichian, S.; Moazzami, B.; Sadoughi, F.; Haddad Kashani, H.; Zaroudi, M.; Asemi, Z. Functional activities of beta-glucans in the prevention or treatment of cervical cancer. *J. Ovarian Res.* 2020, 13, 24.
88. Rop, O.; Mlcek, J.; Jurikova, T. Beta-glucans in higher fungi and their health effects. *Nutr. Rev.* 2009, 67, 624–631.
89. Huang, W.-J. Influences of gut hormones on hepatocellular carcinoma. *Endocrinol. Metab. Syndr.* 2015, 4, 1017–1161.
90. Mo, L.; Chen, Y.; Li, W.; Guo, S.; Wang, X.; An, H.; Zhan, Y. Anti-tumor effects of (1 \rightarrow 3)-beta-D-glucan from *Saccharomyces cerevisiae* in S180 tumor-bearing mice. *Int. J. Biol. Macromol.* 2017, 95, 385–392.
91. Li, X.; Luo, H.; Ye, Y.; Chen, X.; Zou, Y.; Duan, J.; Xiang, D. beta-glucan, a dectin1 ligand, promotes macrophage M1 polarization via NF κ B/autophagy pathway. *Int. J. Oncol.* 2019, 54, 271–282.
92. Morales, D.; Rutckeviski, R.; Villalva, M.; Abreu, H.; Soler-Rivas, C.; Santoyo, S.; Iacomini, M.; Smiderle, F.R. Isolation and comparison of alpha- and beta-D-glucans from shiitake mushrooms (*Lentinula edodes*) with different biological activities. *Carbohydr. Polym.* 2020, 229, 115521.
93. Wagener, J.; Striegler, K.; Wagener, N. alpha- and beta-1,3-Glucan synthesis and remodeling. *Curr. Top. Microbiol. Immunol.* 2020, 425, 53–82.

94. Jin, Y.; Li, P.; Wang, F. beta-glucans as potential immunoadjuvants: A review on the adjuvanticity, structure-activity relationship and receptor recognition properties. *Vaccine* 2018, 36, 5235–5244.
95. Mori, K.; Naganuma, M.; Mizuno, S.; Suzuki, H.; Kitazume, M.T.; Shimamura, K.; Chiba, S.; Sugita, A.; Matsuoka, K.; Hisamatsu, T.; et al. beta-(1,3)-Glucan derived from *Candida albicans* induces inflammatory cytokines from macrophages and lamina propria mononuclear cells derived from patients with Crohn's disease. *Intest. Res.* 2018, 16, 384–392.
96. Wang, J.; Jin, Z.; Zhang, W.; Xie, X.; Song, N.; Lv, T.; Wu, D.; Cao, Y. The preventable efficacy of beta-glucan against leptospirosis. *PLoS Negl. Trop. Dis.* 2019, 13, e0007789.
97. PDQ. Medicinal Mushrooms (PDQ®): Health Professional Version. In PDQ Cancer Information Summaries; Bethesda: Rockville, MD, USA, 2020; pp. 2002–2020.
98. Murphy, E.J.; Rezoagli, E.; Major, I.; Rowan, N.J.; Laffey, J.G. beta-Glucan metabolic and immunomodulatory properties and potential for clinical application. *J. Fungi* 2020, 6, 356.
99. Chen, J.; Zhang, X.D.; Jiang, Z. The application of fungal beta-glucans for the treatment of colon cancer. *Anticancer Agents Med. Chem.* 2013, 13, 725–730.

Retrieved from <https://encyclopedia.pub/entry/history/show/23514>