Cyclosporine A-Implants in Veterinary Ophthalmology

Subjects: Veterinary Sciences

Contributor: Martyna Padjasek , Badr Qasem , Anna Cisło-Pakuluk , Krzysztof Marycz

Cyclosporine A (CsA) is a selective and reversible immunosuppressant agent that is widely used as a medication for a wide spectrum of diseases in humans such as graft versus host disease, non-infectious uveitis, rheumatoid arthritis, psoriasis, and atopic dermatitis. Furthermore, the CsA is used to treat keratoconjunctivitis sicca, chronic superficial keratitis, immune-mediated keratitis and equine recurrent uveitis in animals. The selective activity of Cyclosporine A (CsA) was demonstrated to be an immunomodulation characteristic of T-lymphocyte proliferation and inhibits cytokine gene expression. Moreover, the lipophilic characteristics with poor bioavailability and low solubility in water, besides the side effects, force the need to develop new formulations and devices that will provide adequate penetration into the anterior and posterior segments of the eye.

cyclosporine A keratoconjunctivitis sicca chronic superficial keratitis

immune-mediated keratitis

equine recurrent uveitis

delivery devices

1. Introduction

Cyclosporine A (CsA) is one of the most important transplantation drugs that was discovered and isolated by Jean Borel and co-workers in 1970, from the fungus *Tolypocladium inflatum* ^{[1][2]}. In 1976, their results demonstrated that cyclosporine has immunosuppressive characteristics ^[3], which were crucial in transplantology and immunopharmacology.

Nowadays, it is widely used as human and veterinary medicine in transplantation procedures, and some immunemediated inflammatory diseases (IMIDs)^[4]. Nevertheless, it has several side effects. For instance, a recent study demonstrated that up to 50% of patients have CsA-associated neurotoxicity in both intravenous and oral administrations which makes the CsA mechanism of action remain ambiguous ^[5].

2. CsA-Implants in Veterinary Ophthalmology

Pearson et al. formulated one of the first CsA delivery devices ^[6], based on a sustained-release ganciclovir intravitreal implant ^[7].

The device containing 5 mg of CsA was implanted intravitreally in eighteen New Zealand albino rabbits and an analogous implant containing 6 mg of CsA was used in three cynomolgus monkeys. The study aimed to determine

the toxicity of an intravitreal device that provides long-term delivery of CsA. However, the results showed no evidence of toxicity in the cynomolgus monkeys, but in the rabbits lens opacification in the vicinity of the implant was observed as well as a decrease of the b-wave amplitude in the ERG ^[6].

In addition, a similar study was established by Enyedi et al. ^[8], who investigated the intraocular device containing a combination of dexamethasone (2 mg) and CsA (100 ug) in New Zealand albino rabbits. The 2.5 mm diameter drug pellet of the implant was coated with polyvinyl alcohol (PVA) and ethylene vinyl acetate (EVA) ^{[6][7]}. The highest levels of CsA were detected in the lens compared to low levels in the sclera, cornea, iris and aqueous ^[8]. Moreover, Jaffe et al. have determined the effectiveness of the intravitreal CsA-sustained delivery device, that proposed by Pearson et al., in the treatment of experimental uveitis in rabbits. The inflammation in the treated eye was considerably less than in the control eye. The therapeutic level of CsA in the vitreous was detected 6 months after implantation ^[9].

2.1. Keratoconjunctivitis Sicca (KCS)

Keratoconjunctivitis sicca is an inflammatory disease that affects the gland of the third eyelid and lacrimal glands, causing a decrease of tear film, quantitative or qualitative disorder, that could be the result of a congenital, metabolic, drug-induced, neurogenic or immune-mediated defect ^[10]. The most common signs of KCS are mucopurulent ocular discharge, conjunctivitis and keratitis which can lead to corneal ulcers; some patients develop blepharitis and, in chronic cases, corneal pigmentation and scarring occur ^{[11][12]}. An indispensable element of ophthalmic examination is the Schirmer test (STT-1) which in normal dogs shows tear production around 15 to 25 mm/min; STT-1 in the course of KCS ranges from 9–14 mm/min as mild, >4 to 8 mm/min as moderate and <4 mm/min as a severe stage ^[13].

The histopathology and serologic results suggested that most of the cases may be immune-mediated (more than 30% of dogs with KCS); the confirmation of which is a positive reaction to immunosuppressive drugs ^{[10][11][14][15]}.

Kim et al. formulated three similar silicone-based matrix CsA implants (with 20–30% *wt/wt*), which were used in several subsequent studies. For instance, the study aimed to provide drug delivery devices that were effective in treating lacrimal gland GVHD (graft versus host disease after transplantation of allogeneic stem cells) and assessed the rate of CsA release (in vitro), implant toxicity, pharmacokinetics and pharmacodynamics in normal rabbits, dogs and dogs with KCS. The results after six months revealed the safety of the delivery device; no ocular toxicity and abnormalities in blood examination were observed. The implant provided therapeutic levels of CsA in the lacrimal gland, conjunctiva and cornea; dogs (with clinical signs of KCS and Schirmer test below 5 mm/min) after implantation did not need further local treatment and the Schirmer test results were above 10 mm/min during the study period ^[16].

Acton et al. reported a case of keratoconjunctivitis sicca in a red wolf (*Canis rufus*) ^[17]. After a positive response to topical 2% cyclosporine (initial Schirmer test result at 0 mm/min and after two weeks of combination therapy with triple antibiotic with dexamethasone, the tear production levels were 15 mm/min in the left eye and 16 mm/min in

the right eye) they performed implantation of episcleral sustained-release CsA devices (10% matrix CsA-silicone). After two weeks of implantation the tear production remained at the physiological level above 13 mm/min twelve months after surgery ^[17].

Penetrating keratoplasty (PKP) is a common allograft performed in humans. However, there is one major drawback of this procedure—its high rejection rate (65%). To overcome this obstacle Lee et al. have proposed the use of episcleral CsA implant ^[18] described previously: CsA powder mixed with silicone; *wt/wt* 30% ^[15]. Two implants with different total release were used to determine short (implant B—7.7 mg CsA per implant) and long-term (implant A —12 mg CsA per implant) pharmacokinetics in rabbits and dogs, therefore, the cumulative release observed over the 400-days was approximately 3.8 mg (implant A) and 2.3 mg (implant B). This study showed effective penetration into the cornea and no signs of ocular toxicity. Moreover, CsA concentrations in the cornea were approximately 0.1 µg/mg three hours after implantation and ensured the suppression of T-cell and vascular endothelial cells for over a year. Pharmacokinetics evaluation of CsA in the rabbit model was detected in buccal lymph nodes at 1 h, which suggests that lymphatic vessels in conjunctiva support the rapid dissolution of the drug to the cornea and surrounding tissue ^[18].

Numerous studies have shown the effectiveness and safety of a cyclosporin episcleral implant with a silicone matrix, an implant of 1.9 cm \times 2 mm \times 1 mm, containing 12 mg of CsA and ensuring its release at an average level of 17 µg/day for at least 6 months is particularly useful in anterior segment disease ^{[16][19]}.

Choi et al. proposed hydrogel contact lenses (CLs) loaded with CsA and determined its efficiency in the rabbit model of dry eye ^[20]. Previous studies using drug-soaked lenses showed low efficiency in sustained release of the drugs ^{[21][22][23][24][25]}. Therefore, they used a supercritical fluid (SCF) technique to modify and control the degree and the rate of releasing CsA. The in vivo study showed that adequate concentration of CsA was maintained for over 48 h in the cornea, conjunctiva, and crystalline lens. In comparison with control groups, the CsA-CL group exhibited higher density of the goblet cell, tear volume, lower staining score, and reduction of the inflammatory process through immunomodulatory effects.

Several clinical trials presented new methods for extended-release drug delivery. For Sight Vision, owned by Allergan, proposed a peri-conjunctival ring currently used for delivery bimatoprost in glaucoma patients ^[26]. Work is well under way to deliver a CsA ring based on the same technology. In cooperation with NC State University, they conducted clinical trials on dogs with KCS, in which a conjunctival ring releasing CsA was well tolerated and as long as it rested on the conjunctiva under the upper and lower eyelids, the results were satisfying. The therapeutic effect lasted about a month with 75% retention in the eye (unpublished data, ESVO Webinar, 21 February 2021).

Ocular Therapeutix[™] is working on a group of drug-eluting intracanalicular drug inserts. A study funded by Ocular Therapeutix[™] and conducted by Vanslette et al. evaluated pharmacokinetics of Cyclosporine Intracanalicular Insert (OTX-CSI) in Beagle dogs with surgically induced Dry Eye. Intracanalicular devices combines two treatments of dry eye disease: sustained release delivery of cyclosporine and punctal occlusion which aids tear conservation. OTX-SCI contains 0.36 mg CsA in fully biodegradable polyethylene glycol hydrogel. It was designed to provide

effective therapy for 12 weeks. The study showed successfully released CsA and its higher concentration in tear fluid in dogs with dry eye was probably due to less dilution on the ocular surface. OTX-CSI was well tolerated and assured immunomodulatory levels in tear fluid ^[27]. Although OTX-CSI is a promising device for the treatment of immunological diseases in human and veterinary ophthalmology, Ocular Therapeutix[™] published results of a Phase 2 clinical trial in which OTX-SCI did not meet the primary endpoint of increased tear production at 12 weeks; therefore, more research is required.

2.2. Chronic Superficial Keratitis (CSK)

Chronic superficial keratitis, also known as Pannus, is another immune-mediated disease that affects dogs, with chronic corneal lesions characteristic, mostly in the lateral quadrant: vascularization, progressive pigmentation, and sometimes white opacity ^{[27][28][29]}. The etiology is still not fully understood but CD4+ T lymphocyte infiltration from cornea stroma suggests an immunological background ^{[30][31][32][33]}. The German shepherd dog is a predisposed breed, but it can also occur in the Australian shepherd, collie, border collie, golden retriever, Akita, vizsla, and others ^{[28][34]}. Several studies have found that excessive ultraviolet exposure increases the risk of CSK ^{[28][35][36]}.

Topical immunomodulators such as steroids and calcineurin inhibitors are the standard therapy ^[37]. Dogs that are responsive to topical CsA, and for whom therapy must be continuous, might be good candidates for an episcleral cyclosporine implant proposed for treating keratoconjunctivitis sicca. However, further studies are required ^{[16][18]}.

2.3. Immune-Mediated Keratitis (IMMK)

Immune-mediated keratitis is nonulcerative, primary keratitis (NUK) that occurs in horses ^[38]. Although the etiopathology has not been thoroughly investigated, the absence of microorganisms and significant improvement after implementation of immunosuppressive therapy suggest an immunological background. The common symptoms of IMMK are nonulcerative corneal opacity, corneal edema and neovascularization, cellular infiltration, and no features of uveitis. Horses with IMMK experience no or mild discomfort ^[39]. IMMK is classified into four types based on its location in the cornea: epithelial, superficial stromal, middle stromal, and endothelial, with the superficial stroma being the most frequent site of occurrence ^[38]. CSA topical application is most effective in the case of epithelial and superficial IMMK, but efficiency decreases along with the posterior layers of the cornea ^[39]. Thus, another route of CsA distribution to all layers of the cornea is being investigated.

Gilger et al. proposed the use of a silicone matrix CsA episcleral implant in nineteen horses with different types of IMMK ^[40]. More than two devices, described previously ^{[16][18]}, were implanted per eye in the dorso temporal episcleral space.

The study demonstrated good tolerance of the implants with no significant deviations between the number of devices implanted. Superficial and endothelial immune-mediated keratitis were considered controlled in all treated eyes, although in three cases of endothelial IMMK topical bromfenac was also administered. The worst response was observed in the case of midstromal IMMK. Implants were unable to control inflammation. In vivo, the therapeutic effect of CsA in the case of superficial IMMK was determined for 12–18 months ^[40].

2.4. Equine Recurrent Uveitis (ERU)

Equine recurrent uveitis (moon blindness, periodic ophthalmia, iridocyclitis) is a condition in which immunemediated active episodes of panuveitis reoccur every few weeks to months ^[41]. ERU is still one of the most common causes of horse blindness. Around 30% of horses who presented for examination due to ERU symptoms were unilaterally or bilaterally blind ^{[42][43]}. Horses experience spontaneous relapses similar to humans ^{[44][45][46]}. Initial causes of recurrent uveitis are not always known, but genetic predisposition or microbes such as *Leptospira* sp. might be involved ^{[46][47]}. According to recent research, the retinal expression of neuraminidase 1 (NEU1) plays an important role in ERU. Furthermore, horses with recurrent uveitis had higher levels of NEU1 in Müller glial cells in the retina. Therefore, NEU1 might be a new marker of activated Müller glial cells in uveitis ^[48].

Clinical signs associated with ERU can include anterior segment: blepharospasm, increased lacrimation, photophobia, miosis, edema and vascularization, aqueous flare, cellular infiltration, hypopyon and hyphema, low IOP; posterior segment: vitreous, chorioretinitis, and retinal degeneration ^{[46][49]}.

Gilger et al. used an intravitreal cyclosporine delivery device, previously described by Pearson et al., 1996 ^[6], Enyedi et al., 1996 ^[8], and Jaffe et al., 1998 ^[9], in horses with experimental uveitis ^[50]. The study found that the CsA intravitreal implant reduced the severity and duration of symptoms (but the inflammatory suppression was incomplete), cellular infiltrate was less intense compared to the control eye (PVA/EVA devices without CsA), and the CsA-delivery device was well tolerated. Moreover, the concentration of cyclosporine in the vitreous humor was below therapeutic levels. Nevertheless, tissue levels were not measured ^[50]. Additionally, the long-term study shows that intravitreal sustained-release CsA delivery devices are safe for at least 12 months ^[51].

A similar implant was evaluated in horses with ERU that occurred naturally. Devices releasing 4 μ g of CsA per day (in a previous study 2 μ q/d) were implanted in the eyes of sixteen horses with unilateral uveitis and history of disease recurrence. Follow-up was performed between 6 and 24 months after implantation. After surgery, less than 20% of horses developed uveitis, but as reported by owners, the symptoms were less severe and responded better to anti-inflammatory medication. Complications were noted in four patients, including vision loss due to cataracts or complete retinal detachment as well as glaucoma ^[52].

A different study by Gilger et al. evaluated episcleral and deep scleral bioerodible cyclosporine implants ^[52]. Intravitreal delivery devices showed some good results but also revealed complications after implantation, such as cataracts caused by lens injury, endophthalmitis or increased risk of retinal detachment ^{[6][9][50][51][52][53]}. Thus, the use of implants that do not require entry into the eye has been proposed ^{[16][54][55][56][57][58]}.

Gilger et al. conducted an in vitro study of transscleral diffusion of CsA from a biodegradable matrix-reservoir CsA implant, formulated by Robinson et al. from the National Eye Institute, that suggested the release duration of CsA around 38 months and poor penetration through the sclera. This study aimed to determine the pharmacokinetics and safety of episcleral as well as deep scleral lamellar CsA devices in horses. However, episcleral implantation of the device did not reduce the frequency of relapses due to limited penetration through the sclera.

CsA concentration in retina-choroid and vitreous was below the required minimum to treat inflammation ^[6]. In addition, the deep sclera CsA device was well tolerated, and no toxicity was observed. Therapeutic drug concentration was observed in vitreous and sclera, choroid-retina and optic nerve tissue, although there was no detection of CsA in the aqueous humor, cornea, and samples of peripheral blood. Follow-ups were performed on average after 14 months and a reduction in flare-ups was noted.

Blindness occurred in 15% of the eyes as a result of glaucoma, uncontrolled uveitis, cataract, fungal keratitis, and retinal detachment. At the end of the study period, 68/80 of the eyes had vision after surgery ^[59]. A long-term study on 133 horses (151 eyes) confirmed the promising results from the previous survey but also noted complications such as glaucoma, persistent uveitis, cataracts, and retinal detachment ^{[42][60][61][62]}.

References

- 1. Colombo, D.; Ammirati, E. Cyclosporine in transplantation–A history of converging timelines. J. Biol. Regul. Homeost. Agents 2011, 25, 493–504.
- 2. Borel, J.F. History of the discovery of cyclosporin and of its early pharmacological development. Wien. Klin. Wochenschr. 2002, 114, 433–437.
- 3. Borel, J.F. Comparative study of in vitro and in vivo drug effects on cell-mediated cytotoxicity. Immunology 1976, 31, 631–641.
- Faulds, D.; Goa, K.L.; Benfield, P. Cyclosporin. A review of its pharmacodynamic and pharmacokinetic properties, and therapeutic use in immunoregulatory disorders. Drugs 1993, 45, 953–1040.
- Teimouri, A.; Ahmadi, S.R.; Anavri Ardakani, S.; Foroughian, M. Cyclosporine-A-Based Immunosuppressive Therapy-Induced Neurotoxicity: A Case Report. Open Access Emerg. Med. 2020, 12, 93–97.
- Pearson, P.A.; Jaffe, G.J.; Martin, D.F.; Cordahi, G.J.; Grossniklaus, H.; Schmeisser, E.T.; Ashton,
 P. Evaluation of a delivery system providing long-term release of cyclosporine. Arch. Ophthalmol. 1996, 114, 311–317.
- Smith, T.J.; Pearson, P.A.; Blandford, D.L.; Brown, J.D.; Goins, K.A.; Hollins, J.L.; Schmeisser, E.T.; Glavinos, P.; Baldwin, L.B.; Ashton, P. Intravitreal sustained-release ganciclovir. Arch. Ophthalmol. 1992, 110, 255–258.
- 8. Enyedi, L.B.; Pearson, P.A.; Ashton, P.; Jaffe, G.J. An intravitreal device providing sustained release of cyclosporine and dexamethasone. Curr. Eye Res. 1996, 15, 549–557.
- 9. Jaffe, G.J.; Yang, C.S.; Wang, X.C.; Cousins, S.W.; Gallemore, R.P.; Ashton, P. Intravitreal sustained-release cyclosporine in the treatment of experimental uveitis. Ophthalmology 1998,

105, 46–56.

- Izci, C.; Celik, I.; Alkan, F.; Ogurtan, Z.; Ceylan, C.; Sur, E.; Ozkan, Y. Histologic characteristics and local cellular immunity of the gland of the third eyelid after topical ophthalmic administration of 2% cyclosporine for treatment of dogs with keratoconjunctivitis sicca. Am. J. Vet. Res. 2002, 63, 688–694.
- Kaswan, R.L.; Salisbury, M.A.; Ward, D.A. Spontaneous canine keratoconjunctivitis sicca. A useful model for human keratoconjunctivitis sicca: Treatment with cyclosporine eye drops. Arch. Ophthalmol. 1989, 107, 1210–1216.
- 12. Kaswan, R.L.; Salisbury, M.A. A new perspective on canine keratoconjunctivitis sicca. Treatment with ophthalmic cyclosporine. Vet. Clin. N. Am. Small Anim. Pract. 1990, 20, 583–613.
- Bittencourt, M.K.W.; Barros, M.A.; Martins, J.F.P.; Vasconcellos, J.P.C.; Morais, B.P.; Pompeia, C.; Bittencourt, M.D.; Evangelho, K.D.S.; Kerkis, I.; Wenceslau, C.V. Allogeneic mesenchymal stem cell transplantation in dogs with keratoconjunctivitis sicca. Cell Med. 2016, 8, 63–77.
- 14. Dodi, P.L. Immune-mediated keratoconjunctivitis sicca in dogs: Current perspectives on management. Vet. Med. Res. Rep. 2015, 6, 341–347.
- Williams, D.L.; Tighe, A.A. Immunohistochemical evaluation of lymphocyte populations in the nictitans glands of normal dogs and dogs with keratoconjunctivitis sicca. Open Vet. J. 2018, 8, 47–52.
- Kim, H.; Csaky, K.G.; Gilger, B.C.; Dunn, J.P.; Lee, S.S.; Tremblay, M.; de Monasterio, F.; Tansey, G.; Yuan, P.; Bungay, P.M.; et al. Preclinical evaluation of a novel episcleral cyclosporine implant for ocular graft-versus-host disease. Investig. Ophthalmol. Vis. Sci. 2005, 46, 655–662.
- Acton, A.E.; Beale, A.B.; Gilger, B.C.; Stoskopf, M.K. Sustained release cyclosporine therapy for bilateral keratoconjunctivitis sicca in a red wolf (Canis rufus). J. Zoo Wildl. Med. 2006, 37, 562– 564.
- Lee, S.S.; Kim, H.; Wang, N.S.; Bungay, P.M.; Gilger, B.C.; Yuan, P.; Kim, J.; Csaky, K.G.; Robinson, M.R. A pharmacokinetic and safety evaluation of an episcleral cyclosporine implant for potential use in high-risk keratoplasty rejection. Investig. Ophthalmol. Vis. Sci. 2007, 48, 2023– 2029.
- Barachetti, L.; Rampazzo, A.; Mortellaro, C.M.; Scevola, S.; Gilger, B.C. Use of episcleral cyclosporine implants in dogs with keratoconjunctivitis sicca: Pilot study. Vet. Ophthalmol. 2015, 18, 234–241.
- Choi, J.H.; Li, Y.; Jin, R.; Shrestha, T.; Choi, J.S.; Lee, W.J.; Moon, M.J.; Ju, H.T.; Choi, W.; Yoon, K.C. The Efficiency of Cyclosporine A-Eluting Contact Lenses for the Treatment of Dry Eye. Curr. Eye Res. 2019, 44, 486–496.

- 21. Soluri, A.; Hui, A.; Jones, L. Delivery of ketotifen fumarate by commercial contact lens materials. Optom. Vis. Sci. 2012, 89, 1140–1149.
- 22. Guzman-Aranguez, A.; Colligris, B.; Pintor, J. Contact lenses: Promising devices for ocular drug delivery. J. Ocul. Pharmacol. Ther. 2013, 29, 189–199.
- Maulvi, F.A.; Shaikh, A.A.; Lakdawala, D.H.; Desai, A.R.; Pandya, M.M.; Singhania, S.S.; Vaidya, R.J.; Ranch, K.M.; Vyas, B.A.; Shah, D.O. Design and optimization of a novel implantation technology in contact lenses for the treatment of dry eye syndrome: In vitro and in vivo evaluation. Acta Biomater. 2017, 53, 211–221.
- Chandasana, H.; Prasad, Y.D.; Chhonker, Y.S.; Chaitanya, T.K.; Mishra, N.N.; Mitra, K.; Shukla, P.K.; Bhatta, R.S. Corneal targeted nanoparticles for sustained natamycin delivery and their PK/PD indices: An approach to reduce dose and dosing frequency. Int. J. Pharm. 2014, 477, 317– 325.
- 25. Jung, H.J.; Abou-Jaoude, M.; Carbia, B.E.; Plummer, C.; Chauhan, A. Glaucoma therapy by extended release of timolol from nanoparticle loaded silicone-hydrogel contact lenses. J. Control. Release 2013, 165, 82–89.
- 26. Shirley, M. Bimatoprost implant: First approval. Drugs Aging 2020, 37, 457–462.
- Vanslette, A.; Blizzard, C.D.; Haberman, P.; Tomaszewski, J.; Rosales, C.; Metzinger, J.L.; Goldstein, M.H.; Driscoll, A. Pharmacokinetics of OTX-CSI, a Sustained Release Cyclosporine Intracanalicular Insert in Beagles. Investig. Ophthalmol. Vis. Sci. 2019, 60, 285.
- Slatter, D.H.; Lavach, J.D.; Severin, G.A.; Young, S. Uberreiter's syndrome (chronic superficial keratitis) in dogs in the Rocky Mountain area—A study of 463 cases. J. Small Anim. Pract. 1977, 18, 757–772.
- 29. Andrew, S.E. Immune-mediated canine and feline keratitis. Vet. Clin. N. Am. Small Anim. Pract. 2008, 38, 269–290.
- 30. Bedford, P.G.; Longstaffe, J.A. Corneal pannus (chronic superficial keratitis) in the German shepherd dog. J. Small Anim. Pract. 1979, 20, 41–56.
- Eichenbaum, J.D.; Lavach, J.D.; Gould, D.H.; Severin, G.A.; Paulsen, M.E.; Jones, R.L. Immunohistochemical staining patterns of canine eyes affected with chronic superficial keratitis. Am. J. Vet. Res. 1986, 47, 1952–1955.
- 32. Williams, D.L. Histological and immunohistochemical evaluation of canine chronic superficial keratitis. Res. Vet. Sci. 1999, 67, 191–195.
- 33. Williams, D.L.; Cribb, A.; Scott, J.E. Proteoglycan-collagen interactions in chronic superficial keratitis in the dog. Biochem. Soc. Trans. 1991, 19, 353S.

- 34. Drahovska, Z.; Balicki, I.; Trbolova, A.; Mihalova, M.; Holickova, M. A retrospective study of the occurrence of chronic superficial keratitis in 308 German Shepherd dogs: 1999–2010. Pol. J. Vet. Sci. 2014, 17, 543–546.
- Chavkin, M.J.; Roberts, S.M.; Salman, M.D.; Severin, G.A.; Scholten, N.J. Risk factors for development of chronic superficial keratitis in dogs. J. Am. Vet. Med. Assoc. 1994, 204, 1630– 1634.
- 36. Denk, N.; Fritsche, J.; Reese, S. The effect of UV-blocking contact lenses as a therapy for canine chronic superficial keratitis. Vet. Ophthalmol. 2011, 14, 186–194.
- 37. Williams, D.L.; Hoey, A.J.; Smitherman, P. Comparison of topical cyclosporin and dexamethasone for the treatment of chronic superficial keratitis in dogs. Vet. Rec. 1995, 137, 635–639.
- 38. Gilger, B.C.; Michau, T.M.; Salmon, J.H. Immune-mediated keratitis in horses: 19 cases (1998–2004). Vet. Ophthalmol. 2005, 8, 233–239.
- 39. Matthews, A.; Gilger, B.C. Equine immune-mediated keratopathies. Vet. Ophthalmol. 2009, 12 (Suppl. 1), 10–16.
- 40. Gilger, B.C.; Stoppini, R.; Wilkie, D.A.; Clode, A.B.; Pinto, N.H.; Hempstead, J.; Gerding, J.; Salmon, J.H. Treatment of immune-mediated keratitis in horses with episcleral silicone matrix cyclosporine delivery devices. Vet. Ophthalmol. 2014, 17 (Suppl. 1), 23–30.
- 41. Gilger, B.C.; Michau, T.M. Equine recurrent uveitis: New methods of management. Vet. Clin. N. Am. Equine Pract. 2004, 20, 417–427.
- 42. Sandmeyer, L.S.; Bauer, B.S.; Feng, C.X.; Grahn, B.H. Equine recurrent uveitis in western Canadian prairie provinces: A retrospective study (2002–2015). Can. Vet. J. 2017, 58, 717–722.
- 43. Gerding, J.C.; Gilger, B.C. Prognosis and impact of equine recurrent uveitis. Equine Vet. J. 2016, 48, 290–298.
- 44. Kalsow, C.M.; Dwyer, A.E.; Smith, A.W.; Nifong, T.P. Pinealitis accompanying equine recurrent uveitis. Br. J. Ophthalmol. 1993, 77, 46–48.
- 45. Deeg, C.A.; Ehrenhofer, M.; Thurau, S.R.; Reese, S.; Wildner, G.; Kaspers, B. Immunopathology of recurrent uveitis in spontaneously diseased horses. Exp. Eye Res. 2002, 75, 127–133.
- 46. Malalana, F.; Stylianides, A.; McGowan, C. Equine recurrent uveitis: Human and equine perspectives. Vet. J. 2015, 206, 22–29.
- 47. Allbaugh, R.A. Equine recurrent uveitis: A review of clinical assessment and management. Equine Vet. Educ. 2017, 29, 279–288.
- 48. Lorenz, L.; Amann, B.; Hirmer, S.; Degroote, R.L.; Hauck, S.M.; Deeg, C.A. NEU1 is more abundant in uveitic retina with concomitant desialylation of retinal cells. Glycobiology 2021, 31,

873–883.

- 49. McMullen, R.J.; Fischer, B.M. Medical and surgical management of equine recurrent uveitis. Vet. Clin. N. Am. Equine Pract. 2017, 33, 465–481.
- 50. Gilger, B.C.; Malok, E.; Stewart, T.; Horohov, D.; Ashton, P.; Smith, T.; Jaffe, G.J.; Allen, J.B. Effect of an intravitreal cyclosporine implant on experimental uveitis in horses. Vet. Immunol. Immunopathol. 2000, 76, 239–255.
- 51. Gilger, B.C.; Malok, E.; Stewart, T.; Ashton, P.; Smith, T.; Jaffe, G.J.; Allen, J.B. Long-term effect on the equine eye of an intravitreal device used for sustained release of cyclosporine A. Vet. Ophthalmol. 2000, 3, 105–110.
- 52. Gilger, B.C.; Wilkie, D.A.; Davidson, M.G.; Allen, J.B. Use of an intravitreal sustained-release cyclosporine delivery device for treatment of equine recurrent uveitis. Am. J. Vet. Res. 2001, 62, 1892–1896.
- 53. Shane, T.S.; Martin, D.F.; Endopthalmitis-Gancioclovir Implant Study Group. Endophthalmitis after ganciclovir implant in patients with AIDS and cytomegalovirus retinitis. Am. J. Ophthalmol. 2003, 136, 649–654.
- Sakurai, E.; Nozaki, M.; Okabe, K.; Kunou, N.; Kimura, H.; Ogura, Y. Scleral plug of biodegradable polymers containing tacrolimus (FK506) for experimental uveitis. Investig. Ophthalmol. Vis. Sci. 2003, 44, 4845–4852.
- 55. Miyamoto, H.; Ogura, Y.; Hashizoe, M.; Kunou, N.; Honda, Y.; Ikada, Y. Biodegradable scleral implant for intravitreal controlled release of fluconazole. Curr. Eye Res. 1997, 16, 930–935.
- Okabe, K.; Kimura, H.; Okabe, J.; Kato, A.; Kunou, N.; Ogura, Y. Intraocular tissue distribution of betamethasone after intrascleral administration using a non-biodegradable sustained drug delivery device. Investig. Ophthalmol. Vis. Sci. 2003, 44, 2702–2707.
- 57. Okabe, J.; Kimura, H.; Kunou, N.; Okabe, K.; Kato, A.; Ogura, Y. Biodegradable intrascleral implant for sustained intraocular delivery of betamethasone phosphate. Investig. Ophthalmol. Vis. Sci. 2003, 44, 740–744.
- 58. Kato, A.; Kimura, H.; Okabe, K.; Okabe, J.; Kunou, N.; Ogura, Y. Feasibility of drug delivery to the posterior pole of the rabbit eye with an episcleral implant. Investig. Ophthalmol. Vis. Sci. 2004, 45, 238–244.
- Gilger, B.C.; Salmon, J.H.; Wilkie, D.A.; Cruysberg, L.P.J.; Kim, J.; Hayat, M.; Kim, H.; Kim, S.; Yuan, P.; Lee, S.S.; et al. A novel bioerodible deep scleral lamellar cyclosporine implant for uveitis. Investig. Ophthalmol. Vis. Sci. 2006, 47, 2596–2605.
- 60. Cislo-Pakuluk, A.; Smieszek, A.; Kucharczyk, N.; Bedford, P.G.C.; Marycz, K. Intra-Vitreal Administration of Microvesicles Derived from Human Adipose-Derived Multipotent Stromal Cells

Improves Retinal Functionality in Dogs with Retinal Degeneration. J. Clin. Med. 2019, 8, 510.

- Cislo-Pakuluk, A.; Marycz, K. A Promising Tool in Retina Regeneration: Current Perspectives and Challenges When Using Mesenchymal Progenitor Stem Cells in Veterinary and Human Ophthalmological Applications. Stem Cell Rev. Rep. 2017, 13, 598–602.
- 62. Kulig, D.; Zimoch-Korzycka, A.; Jarmoluk, A.; Marycz, K. Study on Alginate–Chitosan Complex Formed with Different Polymers Ratio. Polymers 2016, 8, 167.

Retrieved from https://encyclopedia.pub/entry/history/show/77973