

# Fatty Acid Profile as Disease Predictor

Subjects: Biochemistry & Molecular Biology

Contributor: Raja Chaaba, Aicha Bouaziz, Asma Ben Amor, Wissem Mnif, Mohamed Hammami, Sounira Mehri

Circulating fatty acids (FA) have an endogenous or exogenous origin and are metabolized under the effect of many enzymes. They play crucial roles in many mechanisms: cell signaling, modulation of gene expression, etc., which leads to the hypothesis that their perturbation could be the cause of disease development. FA in erythrocytes and plasma rather than dietary FA could be used as a biomarker for many diseases. Cardiovascular disease was associated with elevated trans FA and decreased DHA and EPA. Increased arachidonic acid and decreased Docosahexaenoic Acids (DHA) were associated with Alzheimer's disease. Low Arachidonic acid and DHA are associated with neonatal morbidities and mortality. Decreased saturated fatty acids (SFA), increased monounsaturated FA (MUFA) and polyunsaturated FA (PUFA) (C18:2 n-6 and C20:3 n-6) are associated with cancer. Additionally, genetic polymorphisms in genes coding for enzymes implicated in FA metabolism are associated with disease development. FA desaturase (FADS1 and FADS2) polymorphisms are associated with Alzheimer's disease, Acute Coronary Syndrome, Autism spectrum disorder and obesity. Polymorphisms in FA elongase (ELOVL2) are associated with Alzheimer's disease, Autism spectrum disorder and obesity. FA-binding protein polymorphism is associated with dyslipidemia, type 2 diabetes, metabolic syndrome, obesity, hypertension, non-alcoholic fatty liver disease, peripheral atherosclerosis combined with type 2 diabetes and polycystic ovary syndrome. Acetyl-coenzyme A carboxylase polymorphisms are associated with diabetes, obesity and diabetic nephropathy. FA profile and genetic variants of proteins implicated in FA metabolism could be considered as disease biomarkers and may help with the prevention and management of diseases.

Keywords: fatty acids ; diseases ; gene polymorphisms ; red blood cells

---

## 1. Introduction

Fatty acids (FA) belong to the lipid class. They can be free or associated with alcohols to provide triglycerides, phospholipids, cerids or sterides. They can be saturated (SFA) or unsaturated (UFA) depending on the presence or absence of a double bound in their structure. Based on the number of double bounds, it can distinguish between monounsaturated FA (MUFA with only one double bound) or polyunsaturated FA (PUFA with two or more double bounds). According to the carbon chain length, four groups of FA are identified: short-chain FA with 4 to 6 carbon atoms, medium-chain FA with 8 to 12 carbon atoms, long-chain FA with 14 to 20 carbon atoms, and very long-chain FA with 22 or more carbon atoms. Depending on the configuration, a distinction is made between trans FA and cis FA. FA differ from each other in the number of carbon atoms, unsaturations and amount of configuration. They are present in all cells and tissues of the body. SFA and MUFA are synthesized by all organisms. However, not all PUFA are synthesized by mammals, including humans, because they do not have the enzymes ( $\Delta 12$ - and  $\Delta 15$  desaturases) necessary for their synthesis. These are therefore called essential FA. These FA are represented by alpha-linolenic acid (ALA, C18: 3 n-3) <sup>[1]</sup> and linoleic acid (LA, C18: 2 n-6) <sup>[2]</sup>. They will play the role of precursors of other FA with longer chains and of bioactive mediators in the form of oxygenated molecules (eicosanoids, docosanoids, etc.) <sup>[3]</sup>.

FA are highly energetic substrates. In the mitochondria, they provide a large amount of energy through beta oxidation. This quantity depends on the number of carbons that make up the FA, but it is always greater than that provided by a molecule of glucose (for FA with more than four carbons).

FA are the main constituent of cell membranes. They influence membrane fluidity <sup>[4]</sup>. Incorporated into membrane phospholipids, PUFA increase membrane fluidity and can affect cell function. Moreover, the FA composition of the membrane influences cell signaling. Increasing the concentration of docosahexaenoic acid (DHA, C22:6 n-3) in the excitable membranes of the brain and retina acts on cell signaling by altering lipid rafts at this level <sup>[5]</sup>. Impaired insulin signaling by free FA leads to endothelial dysfunction <sup>[6]</sup>. MUFA affect the Intracellular Signaling in Cancer <sup>[7]</sup>.

FA can be precursors of oxygenated molecules. Dihomo-gamma-linolenic (C20:3 n-6), arachidonic (AA, C20:4 n-6), and eicosapentaenoic (EPA, C20:5 n-3) acids enter the enzymatic pathways of oxygenation, hydroxylation and of peroxidation to form eicosanoids (powerful mediators). Membrane FA released under the effect of phospholipase A2 can enter two

different metabolic pathways: (i) the cyclo-oxygenase (COX) pathway which will give rise to prostaglandins, prostacyclins and thromboxanes and (ii) the lipoxygenase (LOX) pathway, which produces leukotrienes and hydroperoxidized FA [3]. In addition, peroxidation of 20-carbon PUFAs through the cytochrome P450 pathway can yield epoxyeicotrienoic acids. AA and DHA can yield isoprostanes and neuroprostanes through non-enzymatic peroxidation [3].

PUFAs have the ability to bind directly or through their derivatives (eicosanoids) to transcription factors, which modulate the expression of certain genes involved in various metabolic pathways [8]. The most important transcription factors are peroxisome proliferator-activated receptors (PPARs), sterol regulatory element binding protein (SREBP-1c for sterol regulatory element binding protein-1c), hepatic nuclear factors (HNF4 for hepatic nuclear factor 4), retinoid receptor (RXRa for retinoid X receptor) and the hepatic X receptor (LXRa for liver X receptor).

## **2. Fatty Acid Profile as Disease Predictor**

### **2.1. Fatty Acids and Diseases**

#### **2.1.1. Fatty Acids and Cardiovascular Disease (CVD)**

FA play a crucial role in CVD. They exert their effect by acting on lipoprotein metabolism [9] and endothelial function [10]. FA-mediated dysregulation of nitric acid and cytokine production, inflammation, oxidative stress, apoptosis, and activation of the renin–angiotensin system, which causes endothelial dysfunction, therefore increases CVD risk [10]. Many FA are associated, whereas others are inversely associated with CVD. A strong inverse correlation was shown between RBC oleic acid and CVD [11]. Another study showed a protective effect of plasma very long-chain SFA (arachidic acid (20:0), behenic acid (22:0) and lignoceric acid (24:0)) in heart failure [12]. However, Hadj Ahmed et al. showed an increased level of RBC C26:0, C24:0, C22:0, EPA and AA in patients with coronary artery disease compared to the normal population and consider them biomarkers of coronary artery disease [13]. The same team showed an association between RBC and plasma trans FA and coronary artery disease severity [14]. Many other studies showed an association between RBC trans FA and coronary heart disease [9]. This association is mediated by increased low-density lipoprotein cholesterol and decreased high-density lipoprotein cholesterol [9]. Acute myocardial infarction is associated with decreased short-chain FA and increased long-chain FA levels [15]. Acute coronary syndrome is inversely associated with DHA/AA ratio [16]. Early onset coronary atherosclerosis is associated with decreased EPA and DHA levels [17] and risk of atherosclerotic plaque rupture is associated with high AA/DHA ratio [18]. The results are different for different studies. For example, cardiovascular disease is associated with elevated n-6 PUFA [19] or elevated SFA and trans FA [20] or increased MUFA [21] and the same is true for coronary artery diseases [13][22]. Such differences could be explained by genetic, lifestyle and dietary differences between the studied populations. One study showed that the association between FA and cardiometabolic risk is modulated by concurrent physical activity [23]. The main conclusion is that the FA profile is modified between healthy subjects and patients. The most consistent association is for increased trans FA and decreased DHA and EPA.

Moreover, the FA profile could predict cardiovascular disease development in unhealthy subjects such as diabetic patients [24] and patients with renal failure [25]. Additionally, it predicts psychiatric disorder at 6 months after acute coronary syndrome [26]. Moreover, it predicts mortality among patients with acute cardiovascular disease [27], coronary artery disease [28] and with myocardial infarction [29].

#### **2.1.2. Fatty Acids and Diabetes**

Many studies have highlighted the association between elevated total plasma free FA and diabetes [30][31][32], whereas other studies examined individual FA. RBC n-3 PUFA was negatively associated with the risk of type 2 diabetes [33]. Serum C22:0 and plasma n-6/n-3 were associated with diabetes [34][35]. Many studies have agreed on elevated plasma palmitic acid, stearic acid and oleic acid in diabetic patients [31][32][36][37][38][39]. In fact, palmitic acid slows down insulin signal transduction [40]. Meanwhile, the association between increased oleic acid concentration and the development of diabetes is not well understood, as studies show that oleic acid may play a role in the protection and treatment of diabetes [39]. Elevated RBC-linolenic acid was associated with high type 2 diabetes incidence [41].

#### **2.1.3. Fatty Acids and Cancer**

A study in patients with any type of cancer (except head and neck cancer) showed that cancer is associated with decreased SFA (C16:0 and C18:0) and increased MUFA (C18:1) and PUFA (LA and C20:3 n-6) [42]. Hepatocellular carcinoma is strongly associated with low levels of very long-chain SFA and n-3 PUFA [43]. Oral cancer is associated with decreased EPA and DHA [44]. Pancreatic cancer is associated with decreased n-3 PUFA and MUFA and increased arachidonic acid [45], whereas the association between FA and breast cancer depends on many variables, such as

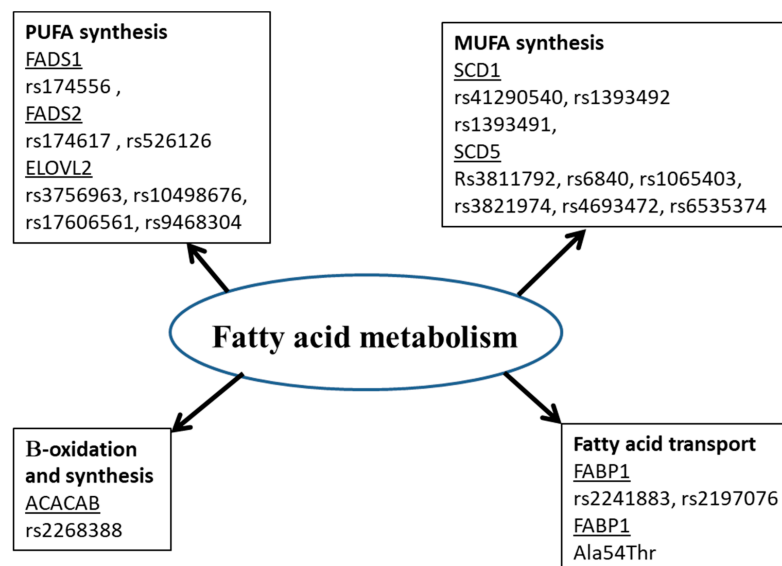
menopause [46] and BMI [47]. In breast cancer, FA (omega 3 and omega 6) modulate the cancer immune response and the fatty acid endogenous synthesis [48]. The concentration of oleic acid is lower in cancer patients, increasing their exposure to the disease [49].

#### 2.1.4. Fatty Acids and Other Diseases

Serum  $\gamma$ -linolenic acid and C9:0 and C19:0 were associated with obesity [50][51] and dihomo- $\gamma$ -linolenic acid and palmitoleic acid were able to predict the future development of metabolic syndrome (MS) in obese subjects [52]. Zarrouk et al. identified hexacosanoic (C26:0) as blood (RBC and plasma) lipid biomarkers of dementia [53], whereas Yamagishi et al. showed that Serum ALA was inversely associated with the risk of disabling dementia [54]. Hammouda et al. showed that AA increased in plasma and RBC and DHA decreased only in plasma of Alzheimer patients [55]. Additionally, Sala-vila et al. found that DHA was inversely associated with Alzheimer's disease [56]. Meanwhile, a study by Tomata et al. did not support the association between PUFA and Alzheimer's disease risks [57].

## 2.2. Polymorphisms of Gene Implicated in Fatty Acid Metabolism and Diseases

Many studies showed that the fatty acid in biological tissues is an indicator of fatty acid intake [58][59]. However, this is not always the case. Amézaga et al. showed that the variation in erythrocyte FA is not linked to dietary intake [42]. FA have an exogenous and endogenous origin. They can be provided by food or synthesized, mainly in hepatocytes. Several proteins are involved in the metabolism of FA (**Figure 1**). Any perturbation in the activity or quantity of those enzymes could be a cause of a fatty acid profile change, then disease development [60].



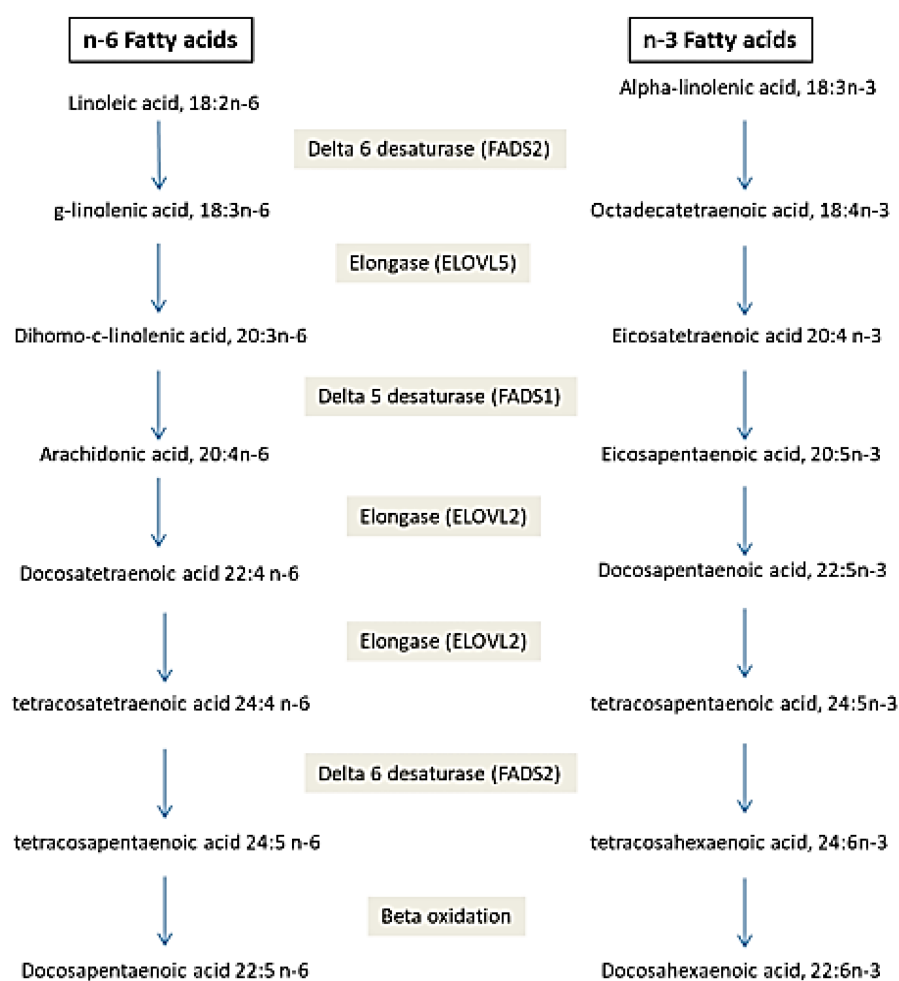
**Figure 1.** Polymorphisms of genes for proteins involved in fatty acid metabolism. ACACB: Acetyl-coenzyme A carboxylase, ELOVL: Elongase, FADS: Fatty Acid Desaturase, FABP: Fatty Acid Binding Protein, MUFA: monounsaturated fatty acids, PUFA: polyunsaturated fatty acids, SCD: stearoyl Co A desaturase.

Delta 9 MUFAs (mainly oleic acid) are synthesized under the control of many enzymes. The key enzyme of desaturation is stearoyl Co A desaturase (SCD: SCD1 and SCD5). This enzyme plays a key role in the development of many diseases. Thus, inhibiting it will have many beneficial effects. SCD1 enzyme was a potential target for cancer [61], non-alcoholic fatty liver disease [62], diabetes, obesity and hepatic steatosis [63]. The activity of SCD1 is usually calculated as C18:1/C18:0 ratio. Single Nucleotide Polymorphisms (SNP) in SCD1 gene (located on chromosome 10q24.31) are associated with many diseases. The rs41290540 SNP in the 3'-untranslated region is associated with decreased risk of coronary artery disease [64].

The association between SCD1 gene polymorphism and disease development depends on many factors, such as diet: oil intake in case of obesity [65] and PUFA intake in case of cancer death [66]. The relationship between SCD1 and disease development could be explained by the variation in SCD1 expression according to gene variant, which influences the fatty acid profile (unsaturated: saturated fatty acid ratio) [67]. Despite the great attention attributed to SCD1 isoform, few studies on SCD5 isoform have been conducted. The rs3811792 SNP in SCD5 promoter is associated with type 1 and type 2 diabetes [68]. This polymorphism decreases SCD5 promoter activity. Zambo explained the association between the polymorphism and type 2 diabetes based on the fact that SCD5 regulates the distribution of fats and accumulation in viscus, which represents a diabetes risk factor. However, the association with type 1 diabetes is due to the overexpression of SCD5 in the pancreas compared to other tissues. Many other polymorphisms in SCD5 gene are described: rs6840,

rs1065403, rs3821974 in 3'-UTR and rs4693472, rs6535374 in intron. All of them are associated with hepatocellular carcinoma [69].

PUFA synthesis requires other enzymes. Desaturase and elongase enzymes involved in fatty acid synthesis from ALA and LA are summarized in **Figure 2**. Two types of desaturases exist: delta-6-desaturase (FA desaturase 2, FADS2) and delta5-desaturase (FA desaturase 1, FADS1). FADS2 converts substrates ALA and LA, respectively at C18:4 n-3 and C18:3 n-6. This enzyme is also involved in a second step leading to the desaturation of the C24:5 n-3 and C24:4 n-6 substrates to C24:6 n-3 and C24:5 n-6, respectively. FADS1 desaturated the FA C20:4 n-3 and C20:3 n-6, respectively into EPA and AA. The activity of elongases (ELOVL) on eighteen- and twenty-carbon FA and n-3 long-chain PUFA is the result of ELOVL5 gene expression. On the other hand, ELOVL2 is the gene allowing the elongation of PUFAs with twenty and twenty-two carbon atoms. The genes coding for desaturases and elongases are the most studied to explore the genetic effect of FA on the development of diseases [55][70][71][72]. Hammouda et al. showed that rs174556 TT genotype of FADS1 is associated with a higher AA level and AA/DGLA (Dihomo-gamma-linolenic acid) index and increased risk of Alzheimer's disease. Additionally, rs3756963 TT genotype of ELOVL2 is associated with increased AA and AA/DGLA index and increased risk of Alzheimer's disease. However, rs174617 of FADS2 is not associated with either the fatty acid profile or Alzheimer's disease. The combination of the two variants increases further the susceptibility to Alzheimer's disease.



**Figure 2.** Desaturase and elongase metabolic pathway in long-chain fatty acid synthesis.

The fatty acid-binding protein (FABP) family includes several FABPs that are abundantly expressed in tissues with active fatty acid metabolism. FABPs would have several roles, including the intracellular transport of fatty acid chains and other lipophilic substances, such as eicosanoids and retinoids, from plasma membranes to sites of metabolism. Members of this family include FABP1 in liver, FABP2 in intestine, FABP3 in muscle and heart and FABP4 in adipocytes. Valizadeh et al. showed that the rs2241883 CC genotype of FABP1 gene is associated with dyslipidemia [73] and Xue et al. found that rs2197076 and rs2241883 of the same protein are associated with polycystic ovary syndrome [74]. According to Xue et al., rs2197076 seemed to have a more important role in the mechanism of polycystic ovary syndrome than rs2241883 because it is closely related to some important clinical features of polycystic ovary syndrome. Moreover, rs2197076 FABP1 is associated with type 2 diabetes [75], rs2241883 is associated with metabolic syndrome [76] and rs2241883 with rs1545224 are associated with non-alcoholic fatty liver [77]. The most FABP-studied polymorphism is located in FABP2 gene. It is an amino acid substitution (Ala54 to Thr54). This polymorphism is associated with diabetes, metabolic

syndrome, obesity and non-alcoholic liver disease [78][79][80][81]. In diabetic patients, the polymorphism is associated with peripheral atherosclerosis, retinopathy and renal disease in diabetic patients [82][83][84]. Another polymorphism in FABP2 is associated with essential hypertension [85] and diabetes [86].

Acetyl-coenzyme A carboxylase (ACACB) catalyzes the synthesis of malonyl-CoA, a metabolite that plays an essential role in the synthesis and oxidation of FA. A meta-analysis showed that rs2268388 C allele in ACACB was inversely associated with susceptibility risk of diabetic nephropathy among Caucasian patients [87], Chinese patients [88] and Asian Indian patients [89]. However, Chan et al. did not find any association between ACACB gene polymorphism and cardiovascular risk susceptibility in type 2 diabetic patients [90]. In Spanish postmenopausal women, the rs2268388 T allele was associated with obesity and diabetes and the rs2239607 C allele was associated with diabetes [91].

Genetics alone cannot confirm the presence of a disease, except in certain cases in which polymorphisms are mutations that directly affect the presence or function of the enzyme. However, genetic studies make it possible to predict diseases and help to manage them. In fact, diseases are usually multifactorial. One disease can be the result of many protein disturbances. Additionally, environmental factors such as diet, temperature and stress can modulate the effect of genetic factors.

---

## References

1. Holman, R.T.; Johnson, S.B.; Hatch, T.F. A case of human linolenic acid deficiency involving neurological abnormalities. *Am. J. Clin. Nutr.* 1982, 35, 617–623.
2. Hansen, A.E.; Haggard, M.E.; Boelsche, A.N.; Adam, D.J.; Wiese, H.F. Essential fatty acids in infant nutrition: III. Clinical manifestations of linoleic acid deficiency. *J. Nutr.* 1958, 66, 565–576.
3. Guesnet, P.; Alessandri, J.-M.; Astorg, P.; Pifferi, F.; Lavialle, M. Les rôles physiologiques majeurs exercés par les acides gras polyinsaturés (AGPI). *Oléagineux Corps Gras Lipides* 2005, 12, 333–343.
4. Maulucci, G.; Cohen, O.; Daniel, B.; Sansone, A.; Petropoulou, P.I.; Filou, S.; Spyridonidis, A.; Pani, G.; De Spirito, M.; Chatgililoglu, C.; et al. Fatty acid-related modulations of membrane fluidity in cells: Detection and implications. *Free. Radic. Res.* 2016, 50, S40–S50.
5. Stillwell, W.; Shaikh, S.R.; Zerouga, M.; Siddiqui, R.; Wassall, S.R. Docosahexaenoic acid affects cell signaling by altering lipid rafts. *Reprod. Nutr. Dev.* 2005, 45, 559–579.
6. Ghosh, A.; Gao, L.; Thakur, A.; Siu, P.M.; Lai, C.W.K. Role of free fatty acids in endothelial dysfunction. *J. Biomed. Sci.* 2017, 24, 50.
7. Scott, J.S.; Nassar, Z.D.; Swinnen, J.V.; Butler, L.M. Monounsaturated Fatty Acids: Key Regulators of Cell Viability and Intracellular Signaling in Cancer. *Mol. Cancer Res.* 2022, 20, 1354–1364.
8. Wahle, K.W.J.; Rotondo, D.; Heys, S.D. Polyunsaturated fatty acids and gene expression in mammalian systems. *Proc. Nutr. Soc.* 2003, 62, 349–360.
9. Sun, Q.; Ma, J.; Campos, H.; Hankinson, S.E.; Manson, J.E.; Stampfer, M.J.; Rexrode, K.M.; Willett, W.C.; Hu, F.B. A Prospective Study of Trans Fatty Acids in Erythrocytes and Risk of Coronary Heart Disease. *Circulation* 2007, 115, 1858–1865.
10. Mallick, R.; Duttaroy, A.K. Modulation of endothelium function by fatty acids. *Mol. Cell. Biochem.* 2021, 477, 15–38.
11. Fan, W.X.; Parker, R.; Parpia, B.; Qu, Y.S.; Cassano, P.; Crawford, M.; Leyton, J.; Tian, J.; Li, J.Y.; Chen, J.S. Erythrocyte fatty acids, plasma lipids, and cardiovascular disease in rural China. *Am. J. Clin. Nutr.* 1990, 52, 1027–1036.
12. Lemaitre, R.N.; McKnight, B.; Sotoodehnia, N.; Fretts, A.M.; Qureshi, W.T.; Song, X.; King, I.B.; Sittlani, C.M.; Siscovick, D.S.; Psaty, B.M.; et al. Circulating Very Long-Chain Saturated Fatty Acids and Heart Failure: The Cardiovascular Health Study. *J. Am. Heart Assoc.* 2018, 7, e010019.
13. Ahmed, S.H.; Koubaa, N.; Kharroubi, W.; Zarrouk, A.; Mnari, A.; Batbout, F.; Gamra, H.; Hammami, S.; Lizard, G.; Hammami, M. Identification of long and very long chain fatty acids, plasmalogen-C16: 0 and phytanic acid as new lipid biomarkers in Tunisian coronary artery disease patients. *Prostaglandins Other Lipid Mediat.* 2017, 131, 49–58.
14. Ahmed, S.H.; Kharroubi, W.; Kaoubaa, N.; Zarrouk, A.; Batbout, F.; Gamra, H.; Najjar, M.F.; Lizard, G.; Hininger-Favier, I.; Hammami, M. Correlation of trans fatty acids with the severity of coronary artery disease lesions. *Lipids Health Dis.* 2018, 17, 52.

15. Guo, M.; Fan, X.; Tuerhongjiang, G.; Wang, C.; Wu, H.; Lou, B.; Wu, Y.; Yuan, Z.; She, J. Targeted metabolomic analysis of plasma fatty acids in acute myocardial infarction in young adults. *Nutr. Metab. Cardiovasc. Dis.* 2021, 31, 3131–3141.
16. Nishizaki, Y.; Shimada, K.; Tani, S.; Ogawa, T.; Ando, J.; Takahashi, M.; Yamamoto, M.; Shinozaki, T.; Miyazaki, T.; Miyauchi, K.; et al. Association between the ratio of serum n-3 to n-6 polyunsaturated fatty acids and acute coronary syndrome in non-obese patients with coronary risk factor: A multicenter cross-sectional study. *BMC Cardiovasc. Disord.* 2020, 20, 160–167.
17. Bittner, D.O.; Goeller, M.; Zopf, Y.; Achenbach, S.; Marwan, M. Early-onset coronary atherosclerosis in patients with low levels of omega-3 fatty acids. *Eur. J. Clin. Nutr.* 2020, 74, 651–656.
18. Bazan, H.A.; Lu, Y.; Jun, B.; Fang, Z.; Woods, T.C.; Hong, S. Circulating inflammation-resolving lipid mediators RvD1 and DHA are decreased in patients with acutely symptomatic carotid disease. *Prostaglandins Leukot. Essent. Fat. Acids* 2017, 125, 43–47.
19. Yang, W.-S.; Chen, Y.-Y.; Chen, P.-C.; Hsu, H.-C.; Su, T.-C.; Lin, H.-J.; Chen, M.-F.; Lee, Y.-T.; Chien, K.-L. Association between Plasma N-6 Polyunsaturated Fatty Acids Levels and the Risk of Cardiovascular Disease in a Community-based Cohort Study. *Sci. Rep.* 2019, 9, 19298.
20. Chien, K.-L.; Lin, H.-J.; Hsu, H.-C.; Chen, P.-C.; Su, T.-C.; Chen, M.-F.; Lee, Y.-T. Comparison of predictive performance of various fatty acids for the risk of cardiovascular disease events and all-cause deaths in a community-based cohort. *Atherosclerosis* 2013, 230, 140–147.
21. Würtz, P.; Havulinna, A.S.; Soininen, P.; Tynkkynen, T.; Prieto-Merino, D.; Tillin, T.; Ghorbani, A.; Artati, A.; Wang, Q.; Tiainen, M.; et al. Metabolite profiling and cardiovascular event risk: A prospective study of 3 population-based cohorts. *Circulation* 2015, 131, 774–785.
22. Dozio, E.; Vianello, E.; Grossi, E.; Menicanti, L.; Schmitz, G.; Romanelli, M.M.C. Plasma fatty acid profile as biomarker of coronary artery disease: A pilot study using fourth generation artificial neural networks. *J. Biol. Regul. Homeost. Agents* 2018, 32, 1007–1013.
23. Muldoon, M.F.; Erickson, K.I.; Goodpaster, B.H.; Jakicic, J.M.; Conklin, S.M.; Sekikawa, A.; Yao, J.K.; Manuck, S.B. Concurrent Physical Activity Modifies the Association between n3 Long-Chain Fatty Acids and Cardiometabolic Risk in Midlife Adults. *J. Nutr.* 2013, 143, 1414–1420.
24. Harris, K.; Oshima, M.; Sattar, N.; Würtz, P.; Jun, M.; Welsh, P.; Hamet, P.; Harrap, S.; Poulter, N.; Chalmers, J.; et al. Plasma fatty acids and the risk of vascular disease and mortality outcomes in individuals with type 2 diabetes: Results from the ADVANCE study. *Diabetologia* 2020, 63, 1637–1647.
25. Shoji, T.; Kakiya, R.; Hayashi, T.; Tsujimoto, Y.; Sonoda, M.; Shima, H.; Mori, K.; Fukumoto, S.; Tahara, H.; Shioi, A.; et al. Serum n-3 and n-6 Polyunsaturated Fatty Acid Profile as an Independent Predictor of Cardiovascular Events in Hemodialysis Patients. *Am. J. Kidney Dis.* 2013, 62, 568–576.
26. Noguchi, H.; Okubo, R.; Hamazaki, K.; Yamashita, A.; Narisawa, T.; Matsuoka, Y.J. Serum polyunsaturated fatty acids and risk of psychiatric disorder at 6 months after acute coronary syndrome: A prospective cohort study. *Prostaglandins Leukot. Essent. Fat. Acids* 2019, 149, 18–23.
27. Ouchi, S.; Miyazaki, T.; Shimada, K.; Sugita, Y.; Shimizu, M.; Murata, A.; Kato, T.; Aikawa, T.; Suda, S.; Shiozawa, T.; et al. Decreased circulating dihomo-gamma-linolenic acid levels are associated with total mortality in patients with acute cardiovascular disease and acute decompensated heart failure. *Lipids Health Dis.* 2017, 16, 150.
28. Li, Z.; Zhang, Y.; Su, D.; Lv, X.; Wang, M.; Ding, D.; Ma, J.; Xia, M.; Wang, D.; Yang, Y.; et al. The opposite associations of long-chain versus very long-chain monounsaturated fatty acids with mortality among patients with coronary artery disease. *Heart* 2014, 100, 1597–1605.
29. Harris, W.S.; Kennedy, K.F.; O'Keefe, J.H.; Spertus, J.A. Red blood cell fatty acid levels improve GRACE score prediction of 2-yr mortality in patients with myocardial infarction. *Int. J. Cardiol.* 2013, 168, 53–59.
30. Salgin, B.; Ong, K.K.; Thankamony, A.; Emmett, P.; Wareham, N.J.; Dunger, D.B. Higher Fasting Plasma Free Fatty Acid Levels Are Associated with Lower Insulin Secretion in Children and Adults and a Higher Incidence of Type 2 Diabetes. *J. Clin. Endocrinol. Metab.* 2012, 97, 3302–3309.
31. Clore, J.N.; Allred, J.; White, D.; Li, J.; Stillman, J. The role of plasma fatty acid composition in endogenous glucose production in patients with type 2 diabetes mellitus. *Metabolism* 2002, 51, 1471–1477.
32. Liu, L.; Li, Y.; Guan, C.; Li, K.; Wang, C.; Feng, R.; Sun, C. Free fatty acid metabolic profile and biomarkers of isolated post-challenge diabetes and type 2 diabetes mellitus based on GC–MS and multivariate statistical analysis. *J. Chromatogr. B* 2010, 878, 2817–2825.

33. Jo, S.; An, W.-S.; Park, Y. Erythrocyte n-3 Polyunsaturated Fatty Acids and the Risk of Type 2 Diabetes in Koreans: A Case-Control Study. *Ann. Nutr. Metab.* 2013, 63, 283–290.
34. Ma, Y.; Xiong, J.; Zhang, X.; Qiu, T.; Pang, H.; Li, X.; Zhu, J.; Wang, J.; Pan, C.; Yang, X.; et al. Potential biomarker in serum for predicting susceptibility to type 2 diabetes mellitus: Free fatty acid 22:6. *J. Diabetes Investig.* 2020, 12, 950–962.
35. Shetty, S.S.; N., S.K.; Shetty, P.K.  $\omega$ -6/ $\omega$ -3 fatty acid ratio as an essential predictive biomarker in the management of type 2 diabetes mellitus. *Nutrition* 2020, 79–80, 110968.
36. Lu, Y.; Wang, Y.; Ong, C.-N.; Subramaniam, T.; Choi, H.W.; Yuan, J.-M.; Koh, W.-P.; Pan, A. Metabolic signatures and risk of type 2 diabetes in a Chinese population: An untargeted metabolomics study using both LC-MS and GC-MS. *Diabetologia* 2016, 59, 2349–2359.
37. Grapov, D.; Adams, S.H.; Pedersen, T.L.; Garvey, W.T.; Newman, J.W. Type 2 Diabetes Associated Changes in the Plasma Non-Esterified Fatty Acids, Oxylipins and Endocannabinoids. *PLoS ONE* 2012, 7, e48852.
38. Yi, L.; He, J.; Liang, Y.; Yuan, D.; Gao, H.; Zhou, H. Simultaneously quantitative measurement of comprehensive profiles of esterified and non-esterified fatty acid in plasma of type 2 diabetic patients. *Chem. Phys. Lipids* 2007, 150, 204–216.
39. Sobczak, A.I.S.; Blindauer, C.A.; Stewart, A.J. Changes in Plasma Free Fatty Acids Associated with Type-2 Diabetes. *Nutrients* 2019, 11, 2022.
40. Ruddock, M.W.; Stein, A.; Landaker, E.; Park, J.; Cooksey, R.C.; McClain, D.; Patti, M.-E. Saturated Fatty Acids Inhibit Hepatic Insulin Action by Modulating Insulin Receptor Expression and Post-receptor Signalling. *J. Biochem.* 2008, 144, 599–607.
41. Miao, Z.; Lin, J.-S.; Mao, Y.; Chen, G.-D.; Zeng, F.-F.; Dong, H.-L.; Jiang, Z.; Wang, J.; Xiao, C.; Shuai, M.; et al. Erythrocyte n-6 Polyunsaturated Fatty Acids, Gut Microbiota, and Incident Type 2 Diabetes: A Prospective Cohort Study. *Diabetes Care* 2020, 43, 2435–2443.
42. Amézaga, J.; Arranz, S.; Urruticoechea, A.; Ugartemendia, G.; Larraioz, A.; Louka, M.; Uriarte, M.; Ferreri, C.; Tueros, I. Altered Red Blood Cell Membrane Fatty Acid Profile in Cancer Patients. *Nutrients* 2018, 10, 1853.
43. Jiao, J.; Kwan, S.-Y.; Sabotta, C.M.; Tanaka, H.; Veillon, L.; Warmoes, M.O.; Lorenzi, P.L.; Wang, Y.; Wei, P.; Hawk, E.T.; et al. Circulating Fatty Acids Associated with Advanced Liver Fibrosis and Hepatocellular Carcinoma in South Texas Hispanics. *Cancer Epidemiol. Biomark. Prev.* 2021, 30, 1643–1651.
44. Chen, Q.; Wang, J.; Lin, J.; Chen, L.; Lin, L.-S.; Pan, L.-Z.; Shi, B.; Qiu, Y.; Zheng, X.-Y.; Chen, F.; et al. Erythrocyte  $\omega$ -3 polyunsaturated fatty acids are inversely associated with the risk of oral cancer: A case-control study. *Nutr. Diabetes* 2020, 10, 35.
45. Shishavan, N.G.; Mohamadkhani, A.; Sepanlou, S.G.; Masoudi, S.; Sharafkhah, M.; Poustchi, H.; Hekmatdoost, A.; Pourshams, A. Circulating plasma fatty acids and risk of pancreatic cancer: Results from the Golestan Cohort Study. *Clin. Nutr.* 2020, 40, 1897–1904.
46. Zaridze, D.G.; Chevchenko, V.E.; Levtschuk, A.A.; Lifanova, Y.E.; Maximovitch, D.M. Fatty acid composition of phospholipids in erythrocyte membranes and risk of breast cancer. *Int. J. Cancer* 1990, 45, 807–810.
47. Hirko, K.A.; Chai, B.; Spiegelman, D.; Campos, H.; Farvid, M.S.; Hankinson, S.E.; Willett, W.C.; Eliassen, A.H. Erythrocyte membrane fatty acids and breast cancer risk: A prospective analysis in the nurses' health study II. *Int. J. Cancer* 2017, 142, 1116–1129.
48. McGee, E.E.; Kim, C.H.; Wang, M.; Spiegelman, D.; Stover, D.G.; Heng, Y.J.; Collins, L.C.; Baker, G.M.; Farvid, M.S.; Schedin, P.; et al. Erythrocyte membrane fatty acids and breast cancer risk by tumor tissue expression of immuno-inflammatory markers and fatty acid synthase: A nested case-control study. *Breast Cancer Res.* 2020, 22, 78.
49. Ruan, X.; Wang, Y.; Zhou, L.; Zheng, Q.; Hao, H.; He, D. Evaluation of Untargeted Metabolomic Strategy for the Discovery of Biomarker of Breast Cancer. *Front. Pharmacol.* 2022, 13, 894099.
50. Kaikkonen, J.E.; Jula, A.; Viikari, J.S.A.; Juonala, M.; Hutri-Kähönen, N.; Kähönen, M.; Lehtimäki, T.; Raitakari, O.T. Associations of Serum Fatty Acid Proportions with Obesity, Insulin Resistance, Blood Pressure, and Fatty Liver: The Cardiovascular Risk in Young Finns Study. *J. Nutr.* 2021, 151, 970–978.
51. Ma, Y.; Qiu, T.; Zhu, J.; Wang, J.; Li, X.; Deng, Y.; Zhang, X.; Feng, J.; Chen, K.; Wang, C.; et al. Serum FFAs profile analysis of Normal weight and obesity individuals of Han and Uygur nationalities in China. *Lipids Health Dis.* 2020, 19, 13.
52. Ni, Y.; Zhao, L.; Yu, H.; Ma, X.; Bao, Y.; Rajani, C.; Loo, L.W.; Shvetsov, Y.B.; Yu, H.; Chen, T.; et al. Circulating Unsaturated Fatty Acids Delineate the Metabolic Status of Obese Individuals. *eBioMedicine* 2015, 2, 1513–1522.



53. Zarrouk, A.; Riedinger, J.-M.; Ahmed, S.H.; Hammami, S.; Chaabane, W.; Debbabi, M.; Ben Ammou, S.; Rouaud, O.; Frih, M.; Lizard, G.; et al. Fatty Acid Profiles in Demented Patients: Identification of Hexacosanoic Acid (C26:0) as a Blood Lipid Biomarker of Dementia. *J. Alzheimer's Dis.* 2015, 44, 1349–1359.
54. Yamagishi, K.; Ikeda, A.; Chei, C.-L.; Noda, H.; Umesawa, M.; Cui, R.; Muraki, I.; Ohira, T.; Imano, H.; Sankai, T.; et al. Serum  $\alpha$ -linolenic and other  $\omega$ -3 fatty acids, and risk of disabling dementia: Community-based nested case–control study. *Clin. Nutr.* 2016, 36, 793–797.
55. Hammouda, S.; Ghzaïel, I.; Khamlaoui, W.; Hammami, S.; Mhenni, S.Y.; Samet, S.; Hammami, M.; Zarrouk, A. Genetic variants in FADS1 and ELOVL2 increase level of arachidonic acid and the risk of Alzheimer's disease in the Tunisian population. *Prostaglandins Leukot. Essent. Fat. Acids* 2020, 160, 102159.
56. Sala-Vila, A.; Satizabal, C.L.; Tintle, N.; van Lent, D.M.; Vasan, R.S.; Beiser, A.S.; Seshadri, S.; Harris, W.S. Red Blood Cell DHA Is Inversely Associated with Risk of Incident Alzheimer's Disease and All-Cause Dementia: Framingham Offspring Study. *Nutrients* 2022, 14, 2408.
57. Tomata, Y.; Larsson, S.C.; Hägg, S. Polyunsaturated fatty acids and risk of Alzheimer's disease: A Mendelian randomization study. *Eur. J. Nutr.* 2019, 59, 1763–1766.
58. Jaček, M.; Hrnčířová, D.; Rambousková, J.; Dlouhý, P.; Tůma, P. Effect of Food with Low Enrichment of N-3 Fatty Acids in a Two-Month Diet on the Fatty Acid Content in the Plasma and Erythrocytes and on Cardiovascular Risk Markers in Healthy Young Men. *Nutrients* 2020, 12, 2207.
59. Seethaler, B.; Basrai, M.; Vetter, W.; Lehnert, K.; Engel, C.; Siniatchkin, M.; Halle, M.; Kiechle, M.; Bischoff, S.C. Fatty acid profiles in erythrocyte membranes following the Mediterranean diet—Data from a multicenter lifestyle intervention study in women with hereditary breast cancer (LIBRE). *Clin. Nutr.* 2019, 39, 2389–2398.
60. Lankinen, M.; Uusitupa, M.; Schwab, U. Genes and Dietary Fatty Acids in Regulation of Fatty Acid Composition of Plasma and Erythrocyte Membranes. *Nutrients* 2018, 10, 1785.
61. Raeisi, M.; Hassanbeigi, L.; Khalili, F.; Kharrati-Shishavan, H.; Yousefi, M.; Mehdizadeh, A. Stearoyl-CoA desaturase 1 as a therapeutic target for cancer: A focus on hepatocellular carcinoma. *Mol. Biol. Rep.* 2022, 49, 8871–8882.
62. Stearoyl-CoA desaturase 1: A potential target for non-alcoholic fatty liver disease?-perspective on emerging experimental evidence. *World J. Hepatol.* 2022, 14, 168–179.
63. Iida, T.; Ubukata, M.; Mitani, I.; Nakagawa, Y.; Maeda, K.; Imai, H.; Ogoshi, Y.; Hotta, T.; Sakata, S.; Sano, R.; et al. Discovery of potent liver-selective stearoyl-CoA desaturase-1 (SCD1) inhibitors, thiazole-4-acetic acid derivatives, for the treatment of diabetes, hepatic steatosis, and obesity. *Eur. J. Med. Chem.* 2018, 158, 832–852.
64. Liu, Z.; Yin, X.; Mai, H.; Li, G.; Lin, Z.; Jie, W.; Li, K.; Zhou, H.; Wei, S.; Hu, L.; et al. SCD rs41290540 single-nucleotide polymorphism modifies miR-498 binding and is associated with a decreased risk of coronary artery disease. *Mol. Genet. Genom. Med.* 2020, 8, e1136.
65. Martín-Núñez, G.M.; Cabrera-Mulero, R.; Rojo-Martínez, G.; Gómez-Zumaquero, J.M.; Chaves, F.J.; de Marco, G.; Soriguer, F.; Castaño, L.; Morcillo, S. Polymorphisms in the SCD1 gene are associated with indices of stearoyl CoA desaturase activity and obesity: A prospective study. *Mol. Nutr. Food Res.* 2013, 57, 2177–2184.
66. Byberg, L.; Kilander, L.; Lemming, E.W.; Michaëlsson, K.; Vessby, B. Cancer death is related to high palmitoleic acid in serum and to polymorphisms in the SCD-1 gene in healthy Swedish men. *Am. J. Clin. Nutr.* 2014, 99, 551–558.
67. Tibori, K.; Orosz, G.; Zámbo, V.; Szelényi, P.; Sarnyai, F.; Tamási, V.; Rónai, Z.; Mátyási, J.; Tóth, B.; Csala, M.; et al. Molecular Mechanisms Underlying the Elevated Expression of a Potentially Type 2 Diabetes Mellitus Associated SCD1 Variant. *Int. J. Mol. Sci.* 2022, 23, 6221.
68. Zámbo, V.; Orosz, G.; Szabó, L.; Tibori, K.; Sipéki, S.; Molnár, K.; Csala, M.; Kereszturi, É. A Single Nucleotide Polymorphism (rs3811792) Affecting Human SCD5 Promoter Activity Is Associated with Diabetes Mellitus. *Genes* 2022, 13, 1784.
69. Yu, G.I.; Mun, K.H.; Yang, S.H.; Shin, D.H.; Hwang, J.S. Polymorphisms in the 3'-UTR of SCD5 gene are associated with hepatocellular carcinoma in Korean population. *Mol. Biol. Rep.* 2018, 45, 1705–1714.
70. Song, Z.; Cao, H.; Qin, L.; Jiang, Y. A Case-Control Study between Gene Polymorphisms of Polyunsaturated Fatty Acid Metabolic Rate-Limiting Enzymes and Acute Coronary Syndrome in Chinese Han Population. *BioMed Res. Int.* 2013, 2013, 928178.
71. Khamlaoui, W.; Mehri, S.; Hammami, S.; Hammouda, S.; Chraeif, I.; Elosua, R.; Hammami, M. Association Between Genetic Variants in FADS1-FADS2 and ELOVL2 and Obesity, Lipid Traits, and Fatty Acids in Tunisian Population. *Clin. Appl. Thromb.* 2020, 26.
72. Sun, C.; Zou, M.; Wang, X.; Xia, W.; Ma, Y.; Liang, S.; Hao, Y.; Wu, L.; Fu, S. FADS1-FADS2 and ELOVL2 gene polymorphisms in susceptibility to autism spectrum disorders in Chinese children. *BMC Psychiatry* 2018, 18, 283.



73. Valizadeh, M.; Aghasizadeh, M.; Nemati, M.; Hashemi, M.; Aghaee-Bakhtiari, S.H.; Zare-Feyzabadi, R.; Esmaily, H.; Ghazizadeh, H.; Sahebi, R.; Ahangari, N.; et al. The association between a Fatty Acid Binding Protein 1 (FABP1) gene polymorphism and serum lipid abnormalities in the MASHAD cohort study. *Prostaglandins Leukot. Essent. Fat. Acids* 2021, 172, 102324.
74. Xue, H.; Zhao, H.; Liu, X.; Zhao, Y.-R.; Chen, Z.-J.; Ma, J. Association of single-nucleotide polymorphisms rs2197076 and rs2241883 of FABP1 gene with polycystic ovary syndrome. *J. Assist. Reprod. Genet.* 2015, 33, 75–83.
75. Mansego, M.L.; Martínez, F.; Martínez-Larrad, M.T.; Zabena, C.; Rojo, G.; Morcillo, S.; Soriguer, F.; Martín-Escudero, J.C.; Serrano-Ríos, M.; Redon, J.; et al. Common Variants of the Liver Fatty Acid Binding Protein Gene Influence the Risk of Type 2 Diabetes and Insulin Resistance in Spanish Population. *PLoS ONE* 2012, 7, e31853.
76. Mozaffari, M.; Ghayour-Mobarhan, M.; Zare-Feyzabadi, R.; Valizadeh, M. FABP1 gene variant associated with risk of metabolic syndrome. *Comb. Chem. High Throughput Screen.* 2022, 25, 1355–1360.
77. Peng, X.-E.; Wu, Y.-L.; Lu, Q.-Q.; Hu, Z.-J.; Lin, X. Two genetic variants in FABP1 and susceptibility to non-alcohol fatty liver disease in a Chinese population. *Gene* 2012, 500, 54–58.
78. Fisher, E.; Li, Y.; Burwinkel, B.; Kühr, V.; Hoffmann, K.; Möhlig, M.; Spranger, J.; Pfeiffer, A.; Boeing, H.; Schrezenmeir, J.; et al. Preliminary Evidence of FABP2 A54T Polymorphism Associated with Reduced Risk of Type 2 Diabetes and Obesity in Women from a German Cohort. *Horm. Metab. Res.* 2006, 38, 341–345.
79. Shabana; Hasnain, S. The fatty acid binding protein 2 (FABP2) polymorphism Ala54Thr and obesity in Pakistan: A population based study and a systematic meta-analysis. *Gene* 2015, 574, 106–111.
80. Liu, Y.; Wu, G.; Han, L.; Zhao, K.; Qu, Y.; Xu, A.; Huang, Q. Association of the FABP2 Ala54Thr polymorphism with type 2 diabetes, obesity, and metabolic syndrome: A population-based case-control study and a systematic meta-analysis. *Genet. Mol. Res.* 2015, 14, 1155–1168.
81. Peng, X.; Zhang, L.; Wang, Q.; Cui, X. Study on the relationship between FABP2 Ala54Thr polymorphism and the risk of non-alcoholic fatty liver diseases. *Wei Sheng Yan Jiu* 2009, 38, 401–404.
82. Khattab, S.A.; Abo-Elmatty, D.; Ghattas, M.H.; Mesbah, N.; Mehanna, E.T. Intestinal fatty acid binding protein Ala54Thr polymorphism is associated with peripheral atherosclerosis combined with type 2 diabetes mellitus. *J. Diabetes* 2016, 9, 821–826.
83. Li, Z.; Ni, C.-L.; Niu, W.-Y.; Chang, B.-C.; Chen, L.-M. The intestinal fatty acid binding protein-2 Ala54Thr polymorphism is associated with diabetic retinopathy in Chinese population. *Diabetol. Metab. Syndr.* 2015, 7, 23.
84. Canani, L.H.; Capp, C.; Ng, D.P.; Choo, S.G.; Maia, A.L.; Nabinger, G.B.; Santos, K.; Crispim, D.; Roisemberg, I.; Krolewski, A.S.; et al. The Fatty Acid-Binding Protein-2 A54T Polymorphism Is Associated With Renal Disease in Patients With Type 2 Diabetes. *Diabetes* 2005, 54, 3326–3330.
85. Abbas, S.; Raza, S.T.; Chandra, A.; Rizvi, S.; Ahmed, F.; Eba, A.; Mahdi, F. Association of ACE, FABP2 and GST genes polymorphism with essential hypertension risk among a North Indian population. *Ann. Hum. Biol.* 2014, 42, 461–469.
86. Sikhayeva, N.; Iskakova, A.; Saigi-Morgui, N.; Zholdybaeva, E.; Eap, C.-B.; Ramanculov, E. Association between 28 single nucleotide polymorphisms and type 2 diabetes mellitus in the Kazakh population: A case-control study. *BMC Med. Genet.* 2017, 18, 76.
87. An, L.; Jiang, H.; Tang, R.-N. The ACACB gene rs2268388 polymorphism is associated with nephropathy in Caucasian patients with diabetes: A meta-analysis. *Ren. Fail.* 2015, 37, 925–928.
88. Tang, S.C.W.; Leung, V.T.M.; Chan, L.Y.Y.; Wong, S.S.H.; Chu, D.W.S.; Leung, J.C.K.; Ho, Y.W.; Lai, K.N.; Ma, L.; Elbein, S.C.; et al. The acetyl-coenzyme A carboxylase beta (ACACB) gene is associated with nephropathy in Chinese patients with type 2 diabetes. *Nephrol. Dial. Transplant.* 2010, 25, 3931–3934.
89. Shah, V.N.; Cheema, B.S.; Sharma, R.; Khullar, M.; Kohli, H.S.; Ahluwalia, T.S.; Mohan, V.; Bhansali, A. ACAC $\beta$  gene (rs2268388) and AGTR1 gene (rs5186) polymorphism and the risk of nephropathy in Asian Indian patients with type 2 diabetes. *Mol. Cell. Biochem.* 2012, 372, 191–198.
90. Chan, G.C.W.; Zhi, H.; Hicks, P.J.; Freedman, B.I.; Tang, S.C.W. Acetyl-coenzyme A carboxylase beta gene polymorphism does not predict cardiovascular risk susceptibility in Chinese type 2 diabetic individuals. *Nephrology* 2022, 27, 404–409.
91. Riancho, J.; Vázquez, L.; García-Pérez, M.; Sainz, J.; Olmos, J.; Hernández, J.; Pérez-López, J.; Amado, J.; Zarrabeitia, M.; Cano, A.; et al. Association of ACACB polymorphisms with obesity and diabetes. *Mol. Genet. Metab.* 2011, 104, 670–676.

