# Applications of Natural Polymers-Based Materials

Subjects: Materials Science, Biomaterials | Materials Science, Composites | Materials Science, Coatings & Films Contributor: Carmen Freire

Natural polymers have emerged as promising candidates for the sustainable development of materials in areas ranging from food packaging and biomedicine to energy storage and electronics.

natural polymers polysaccharides proteins green chemistry

## 1. Introduction

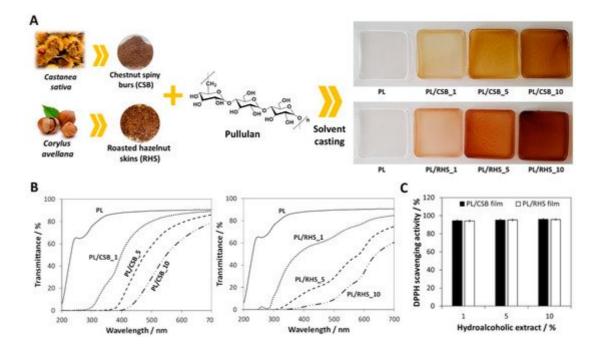
Natural polymers, such as polysaccharides, proteins, and nucleic acids, are components of biological systems responsible for performing a wide range of essential functions <sup>[1]</sup>. For instance, certain natural polymers play a key role in the maintenance of the structural integrity of cells in plants and animals (e.g., cellulose <sup>[2]</sup> and chitin <sup>[3]</sup>), while others offer biological protection against surrounding environments (e.g., lysozyme <sup>[4]</sup>). The diversity in terms of provenance and composition provides these natural polymers with distinct physicochemical and biological properties that can be of interest in various fields <sup>[5][6]</sup>. In fact, natural polymers and their derivatives already find application in numerous sectors, e.g., in the manufacture of paper goods <sup>[2]</sup> and textiles <sup>[8]</sup>, as additives in food products <sup>[9][10][11]</sup>, in the formulation of nutraceuticals and functional foods <sup>[12]</sup>, and in the biomedical field (e.g., in cosmetic treatments <sup>[13]</sup> and drug delivery <sup>[14][15]</sup>). Owing to the natural abundance, renewability, and intrinsically negative carbon footprint of polymers derived from renewable resources <sup>[16]</sup>, their exploitation is favourable and can play a pivotal role in the development of advanced materials in the shape of films <sup>[17][18]</sup>, membranes <sup>[19][20]</sup>, coatings <sup>[21][22]</sup>, hydrogels <sup>[23]</sup>, and micro- and nanoparticle systems <sup>[24][25][26]</sup>.

### 2. Applications of Natural Polymers-Based Materials

#### 2.1. Natural Polymers-Based Films for Active Food Packaging

Biopolymeric films for active food packaging are mostly based on nanocelluloses (bacterial nanocellulose (BNC) <sup>[27]</sup> and cellulose nanofibrils (CNFs) <sup>[28][29]</sup>) or other film-forming polymeric matrices (e.g., pullulan <sup>[30][31][32]</sup>, chitosan <sup>[33]</sup> and agar <sup>[34]</sup>), and are typically loaded with additives with a vast array of functions <sup>[35][36]</sup>, including active components with innate antimicrobial and antioxidant properties, such as silver nanoparticles <sup>[32]</sup>, bioactive ionic liquids <sup>[30]</sup>, lysozyme nanofibrils (LNFs) <sup>[37][38]</sup>, and natural extracts <sup>[31][34][28][29][33]</sup>. As an illustrative example, Esposito et al. <sup>[31]</sup> exploited hydroalcoholic extracts from chestnut spiny burs and roasted hazelnut skins, obtained from industrial processes, to prepare homogeneous pullulan (PL) films (**Figure 1**A). The films revealed good mechanical and UV-barrier properties (**Figure 1**B), allied with high antioxidant activity (ca. 94%, DPPH scavenging

activity, **Figure 1**C), underlining their potential for active food packaging applications. Regarding the topic of active packaging, it is also worth mentioning the recent trends in the design of intelligent packaging options <sup>[35][36]</sup>, which can provide dynamic feedback concerning the condition of packaged food, e.g., monitorization of food humidity levels <sup>[27]</sup>.

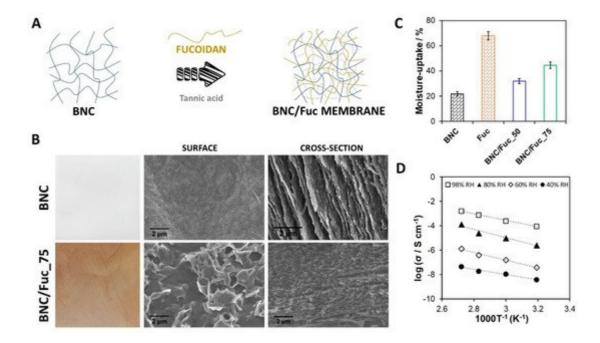


**Figure 1.** (**A**) Schematic illustration of the incorporation of hydroalcoholic extracts from chestnut spiny burs (CSB) and roasted hazelnut skins (RHS) in a pullulan (PL) matrix via a solvent casting technique and photographs of the prepared films. UV–vis spectra (**B**) and antioxidant activity (**C**) of the PL, PL/CSB, and PL/RHS-based films. Reproduced with permission from <sup>[31]</sup>. Copyright Elsevier, 2020.

#### 2.2. Natural Polymers-Based Ion Exchange Membranes for Fuel Cells

Nanoscale forms of cellulose have garnered great attention for the design of polymer electrolyte fuel cell (PEFC) components, as recently reviewed by Vilela et al. <sup>[39]</sup>. In particular, BNC-based materials have been studied for the development of sustainable substitutes of ion-exchange membranes. The lack of intrinsic ionic conductivity of BNC can be surpassed with the introduction of ion-conducting phases in the cellulose nanofibrillar structure <sup>[39]</sup>. In this sense, BNC has been combined with synthetic polyelectrolytes, such as Nafion<sup>TM</sup> <sup>[40]</sup>, poly(4-styrene sulfonic acid) <sup>[41][42][43]</sup>, phosphate bearing monomers <sup>[44][45]</sup>, and poly(ionic liquids) <sup>[46]</sup>, to create partially biobased separators. Particularly relevant is the partnership between BNC and natural polyelectrolytes <sup>[47][48]</sup> for the design of mechanically and thermally robust fully biobased polymer electrolyte membranes (PEMs). Fucoidan and BNC membranes <sup>[47]]</sup> are examples of these materials, prepared via the simple diffusion of aqueous fucoidan solutions into the exopolysaccharide network and subsequent thermal cross-linking with a natural agent, viz. tannic acid (**Figure 2**A). Micrographs of the surface and cross-section of the membranes disclosed the well-dispersed sulfate moieties of the algal polysaccharide in the BNC network (**Figure 2**B). The nanocomposite revealed good dynamic mechanical performance (storage modulus  $\geq$  460 MPa) and thermal-oxidative stability (180–200 °C) in both inert and oxidative atmospheres. Moreover, this fully biobased ion-exchange membrane displayed good moisture-

uptake capacity (**Figure 2**C) and protonic conductivity, with a maximum of 1.6 mS cm<sup>-1</sup> measured at 94 °C and 98% relative humidity conditions (**Figure 2**D). In general, fully biobased membranes display lower ionic conductivity (when compared with partially biobased counterparts) associated with a smaller content of moieties that enable ion motion. Nevertheless, the fabrication methodologies are simpler, and the resulting materials are entirely environmentally friendly.



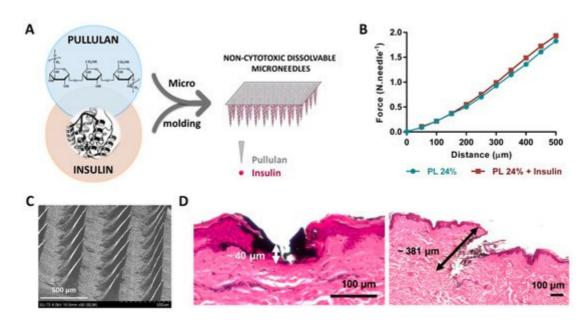
**Figure 2.** (**A**) Schematic illustration of the preparation of fully biobased bacterial nanocellulose (BNC) and fucoidan (Fuc) membranes with tannic acid as a crosslinker. (**B**) Photographs and micrographs of the nanocellulose surface and cross-section before (BNC) and after (BNC/Fuc\_75) inclusion of the sulfated polysaccharide (Fuc) (scale bar: 2 μm). (**C**) Moisture uptake capacity of the membranes. (**D**) Arrhenius-type plot of the through-plane protonic conductivity of the BNC/Fuc\_75 membrane. Reproduced with permission from <sup>[47]</sup>. Copyright Elsevier, 2020.

#### 2.3. Natural Polymers-Based Patches for Drug Delivery and Wound Healing

Works of BNC <sup>[49]</sup> and CNFs-based patches <sup>[50]</sup> have been described as alternatives to non-biodegradable synthetic materials for cutaneous wound healing applications. Aside from the physical protection granted by these systems, the incorporation of antimicrobial agents (e.g., LNFs <sup>[50]</sup>) or active pharmaceutical ingredients that stimulate cell proliferation (e.g., dexpanthenol <sup>[49]</sup>) within the patches prevents skin infections and promotes wound closure. In terms of drug-delivery options, nanocellulose patches offer the possibility of self-administering therapeutic molecules using oral, buccal, sublingual, and transdermal routes, which can significantly improve drug bioavailability and minimize side effects by avoiding the first-pass effect on metabolically active tissues of the body <sup>[51]</sup>.

#### 2.4. Natural Polymers-Based Microneedles for Drug Delivery and Fluid Uptake

Microneedles (MNs) have risen as an efficient, minimally invasive, and painless method for transdermal delivery of an assortment of drugs and bioactive compounds <sup>[52]</sup>. In this field, we report the fabrication of pullulan MNs for the transdermal delivery of insulin <sup>[53]</sup> (**Figure 3**A) and hyaluronic acid MNs with a BNC backing layer for the delivery of rutin <sup>[54]</sup>, using a micromolding technique. In both cases, the casting of the polymeric solutions resulted in needles with good mechanical performance (**Figure 3**B), able to meet the force threshold required for skin insertion ( $\approx$ 0.15 N.needle<sup>-1</sup>). Moreover, penetration studies using both a skin model and human skin (in vitro) proved the ability of the microneedles to successfully perforate the outmost layer of the skin, i.e., the *stratum corneum*, and create pathways across the skin without reaching the nerves (**Figure 3**C,D).



**Figure 3.** (**A**) Schematic illustration of the production of insulin-loaded pullulan microneedles. (**B**) Force– displacement curves of neat (PL 24%) and drug-loaded (PL 24% + insulin) microneedles. (**C**) SEM micrograph (scale bar: 500 µm) and (**D**) histological cross-sections of human abdominal skin after insertion of the pullulan microneedle patch with insulin. Reproduced with permission from <sup>[53]</sup>. Copyright Elsevier, 2020.

Microneedle patches can also be designed for interstitial skin fluid uptake, which is rich in biomarkers of clinical relevance <sup>[52]</sup>. Fonseca et al. <sup>[55]</sup> took advantage of the swelling ability of a gelatin derivative to produce MNs capable of extracting urea, a metabolite of great importance for the management of kidney diseases. When exposed to ex vivo human abdominal skin, a single patch was able to extract  $3.0 \pm 0.7$  mg of fluid. Overall, microneedle systems are associated with quicker diagnosis, the onset of therapeutics, and less pain and discomfort, which can be of significance for low-compliant patients such as the elderly <sup>[52]</sup>.

#### 2.5. Natural Polymers-Based Materials for Other Applications

In addition to the examples listed above, our group has also been exploiting natural polymers for several other emerging fields of application, ranging from electronics to environmental remediation. For example, the formulation of biopolymeric chitosan coatings loaded with corrosion inhibitors <sup>[56][57][58]</sup> was adopted for the protection of metallic substrates in corrosive environments. In a different approach, flexible electroconductive substrates with

potential application in electronic devices, energy storage, or sensors were devised through the inclusion of copper nanowires <sup>[59]</sup> or organic conductive polymer solutions (e.g., poly(3,4-ethylenedioxythiophene): polystyrene sulfonate (PEDOT:PSS) <sup>[60]</sup>) in nanocellulosic substrates. The functionalization of biopolymers can also be exploited to impart luminescence or (bio)imaging functions to the ensuing materials; for instance, the blend of chitosan and pullulan films with lanthanopolyoxometalates <sup>[61]</sup>, the chemical grafting of corrole macrocycles in a chitosan matrix <sup>[62]</sup>, and the adsorption of a chitosan-derivative containing fluorescein isothiocyanate <sup>[63]</sup> on the surface of cellulose nanocrystals (CNCs). In the field of water remediation, works related to the manufacture of bio-sorbent membranes of BNC/poly(2-methacryloyloxyethyl phosphorylcholine) <sup>[64]</sup> and CNFs/LNFs <sup>[65]</sup> have been suggested for the removal of organic dyes (methylene blue, methylene orange) and mercury from contaminated waters, respectively.

#### References

- Yu, L.; Chen, L. Polymeric materials from renewable resources. In Biodegradable Polymer Blends and Composites from Renewable Resources; Yu, L., Ed.; John Wiley & Sons, Ltd.: Hoboken, NJ, USA, 2009; pp. 1–15. ISBN 9780470146835.
- 2. Klemm, D.; Heublein, B.; Fink, H.-P.; Bohn, A. Cellulose: Fascinating Biopolymer and Sustainable Raw Material. Angew. Chem. Int. Ed. 2005, 44, 3358–3393.
- El Knidri, H.; Laajeb, A.; Lahsini, A. Chitin and chitosan: Chemistry, solubility, fiber formation, and their potential applications. In Handbook of Chitin and Chitosan; Gopi, S., Thomas, S., Pius, A., Eds.; Elsevier: Amsterdam, The Netherlands, 2020; pp. 35–57. ISBN 978-0-12-817970-3.
- Silvetti, T.; Morandi, S.; Hintersteiner, M.; Brasca, M. Use of hen egg white lysozyme in the food industry. In Egg Innovations and Strategies for Improvements; Hester, P.Y., Ed.; Elsevier: Amsterdam, The Netherlands, 2017; pp. 233–242. ISBN 9780128011515.
- Vilela, C.; Pinto, R.J.B.; Pinto, S.; Marques, P.A.A.P.; Silvestre, A.J.D.; Freire, C.S.R. Polysaccharide Based Hybrid Materials: Metals and Metal Oxides, Graphene and Carbon Nanotubes, 1st ed.; Springer Briefs in Molecular Science; Springer International Publishing: Cham, Switzerland, 2018; ISBN 9783030003463.
- Silva, N.H.C.S.; Vilela, C.; Marrucho, I.M.; Freire, C.S.R.; Neto, C.P.; Silvestre, A.J.D. Protein-Based materials: From sources to innovative sustainable materials for biomedical applications. J. Mater. Chem. B 2014, 2, 3715–3740.
- Basu, S.; Malik, S.; Joshi, G.; Gupta, P.; Rana, V. Utilization of bio-polymeric additives for a sustainable production strategy in pulp and paper manufacturing: A comprehensive review. Carbohydr. Polym. Technol. Appl. 2021, 2, 100050.

- Silva, A.C.Q.; Silvestre, A.J.D.; Freire, C.S.R.; Vilela, C. Modification of textiles for functional applications. In Fundamentals of Natural Fibres and Textiles; Mondal, M.I.H., Ed.; Woodhead Publishing, Elsevier Ltd.: Amsterdam, The Netherlands, 2021; pp. 303–365. ISBN 9780128214848.
- 9. Hu, C.; Lu, W.; Mata, A.; Nishinari, K.; Fang, Y. Ions-Induced gelation of alginate: Mechanisms and applications. Int. J. Biol. Macromol. 2021, 177, 578–588.
- Mostafavi, F.S.; Zaeim, D. Agar-Based edible films for food packaging applications—A review. Int. J. Biol. Macromol. 2020, 159, 1165–1176.
- Alipal, J.; Pu'Ad, N.M.; Lee, T.; Nayan, N.; Sahari, N.; Basri, H.; Idris, M.; Abdullah, H. A review of gelatin: Properties, sources, process, applications, and commercialisation. Mater. Today Proc. 2021, 42, 240–250.
- Zhao, Y.; Zheng, Y.; Wang, J.; Ma, S.; Yu, Y.; White, W.L.; Yang, S.; Yang, F.; Lu, J. Fucoidan Extracted from Undaria pinnatifida: Source for Nutraceuticals/Functional Foods. Mar. Drugs 2018, 16, 321.
- Bukhari, S.N.A.; Roswandi, N.L.; Waqas, M.; Habib, H.; Hussain, F.; Khan, S.; Sohail, M.; Ramli, N.A.; Thu, H.E.; Hussain, Z. Hyaluronic acid, a promising skin rejuvenating biomedicine: A review of recent updates and pre-clinical and clinical investigations on cosmetic and nutricosmetic effects. Int. J. Biol. Macromol. 2018, 120, 1682–1695.
- Gradishar, W.J.; Tjulandin, S.; Davidson, N.; Shaw, H.; Desai, N.; Bhar, P.; Hawkins, M.; O'Shaughnessy, J. Phase III Trial of Nanoparticle Albumin-Bound Paclitaxel Compared with Polyethylated Castor Oil–Based Paclitaxel in Women with Breast Cancer. J. Clin. Oncol. 2005, 23, 7794–7803.
- 15. Tong, X.; Pan, W.; Su, T.; Zhang, M.; Dong, W.; Qi, X. Recent advances in natural polymer-based drug delivery systems. React. Funct. Polym. 2020, 148, 104501.
- Ibrahim, S.; Riahi, O.; Said, S.M.; Sabri, M.F.M.; Rozali, S. Biopolymers From Crop Plants. In Reference Module in Materials Science and Materials Engineering; Elsevier: Hoboken, NJ, USA, 2019; pp. 1–10.
- 17. Tongdeesoontorn, W.; Rawdkuen, S. Gelatin-Based Films and Coatings for Food Packaging Applications. In Reference Module in Food Science; Elsevier: Hoboken, NJ, USA, 2019; pp. 1–15.
- 18. Priyadarshi, R.; Rhim, J.-W. Chitosan-based biodegradable functional films for food packaging applications. Innov. Food Sci. Emerg. Technol. 2020, 62, 102346.
- 19. Mansoori, S.; Davarnejad, R.; Matsuura, T.; Ismail, A.F. Membranes based on non-synthetic (natural) polymers for wastewater treatment. Polym. Test. 2020, 84, 106381.

- 20. Vedula, S.S.; Yadav, G.D. Chitosan-based membranes preparation and applications: Challenges and opportunities. J. Indian Chem. Soc. 2021, 98, 100017.
- 21. Nechita, P.; Roman, M. Review on Polysaccharides Used in Coatings for Food Packaging Papers. Coatings 2020, 10, 566.
- 22. Song, J.; Winkeljann, B.; Lieleg, O. Biopolymer-Based Coatings: Promising Strategies to Improve the Biocompatibility and Functionality of Materials Used in Biomedical Engineering. Adv. Mater. Interfaces 2020, 7, 2000850.
- 23. Graça, M.F.P.; Miguel, S.P.; Cabral, C.S.D.; Correia, I.J. Hyaluronic acid—Based wound dressings: A review. Carbohydr. Polym. 2020, 241, 116364.
- 24. Carvalho, J.P.F.; Silva, A.C.Q.; Silvestre, A.J.D.; Freire, C.S.R.; Vilela, C. Spherical Cellulose Micro and Nanoparticles: A Review of Recent Developments and Applications. Nanomaterials 2021, 11, 2744.
- 25. Bennacef, C.; Desobry-Banon, S.; Probst, L.; Desobry, S. Advances on alginate use for spherification to encapsulate biomolecules. Food Hydrocoll. 2021, 118, 106782.
- DeFrates, K.; Markiewicz, T.; Gallo, P.; Rack, A.; Weyhmiller, A.; Jarmusik, B.; Hu, X. Protein Polymer-Based Nanoparticles: Fabrication and Medical Applications. Int. J. Mol. Sci. 2018, 19, 1717.
- 27. Vilela, C.; Moreirinha, C.; Domingues, E.M.; Figueiredo, F.M.L.; Almeida, A.; Freire, C.S.R. Antimicrobial and Conductive Nanocellulose-Based Films for Active and Intelligent Food Packaging. Nanomaterials 2019, 9, 980.
- Bastante, C.C.; Silva, N.H.; Cardoso, L.C.; Serrano, C.M.; de la Ossa, E.J.M.; Freire, C.S.; Vilela, C. Biobased films of nanocellulose and mango leaf extract for active food packaging: Supercritical impregnation versus solvent casting. Food Hydrocoll. 2021, 117, 106709.
- 29. Moreirinha, C.; Vilela, C.; Silva, N.H.; Pinto, R.J.; Almeida, A.; Rocha, M.A.M.; Coelho, E.; Coimbra, M.A.; Silvestre, A.J.; Freire, C.S. Antioxidant and antimicrobial films based on brewers spent grain arabinoxylans, nanocellulose and feruloylated compounds for active packaging. Food Hydrocoll. 2020, 108, 105836.
- Tomé, L.C.; Silva, N.H.C.S.; Soares, H.R.; Coroadinha, A.S.; Sadocco, P.; Marrucho, I.M.; Freire, C.S.R. Bioactive transparent films based on polysaccharides and cholinium carboxylate ionic liquids. Green Chem. 2015, 17, 4291–4299.
- Esposito, T.; Silva, N.H.; Almeida, A.; Silvestre, A.J.; Piccinelli, A.L.; Aquino, R.P.; Sansone, F.; Mencherini, T.; Vilela, C.; Freire, C.S. Valorisation of chestnut spiny burs and roasted hazelnut skins extracts as bioactive additives for packaging films. Ind. Crops Prod. 2020, 151, 112491.

- 32. Pinto, R.; Almeida, A.; Fernandes, S.C.; Freire, C.; Silvestre, A.; Neto, C.P.; Trindade, T. Antifungal activity of transparent nanocomposite thin films of pullulan and silver against Aspergillus niger. Colloids Surf. B Biointerfaces 2013, 103, 143–148.
- Vilela, C.; Pinto, R.J.; Coelho, J.; Domingues, M.R.; Daina, S.; Sadocco, P.; Santos, S.A.; Freire, C. Bioactive chitosan/ellagic acid films with UV-light protection for active food packaging. Food Hydrocoll. 2017, 73, 120–128.
- Makhloufi, N.; Chougui, N.; Rezgui, F.; Benramdane, E.; Freire, C.S.R.; Vilela, C.; Silvestre, A.J.D. Bio-based sustainable films from the Algerian Opuntia ficus-indica cladodes powder: Effect of plasticizer content. J. Appl. Polym. Sci. 2021, 138, 50450.
- Carvalho, J.P.F.; Freire, C.S.R.; Vilela, C. Active Packaging. In Sustainable Food Processing and Engineering Challenges; Galanakis, C., Ed.; Elsevier: Amsterdam, The Netherlands, 2021; pp. 315–341. ISBN 9780128227145.
- Vilela, C.; Kurek, M.; Hayouka, Z.; Röcker, B.; Yildirim, S.; Antunes, M.D.C.; Nilsen-Nygaard, J.; Pettersen, M.K.; Freire, C.S.R. A concise guide to active agents for active food packaging. Trends Food Sci. Technol. 2018, 80, 212–222.
- Silva, N.H.; Vilela, C.; Almeida, A.; Marrucho, I.; Freire, C. Pullulan-based nanocomposite films for functional food packaging: Exploiting lysozyme nanofibers as antibacterial and antioxidant reinforcing additives. Food Hydrocoll. 2018, 77, 921–930.
- 38. Silva, N.H.; Vilela, C.; Pinto, R.J.; Martins, M.A.; Marrucho, I.M.; Freire, C.S. Tuning lysozyme nanofibers dimensions using deep eutectic solvents for improved reinforcement ability. Int. J. Biol. Macromol. 2018, 115, 518–527.
- 39. Vilela, C.; Silvestre, A.J.D.; Figueiredo, F.M.L.; Freire, C.S.R. Nanocellulose-based materials as components of polymer electrolyte fuel cells. J. Mater. Chem. A 2019, 7, 20045–20074.
- Gadim, T.D.; Vilela, C.; Loureiro, F.; Silvestre, A.; Freire, C.S.; Figueiredo, F.M. Nafion ® and nanocellulose: A partnership for greener polymer electrolyte membranes. Ind. Crops Prod. 2016, 93, 212–218.
- Gadim, T.D.O.; Figueiredo, A.G.P.R.; Navarro, N.C.R.; Vilela, C.; Gamelas, J.; Timmons, A.B.; Neto, C.P.; Silvestre, A.; Freire, C.; Figueiredo, F.M.L. Nanostructured Bacterial Cellulose–Poly(4styrene sulfonic acid) Composite Membranes with High Storage Modulus and Protonic Conductivity. ACS Appl. Mater. Interfaces 2014, 6, 7864–7875.
- Gadim, T.D.; Loureiro, F.J.; Vilela, C.; Rosero-Navarro, N.; Silvestre, A.J.; Freire, C.S.; Figueiredo, F.M. Protonic conductivity and fuel cell tests of nanocomposite membranes based on bacterial cellulose. Electrochim. Acta 2017, 233, 52–61.
- 43. Vilela, C.; Cordeiro, D.M.; Boas, J.V.; Barbosa, P.; Nolasco, M.; Vaz, P.D.; Rudić, S.; Ribeiro-Claro, P.; Silvestre, A.J.; Oliveira, V.B.; et al. Poly(4-styrene sulfonic acid)/bacterial cellulose

membranes: Electrochemical performance in a single-chamber microbial fuel cell. Bioresour. Technol. Rep. 2019, 9, 100376.

- 44. Vilela, C.; Martins, A.P.C.; Sousa, N.; Silvestre, A.J.D.; Figueiredo, F.M.L.; Freire, C.S.R. Poly(bis phosphate)/Bacterial Cellulose Nanocomposites: Preparation, Characterization and Application as Polymer Electrolyte Membranes. Appl. Sci. 2018, 8, 1145.
- 45. Vilela, C.; Gadim, T.D.O.; Silvestre, A.; Freire, C.; Figueiredo, F.M.L. Nanocellulose/poly(methacryloyloxyethyl phosphate) composites as proton separator materials. Cellulose 2016, 23, 3677–3689.
- Vilela, C.; Sousa, N.; Pinto, R.; Silvestre, A.; Figueiredo, F.M.; Freire, C.S. Exploiting poly(ionic liquids) and nanocellulose for the development of bio-based anion-exchange membranes. Biomass Bioenergy 2017, 100, 116–125.
- 47. Vilela, C.; Silva, A.C.; Domingues, E.; Gonçalves, G.; Martins, M.A.; Figueiredo, F.M.; Santos, S.A.; Freire, C.S. Conductive polysaccharides-based proton-exchange membranes for fuel cell applications: The case of bacterial cellulose and fucoidan. Carbohydr. Polym. 2020, 230, 115604.
- Vilela, C.; Morais, J.D.; Silva, A.C.Q.; Muñoz-Gil, D.; Figueiredo, F.M.L.; Silvestre, A.J.D.; Freire, C.S.R. Flexible Nanocellulose/Lignosulfonates Ion-Conducting Separators for Polymer Electrolyte Fuel Cells. Nanomaterials 2020, 10, 1713.
- 49. Fonseca, D.F.S.; Carvalho, J.P.F.; Bastos, V.; Oliveira, H.; Moreirinha, C.; Almeida, A.; Silvestre, A.J.D.; Vilela, C.; Freire, C.S.R. Antibacterial Multi-Layered Nanocellulose-Based Patches Loaded with Dexpanthenol for Wound Healing Applications. Nanomaterials 2020, 10, 2469.
- 50. Silva, N.H.; Garrido-Pascual, P.; Moreirinha, C.; Almeida, A.; Palomares, T.; Alonso-Varona, A.; Vilela, C.; Freire, C.S. Multifunctional nanofibrous patches composed of nanocellulose and lysozyme nanofibers for cutaneous wound healing. Int. J. Biol. Macromol. 2020, 165, 1198–1210.
- 51. Karki, S.; Kim, H.; Na, S.-J.; Shin, D.; Jo, K.; Lee, J. Thin films as an emerging platform for drug delivery. Asian J. Pharm. Sci. 2016, 11, 559–574.
- 52. Fonseca, D.; Vilela, C.; Silvestre, A.J.; Freire, C.S. A compendium of current developments on polysaccharide and protein-based microneedles. Int. J. Biol. Macromol. 2019, 136, 704–728.
- Fonseca, D.F.; Costa, P.C.; Almeida, I.F.; Dias-Pereira, P.; Correia-Sá, I.; Bastos, V.; Oliveira, H.; Duarte-Araújo, M.; Morato, M.; Vilela, C.; et al. Pullulan microneedle patches for the efficient transdermal administration of insulin envisioning diabetes treatment. Carbohydr. Polym. 2020, 241, 116314.
- 54. Fonseca, D.F.; Vilela, C.; Pinto, R.J.; Bastos, V.; Oliveira, H.; Catarino, J.; Faísca, P.; Rosado, C.; Silvestre, A.J.; Freire, C.S. Bacterial nanocellulose-hyaluronic acid microneedle patches for skin applications: In vitro and in vivo evaluation. Mater. Sci. Eng. C 2021, 118, 111350.

- Fonseca, D.F.S.; Costa, P.C.; Almeida, I.F.; Dias-Pereira, P.; Correia-Sá, I.; Bastos, V.; Oliveira, H.; Vilela, C.; Silvestre, A.J.D.; Freire, C.S.R. Swellable Gelatin Methacryloyl Microneedles for Extraction of Interstitial Skin Fluid toward Minimally Invasive Monitoring of Urea. Macromol. Biosci. 2020, 20, e2000195.
- 56. Carneiro, J.; Tedim, J.; Fernandes, S.; Freire, C.; Gandini, A.; Ferreira, M.; Zheludkevich, M. Functionalized chitosan-based coatings for active corrosion protection. Surf. Coat. Technol. 2013, 226, 51–59.
- 57. Carneiro, J.; Tedim, J.; Fernandes, S.C.M.; Freire, C.; Gandini, A.; Ferreira, M.; Zheludkevich, M. Chitosan as a Smart Coating for Controlled Release of Corrosion Inhibitor 2-Mercaptobenzothiazole. ECS Electrochem. Lett. 2013, 2, C19–C22.
- Carneiro, J.; Tedim, J.; Fernandes, S.; Freire, C.; Silvestre, A.; Gandini, A.; Ferreira, M.; Zheludkevich, M. Chitosan-based self-healing protective coatings doped with cerium nitrate for corrosion protection of aluminum alloy 2024. Prog. Org. Coat. 2012, 75, 8–13.
- Pinto, R.J.B.; Martins, M.A.; Lucas, J.M.F.; Vilela, C.; Sales, A.; Costa, L.C.; Marques, P.; Freire, C.S.R. Highly Electroconductive Nanopapers Based on Nanocellulose and Copper Nanowires: A New Generation of Flexible and Sustainable Electrical Materials. ACS Appl. Mater. Interfaces 2020, 12, 34208–34216.
- Inácio, P.M.; Medeiros, M.D.C.; Carvalho, T.; Félix, R.C.; Mestre, A.; Hubbard, P.C.; Ferreira, Q.; Morgado, J.; Charas, A.; Freire, C.S.; et al. Ultra-low noise PEDOT:PSS electrodes on bacterial cellulose: A sensor to access bioelectrical signals in non-electrogenic cells. Org. Electron. 2020, 85, 105882.
- Pinto, R.J.B.; Granadeiro, C.M.; Freire, C.S.R.; Silvestre, A.J.D.; Neto, C.P.; Ferreira, R.A.S.; Carlos, L.D.; Cavaleiro, A.M.V.; Trindade, T.; Nogueira, H.I.S. Luminescent Transparent Composite Films Based on Lanthanopolyoxometalates and Filmogenic Polysaccharides. Eur. J. Inorg. Chem. 2013, 2013, 1890–1896.
- 62. Barata, J.F.B.; Pinto, R.J.B.; Serra, V.I.R.C.V.; Silvestre, A.J.D.; Trindade, T.; Neves, M.G.P.M.S.; Cavaleiro, J.A.S.; Daina, S.; Sadocco, P.; Freire, C.S.R. Fluorescent Bioactive Corrole Grafted-Chitosan Films. Biomacromolecules 2016, 17, 1395–1403.
- Pinto, R.J.B.; Lameirinhas, N.S.; Guedes, G.; da Silva, G.H.R.; Oskoei, P.; Spirk, S.; Oliveira, H.; Duarte, I.F.; Vilela, C.; Freire, C.S.R. Cellulose Nanocrystals/Chitosan-Based Nanosystems: Synthesis, Characterization, and Cellular Uptake on Breast Cancer Cells. Nanomaterials 2021, 11, 2057.
- 64. Vilela, C.; Moreirinha, C.; Almeida, A.; Silvestre, A.J.D.; Freire, C.S.R. Zwitterionic Nanocellulose-Based Membranes for Organic Dye Removal. Materials 2019, 12, 1404.

65. Silva, N.H.; Figueira, P.; Fabre, E.; Pinto, R.J.; Pereira, M.E.; Silvestre, A.J.; Marrucho, I.M.; Vilela, C.; Freire, C.S. Dual nanofibrillar-based bio-sorbent films composed of nanocellulose and lysozyme nanofibrils for mercury removal from spring waters. Carbohydr. Polym. 2020, 238, 116210.

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