

Polyethyleneimine-Based Drug Delivery Systems for Cancer Theranostics

Subjects: [Materials Science](#), [Biomaterials](#)

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With the development of nanotechnology, various types of polymer-based drug delivery systems have been designed for biomedical applications. Polymer-based drug delivery systems with desirable biocompatibility can be efficiently delivered to tumor sites with passive or targeted effects and combined with other therapeutic and imaging agents for cancer theranostics. As an effective vehicle for drug and gene delivery, polyethyleneimine (PEI) has been extensively studied due to its rich surface amines and excellent water solubility.

polyethylenimine

drug delivery

cancer treatment

cancer imaging

cancer theranostics

1. Introduction

Cancer is a human disease characterized by abnormal cell proliferation and metastasis and poses a very significant threat to human health. Its occurrence is closely related to harmful environment, bad lifestyle, and heredity. Early diagnosis and treatment of cancer is the most important strategy to improve survival rates. Recently, nanotechnology has attracted extensive attention in the biomedical field, particularly in the early diagnosis and treatment of cancer [\[1\]\[2\]](#).

The development of novel multifunctional nanoparticles (NPs) for cancer theranostics is one of the most important trends in the development of nanomedicine [\[3\]\[4\]\[5\]\[6\]](#). Compared with traditional drug delivery systems, NP-based drug delivery systems can not only improve the water solubility and stability of drugs, but also influence the distribution of drugs in vivo owing to the nanosized effects of NPs [\[7\]](#). In addition, the kinetics of drug release can be controlled through material design and surface modifications. More importantly, targeted molecules can be modified on the surface of NPs to specifically target tumor sites, thereby improving the bioavailability of drugs and reducing toxicity to off-target tissues [\[8\]](#). To date, various NPs have been developed for construction of NP-based drug delivery systems including liposomes [\[9\]\[10\]](#), micelles [\[11\]](#), nanogels [\[12\]\[13\]](#), radionuclide-labeled NPs [\[14\]\[15\]](#), and metal NPs [\[16\]\[17\]\[18\]\[19\]](#).

Polyethyleneimine (PEI) is a cationic polymer molecule composed of abundant amine groups and two aliphatic carbons, and because of its specific structure and properties it has been widely used to stabilize or modify various inorganic hybrid NPs [\[20\]](#). As a cationic polyamine, PEI can interact with or bind to anionic residues of DNA templates and polymerase through electrostatic interaction, thus significantly improving their transfection efficiency [\[21\]](#). In addition, the strong positive surface potential of PEI presents obvious cytotoxicity to cells because of its abundant amine groups [\[21\]](#). Therefore, neutralizing the surface potential of PEI through various chemical or

physical modifications can effectively reduce its cytotoxicity and improve biocompatibility. It is worth noting that these surface modifications not only improve the biocompatibility of PEI, but also enable it to acquire other functions, such as biomarker and targeting.

2. Overview of PEI

PEI is a commercially widely used cationic polymer containing primary, secondary, and tertiary amino groups in a ratio of 1:2:1 with strong positive charges [22]. PEI can be synthesized as linear PEI (**Figure 1a**) or branched PEI (**Figure 1b**) with a molecular weight ranging from 700 Da to 1000 kDa according to the degree of polymerization [23]. PEI can be easily prepared using an AB-type monomer via a simple one-step reaction [24]. In addition, PEI can be considered a low-cost option compared to dendrimers with the same molecular weight [25]. PEI has been widely used in different fields because of its unique structure and abundant amino groups. For example, in industry, PEI can be used as a flocculant to remove oil present in synthetically produced water, or as a wet strength agent in paper-making and the manufacture of shampoo [26][27]. In biomedicine, PEI is widely used in enzyme immobilization [28], virus immobilization on cellulose [29], cell adhesion [30], gene transfection [31], and the synthesis of NPs to enhance their stability and anticancer efficacy [32].

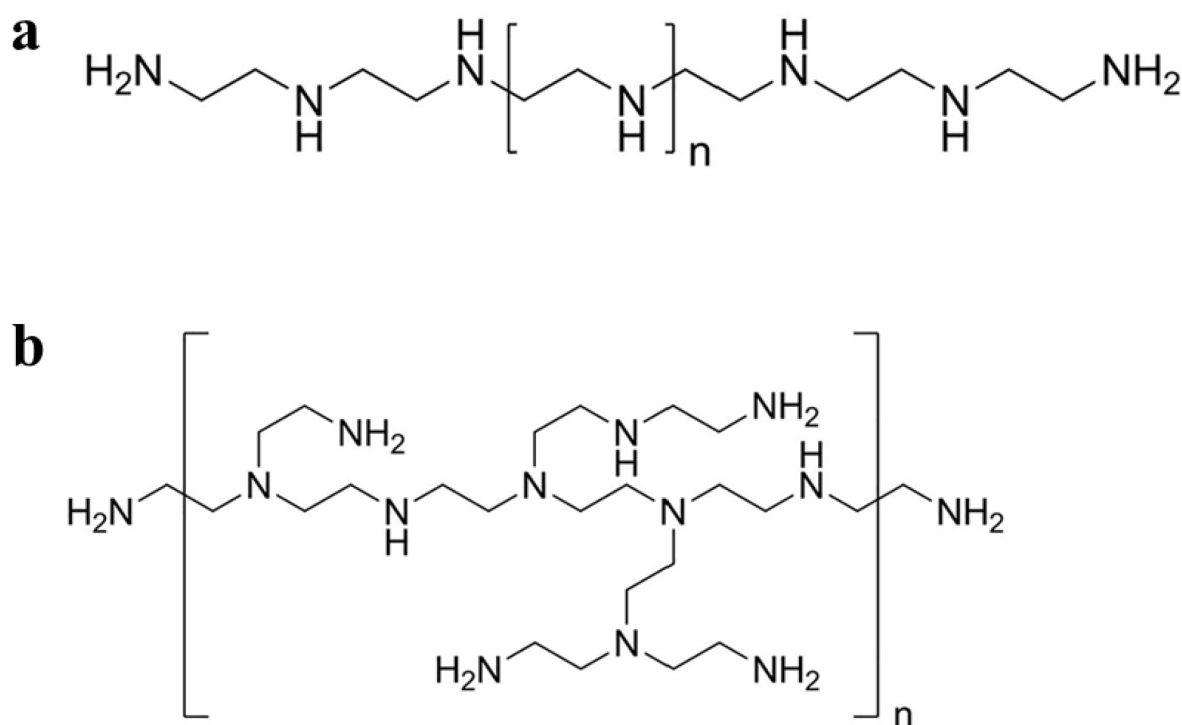


Figure 1. Schematic diagram of the chemical structures of (a) linear and (b) branched PEI.

Branched PEI is a hyperbranched polymer synthesized using the monomer method; that is, the cationic polymer is obtained by acid-catalyzed ring-opening polymerization of aziridine monomers [33]. Each branch of secondary amines in the branched chain of hyperbranched PEI has 3–35 nitrogen atoms on average. This branch distribution can form a spherical internal structure, which can encapsulate NPs, drug molecules, and other small molecules.

Furthermore, the lone pair electrons of nitrogen atoms in branched PEI can stabilize metal ions via coordination interaction. Therefore, branched PEI has a wide range of applications in gene transfection [34][35][36], drug delivery [37][38], and molecular imaging [25][39].

Linear PEI contains only secondary amines, whereas branched PEI contains various types of amines, i.e., primary, secondary, and tertiary. Linear PEI is solid at room temperature, in contrast to branched PEI which is liquid at all molecular weights [20]. Linear PEI is a high-charge cationic polymer and has been widely used in biomedical fields. For example, linear PEI has antibacterial properties against various pathogens and can therefore be used as a bacteriostatic agent [40][41]. Additionally, as a cationic polymer, linear PEI can form a polymer with nucleotides for gene transfer [42]. Compared with branched PEI, linear PEI is an effective nonviral gene vector with higher cell viability and transfection efficiency [43]. A balanced picture of the PEI studies including advantages and limits has been presented in **Figure 2**.

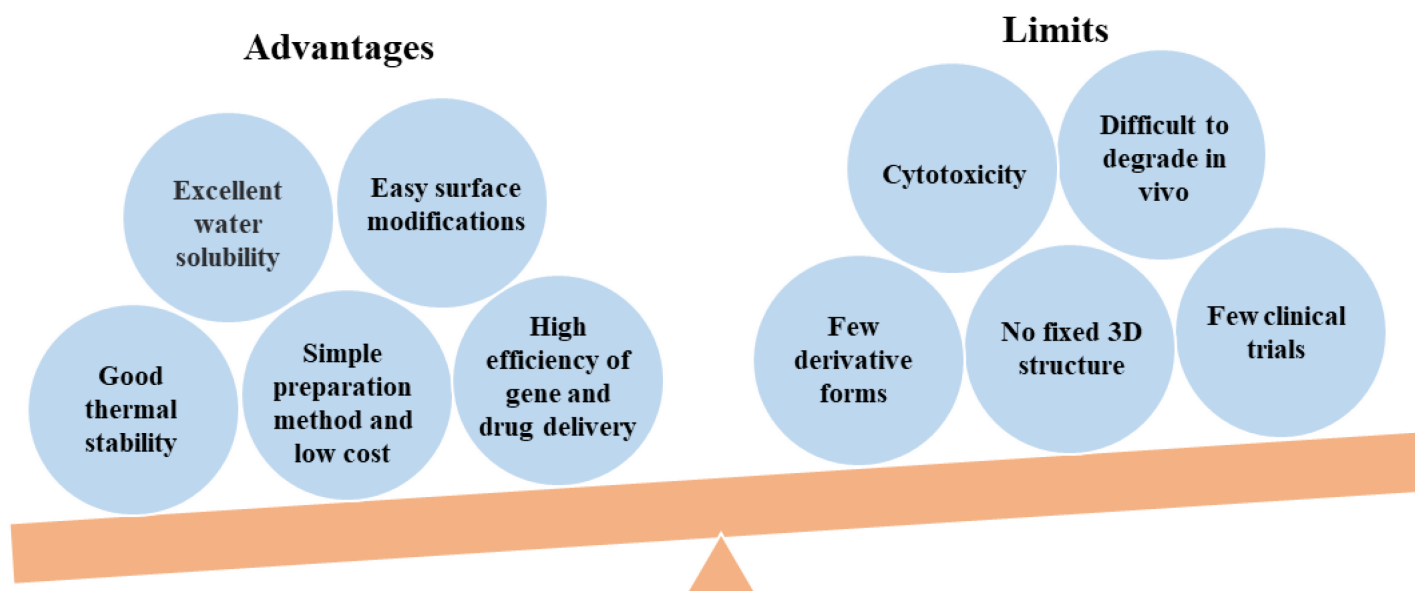


Figure 2. A balanced picture of PEI studies including advantages and limits.

3. PEI Modifications

As a cationic polymer, PEI contains abundant amino groups and as a result has a certain degree of cytotoxicity. Cationic PEI enters cells by adhering to negatively charged transmembrane heparanproteoglycans, which can cause cell damage through membrane destabilization [44]. Additionally, the internalized PEI causes apoptotic cell death by forming pores in the mitochondrial membrane [45][46]. PEI is not well-degraded in organisms, and its cytotoxicity is closely related to its molecular weight and branching degree [47]. Branched PEI with a higher molecular weight has a higher cytotoxicity. The surface amines of PEI can be shielded with simple modifications, thus significantly improving the biocompatibility of PEI [21]. At present, the surface amines of PEI are mainly shielded with covalent bonds such as carboxylation, acetylation, and hydroxylation, or with electrostatic modification of negatively charged proteins. However, currently, there is a lack of systematic research to contrast

the benefits and challenges of these approaches for the surface modifications of PEI. For example, Wen et al. improved the biocompatibility of PEI through carboxylation, acetylation, hydroxylation, and PEGylation [21]. These methods effectively reduced or shielded the positive charge of the PEI, thus reducing cytotoxicity. Various functional groups including polyethylene glycol (PEG), folic acid (FA), hyaluronic acid (HA), fluorescent tags, and protein can be modified with PEI for biomedical applications [24][25][48][49][50][51][52][53][54][55][56][57][58]. PEI modifications for biomedical applications in recent years are summarized in **Table 1**.

Table 1. Summary of PEI modifications carried out in recent years.

Modification Types	Aims	Ref.
Carboxylation modification	Gene delivery, absorption of heavy metals in sewage.	[59][60][61][62]
Acetylation modification	Gene delivery efficiency improvement, cytotoxicity reduction.	[62][63][64]
Hydroxylation modification	Biocompatibility enhancement, gene delivery, transformation improvement of NPs.	[65][66][67]
PEG modification	Stability and transfection efficiency improvement.	[68][69][70]
FA modification	Tumor-targeted delivery.	[71][72]
HA modification	Tumor-targeted gene delivery, stability improvement.	[73][74]
Protein modification	Gene delivery, protein transduction.	[75][76][77]
FI modification	Fluorescence imaging.	[57]

4. Synthesis of PEI-Based NPs

References

NP-based drug delivery systems with high biostability, targeting, and biodegradation have significantly improved clinical efficacy. [78][79][80][81]. Compared with traditional drug delivery systems, NP-based drug delivery systems can Upgrade Cancer Diagnosis and Treatment. Nanoscale Res. Lett. 2021, 16, 88. not only improve the water dispersibility and stability of drugs, but also significantly change the distribution and

2. Aghebiati-Maleki, A.; Dabaei, S.; Ahmadi, M.; Baghbanchi, A.; Asadi, M.; Fotouhi, A.; Yousefi, A.; Aghebiati-Maleki, A. Nanoparticles in cancer therapy: Perspectives for applications of nanoparticles in the treatment of cancers. *J. Cell. Physiol.* 2020, 235, 1962–1972.

NP-based drug delivery systems can be modified with targeted ligands to deliver drugs to specific tumor sites, thereby improving drug bioavailability and reducing toxicity to normal tissues. *Deliv. Rev.* 2002, 54, 631–651.

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hydrophobic cavities in hyperbranched PEI, small molecules, metal ions, and metal oxides can be effectively encapsulated to form stable NPs [25][55][56][58][82][83][84][85][86]. For example, Sun et al. used PEGylated PEI to coat carbon nano-onion clusters (CNOs) for cancer theranostics [87].

Kovács, Z. Lanthanide(III) complexes of tris(amide) PCTA derivatives as potential bimodal magnetic resonance and optical imaging agents. *Chem. Eur. J.* 2009, 15, 13188–13200.

The coating of PEGylated PEI can promote the phagocytosis efficiency of cells to the CNOs. The CNOs–PEI–PEG showed a cell uptake rate of 2.13 pg/cell, which was much higher than that of PBS and free CNOs. The CNOs–PEI–PEG was used as a photothermal and photoacoustic (PA) imaging agent for cancer theranostics because of its excellent photothermal conversion and cell phagocytosis efficiency. *Biomaterials* 2014, 35, 4688–4698.

In addition, the positively charged amino groups on the PEI surface can be bound to organic or inorganic anionic materials using electrostatic interaction. The lone electron of the amino group on the PEI surface can also coordinate with different metal atoms or metal ions to stabilize metal ions, metal oxides, or metal elements. For instance, Liu et al. conjugated PEI to the GO surface via amide bonds, which significantly improved the physiological stability and gene transfection rate of the GO [88].

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AuNPs based on hyperbranched PEI [90]. Note et al. further studied the influencing factors of the size of AuNPs based on hyperbranched PEI, and found that AuNPs with a particle size of less than 10 nm could be obtained in either the aqueous or microemulsion phase when heated to 100 °C [91].

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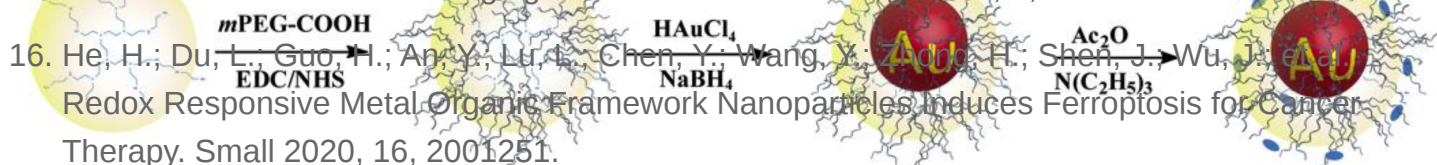
work, the researchers used PEGylated PEI to entrap and stabilize AuNPs (PP-AuNPs, as shown in Figure 3) or iron oxide NPs (PP-IONPs) for biomedical applications [92][93][94][95][96][97][98][99][100].

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Therapeutic Modalities	Therapeutic Agents	Cell Line Models	In Vivo Models	Ref.	sphere
Chemotherapy	DOX	HeLa	HeLa	[57]	;
	MTX	HCT 116	/	[119]	ed virus
	PTX	HepG2	/	[120]	
	DOX	C6	/	[121]	-dose
	DOX	HeLa	HeLa	[122]	25,
	DOX	4T1, HepG2	/	[123]	trices
	DOX	A549	/	[124]	
	DOX, siRNA	MDA-MB-231, HeLa, EAT	EAT	[125]	, T.; 95.
Gene therapy	DOX	SKBR3	SKBR3	[126]	/e geted In
	pDNA	HeLa, 16HBE14o-, HepG2	/	[127]	stry of J.
	pDNA	Huh7	Huh7	[128]	sed on
	DNA	NIH/3T3	/	[45]	217,
	pDNA	HeLa	/	[51]	ncer

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Therapeutic Modalities	Therapeutic Agents	Cell Line Models	In Vivo Models	Ref.
Staphylococcus aureus	DNA	HeLa, CT26	CT26	[129]
	mRNA	B16-OVA	B16-OVA	[130]
	RNase A	MDA-MB-231	/	[131]
	Oxidized mesoporous carbon nanospheres, pDNA	MCF-7	MCF-7	[132]
Other therapies	CAT-Ce6	T24	T24	[133]
	GO, DTX, anti-miRNA21	MDA-MB-231	/	[134]
	CuS, DTX, CpG	4T1	4T1	[135]
	pDNA, 9B9 mAb	SMMC-7721	SMMC-7721	[136]

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Imaging Types	Imaging Agents	Cell Line Models	In Vivo Models	Ref.
CT	AuNPs	A549	A549	[53]
	AuNPs	MCF-7	MCF-7	[139]
	AuNPs	HeLa	HeLa	[140] [141]
	Bi ₂ Se ₃ NPs	A549, U14	U14	[142]
	Gd ions	KB	KB	[143]
	Superparamagnetic iron oxide nanocrystals	MCF-7/Adr	/	[144]
	Superparamagnetic iron oxide NPs	Chondrolyte cells	/	[145]
MR	Ultrasmall iron oxide NPs	4T1	4T1	[85]
	Gd(OH)(3)-doped Fe ₃ O ₄ NPs	KB	/	[146]
	Fe ₃ O ₄ NPs	HepG2	HepG2	[147]
	Fe ₃ O ₄ NPs	U87MG, HeLa	U87MG, HeLa	[148]
SPECT	¹³¹ I	4T1	4T1	[149]
	^{99m} Tc	C6	C6	[150]

biosorbent for Ru biosorption from Ru-bearing acetic acid wastewater. Chem. Eng. J. 2013, 230, 303–307.

Imaging Types	Imaging Agents	Cell Line Models	In Vivo Models	Ref.	Line
MR/CT	AuNPs, Gd ₂ O ₃	HeLa	HeLa	[151]	derisi,
	Fe ₃ O ₄ @Au nanostars	HeLa	HeLa	[152]	
	Fe ₃ O ₄ @Au nanocomposites	KB	/	[49]	EI
	Au-Gd NPs	HeLa	HeLa	[54] [153]	roxyl- 3, 512–
MR/PA	Gd/CuS	KB	KB	[154]	Effects
SPECT/CT	^{99m} Tc, AuNPs	HCC-LM3	HCC-LM3	[58]	ethylene
	^{99m} Tc, AuNPs	SKOV-3	/	[86]	
	AuNPs, ¹³¹ I	C6	C6	[102]	
MR/CT/PA	Fe ₃ O ₄ NPs, Au nanostars	HeLa	HeLa	[155]	ect of ;
MR/SPECT/PA	¹⁹ F, ^{99m} Tc, ICG	HepG2	HepG2	[156]	
CT/MR/upconversion luminescence	Yb ³⁺ - and Gd ³⁺ -doped UCNPs	A2780	A2780	[157]	from

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The development of nanotechnology provides new strategies with regard to the combination of therapeutic drugs and imaging agents for imaging-guided cancer therapy, namely cancer theranostics [18][158][159]. As a highly cationic polymer, PEI has the advantages of low cost, easy surface functionalization, stable chemical properties, and high loading of small molecules and NPs, enabling it to be used to construct PEI-based drug delivery systems for gene carrier based on thermophilic histone and polyethylenimine. *New J. Chem.* 2015, 39, 6718–6728.

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cancer theranostics. As an example, Shi et al. used an inverse mini-emulsion method to prepare PEI-based hybrid

- nanogels for incorporation with ultrasmall iron oxide NPs and the anticancer drug DOX for T₁ MR imaging-guided chemotherapy of tumors [85]. The nanogels displayed excellent water solubility and colloidal stability, high DOX loading efficiency (51.4%), and a pH-dependent release of the DOX with an accelerated release rate under acidic content on the regulation of transgene expression by protein kinase C α -responsive linear pH. Compared to free ultrasmall iron oxide NPs, the nanogels showed a much higher r_1 relaxivity at 2.29 mM⁻¹ s⁻¹. polyethylenimine-peptide conjugates. *Colloids Surf. B Biointerfaces* 2014, 123, 123–129.
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