

HDAC6

Subjects: **Oncology**

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Histone deacetylase (HDAC) 6 is a zinc-dependent enzyme of HDAC class IIb. HDAC6 is unique within the HDAC family due to a particular structure giving it unique biological functions implicated in all major cell pathways. This isoenzyme is mainly active in the cytoplasm and possesses two functional catalytic sites and an ubiquitin-binding domain. The deacetylase functions of HDAC6 targets multiple substrates including essentially α -tubulin and heat shock protein (HSP)90 α which are key factors in cell regulatory networks through the regulation of the microtubule network and many protein functions, respectively. Accordingly, several studies have highlighted the role of HDAC6 in various pathological conditions. For instance HDAC6 overexpression frequently correlates with tumorigenesis and favor cell survival and metastasis. Therefore, HDAC6 represents an interesting potential therapeutic target.

histone deacetylase 6 inhibitor

personalized treatment

heat shock protein 90 α

leukemia stem cells

imatinib resistance

targeted therapy

1. Introduction

Carcinogenesis is a multistep process whereby normal cells are transformed into malignant cells. The process is characterized by major biological changes shared by most neoplastic cells called hallmarks of cancer. These transformational events relies on multiple alterations at genetic and epigenetic levels leading to abnormal cell growth ^[1].

Over the past years protein lysine acetylation has emerged as a key post-translational modification in the coordination of tightly regulated biological functions and alterations of the acetylome profiles are associated with various pathological conditions such as cancer.

The acetylation status of lysine residues within histone and non-histone proteins is finely tuned by the concert action of histone acetyltransferases (HATs) and histone deacetylases (HDACs) catalyzing the addition and removal of the acetyl groups, respectively. Recently, there was a particular focus on HDAC6 coming from its unique properties to control multiple cellular pathways linked to cell growth, survival, and migration. Accordingly, the use of HDAC6 inhibitors alone or in combination with additional chemotherapeutic agents appear as a promising strategy to treat various cancers.

2. Histone Deacetylase 6

The HDAC6 protein is part of the HDAC family, which are enzymes catalyzing the deacetylation of proteins, which corresponds to the removal of an acetyl group from lysine residues [2]. The 18 HDACs found in mammals are divided into four classes according to their sequence homology. For classes I (HDAC1, 2, 3, and 8), IIa (HDAC4, 5, 7, and 9), IIb (HDAC6 and 10), and IV (HDAC11), the deacetylation of lysine occurs through a transfer of charge, and their essential component is a zinc ion (Zn^{2+}) present at the bottom of the catalytic pocket of HDAC enzymes [3]. For class III [sirtuins (SIRT) 1-7] HDACs, the presence of a cofactor, nicotinamide adenine dinucleotide (NAD^{+}), is essential for the reaction [4][5].

Class I HDACs are ubiquitously present in many human cell lines and tissues, while class II HDACs exhibit a specific expression profile in certain human tissues such as the heart (HDAC5), the breast (HDAC6), the ovary (HDAC7 and 9), and the kidney (HDAC10) [6][7].

2.1. Structure

Here, we will focus more specifically on HDAC6 belonging to class IIb. This enzyme is the only HDAC to possess two functional active catalytic sites, and has a nuclear localization sequence, a nuclear export sequence, and a repetitive region of eight consecutive serine-glutamic acid tetradecapeptides, a cytoplasmic retention signal, and is mainly present in the cytoplasm [8]. HDAC6 also has a C-terminal ubiquitin-binding domain required when binding to poly-ubiquitinated proteins (Figure 1).

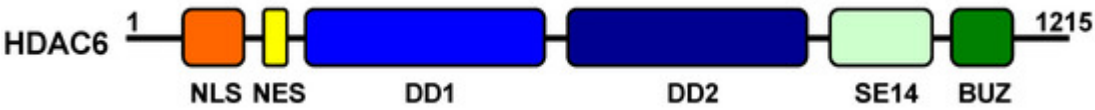


Figure 1. Protein structure of histone deacetylase 6 (HDAC6). The HDAC6 protein consists of 1215 amino acids. It has a nuclear localization sequence (NLS), a nuclear export sequence (NES), two functional active catalytic (SC) sites, a cytoplasmic retention signal of eight consecutive serine-glutamic acid tetradecapeptides (SE14), and a ubiquitin-binding domain (BUZ) at the C-terminal.

2.2. Function

The HDAC6 protein deacetylates many substrates [9] (Table 1) including α -tubulin, cortactin, and heat shock protein (HSP)90 α , and is thus involved in many cell processes, some of which are described below [10].

Table 1. List of substrates specifically deacetylated by HDAC6.

Substrates	Localization of the Substrate	Deacetylated Lysine(s)	Function of the Deacetylated Substrate	Interaction Domains of HDAC6	Reference
14-3-3 ζ	Cytoplasm and nucleus	49, 120	Regulation of protein binding Bad and AS160	ND	[11]

Substrates	Localization of the Substrate	Deacetylated Lysine(s)	Function of the Deacetylated Substrate	Interaction Domains of HDAC6	Reference
β-catenin	Cytoplasm and nucleus	49	Epidermal growth factor-induced nuclear localization and decreased expression of c-Myc	ND	[9]
Cortactin *	Cytoplasm	87, 124, 161, 189, 198, 235, 272, 309, 319	Regulation of cell migration and actin filament binding	DD1 and DD2	[9]
DNAJA1	Cytoplasm	ND	Protein folding	ND	[12]
ERK1	Cytoplasm and nucleus	72	Proliferation, mobility, and cell survival	ND	[13]
Foxp3 *	Nucleus	ND	ND	ND	[14]
HDAC9	Cytoplasm and nucleus	ND	Modulation of cell survival and arrest of cellular movement	DD2	[15]
HDAC11	Nucleus	ND	Transcriptional activation of interleukin 10	ND	[16]
HMG2	Nucleus	2	Increased transcription of STAT5	ND	[17]
HSC70	Cytoplasm	ND	Protein folding	ND	[12]
HSPA5	Cytoplasm	353	Ubiquitination of HSPA5 mediated by GP78	ND	[18]
HSP90α	Cytoplasm	294	Degradation and elimination of misfolded proteins and regulation of glucocorticoid receptors	DD1, DD2 et BUZ	[9]
K-RAS *	Cytoplasm	104	Cell proliferation	ND	[19]
Ku70	Cytoplasm	539, 542	Suppression of apoptosis	ND	[9]
LC3B-II*	Cytoplasm	ND	Regulation of autophagy	ND	[20]
MSH2	Cytoplasm and nucleus	845, 847, 871, 892	Reduced cellular sensitivity to DNA damaging agents and reduced DNA mismatch repair activities by downregulation of MSH2	DD1	[21]
MYH9	Cytoplasm	ND	Regulation of binding to actin	ND	[12]

Substrates	Localization of the Substrate	Deacetylated Lysine(s)	Function of the Deacetylated Substrate	Interaction Domains of HDAC6	Reference
			filaments		
PrxI	Cytoplasm and nucleus	197	Antioxidant activity	ND	[22][23]
PrxII	Cytoplasm and nucleus	196	Antioxidant activity	ND	[22][23]
RIG-I	Cytoplasm	858, 909	Recognition of viral RNA	ND	[24]
Sam68	Nucleus	ND	Alternative splicing	ND	[25]
Survivin	Nucleus	129	Anti-apoptotic function	DD2	[9]
Tat	Cytoplasm	28	Suppression of HIV transactivation	DD2 and BUZ	[26]
α -tubulin*	Cytoplasm	40	Formation of immune synapses, viral infection, cell migration and chemotaxis	DD1 or DD2	[9][27]

with lysosomes via autophagy ([Figure 2C](#)). A decrease in the acetylation of microtubule-associated protein 1 light chain 3 by HDAC6 was observed during autophagic degradation [[28](#)][[29](#)]. The HDAC6 protein is also involved in proteasome-dependent protein degradation by SIRT1, KAT5 and HSP90 are also phosphorylated by Src [2], and F-actin is also deacetylated by HDAC6 and SIRT1. Akt/HSP90 cascade of 16S kDa pBcr/Bcl-2 translocated leads to the degradation of BCL-2 binding proteins by the proteasome ([Figure 2D](#)). Anacardic acid inhibited p70S6 kinase dissociation, ERK1/2 extracellular signal-regulated GSK3 α , kinase shc/Foxo3 factor (Hsp)ok, P3ap/Glycylalysine site in HDACg histone/AceAcyase; HDAC6 butano complex in defective, forming, p11MG N2 sorting, inhibitory release of HSF1 includes the transcription of HSC gene at Spn ([Figure 2E](#)); HSP(HDAC) heat shock proteins (family of Ac(HSP70)ns) ([number 5](#)); LC3B-II: microtubule-associated protein 1 light chain 3; MSH2: MutS protein homolog 2; MYH9: myosin heavy chain 9; ND: non determined; Prx: peroxiredoxin; RIG-I: retinoic acid-inducible gene I protein; Sam: Src-associated substrate in mitosis; STAT: signal transducer and transcriptional activator; Tat: twin-arginine translocation protein.

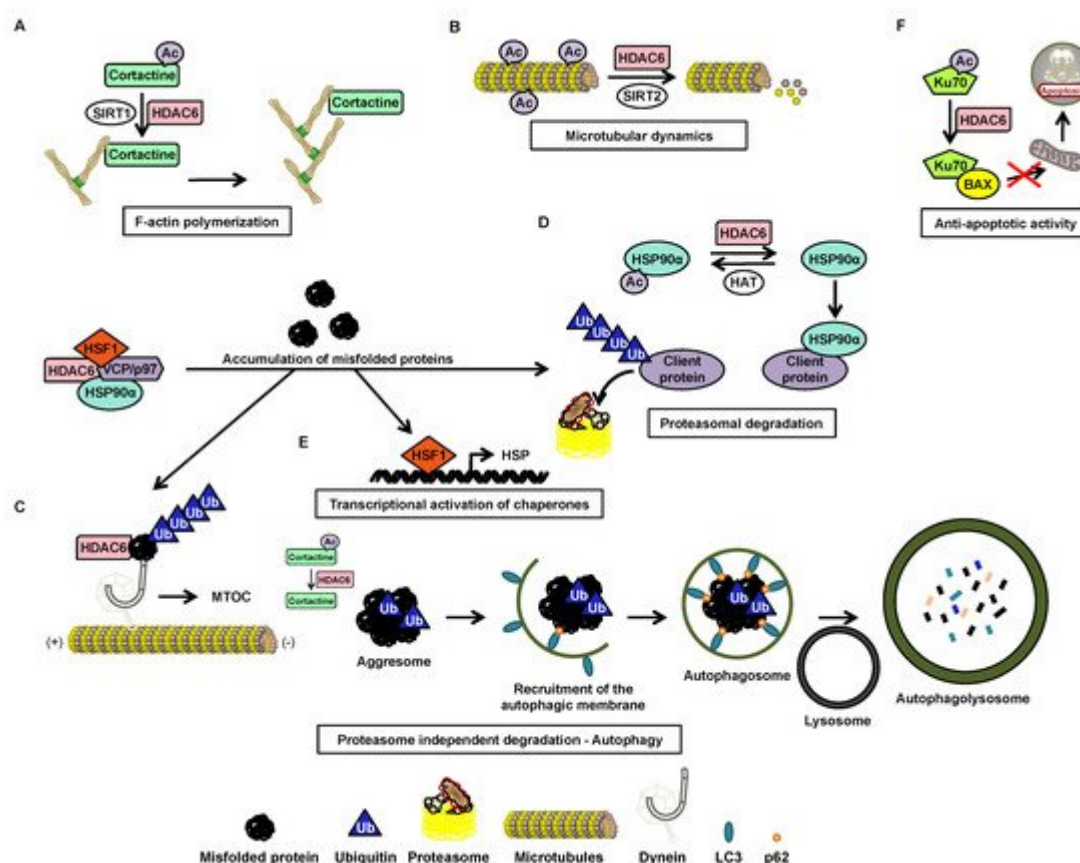


Figure 2. The HDAC6 protein is involved in many cellular processes. HDAC6 is involved in F-actin polymerization (A), microtubule dynamics (B), anti-apoptotic activity (C), proteasome-dependent and -independent degradation (D), transcriptional activation of chaperone proteins (E), and autophagy (F). Ac: acetylated; HAT: histone acetyltransferase; HDAC: histone deacetylase; HSF: heat shock factor; HSP: heat shock protein; MTOC: microtubule organizing center; SIRT: sirtuin; VCP: valosin-containing protein/ATPase.

HDAC6 is also involved in apoptosis by deacetylating the Ku70 protein, which then forms a complex with BAX, a proapoptotic protein, allowing the inhibition of apoptosis (Figure 2F). Similarly, inhibition of the catalytic activity of HDAC6 promotes the dephosphorylation of AKT and ERK, associated with decreased cell proliferation and death of cancer cells [28].

Furthermore, HDAC6 regulates endocytosis and exocytosis vesicles. When the epidermal growth factor receptor (EGFR) receptor is bound to its ligand, it interacts with HDAC6 and inactivates it by phosphorylation, which then leads to the hyperacetylation of microtubules and finally the internalization of the receptor. The inhibition of HDAC6 induces the increase of the acetylation of peroxiredoxins 1 and 2, which are antioxidant enzymes, increasing their activity and causing a reduction in cell resistance to chemotherapy [28]. HDAC6 is involved in the process of autophosphorylation of tau protein, giving it the ability to form aggregates called neurofibrillary tangles that can cause neurotoxicity [30].

2.3. Post-Transcriptional Regulation

There is a lack of current data explaining the post-transcriptional regulation of HDAC6 protein. Nevertheless, some microRNAs stimulating cancer cell proliferation and metastasis formation (miR-22, miR-221, miR-433, and miR-548) [10], and stem cell differentiation (miR-26a) [31], are predicted to interact with HDAC6 protein, thus inducing a destabilization or repression of the translation of its mRNA.

2.4. Post-Translational Regulation

Post-translational modifications such as phosphorylation and acetylation have a significant impact on HDAC6 functions. Indeed, although EGFR induces an inhibitory phosphorylation of HDAC6, in the majority of cases it is established that the phosphorylation of HDAC6 improves its deacetylase activity, whereas acetylation decreases its enzymatic activity, preventing the deacetylation of α -tubulin. Examples of post-translational modifications of the HDAC6 protein influencing its activity are shown in Table 2.

Table 2. Post-translational modifications regulating the activity of HDAC6.

Post-Translational Modification	Enzyme	Target Site	Consequences	Reference
Phosphorylation	GSK3 β	Ser-22	Increased deacetylation activity of α -tubulin	[10]
	ERK1	Ser-1035	Regulation of cellular motility	[10]
	GRK2	ND	Increased deacetylation activity of α -tubulin	[32]
	Aurora	ND	Increased deacetylation activity of α -tubulin	[10]
	PKC ζ	ND	Increased deacetylation activity of α -tubulin	[10]
	CK2	Ser-458	Improved formation and elimination of aggresomes	[10]
	EGFR	Tyr-570	Inhibition of deacetylation activity	[33]
Acetylation	p300	Lys-16	Inhibition of deacetylation activity	[10]

In addition to these known post-translational modifications, there are proteins interacting directly with the HDAC6 protein and inducing its inhibition by direct interaction (Table 3).

CK2: casein kinase 2; EGFR: epidermal growth factor receptor; ERK1: extracellular signal-regulated kinase; GRK2: G protein-coupled receptor kinase 2; GSK3: glycogen synthase kinase 3; Lys: lysine; ND: non determined; PKC ζ : protein kinase C isoform ζ ; Ser: serine; Thr: threonine.

Protein Inhibiting HDAC6 by Direct Interaction	Protein Function	Protein Region Required for Interaction with HDAC6	HDAC6 Domain Interacting with the Protein	Cellular Impact	References
CYLD	Deubiquitinase	ND	DD1/DD2	Cell proliferation, ciliogenesis	[10]
Dysferlin	Skeletal muscle membrane repair, myogenesis, cell adhesion, intercellular calcium signaling	Domain C2	ND	Myogenesis	[34]
Mdp3	Stabilization factor of microtubules	Amino-terminal region	ND	Cell motility	[35]
Paxillin	Focal adhesion	Region rich in proline	ND	Polarization and cell migration	[10]
p62	Transport of misfolded proteins	Between the ZZ domain and the TRAF6 link area	DD2	Aggresome formation	[36]
RanBPM	Apoptosis, proliferation and cell migration		ND	Aggresome formation	[37]
Tau	Stabilization factor of microtubules	Tubulin binding region	SE14 domain	Aggresome formation	[36][38]
TPPP1	Polymerization and acetylation of microtubules		ND	Regulation of microtubule acetylation and β -catenin expression	[39]

and renal diseases, as well as in inflammation [40] and viral response [10]. The role of the HDAC6 protein in cancer is also now well better understood. Although its oncogenic or tumor suppressor potential is dependent on the type of cancer [28], its involvement in oncogenic cell transformation, tumor development, and cancer immunity regulation makes a strong therapeutic candidate [41].

DD: deacetylase domain; Mdp3: microtubule-associated protein (MAP) 7 domain-containing protein 3; ND: non determined; RanBPM: Ran-binding protein microtubule-organizing center; tau: tubulin-associated unit; TPPP1: tubulin polymerization-promoting protein-1.

Table 4. Deregulation of HDAC6 expression in different types of cancers.

Cancer Type	Cancers	Expression of HDAC6-Comments	References
Solid tumors	Bladder	Overexpressed	[41]
	Melanoma	Overexpressed	[41]
	Lung	Overexpressed	[41]

Cancer Type	Cancers	Expression of HDAC6-Comments	References
Hematological	Oral squamous cell carcinoma	Overexpressed-Enhanced expression in advanced stages	[28][42]
	Ovarian carcinoma	Overexpressed-Enhanced expression in advanced stages	[28][42]
	Breast	Overexpressed-Prediction of a good or bad prognosis	[28][43]
	Hepatocytic carcinoma	Overexpressed-Enhanced expression in advanced stages	[28]
		Under-expressed-HDAC6 suggested as a tumor suppressor	[28][44]
	Chronic lymphocytic leukemia	Overexpressed-Observation on patient samples, cell lines and a transgenic mouse model	[42]
	Acute myeloid leukemia	Overexpressed	[28][42]
	Acute lymphoblastic leukemia	Overexpressed-Enhanced expression in advanced stages	[28]
	Chronic lymphocytic leukemia	Overexpressed-Correlated with longer survival	[28]
	T-cell cutaneous lymphoma	Overexpressed-Correlated with longer survival	[28]
	Chronic myeloid leukemia	Overexpressed-Increased expression in CD34 ⁺ cells	[45]
	Multiple myeloma	Overexpressed	[46]
	Mantle cell lymphoma	Overexpressed	[46]
	Diffuse large B cell lymphoma	Overexpressed	[46]
	Peripheral T-cell lymphoma	Overexpressed	[46]

The ability to specifically target HDAC6 would have valuable clinical utility in the treatment of these cancers. However, despite a large number of pan-HDAC inhibitors, very few compounds are capable of selectively inhibiting HDAC6 (Table 5). This type of inhibitor can be divided into 2 groups according to their chemical structure: benzamides and hydroxamates [28].

CD: cluster of differentiation, HDAC6: histone deacetylase 6.

Table 5. List of HDAC6 inhibitors.

Class	HDAC6 Inhibitor	Binding Domain	CI ₅₀ (nM) of the HDAC6 Activity <i>in Vitro</i>	Selectivity Ratio for HDAC6 Compared to (Other HDACs)	Inhibition of HDAC6 <i>in Cellulo</i> (μM) [§]	Effect on Cancer Cell Lines or Cancer Type	References
Benzamides	Trithiocarbonate derivative (12ac)	ND	65	19 (HDAC1)	10 (lung cancer)	CI ₅₀ = 8.2 μM (cervical cancer)	[47]
	NQN-1 (2-benzyl-amino-naphthoquinone)	ND	5540	Values non available (HDAC1, 2, 3, 4, 5, 7, 8, 9, 10, 11)	4 (chronic myeloid leukemia)	CI ₅₀ = 0.8 μM (leukemia)	[48]
Hydroxamates	Hydroxamic acid containing a phenylalanine (4n)	His215, His216, Tyr386, Phe283, and Tyr255 of DD1 and His610, His611, Tyr782, Phe620, and Phe680 of an HDAC6 homology model	1690	14 (HDAC1)	1 (colorectal carcinoma)	IC ₅₀ : 3 to > 50 μM (various cancer cell lines)	[49]
	Hydroxamic acid containing a pyridylalanine (5a)	Phe566 of DD2 of an HDAC6 homology model	3970	25 (HDAC1)	ND	IC ₅₀ : 104 μM (breast cancer)	[50]
	ACY-738	ND	1.7	55 (HDAC1), 75 (HDAC2), 128 (HDAC3)	2.5 (neural cells)	ND	[51]
	ACY-775	ND	7.5	283 (HDAC1), 343	2.5 (neural cells)	ND	[51]

Class	HDAC6 Inhibitor	Binding Domain	Cl ₅₀ (nM) of the HDAC6 Activity <i>in Vitro</i>	Selectivity Ratio for HDAC6 Compared to (Other HDACs)	Inhibition of HDAC6 <i>in Cellulo</i> (μM) ^{\$}	Effect on Cancer Cell Lines or Cancer Type	References
				(HDAC2), 1496 (HDAC3)			
	ACY-1083	His573 and His574 of DD2	3	260 (HDAC1)	0.03 (neuroblastoma)	ND	[52][53]
	Bavarostat	Ser568 of DD2	60	>10000 (HDAC1, 2, 3), 188 (HDAC4), 317 (HDAC5), 78 (HDAC7), 142 (HDAC8), 87 (HDAC9), >17 (HDAC10), 167 (HDAC11)	10 (neural progenitor cells derived from induced pluripotent stem cells)	ND	[54]
	BRD9757	ND	30	21 (HDAC1), 60 (HDAC2), 23 (HDAC3), 727 (HDAC4), 611 (HDAC5), 420 (HDAC7), 36 (HDAC8), >1000 (HDAC9)	10 (cervical cancer)	ND	[55]
	Cay10603	His499 of DD2 of an	0.002	ND	<1 to 1 μM (several pancreatic cancer cell lines)	ND	[56][57]

Class	HDAC6 Inhibitor	Binding Domain	Cl ₅₀ (nM) of the HDAC6 Activity <i>in Vitro</i>	Selectivity Ratio for HDAC6 Compared to (Other HDACs)	Inhibition of HDAC6 <i>in Cellulo</i> (μM) [§]	Effect on Cancer Cell Lines or Cancer Type	References
		HDAC6 homology model					
	Citarinostat (ACY-241)	ND	2.6	14 (HDAC1), 17 (HDAC2), 18 (HDAC3 and 4), >7000 (HDAC4, 5,9), 2808 (HDAC7), 53 (HDAC8),	0.3 (ovarian cancer)	Cl ₅₀ : 4.6 to 6.1 μM (ovarian and breast cancer)	[58]
	α3β-cyclic tetrapeptide (23)	ND	39	3 (HDAC1), 4 (HDAC3), 6 (HDAC8)	2 (acute lymphoblastic leukemia)	IC ₅₀ : 9 to > 20 μM (various cancer cell lines)	[59]
	Compound containing a phenylisoxazole group as a surface recognition group (7)	His499 of HDAC7	0.002	>100000 (HDAC1), >100000 (HDAC2), 210 (HDAC3), >3000000 (HDAC8), 45350 (HDAC10)	ND	IC ₅₀ : 0.1 to 1 μM (various prostate cancer cell lines)	[56]
	Compound containing a triazolylphenyl group (6b)	ND	1.9	52 (HDAC1), 155 (HDAC2), 7 (HDAC3), 420 (HDAC8), 59 (HDAC10)	ND	IC ₅₀ : <0.5 to 22 μM (several prostate cancer lines)	[60]

Class	HDAC6 Inhibitor	Binding Domain	CI ₅₀ (nM) of the HDAC6 Activity <i>in Vitro</i>	Selectivity Ratio for HDAC6 Compared to (Other HDACs)	Inhibition of HDAC6 <i>in Cellulo</i> (μM) [§]	Effect on Cancer Cell Lines or Cancer Type	References
	Compound containing a peptoid (2i)	Tyr301 of DD2 of an HDAC6 homology model	1.59	126 (HDAC2), >6000 (HDAC4), 40 (HDAC11)	N	IC ₅₀ : 0.34 to 2.7 μM (various cancer cell lines)	[61]
	3-aminopyrrolidinone derivative (33)	ND	17	4359 (HDAC1), 11 (HDAC8)	0.3 (multiple myeloma)	Good oral bioavailability	[62]
	4-aminomethylaryl acid derivative (1a)	ND	19	305 (HDAC1), 842 (HDAC2), 237 (HDAC3), 790 (HDAC4), 174 (HDAC5), 242 (HDAC7), 36 (HDAC8), 195 (HDAC0)	0.46 (cervical cancer)	ND	[63]
	4-hydroxybenzoic acid derivative (7b)	ND	200	>50000 (HDAC1, 2, 8), >500000 (HDAC3, 10, 11)	50 (prostate cancer)	IC ₅₀ : 41 to 130 (several prostate and breast cancer cell lines)	[64]
	4-hydroxybenzoic acid derivative (13a)	ND	20000	25 (HDAC1), >5000 (HDAC2, 3, 4, 8, 10), >2500 (HDAC11)	50 (prostate cancer)	IC ₅₀ : 19 to 127 (several prostate and breast cancer cell lines)	[64]
	Aminoteralline derivative (32)	Phe620 and	50	126 (HDAC1),	2 (neuroblastoma)	IC ₅₀ = 5.4 μM (neuroblastoma)	[65]

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		Phe680 of an HDAC6 homology model		2 (HDAC8)			
	Benzothiophene derivative (39)	ND	14	ND	Same effect as tubastatin A	Does not target NF-κB and AP-1 at the transcriptional level	[66]
	2,4-imidazolidinedione derivative (10c)	ND	4.4	218 (HDAC1), 63 (HDAC2), 53 (HDAC3), > 20000 (HDAC4, 7, 8, 9, 11), 3386 (HDAC5), 37 (HDAC10)	1.6 (acute myeloid leukemia)	IC ₅₀ : 0.2 to 0.8 μM (various cancer cell lines)	[67]
	Mercaptoacetamide derivative (2)	ND	95.3	34 (HDAC1), 77 (HDAC2), 64 (HDAC8), 112 (HDAC10)	ND	At 10 μM protects cortical neurons from oxidative stress inducing death	[68]
	N-Hydroxycarbonylbenzylamino quinoline derivative (13)	ND	0.291	32817 (HDAC1), 42955 (HDAC2), 26632 (HDAC3), 15250 (HDAC4), 10694 (HDAC5), 2436 (HDAC7), 4089	0.1 (multiple myeloma)	IC ₅₀ : 9.1 to 40.6 μM (multiple myeloma)	[69]

Class	HDAC6 Inhibitor	Binding Domain	Cl ₅₀ (nM) of the HDAC6 Activity <i>in Vitro</i>	Selectivity Ratio for HDAC6 Compared to (Other HDACs)	Inhibition of HDAC6 <i>in Cellulo</i> (μM) [§]	Effect on Cancer Cell Lines or Cancer Type	References
				(HDAC8), 5258 (HDAC9), 33646 (HDAC10), 1292 (HDAC11)			
	Isoxazole-3-hydroxamate derivative (SS-208)	His463, Pro464, Phe583, and Leu712 of DD2	12	116 (HDAC1), 1625 (HDAC4), 576 (HDAC5), 695 (HDAC7), 103 (HDAC8), 3183 (HDAC9), 427 (HDAC11)	5 (melanoma)	ND	[70]
	Phenothiazine derivative (7i)	Phe620 and Phe680 of DD2	5	538 (HDAC1)	0.1 (acute myeloid leukemia)	ND	[71]
	Phenylhydroxamate derivative (2)	Phe464 and His614 of DD2	3	27 (HDAC1)	ND	Cl ₅₀ : 0.65 to 2.77 (ovarian cancer and squamous cell carcinoma of the tongue)	[61][72]
	Phenylsulfonylfuroxan derivative (5c)	ND	7.4	33 (HDAC1), 51 (HDAC2), 45 (HDAC3), 4 (HDAC4), 46 (HDAC8), 82 (HDAC11)	0.013 (acute myeloid leukemia)	IC ₅₀ : 0.4 to 5.8 μM (various cancer cell lines)	[73]

The compounds ACY-241 (Citarinostat) and ACY-1215 (Ricolinostat) are derivatives of hydroxamic acid, which shows a specific inhibitory activity against HDAC6 with IC₅₀ values of 2.6 and 5 nM, respectively. They are the only

Class	HDAC6 Inhibitor	Binding Domain	Cl ₅₀ (nM) of the HDAC6 Activity in Vitro	Selectivity Ratio for HDAC6 Compared to (Other HDACs)	Inhibition of HDAC6 in Cellulo (μM) [§]	Effect on Cancer Cell Lines or Cancer Type	References
	Pyridone derivative (11e)	Phe155 and Phe210 of HDAC2	2.46	8 (HDAC1), 52 (HDAC2), 127 (HDAC3), 2329 (HDAC4), 785 (HDAC5), 1512 (HDAC7), 77	ND	IC ₅₀ : 0.14 to 0.38 μM (various cancer cell lines)	[74]
HDAC6 Inhibitor	Clinical Trial Identification	Phase of the Clinical Trial		Pathology			
ACY-241	NCT02400242	Ia/Ib		Multiple myeloma			
	NCT02935790	Ib		Stage III and IV unresectable melanoma			
	NCT02551185	Ib		Advanced solid tumors			
	NCT02635061	Ib		Non-resectable non-small cell lung cancer			
ACY-1215	NCT02632071	Ib		Unresectable or metastatic breast cancer			
	NCT02787369	Ib		Relapsed chronic lymphocytic leukemia			
	NCT02091063	Ib/II		Relapsed or refractory lymphoid malignancies			
	NCT01997840	Ib/II		Recurrent and refractory multiple myeloma			
	NCT01583283	I/II		Multiple myeloma recurrent or recurrent and refractory			
	NCT02189343	Ib		Recurrent and refractory multiple myeloma			
	NCT01323751	I/II		Multiple myeloma recurrent or recurrent and refractory			
	NCT02856568 [94]	Ib		Unresectable or metastatic cholangiocarcinoma			
	NCT02661815	Ib		Ovarian cancer, primary peritoneal cancer or platinum-resistant fallopian tubes			

Despite the observation of a moderate overexpression of the HDAC6 protein in urothelial cancerous tissues, the inhibition of the protein had limited efficacy compared to the use of inhibitors targeting several HDACs [102]. On the other hand, HDAC6 inhibitors have notable anti-cancer properties in prostate cancer [64], breast cancer [103], melanoma [66], and ovarian cancer [29]. These effects could be explained by the implication of HDAC6 in metastasis formation by epithelial-mesenchymal transition induction via its recruitment by TGFβ [104], in cell migration via α-tubulin deacetylation and in angiogenesis via cortactin deacetylation [105]. In contrast to some selective HDAC6 inhibitors, currently approved pan-HDAC inhibitors failed to show any clinical benefits in solid tumors [94]. The

Class	HDAC6 Inhibitor	Binding Domain	Cl ₅₀ (nM) of the HDAC6 Activity in Vitro	Selectivity Ratio for HDAC6 Compared to (Other HDACs)	Inhibition of HDAC6 in Cellulo (μM) [§]	Effect on Cancer Cell Lines or Cancer Type	References
		Arg196 of HDAC7				1.5 to 4.7 μM (multiple myeloma cell lines)	
[108]	Tubastatin A derivative (Marbostat-100)	Asp649, His651 et Asp742 of DD2	0.7	[107] 1106 (HDAC2), 247 (HDAC8)	0.05 (acute monocytic leukemia)	Non-cytotoxic	[79]
	Indolylsulfonylcinnamic hydroxamate (12)	ND	5.2	60 (HDAC1), 223 (HDAC2)	0.1 (colon cancer)	IC ₅₀ : 0.4 to 2.5 μM (multiple cancer cell lines)	[80]
	[109] MAIP-032	DD2	58	38 (HDAC1)	ND	Cl ₅₀ : 3.87 μM (squamous cell carcinoma line of the tongue)	[81]
	MPT0G211	ND	0.291	ND	0.1 (neuroblastoma)	ND	[30]
	N-hydroxy-4-[(N(2-hydroxyethyl)-2-phenylacetamido)methyl]-benzamide]] (HPB)	His573 and His574 of DD2	31	37 (HDAC1)	8 (prostate cancer)	ND	[52][82]
[42]	[112] N-hydroxy-4-(2-[(2-hydroxyethyl) (phenyl)amino]-2-oxoethyl)benzamide (HPOB)	Binding to zinc ion only via its OH group but does not displace the zinc-bound water molecule	56	[111] 52 (HDAC1)	16 (prostate cancer, adenocarcinoma, glioblastoma)	Increases the effect on cell viability in combination with etoposide, dexamethasone or SAHA	[83][84]
	N-hydroxy-4-(2-methoxy-5-(methyl(2-methylquinazolin-4-yl)-amino)phenoxy)butanamide (23bb)	Tyr298 and Glu255 of an HDAC6 homology model	[113] 17	25 (HDAC1), 200 (HDAC8)	0.051 (cervical cancer)	IC ₅₀ : 14 to 104 nM (various cancer cell lines)	[85]
	Nexturastat A	DD2 of an	5	604 (HDAC1)	0.01 (murine melanoma)	IC ₅₀ = 14.3 μM (melanoma)	[57][86]

apoptotic protein BIM in acute myeloid leukemia cells [114]. Moreover, in a model exhibiting significant nuclear HDAC6 levels, chemical HDAC6 inhibition reduces its nuclear localization and p53-HDAC6 interactions inducing cell cycle arrest and apoptosis via changes of p53 target gene expression [115]. The specific nuclear localization of HDAC6 in leukemia cells might offer a therapeutic advantage to specifically target those cells.

3.3.2. HDAC6 in CML

3.3.2.1.Degradation of BCR-ABL via Deacetylation of HSP90α by HDAC6 in the Cytoplasm

Although little research exists on HDAC6 in the context of CML, this protein has a function that makes it particularly interesting in the context of such pathology. HDAC6 deacetylates heat shock protein (HSP90)α, which is involved in the stabilization of the oncogenic tyrosine kinase *breakpoint cluster region-Abelson* (BCR-ABL) protein [114] protein. In the acetylated form, HSP90α loses its chaperone function, which leads to the degradation of its client proteins by the proteasome (Figure 3A). The importance of the acetylation status of HSP90α in the protein degradation of BCR-ABL

Class	HDAC6 Inhibitor	Binding Domain	Cl ₅₀ (nM) of the HDAC6 Activity <i>in Vitro</i>	Selectivity Ratio for HDAC6 Compared to (Other HDACs)	Inhibition of HDAC6 <i>in Cellulo</i> (μM) ^{\$}	Effect on Cancer Cell Lines or Cancer Type	References
		HDAC6 homology model			[116][117]		
	Oxazole hydroxamate (4g)	Phe620, Phe680, Leu749, and Tyr782 of DD2 of an HDAC6 homology model	59	237 (HDAC1, 8)	10 (cervical cancer)	IC ₅₀ = 10.2 μM (acute myeloid leukemia)	[87]
	Ricolinostat (ACY-1215)	DD2 of an HDAC6 homology model	4.7	12 (HDAC1), 10 (HDAC2), 11 (HDAC3), 1490 (HDAC4), 1064 (HDAC5), 298 (HDAC7), 21 (HDAC8), >2000 (HDAC9, 11)	0.62 (multiple myeloma)	Cl ₅₀ : 2 to 8 μM (multiple myeloma cell lines)	[57][88][89]
	Sahaquine	ND	ND	ND	0.1 (glioblastoma)	Cl ₅₀ : 10 μM (glioblastoma)	[90]
	TC24	Ser568, His610, Phe679 and Tyr782 of HDAC6	ND	ND	1 et 10 (gastric cancer)	Cl ₅₀ : 10.2 to 17.2 μM (several gastric cancer cell lines)	[91]
	Tetrahydroisoquinoline (5a)	ND	36	1250 (HDAC1), >1000 (HDAC2,	0.21 (cervical cancer)	ND	[63]

3.3.2.2. OverExpression of HDAC6 in CML Stem Cells

LSCs that are not targeted by TKI and are characterized by a capacity for self-renewal play a crucial role in CML relapse. Although HDAC6 is necessary for the repression of genes involved in the differentiation targeted by the Tip60-p400 complex in embryonic stem cells (ESCs) [118], no study has provided evidence for this in LSCs, more differentiated. In contrast, studies have shown that several proteins in the HDAC family are overexpressed in LCSs of CML. Indeed, SIRT1 is activated by BCR-ABL via STAT5 and its expression is increased in LSCs compared to in CML cells [119]. Finally, overexpression of isoforms of HDAC (HDAC1, HDAC2, HDAC3, HDAC4, and HDAC5) and in particular HDAC6 was more frequently observed in LSCs (CD34⁺ CD38⁻) isolated from patients with CML than in K562 cells [45] (Figure 3B), making it a protein of interest in the search for treatments to prevent relapse in patients with CML.

3.3.3 HDAC6 inhibitors in CLL

Class	HDAC6 Inhibitor	Binding Domain	Cl ₅₀ (nM) of the HDAC6 Activity in Vitro	Selectivity Ratio for HDAC6 Compared to (Other HDACs)	Inhibition of HDAC6 in Cellulo (μM) [§]	Effect on Cancer Cell Lines or Cancer Type	References
			[120]	4, 5, 7, 10, 11), 1278 (HDAC3), 58 (HDAC8)			
	Thiazole	ND	52	ND	ND	ND	[63]
	Tubacin	DD2 of an HDAC6 homology model	4	350 (HDAC1)	5 (prostate cancer)SangtingTao2.5 (acute lymphoblastic leukemia)	IC ₅₀ : 1.2 to 2 μM (acute lymphoblastic leukemia)	[57][92][93]
	Tubastatin A	His610, His611, Phe679, Phe680 and Tyr1821 of HDAC6	15	1093 (HDAC1)	2.5 (unspecified)	ND	[91][92]
	Tubathian A	ND	1.9	5790 (HDAC1)	0.1 (ovarian cancer)	ND	[94]
Other	[122] 3-hydroxypyridine-2-thione (3-HPT)	Tyr306 of HDAC8	681	5 (HDAC8)	ND	Inactive against two prostate cancer cell lines and one acute T cell leukemia cell line	[95]
	[124] 1-hydroxypyridine-2-thione (1HPT)-6-carboxylic acid	DD	150	287 (HDAC1), 4733 (HDAC2), 473 (HDAC4), 233 (HDAC5), 1933 (HDAC7), 22 (HDAC8), 313 (HDAC9)	ND	Cl ₅₀ : 18 to 75 μM (leukemia)	[96]
	[126]						

cytotoxicity against non-malignant cells [127].

In AML, inhibition of HDAC6 was essentially investigated in combination with other pharmacologically active compounds at a pre-clinical level. For instance, a combination of 17-(allylamino)-17-demethoxygeldanamycin (17-AAG), a synthetic derivative of the ansamycin benzoquinone antibiotic geldanamycin, with the HDAC6 inhibitor tubacin reduces the viability of primary AML samples, validating HDAC6 as a HSP90 client protein also in AML and that its hyperacetylation facilitates the anticancer potential of 17-AAG [128]. LBH-589 and PXD101 inhibit HDAC1 and HDAC6 and synergize with cytarabine to induce cell death in pediatric AML, accompanied by DNA damage induction and increased Bim expression levels [129]. Similarly, Bim protein induction and inhibition of nuclear factor-kappa B (NF-kB) pathway were identified as a mechanistic basis for the synergistic anti-cancer effects of belinostat in combination with the proteasome inhibitor bortezomib in AML and ALL cells [130]. The selective JAK2/HDAC6 dual inhibitor 20a shows excellent in vivo antitumor efficacy in HEL AML mouse xenograft assays and synergizes with the antifungal drug fluconazole [131]. The selective HDAC6 inhibitor MPT0G211 combined with doxorubicin displays anti-cancer effect by inducing a DNA damage response associated with increased Ku70 acetylation and

Class	HDAC6 Inhibitor	Binding Domain	Cl ₅₀ (nM) of the HDAC6 Activity <i>in Vitro</i>	Selectivity Ratio for HDAC6 Compared to (Other HDACs)	Inhibition of HDAC6 <i>in Cellulo</i> (μM) [§]	Effect on Cancer Cell Lines or Cancer Type	References
	Adamantylamino derivative (20a)	ND	82	46 (HDAC1), 51 (HDAC4)	ND	ND	[97]
	Mercaptoacetamide derivative (2b)	ND	1.3	3615 (HDAC1)	10 (primary rat cortical culture)	ND	[98]
	Sulfamide derivative (13e)	[133] ND	440	>23 (HDAC1)	1 (bladder cancer)	ND	[99]
Undefined structure	CKD-506	ND	5	>400 (HDAC1, 2, 7, 8)	0.03 (Human PBMCs)	ND	[100]

general and efficient mechanism explaining the synergistic effect observed [130]. MPT0G211 combined with vincristine interrupts ALL mitosis via interference with microtubular dynamics leading to apoptosis. In vivo, MPT0G211 plus doxorubicin or vincristine reduces tumor growth xenograft models [132].

Remarkably, it has been shown that the inhibition of HDAC6 using either the pan-HDAC inhibitor trichostatin, the selective HDAC6 inhibitor tubacin, or a genetic knock-down efficiently reduces Notch3 signaling through a post-translational, Aspartate protease (Dab1)-regulated pathway leading to inhibiting 50% of cell apoptosis; DDT-41 acetylase add mainly on Glu-leukemia growth HDAC6 histone deacetylase; Ala: alanine; Arg: arginine; Asn: asparagine; Asp: aspartate; Cys: cysteine; Glu: glutamate; Gly: glycine; His: histidine; Ile: isoleucine; Leu: leucine; Lys: lysine; Met: methionine; Phe: phenylalanine; Pro: proline; SAHA: suberoylanilide hydroxamic acid; Ser: serine; Tyr: tyrosine. [134]

3.3.6 HDAC6 inhibitors in other hematological malignancies

Inhibition of HDAC6 activity increases CD20 levels in B-cell tumor cell lines and malignant patient cells, potentializing the in vivo effect of anti-CD20 monoclonal antibodies like rituximab. Translation of CD20 mRNA is significantly enhanced after HDAC6 inhibition as CD20 mRNA was abundant within the polysomal fraction, indicating a post-transcriptional function of HDAC6. Collectively, these findings suggest HDAC6 inhibition is a rational therapeutic strategy to be implemented in combination therapies with anti-CD20 monoclonal antibodies and open up novel avenues for the clinical use of HDAC6 inhibitors [135].

The HDAC6 inhibitor A452 combined with the Bruton's tyrosine kinase inhibitor ibrutinib efficiently kills non-Hodgkin lymphoma cells, including follicular lymphoma [135].

The HDAC6 inhibitor KT-531 displays the highest anti-cancer potency against T-cell prolymphocytic leukemia (T-PLL) cells compared to other hematological neoplasms, together with safe differential toxicity compared to non-transformed cell lines. Accordingly, HDAC6 is overexpressed in primary T-PLL patient samples in which KT-531 exerts a potent anti-cancer activity. Moreover, a combination of KT-531 with various approved drugs including bendamustine, idasanutlin, and venetoclax shows promising synergistic effects against T-PLL patient cells [136].

References

1. Michael Schneckengerber; Cristina Florean; Mario Dicato; Marc Diederich; Epigenetic alterations as a universal feature of cancer hallmarks and a promising target for personalized treatments. *Current Topics in Medicinal Chemistry* **2015**, 16, 745-776, 10.2174/1568026615666150825141330.
2. Losson, H.; Schneckengerber, M.; Dicato, M.; Diederich, M. Natural Compound Histone Deacetylase Inhibitors (HDACi): Synergy with Inflammatory Signaling Pathway Modulators and Clinical Applications in Cancer. *Molecules* 2016, 21, 1608.
3. Finnin, M.S.; Donigian, J.R.; Cohen, A.; Richon, V.M.; Rifkind, R.A.; Marks, P.A.; Breslow, R.; Pavletich, N.P. Structures of a histone deacetylase homologue bound to the TSA and SAHA inhibitors. *Nature* 1999, 401, 188–193.
4. Carafa, V.; Rotili, D.; Forgione, M.; Cuomo, F.; Serretiello, E.; Hailu, G.S.; Jarho, E.; Lahtela-Kakkonen, M.; Mai, A.; Altucci, L. Sirtuin functions and modulation: From chemistry to the clinic. *Clin. Epigenetics* 2016, 8, 61.
5. Mei, Z.; Zhang, X.; Yi, J.; Huang, J.; He, J.; Tao, Y. Sirtuins in metabolism, DNA repair and cancer. *J. Exp. Clin. Cancer Res.* 2016, 35, 182.
6. Thiagalingam, S.; Cheng, K.H.; Lee, H.J.; Mineva, N.; Thiagalingam, A.; Ponte, J.F. Histone deacetylases: Unique players in shaping the epigenetic histone code. *Ann. N. Y. Acad. Sci.* 2003, 983, 84–100.
7. De Ruijter, A.J.; van Gennip, A.H.; Caron, H.N.; Kemp, S.; van Kuilenburg, A.B. Histone deacetylases (HDACs): Characterization of the classical HDAC family. *Biochem. J.* 2003, 370, 737–749.
8. Bertos, N.R.; Gilquin, B.; Chan, G.K.; Yen, T.J.; Khochbin, S.; Yang, X.J. Role of the tetradecapeptide repeat domain of human histone deacetylase 6 in cytoplasmic retention. *J. Biol. Chem.* 2004, 279, 48246–48254.
9. Li, Y.; Shin, D.; Kwon, S.H. Histone deacetylase 6 plays a role as a distinct regulator of diverse cellular processes. *FEBS J.* 2013, 280, 775–793.
10. Zheng, K.; Jiang, Y.; He, Z.; Kitazato, K.; Wang, Y. Cellular defence or viral assist: The dilemma of HDAC6. *J. Gen. Virol.* 2017, 98, 322–337.
11. Mortenson, J.B.; Heppler, L.N.; Banks, C.J.; Weerasekara, V.K.; Whited, M.D.; Piccolo, S.R.; Johnson, W.E.; Thompson, J.W.; Andersen, J.L. Histone deacetylase 6 (HDAC6) promotes the pro-survival activity of 14-3-3zeta via deacetylation of lysines within the 14-3-3zeta binding pocket. *J. Biol. Chem.* 2015, 290, 12487–12496.
12. Zhang, L.; Liu, S.; Liu, N.; Zhang, Y.; Liu, M.; Li, D.; Seto, E.; Yao, T.P.; Shui, W.; Zhou, J. Proteomic identification and functional characterization of MYH9, Hsc70, and DNAJA1 as novel substrates of HDAC6 deacetylase activity. *Protein Cell* 2015, 6, 42–54.

13. Wu, J.Y.; Xiang, S.; Zhang, M.; Fang, B.; Huang, H.; Kwon, O.K.; Zhao, Y.; Yang, Z.; Bai, W.; Bepler, G.; et al. Histone deacetylase 6 (HDAC6) deacetylates extracellular signal-regulated kinase 1 (ERK1) and thereby stimulates ERK1 activity. *J. Biol. Chem.* 2018, 293, 1976–1993.
14. Beier, U.H.; Wang, L.; Han, R.; Akimova, T.; Liu, Y.; Hancock, W.W. Histone deacetylases 6 and 9 and sirtuin-1 control Foxp3+ regulatory T cell function through shared and isoform-specific mechanisms. *Sci. Signal.* 2012, 5, ra45.
15. Salian-Mehta, S.; Xu, M.; McKinsey, T.A.; Tobet, S.; Wierman, M.E. Novel Interaction of Class IIb Histone Deacetylase 6 (HDAC6) with Class IIa HDAC9 Controls Gonadotropin Releasing Hormone (GnRH) Neuronal Cell Survival and Movement. *J. Biol. Chem.* 2015, 290, 14045–14056.
16. Gao, L.; Cueto, M.A.; Asselbergs, F.; Atadja, P. Cloning and functional characterization of HDAC11, a novel member of the human histone deacetylase family. *J. Biol. Chem.* 2002, 277, 25748–25755.
17. Medler, T.R.; Craig, J.M.; Fiorillo, A.A.; Feeney, Y.B.; Harrell, J.C.; Clevenger, C.V. HDAC6 Deacetylates HMGN2 to Regulate Stat5a Activity and Breast Cancer Growth. *Mol. Cancer Res.* 2016, 14, 994–1008.
18. Chang, Y.W.; Tseng, C.F.; Wang, M.Y.; Chang, W.C.; Lee, C.C.; Chen, L.T.; Hung, M.C.; Su, J.L. Deacetylation of HSPA5 by HDAC6 leads to GP78-mediated HSPA5 ubiquitination at K447 and suppresses metastasis of breast cancer. *Oncogene* 2016, 35, 1517–1528.
19. Yang, M.H.; Laurent, G.; Bause, A.S.; Spang, R.; German, N.; Haigis, M.C.; Haigis, K.M. HDAC6 and SIRT2 regulate the acetylation state and oncogenic activity of mutant K-RAS. *Mol. Cancer Res.* 2013, 11, 1072–1077.
20. Liu, K.P.; Zhou, D.; Ouyang, D.Y.; Xu, L.H.; Wang, Y.; Wang, L.X.; Pan, H.; He, X.H. LC3B-II deacetylation by histone deacetylase 6 is involved in serum-starvation-induced autophagic degradation. *Biochem. Biophys. Res. Commun.* 2013, 441, 970–975.
21. Zhang, M.; Xiang, S.; Joo, H.Y.; Wang, L.; Williams, K.A.; Liu, W.; Hu, C.; Tong, D.; Haakenson, J.; Wang, C.; et al. HDAC6 deacetylates and ubiquitinates MSH2 to maintain proper levels of MutSalpha. *Mol. Cell* 2014, 55, 31–46.
22. Perkins, A.; Nelson, K.J.; Parsonage, D.; Poole, L.B.; Karplus, P.A. Peroxiredoxins: Guardians against oxidative stress and modulators of peroxide signaling. *Trends Biochem. Sci.* 2015, 40, 435–445.
23. Parmigiani, R.B.; Xu, W.S.; Venta-Perez, G.; Erdjument-Bromage, H.; Yaneva, M.; Tempst, P.; Marks, P.A. HDAC6 is a specific deacetylase of peroxiredoxins and is involved in redox regulation. *Proc. Natl. Acad. Sci. USA* 2008, 105, 9633–9638.

24. Moreno-Gonzalo, O.; Mayor, F., Jr.; Sanchez-Madrid, F. HDAC6 at Crossroads of Infection and Innate Immunity. *Trends Immunol.* 2018, 39, 591–595.
25. Nakka, K.K.; Chaudhary, N.; Joshi, S.; Bhat, J.; Singh, K.; Chatterjee, S.; Malhotra, R.; De, A.; Santra, M.K.; Dilworth, F.J.; et al. Nuclear matrix-associated protein SMAR1 regulates alternative splicing via HDAC6-mediated deacetylation of Sam68. *Proc. Natl. Acad. Sci. USA* 2015, 112, E3374–E3383.
26. Huo, L.; Li, D.; Sun, X.; Shi, X.; Karna, P.; Yang, W.; Liu, M.; Qiao, W.; Aneja, R.; Zhou, J. Regulation of Tat acetylation and transactivation activity by the microtubule-associated deacetylase HDAC6. *J. Biol. Chem.* 2011, 286, 9280–9286.
27. Matsuyama, A.; Shimazu, T.; Sumida, Y.; Saito, A.; Yoshimatsu, Y.; Seigneurin-Berny, D.; Osada, H.; Komatsu, Y.; Nishino, N.; Khochbin, S.; et al. In vivo destabilization of dynamic microtubules by HDAC6-mediated deacetylation. *EMBO J.* 2002, 21, 6820–6831.
28. Seidel, C.; Schnekenburger, M.; Dicato, M.; Diederich, M. Histone deacetylase 6 in health and disease. *Epigenomics* 2015, 7, 103–118.
29. Haakenson, J.; Zhang, X. HDAC6 and ovarian cancer. *Int. J. Mol. Sci.* 2013, 14, 9514–9535.
30. Fan, S.J.; Huang, F.I.; Liou, J.P.; Yang, C.R. The novel histone de acetylase 6 inhibitor, MPT0G211, ameliorates tau phosphorylation and cognitive deficits in an Alzheimer's disease model. *Cell Death Dis.* 2018, 9, 655.
31. Lee, S.W.; Yang, J.; Kim, S.Y.; Jeong, H.K.; Lee, J.; Kim, W.J.; Lee, E.J.; Kim, H.S. MicroRNA-26a induced by hypoxia targets HDAC6 in myogenic differentiation of embryonic stem cells. *Nucleic Acids Res.* 2015, 43, 2057–2073.
32. Lafarga, V.; Aymerich, I.; Tapia, O.; Mayor, F., Jr.; Penela, P. A novel GRK2/HDAC6 interaction modulates cell spreading and motility. *EMBO J.* 2012, 31, 856–869.
33. Williams, K.A.; Zhang, M.; Xiang, S.; Hu, C.; Wu, J.Y.; Zhang, S.; Ryan, M.; Cox, A.D.; Der, C.J.; Fang, B.; et al. Extracellular signal-regulated kinase (ERK) phosphorylates histone deacetylase 6 (HDAC6) at serine 1035 to stimulate cell migration. *J. Biol. Chem.* 2013, 288, 33156–33170.
34. Di Fulvio, S.; Azakir, B.A.; Therrien, C.; Sinnreich, M. Dysferlin interacts with histone deacetylase 6 and increases alpha-tubulin acetylation. *PLoS ONE* 2011, 6, e28563.
35. Tala, S.X.; Chen, J.; Zhang, L.; Liu, N.; Zhou, J.; Li, D.; Liu, M. Microtubule stabilization by Mdp3 is partially attributed to its modulation of HDAC6 in addition to its association with tubulin and microtubules. *PLoS ONE* 2014, 9, e90932.
36. Yan, J.; Seibenhener, M.L.; Calderilla-Barbosa, L.; Diaz-Meco, M.T.; Moscat, J.; Jiang, J.; Wooten, M.W.; Wooten, M.C. SQSTM1/p62 interacts with HDAC6 and regulates deacetylase activity. *PLoS ONE* 2013, 8, e76016.

37. Salemi, L.M.; Almawi, A.W.; Lefebvre, K.J.; Schild-Poulter, C. Aggresome formation is regulated by RanBPM through an interaction with HDAC6. *Biol. Open* 2014, 3, 418–430.
38. Perez, M.; Santa-Maria, I.; Gomez de Barreda, E.; Zhu, X.; Cuadros, R.; Cabrero, J.R.; Sanchez-Madrid, F.; Dawson, H.N.; Vitek, M.P.; Perry, G.; et al. Tau--an inhibitor of deacetylase HDAC6 function. *J. Neurochem.* 2009, 109, 1756–1766.
39. Schofield, A.V.; Gamell, C.; Bernard, O. Tubulin polymerization promoting protein 1 (TPPP1) increases beta-catenin expression through inhibition of HDAC6 activity in U2OS osteosarcoma cells. *Biochem. Biophys. Res. Commun.* 2013, 436, 571–577.
40. Batchu, S.N.; Brijmohan, A.S.; Advani, A. The therapeutic hope for HDAC6 inhibitors in malignancy and chronic disease. *Clin. Sci. (Lond.)* 2016, 130, 987–1003.
41. Li, T.; Zhang, C.; Hassan, S.; Liu, X.; Song, F.; Chen, K.; Zhang, W.; Yang, J. Histone deacetylase 6 in cancer. *J. Hematol. Oncol.* 2018, 11, 111.
42. Maharaj, K.; Powers, J.J.; Achille, A.; Deng, S.; Fonseca, R.; Pabon-Saldana, M.; Quayle, S.N.; Jones, S.S.; Villagra, A.; Sotomayor, E.M.; et al. Silencing of HDAC6 as a therapeutic target in chronic lymphocytic leukemia. *Blood Adv.* 2018, 2, 3012–3024.
43. Li, A.; Chen, P.; Leng, Y.; Kang, J. Histone deacetylase 6 regulates the immunosuppressive properties of cancer-associated fibroblasts in breast cancer through the STAT3-COX2-dependent pathway. *Oncogene* 2018, 37, 5952–5966.
44. Qian, H.; Chen, Y.; Nian, Z.; Su, L.; Yu, H.; Chen, F.J.; Zhang, X.; Xu, W.; Zhou, L.; Liu, J.; et al. HDAC6-mediated acetylation of lipid droplet-binding protein CIDEC regulates fat-induced lipid storage. *J. Clin. Invest.* 2017, 127, 1353–1369.
45. Bamodu, O.A.; Kuo, K.T.; Yuan, L.P.; Cheng, W.H.; Lee, W.H.; Ho, Y.S.; Chao, T.Y.; Yeh, C.T. HDAC inhibitor suppresses proliferation and tumorigenicity of drug-resistant chronic myeloid leukemia stem cells through regulation of hsa-miR-196a targeting BCR/ABL1. *Exp. Cell Res.* 2018, 370, 519–530.
46. Cosenza, M.; Pozzi, S. The Therapeutic Strategy of HDAC6 Inhibitors in Lymphoproliferative Disease. *Int. J. Mol. Sci.* 2018, 19, 2337.
47. Dehmel, F.; Weinbrenner, S.; Julius, H.; Ciossek, T.; Maier, T.; Stengel, T.; Fettis, K.; Burkhardt, C.; Wieland, H.; Beckers, T. Trithiocarbonates as a novel class of HDAC inhibitors: SAR studies, isoenzyme selectivity, and pharmacological profiles. *J. Med. Chem.* 2008, 51, 3985–4001.
48. Inks, E.S.; Josey, B.J.; Jesinkey, S.R.; Chou, C.J. A novel class of small molecule inhibitors of HDAC6. *ACS Chem. Biol.* 2012, 7, 331–339.
49. Schafer, S.; Saunders, L.; Eliseeva, E.; Velena, A.; Jung, M.; Schwienhorst, A.; Strasser, A.; Dickmanns, A.; Ficner, R.; Schlimme, S.; et al. Phenylalanine-containing hydroxamic acids as

- selective inhibitors of class IIb histone deacetylases (HDACs). *Bioorg. Med. Chem.* 2008, 16, 2011–2033.
50. Schafer, S.; Saunders, L.; Schlimme, S.; Valkov, V.; Wagner, J.M.; Kratz, F.; Sippl, W.; Verdin, E.; Jung, M. Pyridylalanine-containing hydroxamic acids as selective HDAC6 inhibitors. *ChemMedChem* 2009, 4, 283–290.
51. Jochems, J.; Boulden, J.; Lee, B.G.; Blendy, J.A.; Jarpe, M.; Mazitschek, R.; Van Duzer, J.H.; Jones, S.; Berton, O. Antidepressant-like properties of novel HDAC6-selective inhibitors with improved brain bioavailability. *Neuropsychopharmacology* 2014, 39, 389–400.
52. Porter, N.J.; Mahendran, A.; Breslow, R.; Christianson, D.W. Unusual zinc-binding mode of HDAC6-selective hydroxamate inhibitors. *Proc. Natl. Acad. Sci. USA* 2017, 114, 13459–13464.
53. Krukowski, K.; Ma, J.; Golonzhka, O.; Laumet, G.O.; Gutti, T.; van Duzer, J.H.; Mazitschek, R.; Jarpe, M.B.; Heijnen, C.J.; Kavelaars, A. HDAC6 inhibition effectively reverses chemotherapy-induced peripheral neuropathy. *Pain* 2017, 158, 1126–1137.
54. Strebl, M.G.; Campbell, A.J.; Zhao, W.N.; Schroeder, F.A.; Riley, M.M.; Chindavong, P.S.; Morin, T.M.; Haggarty, S.J.; Wagner, F.F.; Ritter, T.; et al. HDAC6 Brain Mapping with [(18)F]Bavarostat Enabled by a Ru-Mediated Deoxyfluorination. *ACS Cent. Sci.* 2017, 3, 1006–1014.
55. Wagner, F.F.; Olson, D.E.; Gale, J.P.; Kaya, T.; Weiwer, M.; Aidoud, N.; Thomas, M.; Davoine, E.L.; Lemercier, B.C.; Zhang, Y.L.; et al. Potent and selective inhibition of histone deacetylase 6 (HDAC6) does not require a surface-binding motif. *J. Med. Chem.* 2013, 56, 1772–1776.
56. Kozikowski, A.P.; Tapadar, S.; Luchini, D.N.; Kim, K.H.; Billadeau, D.D. Use of the nitrile oxide cycloaddition (NOC) reaction for molecular probe generation: A new class of enzyme selective histone deacetylase inhibitors (HDACIs) showing picomolar activity at HDAC6. *J. Med. Chem.* 2008, 51, 4370–4373.
57. Sixto-Lopez, Y.; Bello, M.; Rodriguez-Fonseca, R.A.; Rosales-Hernandez, M.C.; Martinez-Archundia, M.; Gomez-Vidal, J.A.; Correa-Basurto, J. Searching the conformational complexity and binding properties of HDAC6 through docking and molecular dynamic simulations. *J. Biomol. Struct. Dyn.* 2017, 35, 2794–2814.
58. Huang, P.; Almeciga-Pinto, I.; Jarpe, M.; van Duzer, J.H.; Mazitschek, R.; Yang, M.; Jones, S.S.; Quayle, S.N. Selective HDAC inhibition by ACY-241 enhances the activity of paclitaxel in solid tumor models. *Oncotarget* 2017, 8, 2694–2707.
59. Olsen, C.A.; Ghadiri, M.R. Discovery of potent and selective histone deacetylase inhibitors via focused combinatorial libraries of cyclic alpha3beta-tetrapeptides. *J. Med. Chem.* 2009, 52, 7836–7846.
60. Chen, Y.; Lopez-Sanchez, M.; Savoy, D.N.; Billadeau, D.D.; Dow, G.S.; Kozikowski, A.P. A series of potent and selective, triazolyphenyl-based histone deacetylases inhibitors with activity against

- pancreatic cancer cells and *Plasmodium falciparum*. *J. Med. Chem.* 2008, 51, 3437–3448.
61. Diedrich, D.; Hamacher, A.; Gertzen, C.G.; Alves Avelar, L.A.; Reiss, G.J.; Kurz, T.; Gohlke, H.; Kassack, M.U.; Hansen, F.K. Rational design and diversity-oriented synthesis of peptoid-based selective HDAC6 inhibitors. *Chem. Commun. (Camb.)* 2016, 52, 3219–3222.
 62. Lin, X.; Chen, W.; Qiu, Z.; Guo, L.; Zhu, W.; Li, W.; Wang, Z.; Zhang, W.; Zhang, Z.; Rong, Y.; et al. Design and synthesis of orally bioavailable aminopyrrolidinone histone deacetylase 6 inhibitors. *J. Med. Chem.* 2015, 58, 2809–2820.
 63. Blackburn, C.; Barrett, C.; Chin, J.; Garcia, K.; Gigstad, K.; Gould, A.; Gutierrez, J.; Harrison, S.; Hoar, K.; Lynch, C.; et al. Potent histone deacetylase inhibitors derived from 4-(aminomethyl)-N-hydroxybenzamide with high selectivity for the HDAC6 isoform. *J. Med. Chem.* 2013, 56, 7201–7211.
 64. Seidel, C.; Schnekenburger, M.; Mazumder, A.; Teiten, M.H.; Kirsch, G.; Dicato, M.; Diederich, M. 4-Hydroxybenzoic acid derivatives as HDAC6-specific inhibitors modulating microtubular structure and HSP90 α chaperone activity against prostate cancer. *Biochem. Pharmacol.* 2016, 99, 31–52.
 65. Tang, G.; Wong, J.C.; Zhang, W.; Wang, Z.; Zhang, N.; Peng, Z.; Zhang, Z.; Rong, Y.; Li, S.; Zhang, M.; et al. Identification of a novel aminotetralin class of HDAC6 and HDAC8 selective inhibitors. *J. Med. Chem.* 2014, 57, 8026–8034.
 66. Wang, X.X.; Wan, R.Z.; Liu, Z.P. Recent advances in the discovery of potent and selective HDAC6 inhibitors. *Eur. J. Med. Chem.* 2018, 143, 1406–1418.
 67. Liang, T.; Hou, X.; Zhou, Y.; Yang, X.; Fang, H. Design, Synthesis, and Biological Evaluation of 2,4-Imidazolinedione Derivatives as HDAC6 Isoform-Selective Inhibitors. *ACS Med. Chem. Lett.* 2019, 10, 1122–1127.
 68. Kozikowski, A.P.; Chen, Y.; Gaysin, A.; Chen, B.; D’Annibale, M.A.; Suto, C.M.; Langley, B.C. Functional differences in epigenetic modulators-superiority of mercaptoacetamide-based histone deacetylase inhibitors relative to hydroxamates in cortical neuron neuroprotection studies. *J. Med. Chem.* 2007, 50, 3054–3061.
 69. Lee, H.Y.; Nepali, K.; Huang, F.I.; Chang, C.Y.; Lai, M.J.; Li, Y.H.; Huang, H.L.; Yang, C.R.; Liou, J.P. (N-Hydroxycarbonylbenzylamino)quinolines as Selective Histone Deacetylase 6 Inhibitors Suppress Growth of Multiple Myeloma in Vitro and in Vivo. *J. Med. Chem.* 2018, 61, 905–917.
 70. Shen, S.; Hadley, M.; Ustinova, K.; Pavlicek, J.; Knox, T.; Noonepalle, S.; Tavares, M.T.; Zimprich, C.A.; Zhang, G.; Robers, M.B.; et al. Discovery of a New Isoxazole-3-hydroxamate-Based Histone Deacetylase 6 Inhibitor SS-208 with Antitumor Activity in Syngeneic Melanoma Mouse Models. *J. Med. Chem.* 2019, 62, 8557–8577.

71. Vogerl, K.; Ong, N.; Senger, J.; Herp, D.; Schmidtkunz, K.; Marek, M.; Muller, M.; Bartel, K.; Shaik, T.B.; Porter, N.J.; et al. Synthesis and Biological Investigation of Phenothiazine-Based Benzhydroxamic Acids as Selective Histone Deacetylase 6 Inhibitors. *J. Med. Chem.* 2019, 62, 1138–1166.
72. Porter, N.J.; Osko, J.D.; Diedrich, D.; Kurz, T.; Hooker, J.M.; Hansen, F.K.; Christianson, D.W. Histone Deacetylase 6-Selective Inhibitors and the Influence of Capping Groups on Hydroxamate-Zinc Denticity. *J. Med. Chem.* 2018, 61, 8054–8060.
73. Duan, W.; Li, J.; Inks, E.S.; Chou, C.J.; Jia, Y.; Chu, X.; Li, X.; Xu, W.; Zhang, Y. Design, synthesis, and antitumor evaluation of novel histone deacetylase inhibitors equipped with a phenylsulfonylfuroxan module as a nitric oxide donor. *J. Med. Chem.* 2015, 58, 4325–4338.
74. Cho, M.; Choi, E.; Yang, J.S.; Lee, C.; Seo, J.J.; Kim, B.S.; Oh, S.J.; Kim, H.M.; Lee, K.; Park, S.K.; et al. Discovery of pyridone-based histone deacetylase inhibitors: Approaches for metabolic stability. *ChemMedChem* 2013, 8, 272–279.
75. Liu, Y.M.; Lee, H.Y.; Lai, M.J.; Pan, S.L.; Huang, H.L.; Kuo, F.C.; Chen, M.C.; Liou, J.P. Pyrimidinedione-mediated selective histone deacetylase 6 inhibitors with antitumor activity in colorectal cancer HCT116 cells. *Org. Biomol. Chem.* 2015, 13, 10226–10235.
76. Yu, C.W.; Chang, P.T.; Hsin, L.W.; Chern, J.W. Quinazolin-4-one derivatives as selective histone deacetylase-6 inhibitors for the treatment of Alzheimer's disease. *J. Med. Chem.* 2013, 56, 6775–6791.
77. Heltweg, B.; Dequiedt, F.; Marshall, B.L.; Brauch, C.; Yoshida, M.; Nishino, N.; Verdin, E.; Jung, M. Subtype selective substrates for histone deacetylases. *J. Med. Chem.* 2004, 47, 5235–5243.
78. Hideshima, T.; Qi, J.; Paranal, R.M.; Tang, W.; Greenberg, E.; West, N.; Colling, M.E.; Estiu, G.; Mazitschek, R.; Perry, J.A.; et al. Discovery of selective small-molecule HDAC6 inhibitor for overcoming proteasome inhibitor resistance in multiple myeloma. *Proc. Natl. Acad. Sci. USA* 2016, 113, 13162–13167.
79. Sellmer, A.; Stangl, H.; Beyer, M.; Grunstein, E.; Leonhardt, M.; Pongratz, H.; Eichhorn, E.; Elz, S.; Striegl, B.; Jenei-Lanzl, Z.; et al. Marbostat-100 Defines a New Class of Potent and Selective Antiinflammatory and Antirheumatic Histone Deacetylase 6 Inhibitors. *J. Med. Chem.* 2018, 61, 3454–3477.
80. Lee, H.Y.; Tsai, A.C.; Chen, M.C.; Shen, P.J.; Cheng, Y.C.; Kuo, C.C.; Pan, S.L.; Liu, Y.M.; Liu, J.F.; Yeh, T.K.; et al. Azaindolylysulfonamides, with a more selective inhibitory effect on histone deacetylase 6 activity, exhibit antitumor activity in colorectal cancer HCT116 cells. *J. Med. Chem.* 2014, 57, 4009–4022.
81. Mackwitz, M.K.W.; Hamacher, A.; Osko, J.D.; Held, J.; Scholer, A.; Christianson, D.W.; Kassack, M.U.; Hansen, F.K. Multicomponent Synthesis and Binding Mode of Imidazo[1,2 -a]pyridine-

- Capped Selective HDAC6 Inhibitors. *Org. Lett.* 2018, 20, 3255–3258.
82. Lee, J.H.; Yao, Y.; Mahendran, A.; Ngo, L.; Venta-Perez, G.; Choy, M.L.; Breslow, R.; Marks, P.A. Creation of a histone deacetylase 6 inhibitor and its biological effects [corrected]. *Proc. Natl. Acad. Sci. USA* 2015, 112, 12005–12010.
83. Lee, J.H.; Mahendran, A.; Yao, Y.; Ngo, L.; Venta-Perez, G.; Choy, M.L.; Kim, N.; Ham, W.S.; Breslow, R.; Marks, P.A. Development of a histone deacetylase 6 inhibitor and its biological effects. *Proc. Natl. Acad. Sci. USA* 2013, 110, 15704–15709.
84. Hai, Y.; Christianson, D.W. Histone deacetylase 6 structure and molecular basis of catalysis and inhibition. *Nat. Chem. Biol.* 2016, 12, 741–747.
85. Yang, Z.; Wang, T.; Wang, F.; Niu, T.; Liu, Z.; Chen, X.; Long, C.; Tang, M.; Cao, D.; Wang, X.; et al. Discovery of Selective Histone Deacetylase 6 Inhibitors Using the Quinazoline as the Cap for the Treatment of Cancer. *J. Med. Chem.* 2016, 59, 1455–1470.
86. Bergman, J.A.; Woan, K.; Perez-Villarroel, P.; Villagra, A.; Sotomayor, E.M.; Kozikowski, A.P. Selective histone deacetylase 6 inhibitors bearing substituted urea linkers inhibit melanoma cell growth. *J. Med. Chem.* 2012, 55, 9891–9899.
87. Senger, J.; Melesina, J.; Marek, M.; Romier, C.; Oehme, I.; Witt, O.; Sippl, W.; Jung, M. Synthesis and Biological Investigation of Oxazole Hydroxamates as Highly Selective Histone Deacetylase 6 (HDAC6) Inhibitors. *J. Med. Chem.* 2016, 59, 1545–1555.
88. Santo, L.; Hideshima, T.; Kung, A.L.; Tseng, J.C.; Tamang, D.; Yang, M.; Jarpe, M.; van Duzer, J.H.; Mazitschek, R.; Ogier, W.C.; et al. Preclinical activity, pharmacodynamic, and pharmacokinetic properties of a selective HDAC6 inhibitor, ACY-1215, in combination with bortezomib in multiple myeloma. *Blood* 2012, 119, 2579–2589.
89. Wang, F.; Zhong, B.W.; Zhao, Z.R. ACY 1215, a histone deacetylase 6 inhibitor, inhibits cancer cell growth in melanoma. *J. Biol. Regul. Homeost. Agents* 2018, 32, 851–858.
90. Zhang, I.; Beus, M.; Stochaj, U.; Le, P.U.; Zorc, B.; Rajic, Z.; Petrecca, K.; Maysinger, D. Inhibition of glioblastoma cell proliferation, invasion, and mechanism of action of a novel hydroxamic acid hybrid molecule. *Cell Death Discov* 2018, 4, 41.
91. Dong, J.; Zheng, N.; Wang, X.; Tang, C.; Yan, P.; Zhou, H.B.; Huang, J. A novel HDAC6 inhibitor exerts an anti-cancer effect by triggering cell cycle arrest and apoptosis in gastric cancer. *Eur. J. Pharmacol.* 2018, 828, 67–79.
92. Butler, K.V.; Kalin, J.; Brochier, C.; Vistoli, G.; Langley, B.; Kozikowski, A.P. Rational design and simple chemistry yield a superior, neuroprotective HDAC6 inhibitor, tubastatin A. *J. Am. Chem. Soc.* 2010, 132, 10842–10846.

93. Aldana-Masangkay, G.I.; Rodriguez-Gonzalez, A.; Lin, T.; Ikeda, A.K.; Hsieh, Y.T.; Kim, Y.M.; Lomenick, B.; Okemoto, K.; Landaw, E.M.; Wang, D.; et al. Tubacin suppresses proliferation and induces apoptosis of acute lymphoblastic leukemia cells. *Leuk. Lymphoma* 2011, 52, 1544–1555.
94. Depetter, Y.; Geurs, S.; De Vreese, R.; Goethals, S.; Vandoorn, E.; Laevens, A.; Steenbrugge, J.; Meyer, E.; de Tullio, P.; Bracke, M.; et al. Selective pharmacological inhibitors of HDAC6 reveal biochemical activity but functional tolerance in cancer models. *Int. J. Cancer* 2019, 145, 735–747.
95. Patil, V.; Sodji, Q.H.; Kornacki, J.R.; Mrksich, M.; Oyeler, A.K. 3-Hydroxypyridin-2-thione as novel zinc binding group for selective histone deacetylase inhibition. *J. Med. Chem.* 2013, 56, 3492–3506.
96. Muthyala, R.; Shin, W.S.; Xie, J.; Sham, Y.Y. Discovery of 1-hydroxypyridine-2-thiones as selective histone deacetylase inhibitors and their potential application for treating leukemia. *Bioorg. Med. Chem. Lett.* 2015, 25, 4320–4324.
97. Itoh, Y.; Suzuki, T.; Kouketsu, A.; Suzuki, N.; Maeda, S.; Yoshida, M.; Nakagawa, H.; Miyata, N. Design, synthesis, structure--selectivity relationship, and effect on human cancer cells of a novel series of histone deacetylase 6-selective inhibitors. *J. Med. Chem.* 2007, 50, 5425–5438.
98. Segretti, M.C.; Vallerini, G.P.; Brochier, C.; Langley, B.; Wang, L.; Hancock, W.W.; Kozikowski, A.P. Thiol-Based Potent and Selective HDAC6 Inhibitors Promote Tubulin Acetylation and T-Regulatory Cell Suppressive Function. *ACS Med. Chem. Lett.* 2015, 6, 1156–1161.
99. Wahhab, A.; Smil, D.; Ajamian, A.; Allan, M.; Chantigny, Y.; Therrien, E.; Nguyen, N.; Manku, S.; Leit, S.; Rahil, J.; et al. Sulfamides as novel histone deacetylase inhibitors. *Bioorg. Med. Chem. Lett.* 2009, 19, 336–340.
100. Choi, E.W.; Song, J.W.; Ha, N.; Choi, Y.I.; Kim, S. CKD-506, a novel HDAC6-selective inhibitor, improves renal outcomes and survival in a mouse model of systemic lupus erythematosus. *Sci. Rep.* 2018, 8, 17297.
101. Schnekenburger, M.; Florean, C.; Dicato, M.; Diederich, M. Epigenetic alterations as a universal feature of cancer hallmarks and a promising target for personalized treatments. *Curr. Top. Med. Chem.* 2016, 16, 745–776.
102. Rosik, L.; Niegisch, G.; Fischer, U.; Jung, M.; Schulz, W.A.; Hoffmann, M.J. Limited efficacy of specific HDAC6 inhibition in urothelial cancer cells. *Cancer Biol. Ther.* 2014, 15, 742–757.
103. Hsieh, Y.L.; Tu, H.J.; Pan, S.L.; Liou, J.P.; Yang, C.R. Anti-metastatic activity of MPT0G211, a novel HDAC6 inhibitor, in human breast cancer cells in vitro and in vivo. *Biochim. Biophys. Acta Mol. Cell Res.* 2019, 1866, 992–1003.
104. Shan, B.; Yao, T.P.; Nguyen, H.T.; Zhuo, Y.; Levy, D.R.; Klingsberg, R.C.; Tao, H.; Palmer, M.L.; Holder, K.N.; Lasky, J.A. Requirement of HDAC6 for transforming growth factor-beta1-induced epithelial-mesenchymal transition. *J. Biol. Chem.* 2008, 283, 21065–21073.

105. Kaluza, D.; Kroll, J.; Gesierich, S.; Yao, T.P.; Boon, R.A.; Hergenreider, E.; Tjwa, M.; Rossig, L.; Seto, E.; Augustin, H.G.; et al. Class IIb HDAC6 regulates endothelial cell migration and angiogenesis by deacetylation of cortactin. *EMBO J.* 2011, 30, 4142–4156.
106. Morel, D.; Jeffery, D.; Aspeslagh, S.; Almouzni, G.; Postel-Vinay, S. Combining epigenetic drugs with other therapies for solid tumours—Past lessons and future promise. *Nat. Rev. Clin. Oncol.* 2020, 17, 91–107.
107. Valdespino, V.; Valdespino, P.M. Potential of epigenetic therapies in the management of solid tumors. *Cancer Manag. Res.* 2015, 7, 241–251.
108. Grassadonia, A.; Cioffi, P.; Simiele, F.; Iezzi, L.; Zilli, M.; Natoli, C. Role of Hydroxamate-Based Histone Deacetylase Inhibitors (Hb-HDACIs) in the Treatment of Solid Malignancies. *Cancers* 2013, 5, 919–942.
109. Wang, E.C.; Min, Y.; Palm, R.C.; Fiordalisi, J.J.; Wagner, K.T.; Hyder, N.; Cox, A.D.; Caster, J.M.; Tian, X.; Wang, A.Z. Nanoparticle formulations of histone deacetylase inhibitors for effective chemoradiotherapy in solid tumors. *Biomaterials* 2015, 51, 208–215.
110. Brindisi, M.; Saraswati, A.P.; Brogi, S.; Gemma, S.; Butini, S.; Campiani, G. Old but Gold: Tracking the New Guise of Histone Deacetylase 6 (HDAC6) Enzyme as a Biomarker and Therapeutic Target in Rare Diseases. *J. Med. Chem.* 2020, 63, 23–39.
111. Hackanson, B.; Rimmele, L.; Benkisser, M.; Abdelkarim, M.; Fliegauf, M.; Jung, M.; Lubbert, M. HDAC6 as a target for antileukemic drugs in acute myeloid leukemia. *Leuk. Res.* 2012, 36, 1055–1062.
112. Hélène Losson; Sruthi Reddy Gajulapalli; Manon Lernoux; Jin-Young Lee; Aloran Mazumder; Déborah Gérard; Carole Seidel; HyungGu Hahn; Christo Christov; Mario Dicato; et al. Gilbert KirschByung Woo HanMichael SchneckengerMarc Diederich The HDAC6 inhibitor 7b induces BCR-ABL ubiquitination and downregulation and synergizes with imatinib to trigger apoptosis in chronic myeloid leukemia. *Pharmacological Research* **2020**, 160, 105058, 10.1016/j.phrs.2020.105058.
113. Liu, Y.; Peng, L.; Seto, E.; Huang, S.; Qiu, Y. Modulation of histone deacetylase 6 (HDAC6) nuclear import and tubulin deacetylase activity through acetylation. *J. Biol. Chem.* 2012, 287, 29168–29174.
114. Kramer, O.H.; Mahboobi, S.; Sellmer, A. Drugging the HDAC6-HSP90 interplay in malignant cells. *Trends Pharmacol. Sci.* 2014, 35, 501–509.
115. Ryu, H.W.; Shin, D.H.; Lee, D.H.; Choi, J.; Han, G.; Lee, K.Y.; Kwon, S.H. HDAC6 deacetylates p53 at lysines 381/382 and differentially coordinates p53-induced apoptosis. *Cancer Lett.* 2017, 391, 162–171.

116. Bali, P.; Pranpat, M.; Bradner, J.; Balasis, M.; Fiskus, W.; Guo, F.; Rocha, K.; Kumaraswamy, S.; Boyapalle, S.; Atadja, P.; et al. Inhibition of histone deacetylase 6 acetylates and disrupts the chaperone function of heat shock protein 90: A novel basis for antileukemia activity of histone deacetylase inhibitors. *J. Biol. Chem.* 2005, 280, 26729–26734.
117. Rao, R.; Fiskus, W.; Yang, Y.; Lee, P.; Joshi, R.; Fernandez, P.; Mandawat, A.; Atadja, P.; Bradner, J.E.; Bhalla, K. HDAC6 inhibition enhances 17-AAG--mediated abrogation of hsp90 chaperone function in human leukemia cells. *Blood* 2008, 112, 1886–1893.
118. Chen, P.B.; Hung, J.H.; Hickman, T.L.; Coles, A.H.; Carey, J.F.; Weng, Z.; Chu, F.; Fazzio, T.G. Hdac6 regulates Tip60-p400 function in stem cells. *Elife* 2013, 2, e01557.
119. Kuo, Y.H.; Qi, J.; Cook, G.J. Regain control of p53: Targeting leukemia stem cells by isoform-specific HDAC inhibition. *Exp. Hematol.* 2016, 44, 315–321.
120. Kamira Maharaj; John J. Powers; Melanie Mediavilla-Varela; Alex Achille; Wael Gamal; Steven Quayle; Simon S. Jones; Eva Sahakian; Javier Pinilla-Ibarz; HDAC6 Inhibition Alleviates CLL-Induced T-Cell Dysfunction and Enhances Immune Checkpoint Blockade Efficacy in the Eμ-TCL1 Model. *Frontiers in Immunology* **2020**, 11, -, 10.3389/fimmu.2020.590072.
121. Björn Hackanson; Leander Rimmele; Marco Benkiser; Mahmoud Abdelkarim; Manfred Fliegau; Manfred Jung; Michael Lübbert; HDAC6 as a target for antileukemic drugs in acute myeloid leukemia. *Leukemia Research* **2012**, 36, 1055-1062, 10.1016/j.leukres.2012.02.026.
122. Elizabeth S. Inks; Benjamin J. Josey; Sean R. Jesinkey; C. James Chou; A Novel Class of Small Molecule Inhibitors of HDAC6. *ACS Chemical Biology* **2011**, 7, 331-339, 10.1021/cb200134p.
123. Youxuan Li; Patrick M. Woster; Discovery of a new class of histone deacetylase inhibitors with a novel zinc binding group. *MedChemComm* **2014**, 6, 613-618, 10.1039/c4md00401a.
124. Zhuang Yang; Taijin Wang; Fang Wang; Ting Niu; Zhuowei Liu; Xiaoxin Chen; Chaofeng Long; Minghai Tang; Dong Cao; Xiaoyan Wang; et al. Wei XiangYuyao YiLiang MaJingsong YouLijuan Chen Discovery of Selective Histone Deacetylase 6 Inhibitors Using the Quinazoline as the Cap for the Treatment of Cancer. *Journal of Medicinal Chemistry* **2015**, 59, 1455-1470, 10.1021/acs.jmedchem.5b01342.
125. Xiaoyang Li; Yuri K. Peterson; Elizabeth S. Inks; Richard A. Himes; Jiaying Li; Yingjie Zhang; Xiujie Kong; C. James Chou; Class I HDAC Inhibitors Display Different Antitumor Mechanism in Leukemia and Prostatic Cancer Cells Depending on Their p53 Status. *Journal of Medicinal Chemistry* **2018**, 61, 2589-2603, 10.1021/acs.jmedchem.8b00136.
126. Weizhi Ge; Zhongquan Liu; Yu Sun; Tianpeng Wang; Hongyu Guo; Xinyi Chen; Shengzu Li; Mengmeng Wang; Yue Chen; Yahui Ding; et al. Quan Zhang Design and synthesis of parthenolide-SAHA hybrids for intervention of drug-resistant acute myeloid leukemia. *Bioorganic Chemistry* **2019**, 87, 699-713, 10.1016/j.bioorg.2019.03.056.

127. Justyna M. Gawel; Andrew E. Shouksmith; Yasir S. Raouf; Nabanita Nawar; Krime Toutah; Shazreh Bukhari; Pimyupa Manaswiyoungkul; Olasunkanmi O. Olaoye; Johan Israelian; Tudor B. Radu; et al. Aaron D. Cabral Diana Sina Abootaleb Sedighi Elvin D. de Araujo Patrick T. Gunning PTG-0861: A novel HDAC6-selective inhibitor as a therapeutic strategy in acute myeloid leukaemia. *European Journal of Medicinal Chemistry* **2020**, 201, 112411, 10.1016/j.ejmech.2020.112411.
128. Rekha Rao; Warren Fiskus; Yonghua Yang; Pearl Lee; Rajeshree Joshi; Pravina Fernandez; Aditya Mandawat; Peter Atadja; James E. Bradner; Kapil Bhalla; et al. HDAC6 inhibition enhances 17-AAG-mediated abrogation of hsp90 chaperone function in human leukemia cells. *Blood* **2008**, 112, 1886-1893, 10.1182/blood-2008-03-143644.
129. Xuelian Xu; Chengzhi Xie; Holly Edwards; Hui Zhou; Steven A. Buck; Yubin Ge; Inhibition of Histone Deacetylases 1 and 6 Enhances Cytarabine-Induced Apoptosis in Pediatric Acute Myeloid Leukemia Cells. *PLOS ONE* **2011**, 6, e17138, 10.1371/journal.pone.0017138.
130. Yun Dai; Shuang Chen; Li Wang; Xin-Yan Pei; Lora B. Kramer; Paul Dent; Steven Grant; Bortezomib interacts synergistically with belinostat in human acute myeloid leukaemia and acute lymphoblastic leukaemia cells in association with perturbations in NF- κ B and Bim. *British Journal of Haematology* **2011**, 153, 222-235, 10.1111/j.1365-2141.2011.08591.x.
131. Yahui Huang; Guoqiang Dong; Huanqiu Li; Na Liu; Wannian Zhang; Chunquan Sheng; Discovery of Janus Kinase 2 (JAK2) and Histone Deacetylase (HDAC) Dual Inhibitors as a Novel Strategy for the Combinational Treatment of Leukemia and Invasive Fungal Infections. *Journal of Medicinal Chemistry* **2018**, 61, 6056-6074, 10.1021/acs.jmedchem.8b00393.
132. Huang-Ju Tu; Yi-Jyun Lin; Min-Wu Chao; Ting-Yi Sung; Yi-Wen Wu; Yi-Ying Chen; Mei-Hsiang Lin; Jing-Ping Liou; Shiow-Lin Pan; Chia-Ron Yang; et al. The anticancer effects of MPT0G211, a novel HDAC6 inhibitor, combined with chemotherapeutic agents in human acute leukemia cells. *Clinical Epigenetics* **2018**, 10, 1-13, 10.1186/s13148-018-0595-8.
133. Grace I. Aldana-Masangkay; Agustin Rodriguez-Gonzalez; Tara Lin; Alan K. Ikeda; Yao-Te Hsieh; Yong-Mi Kim; Brett Lomenick; Kazuo Okemoto; Elliot M. Landaw; Dongpeng Wang; et al. Ralph Mazitschek James E. Bradner Kathleen M. Sakamoto Tubacin suppresses proliferation and induces apoptosis of acute lymphoblastic leukemia cells. *Leukemia & Lymphoma* **2011**, 52, 1544-1555, 10.3109/10428194.2011.570821.
134. Marica Pinazza; Margherita Ghisi; Sonia Anna Minuzzo; Valentina Agnusdei; Gianluca Fossati; Vincenzo Ciminale; Laura Pezzè; Yari Ciribilli; Giorgia Pilotto; Carolina Venturoli; et al. Alberto Amadori Stefano Indraccolo Histone deacetylase 6 controls Notch3 trafficking and degradation in T-cell acute lymphoblastic leukemia cells. *Oncogene* **2018**, 37, 3839-3851, 10.1038/s41388-018-0234-z.

135. Ng Hoon Lee; Go Woon Kim; So Hee Kwon; The HDAC6-selective inhibitor is effective against non-Hodgkin lymphoma and synergizes with ibrutinib in follicular lymphoma. *Molecular Carcinogenesis* **2019**, 58, 944-956, 10.1002/mc.22983.
136. Krimo Toutah; Nabanita Nawar; Sanna Timonen; Helena Sorger; Yasir S. Raouf; Shazreh Bukhari; Jana von Jan; Aleksandr Ianevski; Justyna M. Gawel; Olasunkanmi O. Olaoye; et al. Mulu Geletu Ayah Abdeldayem Johan Israelian Tudor B. Radu Abootaleb Sedighi Muzaffar N. Bhatti Muhammad Murtaza Hassan Pimyupa Manaswiyoungkul Andrew E. Shouksmith Heidi A. Neubauer Elvin D. de Araujo Tero Aittokallio Oliver H. Krämer Richard Moriggi Satu Mustjoki Marco Herling Patrick T. Gunning Development of HDAC Inhibitors Exhibiting Therapeutic Potential in T-Cell Prolymphocytic Leukemia. *Journal of Medicinal Chemistry* **2021**, 64, 8486-8509, 10.1021/acs.jmedchem.1c00420.
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