

# Antioxidant Activities of *Mentha* spp. Essential Oils

Subjects: Plant Sciences

Contributor: Annarita Stringaro, Marisa Colone, Letizia Angiolella

Plant essential oils (EOs) are produced predominantly using steam distillation, but can also be generated using fermentation, crushing, extraction, hydrolysis, and airing. EOs are used extensively in cosmetics in many different aspects as perfumes, in antiseptic applications, and in domestic cleaning products. The essential oils of *Mentha* (the Lamiaceae family) have been extensively studied for their biological actions.

Keywords: essential oil ; *Mentha* spp. ; antioxidant

---

## 1. Introduction

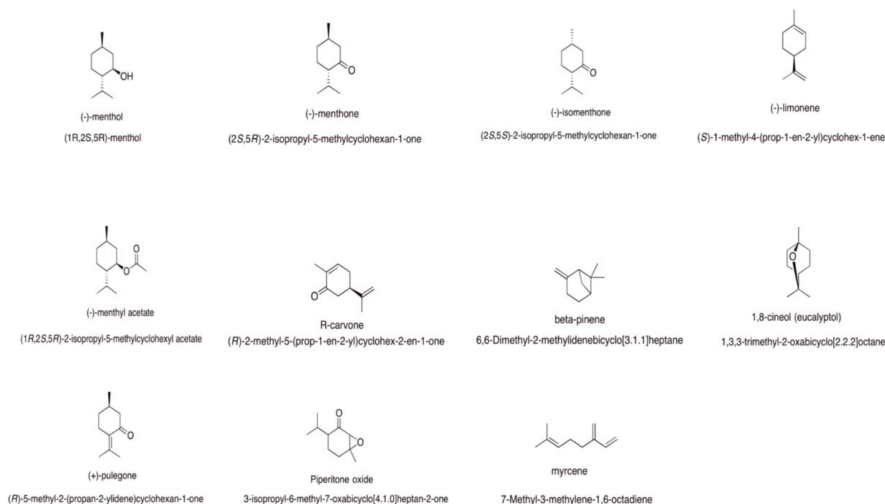
Plant essential oils (EOs) are produced predominantly using steam distillation, but can also be generated using fermentation, crushing, extraction, hydrolysis, and airing <sup>[1]</sup>. EOs are used extensively in cosmetics in many different aspects as perfumes, in antiseptic applications, and in domestic cleaning products <sup>[2][3]</sup>. They are volatile liquids or semi-liquids <sup>[4]</sup> that are limpid, but are rarely colored, and are soluble in organic solvents. All of the organs of the plants can synthesize EOs, which are stored in secretory compartments as cavities, canals, epidermic cells, or glandular trichomes. EOs are complex mixtures of terpenoides containing sesquiterpene and monoterpene, and their oxygenated derivatives. EOs may also incorporate a variety of other molecules such as fatty acids, oxides, and sulfur derivatives <sup>[5]</sup>. Both the terpenoid and phenylpropanoid families, which are sometimes identified as the principal constituents of several EOs, can constitute 85% of the total concentration of the oil. There are about 3000 well-recognized EOs, of which 300 are widely sold <sup>[6]</sup>. Various factors influence the chemical compositions of EOs, such as their geographic location, the seasonal period in which they are collected, the soil composition and cultivation method, their storage, and the oil extraction method <sup>[7][8]</sup>. The high level of interest in research regarding EOs is due to their many biological and medical properties <sup>[9]</sup>. They are generally recognized as safe, and they can act synergistically with other compounds, which are promising factors for their use as bioactive compounds <sup>[10]</sup>.

Based on the number of search results in the PubMed database, the published studies on their antimicrobial, antioxidant, and anti-tumoral activities are 2671, 1186, and 108, respectively <sup>[11]</sup>.

### *Mentha* spp. Essential Oils

Many aromatic plants used in medicine, food, and pharmaceutical industries belong to the Lamiaceae family. In this family, *Mentha* is a well-known genus that includes 25–30 species that are generally grown in temperate areas around the world, particularly in Europe, North America, North Africa, Asia Minor, the northern parts of Iran, and near the east (Syria, Ethiopia). *Mentha* spp. includes plants that exhibit important biological activities and have high morphological variability and a great chemical diversity with respect to their EOs <sup>[10]</sup>.

For this reason, *Mentha*-derived EOs have been used as a folk remedy for respiratory diseases such as bronchitis, sinusitis, tuberculosis, and the common cold <sup>[12]</sup>. *Mentha* acts as a good expectorant. The chemistry of *Mentha* EOs is complex and high variable. The main constituents of the most commonly used *Mentha* EOs revealed by gas chromatography–mass spectrometry (GC-MS) analysis show the presence of menthol, menthone, limonene, isomenthone, menthyl acetate, carvone,  $\beta$ -pinene, 1,8-cineole, pulegone, piperitone oxide, and micene. Each species has a characteristic prevalent compound (**Figure 1**). Studies that have already been carried out with *Mentha* spp. have shown antimicrobial activity related to some species of this genus. The most cited activities of the plant are its antiviral, antibacterial, antifungal <sup>[13]</sup>, high antioxidant, and cytotoxic properties <sup>[14]</sup>, and also other properties such as its antinociceptive, anti-inflammatory, and antiallergic qualities <sup>[15]</sup>.



**Figure 1.** Chemical structures of the main components of *Mentha* spp. essential oils (EOs).

## 2. Antioxidant Properties

In recent years, there has been an increasing interest in the consumption of EOs as natural antioxidants [16]. It is well-known that reactive oxygen species (ROS) cause damage to cellular macromolecules, and they are implicated in the development of many human diseases. Under many pathological conditions the oxidation–reduction (redox) potential imbalance cannot remove excessive amounts of ROS [17]. Oxidant species such as hydrogen peroxide ( $\text{H}_2\text{O}_2$ ) and superoxide ( $\text{O}_2^-$ ) are produced following the phagocytosis of the pathogen by these cells as part of their machinery to respond to harmful insults [18]. Excessive nitric oxide (NO) production and increased levels of prooxidant species may lead to damage and poor perfusion of the vital organs of the host, contributing to multiple organ failure; thus, to counteract this response, antioxidant pathways are activated [19]. Free radicals generated by damaged membranes, when combined with EOs, produce radicals with scavenging activity.

Natural antioxidants such as phenolic compounds can be found in many plants. The antioxidant activity of essential oils does not always depend on its main component but can be modulated by other components [20].

Antioxidant activity can be evaluated using various methods, and analytical tools are utilized for measuring antioxidant content and total antioxidant capacity evaluation. The methods of antioxidant capacity evaluation include counting spectrometry, chromatography, and electrochemical techniques [21]. Generally, the most frequently used methods are 2,2-diphenyl-1-picrylhydrazyl, (DPPH), 2,2'-azino-bis (3-ethylbenzthiazoline-6-sulphonic acid), (ABTS), and others [22].

The main reaction occurring in spectrometric techniques is between a radical, radical cation, or a complex and an antioxidant molecule donor of a hydrogen atom. DPPH is a stable free radical, and the assay is based on electron transfer that produces a purple solution in ethanol, with an absorption band with a maximum of around 520 nm. When DPPH reacts with a hydrogen donor, it generates the reduced molecular form (DPPH), and the purple color disappears. The antioxidant concentration is linearly correlated with the absorbance diminution. The standard antioxidant used is Trolox. The standard curve was linear between 25–800 mM Trolox [23].

The bioluminescent antioxidant capacity assay also uses the redox couple ABTS+/ABTS. The ABTS cation radical (ABTS+) [24], which absorbs at 743 nm (giving a green color), is formed by the loss of an electron by the nitrogen atom of ABTS. In the presence of Trolox (or another hydrogen-donating antioxidant), the nitrogen atom quenches the hydrogen atom, causing the solution's decolorization. ABTS can be oxidized by potassium persulfate [24][25] or manganese dioxide [26]. The standard curve was linear between 25–600  $\mu\text{M}$  of Trolox. Unfortunately, the evaluation of the antioxidant activity of EOs remains a critical issue, because many “tests” are unsuitable and provide contradictory results [27].

*Mentha piperita* L. is a plant native to the Mediterranean region that is popularly known as peppermint; it is used medicinally for its antiproliferative and antioxidant actions. The plant is also used worldwide, especially in the perfume and food industries, for its taste and fragrance.

*Mentha piperita* EO possesses antiradical activity with respect to DPPH and hydroxyl ( $\text{OH}^\cdot$ ) radicals; indeed, Schmidt et al. [28] have reported the antiradical activity of this EO for DPPH as ( $\text{IC}_{50}$ ), while some authors have reported its radical scavenger activity against the ABTS radical [29].

Recently, similar results were confirmed by Sun et al. [30], specifically the action of peppermint EO as a scavenger of hydroxyl radicals, and the potential for it to be an antioxidant at concentrations  $\geq 200$   $\mu\text{g/mL}$ . da Silva Ramos et al. [31] reported an antioxidant activity of  $79.9 \pm 1.6\%$  and  $\text{IC}_{50} = 414.6$   $\mu\text{g/mL}$ . On the contrary, other researchers [32] have described low antioxidant activity. More probably, the antioxidant actions of *M. piperita* may be due to the presence of phenolic constituents in its leaves, including rosmarinic acid and different flavonoids such as rutin, naringin, eriocitrin, luteolin, and hesperidin, which are present in aqueous extracts [33][34], but not in the essential oil. Furthermore, *M. piperita* EO is associated with increased levels of intracellular ROS, which is indicative of an apoptotic process [35] without the loss of the plasma membrane integrity.

*M. pulegium*, another species of *Mentha*, has been used in traditional medicine to treat numerous illnesses, such as microbial infections and oxidative stress. Kamkar et al. and Cherrat et al. [36][37] have described the lower antioxidative activity of the *M. pulegium* EO with respect to aqueous or methanol extracts. This difference could be due to a lack of diverse antioxidants in the EO. On the contrary, some authors [38][39] have observed a good radical scavenging ability of *M. pulegium* EO compared with ascorbic acid and Trolox.

Regarding other species of the genus *Mentha* used as antioxidants, *Mentha spicata* EO has been used. In this case, different results were reported; some authors described the antioxidant activity of *M. spicata* EO [40], while others [41] described a weak antioxidant activity.

*Mentha longifolia* L. (*M. longifolia*) is known as a wild mint named Puneh, and is a fast-growing and perennial herb that creeps along an underground rootstock, which can grow to 1–2 m tall. Eissa et al. [42] revealed that *M. longifolia* EOs possesses the highest scavenging activity against peroxy radicals.

*Mentha suaveolens* Ehrh is a communal wild plant that is found near streams, bogs, and humid places. There are different subspecies, each including several varieties. El-Askary et al. [43] reported a potent antioxidant activity in vivo, which was about 96% relative to vitamin E, while Ferreira et al. [44] described the AChE inhibitory capacity as higher than 50% in the essential oil fraction of *M. suaveolens*. The antioxidant capacity in this case is due to piperitone oxide being present at 88% [45]. Other authors have reported no relevant antioxidant activity for this species [46]. Some *Mentha* spp. have not been assessed for antioxidant activity as yet. **Table 1** reports antioxidant information about *Mentha* spp. EOs.

**Table 1.** Radical scavenging activity of *Mentha* essential oils (EOs). ABTS: 2,2'-azino-bis 3-ethylbenzthiazoline-6-sulphonic acid, DPPH: 2,2-diphenyl-1-picrylhydrazyl.

EOs	DPPH Activity	ABTS Activity	References
<i>M. piperita</i>	860 $\mu\text{g/mL}$	-	[28]
	$57.9 \pm 1.34\%$	$80.6 \pm 1.45\%$	[29]
	600 $\mu\text{g/mL}$	-	[30]
	540 $\mu\text{g/mL}$	-	[31]
	$11.289 \pm 0.514$ $\mu\text{g/g}$	$0.154 \pm 0.006$ $\text{mmol/g}$	[32]
	$14736 \pm 156$ $\mu\text{g/mL}$	-	[36]
<i>M. pulegium</i>	$30.38 \pm 0.8\%$	-	[37]
	69.60 $\mu\text{g/mL}$	-	[38]
	$321.41 \pm 2.53$ $\mu\text{g/mL}$	-	[39]
<i>M. spicata</i>	3 $\mu\text{g/mL}$	-	[40]

EOs	DPPH Activity	ABTS Activity	References
	3450 ± 172.5 µg/mL	40.2 ± 0.2 µg/mL	[41]
<b><i>M. longifolia</i></b>	57.4 µg/mL	-	[42]
<b><i>M. suaveolens</i></b>	31 µg/mL	-	[43]
	52.4 ± 2.5%	-	[44]

## References

1. Prabuseenivasan, S.; Jayakumar, M.; Ignacimuthu, S. In vitro antibacterial activity of some plant essential oils. *BMC Complement. Altern. Med.* 2006, 6, 39–50.
2. Aburjai, T.; Natsheh, F.M. Plants used in cosmetics. *Phytother. Res.* 2003, 17, 987–1000.
3. Wallace, R.J. Antimicrobial properties of plant secondary metabolites. *Proc. Nutr. Soc.* 2004, 63, 621–629.
4. Bakkali, F.; Averbeck, S.; Averbeck, D.; Idaomar, M. Biological effects of essential oils—A review. *Food Chem. Toxicol.* 2008, 46, 446–475.
5. Nakatsu, T.; Lupo, A.T.; Chinn, J.W.; Kang, R.K.L. *Studies in Natural Products Chemistry*; Elsevier: New York, NY, USA, 2000; Volume 21, pp. 571–631.
6. Burt, S. Essential oils: Their antibacterial properties and potential applications in foods—A review. *Int. J. Food Microbiol.* 2004, 94, 223–253.
7. Edris, A. Pharmaceutical and therapeutic potentials of essential oils and their individuals' volatile constituents. A review. *Phytother. Res.* 2007, 21, 308–323.
8. Garzoli, S.; Pirolli, A.; Vavala, E.; Di Sotto, A.; Sartorelli, G.; Bozovic, M.; Angiolella, L.; Mazzanti, G.; Pepi, F.; Ragno, R. Multidisciplinary approach to determine the optimal time and period to extract the essential oil from *Mentha suaveolens* ehrh. *Molecules* 2015, 20, 9640–9655.
9. Nieto, G.; Ros, G.; Castillo, J. Antioxidant and Antimicrobial Properties of Rosemary (*Rosmarinus officinalis*, L.): A Review. *Medicines* 2018, 5, 98.
10. Tucker, A.O.; Naczi, R.F.C. *Mentha: An overview of its classification and relationships*. In *Mint: The Genus Mentha: Medicinal and Aromatic Plants-Industrial Profiles*; Lawrence, B.M., Ed.; CRC Press: Boca Raton, FL, USA, 2006; p. 3.
11. Sakkas, H.; Papadopoulou, C. Antimicrobial Activity of Basil, Oregano, and Thyme Essential Oils. *J. Microbiol. Biotechnol.* 2017, 27, 429–438.
12. Peixoto, I.T.A.; Furlanetti, V.F.; Anibal, P.C.; Duarte, M.C.T.; Höfling, J.F. Potential pharmacological and toxicological basis of the essential oil from *Mentha* spp. *Rev. Ciênc. Farm. Básica Apl.* 2009, 30, 235–239.
13. Chávez-González, M.L.; Rodríguez-Herrera, R.; Aguilar, C.N. Essential oils: A natural alternative to combat antibiotics resistance antibiotic resistance in mechanisms and new antimicrobial approaches. In *Antibiotic Resistance*; Kateryna, K., Mahendra, R., Eds.; Academic Press: Cambridge, MA, USA, 2016; pp. 227–237.
14. Sharma, V.; Hussain, S.; Gupta, M.; Saxena, A. In vitro anticancer activity of extracts of *Mentha* spp. against human cancer cells. *Indian J. Biochem. Biophys.* 2014, 51, 416–419.
15. Amabeoku, G.J.; Erasmus, S.J.; Ojewole, J.A.; Mukinda, J.T. Antipyretic and antinociceptive properties of *Mentha longifolia* Huds. (Lamiaceae) leaf aqueous extract in rats and mice. *Meth. Find. Exp. Clin. Pharmacol.* 2009, 31, 645–649.
16. Yadegarinia, D.; Gachkar, L.; Rezaei, M.B.; Taghizadeh, M.; Astaneh, S.A.; Rasooli, I. Biochemical activities of Iranian *Mentha piperita* L. and *Myrtus communis* L. essential oils. *Phytochemistry* 2006, 67, 1249–1255.
17. Gonzales-Burgos, E.; Gomez-Serranillos, M.P. Terpene compounds in nature. A review of their potential antioxidant activity. *Curr. Med. Chem.* 2012, 19, 5319–5341.
18. Kaufmann, H.; Dorhoi, A. Molecular determinants in phagocyte-bacteria interactions. *Immunity* 2016, 44, 476–491.

19. Nathan, C.; Cunningham-Bussell, A. Beyond oxidative stress: An immunologist's guide to reactive oxygen species. *Nat. Rev. Immunol.* 2013, 13, 349–361.
20. Dawidowicz, A.L.; Olszowy, M. Does antioxidant properties of the main component of essential oil reflect its antioxidant properties? The comparison of antioxidant properties of essential oils and their main components. *Nat. Prod. Res.* 2014, 28, 1952–1963.
21. Pisoschi, A.M.; Negulescu, G.P. Methods for total antioxidant activity determination: A review. *Biochem. Anal. Biochem.* 2011, 1.
22. Olszowy, M.; Dawidowicz, A.L. Essential oils as antioxidants: Their evaluation by DPPH, ABTS, FRAP, CUPRAC, and  $\beta$ -carotene bleaching methods. *Monatsh. Chem.* 2016, 147, 2083–2091.
23. Thaipong, K.; Boonprakob, U.; Crosby, K.; Cisneros-Zevallos, L.; Byrne, D.H. Comparison of ABTS, DPPH, FRAP, and ORAC assays for estimating from guava fruit extracts. *J. Food Compos. Anal.* 2006, 19, 669–675.
24. Marc, F.; Davin, A.; Deglène-Benbrahim, L.; Ferrand, C.; Baccaunaud, M.; Fritsch, P. Studies of several analytical methods for antioxidant potential evaluation in food. *Med. Sci.* 2004, 20, 458–463.
25. Pellegrini, N.; Serafini, M.; Colombi, B.; Del Rio, D.; Salvatore, S.; Bianchi, M.; Brighenti, F. Total antioxidant capacity of plant foods, beverages and oils consumed in Italy assessed by three different in vitro assays. *J. Nutr.* 2003, 133, 2812–2819.
26. Su, L.; Yin, J.J.; Charles, D.; Zhou, K.; Moore, J.; Yu, L.L. Total phenolic contents, chelating capacities, and radical-scavenging properties of black peppercorn, nutmeg, rosehip, cinnamon and oregano leaf. *Food Chem.* 2007, 100, 990–997.
27. Amorati, R.; Foti, M.C.; Valgimigli, L. Antioxidant activity of essential oils. *J. Agric. Food Chem.* 2013, 61, 10835–10847.
28. Schmidt, E.; Bail, S.; Buchbauer, G.; Stoilova, I.; Atanasova, T.; Stoyanova, A.; Krastanov, A.; Jirovetz, L. Chemical composition, olfactory evaluation and antioxidant effects of essential oil from *Mentha x piperita*. *Nat. Prod. Commun.* 2009, 4, 1107–1112.
29. Yang, S.A.; Jeon, S.K.; Lee, E.J.; Shim, C.H.; Lee, I.S. Comparative study of the chemical composition and antioxidant activity of six essential oils and their components. *Nat. Prod. Res.* 2010, 24, 140–151.
30. Sun, Z.; Wang, H.; Wang, J.; Zhou, L.; Yang, P. Chemical composition and anti-inflammatory, cytotoxic and antioxidant activities of essential oil from leaves of *Mentha piperita* grown in China. *PLoS ONE* 2014, 9, e114767.
31. da Silva Ramos, R.; Lobato Rodrigues, A.B.; Ferreira Farias, A.L.; Simões, R.C.; Pinheiro, M.T.; dos Anjos Ferreira, R.M.; Costa Barbosa, L.M.; Picanço Souto, R.N.; Fernandes, J.B.; da Silvas Santos, L.; et al. Chemical composition and in vitro antioxidant, cytotoxic, antimicrobial, and larvicidal activities of the essential oil of *Mentha piperita* L. (Lamiaceae). *Sci. World J.* 2017, 2017.
32. Pellegrini, M.; Ricci, A.; Serio, A.; Chaves-López, C.; Mazzarrino, G.; D'Amato, S.; Lo Sterzo, C.; Paparella, A. Characterization of Essential Oils Obtained from Abruzzo Autochthonous Plants: Antioxidant and Antimicrobial Activities Assessment for Food Application. *Foods* 2018, 7, 19.
33. Dorman, H.; Koşar, M.; Başer, K.; Hiltunen, R. Phenolic profile and antioxidant evaluation of *Mentha x piperita* L. (peppermint) extracts. *Nat. Prod. Commun.* 2009, 4, 535–542.
34. Sroka, Z.; Fecka, I.; Cisowski, W. Antiradical and Anti-H<sub>2</sub>O<sub>2</sub> properties of polyphenolic compounds from an aqueous peppermint extract. *Z. Naturforsch. C* 2005, 60, 826–832.
35. Ferreira, P.; Cardoso, T.; Ferreira, F.; Fernandes-Ferreira, M.; Piper, P.; Sousa, M.J. *Mentha piperita* essential oil induces apoptosis in yeast associated with both cytosolic and mitochondrial ROS-mediated damage. *FEMS Yeast Res.* 2014, 14, 1006–1014.
36. Kamkar, A.; Javan, A.J.; Asadi, F.; Kamalinejad, M. The antioxidative effect of Iranian *Mentha pulegium* extracts and essential oil in sunflower oil. *Food Chem. Toxicol.* 2010, 48, 1796–1800.
37. Cherrat, L.; Espina, L.; Bakkali, M.; Pagan, R.; Laglaoui, A. Chemical composition, antioxidant and antimicrobial properties of *Mentha pulegium*, *Lavandula stoechas* and *Satureja calamintha* Scheele essential oils and an evaluation of their bactericidal effect in combined processes. *Innov. Food Sci. Emerg. Technol.* 2014, 22, 221–229.
38. Abdelli, M.; Moghrani, H.; Aboun, A.; Maachi, R. Algerian *Mentha pulegium* L. leaves essential oil: Chemical composition, antimicrobial, insecticidal and antioxidant activities. *Ind. Crops Prod.* 2016, 94, 197–205.
39. Bouyahya, A.; Et-Touys, A.; Bakri, Y.; Talbaui, A.; Fellah, H.; Abrini, J.; Dakka, N. Chemical composition of *Mentha pulegium* and *Rosmarinus officinalis* essential oils and their antileishmanial, antibacterial and antioxidant activities. *Microb. Pathog.* 2017, 111, 41–49.

40. Snoussi, M.; Noumi, E.; Trabelsi, N.; Flamini, G.; Papetti, A.; De Feo, V. *Mentha spicata* Essential Oil: Chemical Composition, Antioxidant and Antibacterial Activities against Planktonic and Biofilm Cultures of *Vibrio* spp. Strains. *Molecules* 2015, 20, 14402–14424.
41. Bardaweel, S.K.; Bakchiche, B.; AL-Salamat, H.A.; Rezzoug, M.; Gherib, A.; Flamini, G. Chemical composition, antioxidant, antimicrobial and Antiproliferative activities of essential oil of *Mentha spicata* L. (Lamiaceae) from Algerian Saharan atlas. *BMC Complement. Altern. Med.* 2018, 18.
42. Eissa, T.F.; González-Burgos, E.; Carretero, M.E.; Gómez-Serranillos, M.P. Compositional analysis and in vitro protective activity against oxidative stress of essential oils from egyptian plants used in traditional medicine. *Nat. Prod. Commun.* 2014, 9, 1377–1382.
43. El-Askary, H.I.; El-Kashoury, E.A.; Kandil, Z.A.; Salem, M.A.; Ezzat, S.M. Biological activity and standardization of the ethanolic extract of the aerial parts of *Mentha suaveolens* Ehrh. *World J. Pharm. Pharm. Sci.* 2014, 3, 223–241.
44. Ferreira, A.; Proenc, C.; Serralheiro, M.L.M.; Araújo, M.E.M. The in vitro screening for acetylcholinesterase inhibition and antioxidant activity of medicinal plants from Portugal. *J. Ethnopharmacol.* 2006, 108, 31–37.
45. Sitzmann, J.; Habegger, R.; Schnitzler, W.H.; Grassmann, J. Comparative analysis of antioxidant activities of fourteen *Mentha* essential oils and their components. *Chem. Biodivers.* 2014, 11, 1978–1989.
46. Spagnoletti, A.; Guerrini, A.; Tacchini, M.; Vinciguerra, V.; Leone, C.; Maresca, I.; Simonetti, G.; Sacchetti, G.; Angiolella, L. Chemical Composition and Bio-efficacy of Essential Oils from Italian Aromatic Plants: *M. suaveolens*, *C. capitatus*, *O. hirtum* and *R. officinalis*. *Nat. Prod. Commun.* 2016, 11, 1517–1520.

---

Retrieved from <https://encyclopedia.pub/entry/history/show/126520>