HDPs in Skin Wound Healing

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Cutaneous wound healing is a vital biological process that aids skin regeneration upon injury. Wound healing failure results from persistent inflammatory conditions observed in diabetes, or autoimmune diseases like psoriasis. Chronic wounds are incurable due to factors like poor oxygenation, aberrant function of peripheral sensory nervature, inadequate nutrients and blood tissue supply. The most significant hallmark of chronic wounds is heavily aberrant immune skin function. As the first line of defence against pathogenic bacterial infection, host defence peptides (HDPs) are a critical element in preventing biofilm-associated infections. A decline in sufficiently effective antibiotic treatments due to growing problems with antibiotic resistance may be ameliorated with alternatives to conventional antibiotics like peptides with antimicrobial properties. Aside from bacterial pathogen inactivation by disrupting their cell membranes, host defence peptides also have immunomodulatory properties, due to their ability to stimulate the cross-talk between immune cells promoting cutaneous wound healing in a healthier manner.

Keywords: antimicrobial peptides ; host defence peptides ; chronic wounds ; skin wound healing ; inflammation ; skin immune response

1. Host Defence Peptides in Wound Healing

Cationic peptides play a primarily role in maintaining the skin barrier's integrity and cutaneous tissue restoration during injury ^[1]. Due to expanding the effects of antimicrobial peptides on bacteria, HDP have been assigned the fundamental biological role in innate immunity ^[2]. Due to their lack of adaptive immune systems, arthropods and plants rely on their HDPs' primary defence response ^{[3][4]}. In higher eukaryotes, the levels of the host's defence peptides patrolling through signalling networks of the immune response are significantly more abundant, as their multifaceted role is more complex ^[5].

Skin resident HDPs are crucial participants of each step of the wound healing process: inflammation (neutrophil and macrophage infiltration), wound site regeneration (angiogenesis and re-epithelialization) and remodelling (restoration of tensile strength) $\frac{[1][G]}{2}$.

In humans, two main classes of host defence peptides have been identified: defensins and cathelicidins (**Figure 1**). However, there are also a variety of other small peptides expressed by epithelial cells like Substance P ^[Z], neurotensin ^[B], granulysin ^[9], calprotectin ^[10], adrenomedullin ^[11], MRP8/MRP-14 ^[12] and RNase A superfamily ^[13], which are also important wound inflammatory biomarkers.



Figure 1. Host defence peptides expressed by skin cells are induced by immune response, and act via signalling proteins belonging to growth factors like Vascular Endothelial Growth Factor (VEGF), Transforming Growth Factor beta (TGF β), Epidermal Growth Factor (EGF) and cytokines like Tumor Necrosis factor alpha (TNF α). Immune cells produce LL-37 from the cathelicidin family (Pdb code: 2K6O) ^[14] and defensin family: Human defensin 1 (Pdb code: 1IJU) ^[15], Human defensin 2 (Pdb code: 1FD4) ^[16], Human defensin 3 (Pdb code: 1KJ6) ^[17].

1.1. Defensins

Peptides from the defensin family are small peptides that are widely distributed across species, including humans. Human defensins (HDs) are produced in leukocytes and are also secreted by different epithelial cells and mucosal tissues $^{[15]}$. The mature defensins are described as short (28–42 amino acids length), cationic (net charge +1 to +11), amphipathic peptides with a highly conserved tertiary structure of a triple antiparallel β -sheet fold arrangement accommodating the six cysteine residues connected with three disulphide bridges $^{[16]}$.

Moreover, depending on the size, location, spatial conformation, and spot where the cysteines lay within the peptide chains, they are categorised as α -defensins, β -defensins and θ -defensins $\frac{[16][17]}{1}$. In humans, only α and β -defensins are present, while structurally different, cyclic θ -defensins have been identified in rhesus macaques $\frac{[16][18]}{1}$.

Additionally, the HDs have antimicrobial activity against various strains of Gram-positive and negative bacteria $^{[19]}$, fungi $^{[20]}$ and viruses such as the herpes simplex virus $^{[21]}$.

In humans, there are 6 α -defensins: HNPs1–4 and human α -defensins 5 and 6 (HD5 and HD6) ^{[19][22]}. HNPs 1–3 (human neutrophil peptides) differ by single amino acid substitutions and are predominantly produced by neutrophils.

Moreover, β -defensins abundance is tightly governed by transcriptional controls assigned to epithelial and epidermal cells ^[23]. One representative of the β -defensin family such as human β -defensin 1 (HBD1) is continuously transcribed in skin cells, while the transcription of human β -defensin 2 (HBD2) and human β -defensin 3 (HBD3) as the others is triggered in response to a microbial or pro-inflammatory cytokines presence ^[24].

1.2. Cathelicidins

Endogenous cathelicidins are stored at high concentrations as inactive precursors in granules of mammalian neutrophils and mast cells. In contrast to defensins, cathelicidins are predominantly α -helical, amphipathic, cationic (possessing a net positive charge of 2–9) and consist of 23–37 amino acids ^[25].

Cathelicidins are named after the conserved cathelin-like domain, and are produced as protein precursors, containing a conserved amino-terminal (N-terminal) signal peptide domain, an antimicrobial C-terminal domain mature peptide and cathelin-like domain ^[26].

The active form of peptide is formed once the cathelin domain is cleaved by serine proteases upon neutrophils degranulation and secretion of peptides ^[27].

The gene for hCAP-18, named *CAMP*, encodes the human cathelicidin. LL-37, the only member of the cathelicidin family identified so far in humans, is present in the secondary (specific) granules of neutrophils, macrophages, and epithelial cells ^[28]. Human cathelicidin hCAP-18, is processed by proteinases into several extracellular cleavage products, with LL-37 being one of them ^[29]. Cathelicidins play an important regulatory role in the inflammatory response ^[30]. Many studies support immunomodulatory actions over the antimicrobial actions of cathelicidins ^{[2][25]}. However, the constitutive generation of the cathelicidins across the various species provides them with strong antimicrobial protection from bacteria ^[31], viruses ^[32] or fungi ^[33].

2. Cellular Regulation and Immunomodulatory Actions of Host Defence Peptides during Wound Healing

Factors affecting wound healing can be categorised into systemic and local factors. Systemic factors are present at all times in the individual, as seen in diseases such as diabetes, stress, obesity and age, while local factors include factors, such as infection, oxygenation and foreign bodies, which directly influence the wound locally ^[34]. The local factors lead to high levels of pro-inflammatory cytokines and ROS, impaired cell and protease function and a lack of growth factors ^[35]. HDPs have been demonstrated to influence many local factors, such as regulating expression of cytokines ^[36], chemokines ^[37], proteases ^[38], growth factors ^[39] and immune cell activity ^{[40][41][42]} (**Table 1**).

Human Host Defence Peptide	Immunomodulatory Action	Reference
Inflammatory phase		
LL-37	Cell migration mediated by EGFR and ERK1/2	<u>[43]</u>
	Recruitment of leukocytes	[44]
	Secretion of anti-inflammatory cytokines like IL-1RA	[<u>45</u>]
	Enhanced SMAD2/3 and STAT3 phosphorylation	[46]
LL-37, hBD-2, hBD-3, hBD-4	Activation of the p38 and ERK1/2 MAPK pathways	[<u>47</u>]
hBD-2, hBD-3	Activate plasmacytoid dendritic cells (pDCs)	<u>[48]</u>
S100A7	Increased NFkB/p38MAPK/Caspase-1/IL-1a signalling	[49]
Proliferative phase		
LL-37	Induction of growth factors such as EGF and VEGF	[<u>43</u>]
	Neovascularization	[50]
hBD-2, hBD-3	Keratinocyte migration and proliferation	<u>[51]</u>
Remodelling phase		
LL-37	Polarisation to M1 macrophages	[30]
Cathelicidin-WA (CWA)	Suppressed phosphorylation of STAT1 and NF-ĸB, repressed phosphorylation of STAT6	[<u>52</u>]
S100A7	Aberrant response of fibroblasts and endothelial cells	[<u>53]</u>

Table 1. Host defence peptides immunomodulatory actions in wound healing phases.

2.1. Host Defense Peptides Triggered by Inflammation

Many acute and chronic inflammatory disorders have been correlated with dysregulation of the natural HDP response ^[2] ^[54]. Although the reciprocity of either deficiency or overproduction of HDPs in the presence of inflammation and thus balance between pro- or anti-inflammatory effects is not so easy to define directly, this can lead to a pathological inflammatory response ^{[55][56]}. Endogenous host defence peptides, stored intracellularly at high concentrations as inactive precursors in granules, are released locally at inflection and inflammation sites, whereas the expression of others is induced in response to pathogen-associated molecules ^[57].

The role of the immune cells in the inflammatory phase is intended to eliminate intracellular pathogens, and this is achieved by producing host defence peptides, which in turn promote the robust recruitment of immune cells such as neutrophils, monocytes/macrophages, dendritic cells, and T cells ^[2]. Altered levels of either pro- $\frac{[58][59]}{2}$ or anti-

inflammatory ^{[37][60][61]} cytokines have been reported in inflammatory conditions. A persistent inflammatory environment has been demonstrated to affect not only the function of immune cells ^[62], but also the composition of the host defence peptides detected at the wound site ^[63] (**Figure 2**).



Figure 2. Immune response triggers skin cells to secrete host defence peptides. The secreted peptides are involved in antibacterial killing but also the recruitment of other immune cells to the wounded area.

Defensins and cathelicidins family members have distinct roles in various disease states, and many studies support the potential synergistic roles of defensins and cathelicidins in cytokines chemoattraction in skin immune responses ^[47]. Tightly coordinated actions of hBD-2, -3 and -4 along with LL-37 induce secretion of IL-18 by keratinocytes through activation of the p38 and ERK1/2 MAPK pathways ^[47]. Under inflammatory skin conditions, high expression levels of keratinocyte hBD-2 induced by IL-17A and TNF α indicate that it may be considered as a marker for disease severity ^[64].

Inflammatory skin conditions like psoriasis are linked to elevated levels of both IL-1a and S100A7 (also known as psoriasin) and an increased signalling axis composed of NF- κ B/p38MAPK/Caspase-1/IL-1a, which regulates S100A7 ^[49]. Moreover, the increased expression of LL-37 is associated with overproduction of IFNy in psoriatic lesions, while hBD-2, hBD-3 and lysozyme can activate plasmacytoid dendritic cells (pDCs) in co-action with LL-37 ^[48]. LL-37 evokes pro-inflammatory chemotaxis through binding to the formyl peptide-like receptor-2 (Fpr-2), a cell sensing element for microbial products ^[44]. The human host defence peptide LL-37 binds to these G protein–coupled receptors and activates mucosal immune response through recruitment of leukocytes ^[44]. In contrast, LL-37 can mediate anti-inflammatory responses such as secretion of anti-inflammatory cytokines like IL-1RA ^[45]. Another prominent feature of the neutrophil released cathelicidins is enhanced SMAD2/3 and STAT3 phosphorylation in the presence of transforming growth factor- β (TGF β), shifting the T-cells subset towards Th17 rather than the Th1 phenotype, resulting in Th17 but not Th1 cells protection from apoptosis ^[46].

Atopic dermatitis (eczema) is another pathological skin condition ^[65]. Unlike the psoriasis which is characterised by elevated levels of innate immunity peptides, in atopic dermatitis, host defence peptide levels are not abundant and together with skin dryness make affected areas prone to infections ^[66].

2.2. Host Defense Peptides Involved in Proliferative Phase of Wound Healing

It has been shown that LL-37 peptide induces growth factors, such as EGF and VEGF, but also binds their receptors ^[43]. Aberrant vascularization during wound restoration in mice lacking the CRAMP (the murine homologue of LL-37/hCAP-18), shows that the peptide can stimulate endothelial cells, increasing the proliferation rate and thus enhancing the dermal neovascularization ^[50].

Furthermore, high levels of hBD-2 and hBD-3 detected at wound sites promote keratinocyte migration and proliferation, indicating their involvement in the re-epithelialization of the healing epithelium $\frac{[51][68]}{100}$. A relatively recent study demonstrated that topical insulin delivery to the wound enhanced the levels of extracellular-signal regulated kinase (ERK) and protein kinase B (Akt) $\frac{[69]}{100}$. ERK is a part of a phosphorylation pathway that activates gene transcription leading to cell $\frac{[70][71]}{100}$ and increased Akt stimulated angiogenesis through activation VEGF signalling $\frac{[72][73]}{[Figure 3]}$.





2.3. Host Defense Peptides in Tissue Remodelling Phase

The direction of macrophages polarisation into relevant phenotypes during different phases of wound healing (i.e., polarisation to a pro-inflammatory phenotype during the inflammatory phase and anti-inflammatory phenotype during the subsequent proliferation and remodelling phase) $^{[74][75]}$ is controlled by many factors, while different HDPs (like LL-37) promote polarisation to M1 macrophages $^{[30]}$. Similarly, elevated levels of interleukin-6 (IL-6), IL-1 β , tumour necrosis factor- α (TNF α) and chemokine CCL3 in RAW264.7 cells can be dampened by Cathelicidin-WA (CWA) $^{[52]}$. CWA peptide suppresses the expression of TLR-4 and the phosphorylation of STAT1 and NF- κ B, downregulating the activity of pro-inflammatory macrophages while stimulating the phosphorylation of STAT6 and activating *E. coli* K88-induced anti-inflammatory macrophages $^{[52]}$.

An aberrant response described as overgrowth of fibroblasts and endothelial cells may be due to high concentrations of psoriasin in wound fluid and granulation tissue ^[53]. Psoriasin stimulates the tissue remodelling by inhibiting the excessive production of collagen, fibronectin and TGF β formation in fibroblasts. In keloid-derived fibroblasts, psoriasin production is decreased ^[53] (**Figure 3**).

References

- 1. Otvos, L.; Ostorhazi, E. Therapeutic utility of antibacterial peptides in wound healing. Expert Rev. Anti. Infect. Ther. 201 5, 13, 871–881.
- 2. Hancock, R.E.W.; Haney, E.F.; Gill, E.E. The immunology of host defence peptides: Beyond antimicrobial activity. Nat. Rev. Immunol. 2016, 16, 321–334.
- Campos, M.L.; De Souza, C.M.; De Oliveira, K.B.S.; Dias, S.C.; Franco, O.L. The role of antimicrobial peptides in plant immunity. J. Exp. Bot. 2018, 69, 4997–5011.
- 4. Mylonakis, E.; Podsiadlowski, L.; Muhammed, M.; Vilcinskas, A. Diversity, evolution and medical applications of insect antimicrobial peptides. Philos. Trans. R. Soc. B Biol. Sci. 2016, 371, 20150290.
- 5. Browne, K.; Chakraborty, S.; Chen, R.; Willcox, M.D.P.; Black, D.S.; Walsh, W.R.; Kumar, N. A new era of antibiotics: T he clinical potential of antimicrobial peptides. Int. J. Mol. Sci. 2020, 21, 7047.
- 6. Steinstraesser, L.; Koehler, T.; Jacobsen, F.; Daigeler, A.; Goertz, O.; Langer, S.; Kesting, M.; Steinau, H.; Eriksson, E.; Hirsch, T. Host defense peptides in wound healing. Mol. Med. 2008, 14, 528–537.
- Choi, H.; Kim, D.; Nam, S.; Lim, S.; Hwang, J.S.; Park, K.S.; Hong, H.S.; Won, Y.; Shin, M.K.; Chung, E.; et al. Substan ce P restores normal skin architecture and reduces epidermal infiltration of sensory nerve fiber in TNCB-induced atopic dermatitis-like lesions in NC/Nga mice. J. Dermatol. Sci. 2018, 89, 248–257.
- Zheng, Z.; Liu, Y.; Huang, W.; Mo, Y.; Lan, Y.; Guo, R.; Cheng, B. Neurotensin-loaded PLGA/CNC composite nanofiber membranes accelerate diabetic wound healing. Artif. Cells Nanomed. Biotechnol. 2018, 46, 493–501.
- Chung, W.H.; Hung, S.I.; Yang, J.Y.; Su, S.C.; Huang, S.P.; Wei, C.Y.; Chin, S.W.; Chiou, C.C.; Chu, S.C.; Ho, H.C.; et al. Granulysin is a key mediator for disseminated keratinocyte death in Stevens-Johnson syndrome and toxic epidermal necrolysis. Nat. Med. 2008, 14, 1343–1350.
- Ingram, J.R.; Cawley, S.; Coulman, E.; Gregory, C.; Thomas-Jones, E.; Pickles, T.; Cannings-John, R.; Francis, N.A.; H arding, K.; Hood, K.; et al. Levels of wound calprotectin and other inflammatory biomarkers aid in deciding which patien ts with a diabetic foot ulcer need antibiotic therapy (INDUCE study). Diabet. Med. 2018, 35, 255–261.

- 11. Hans, M.; Madaan Hans, V. Epithelial antimicrobial peptides: Guardian of the oral cavity. Int. J. Pept. 2014, 2014, 3702 97.
- 12. Croce, K.; Gao, H.; Wang, Y.; Mooroka, T.; Sakuma, M.; Shi, C.; Sukhova, G.K.; Packard, R.R.S.; Hogg, N.; Libby, P.; et al. Myeloid-related protein-8/14 is critical for the biological response to vascular injury. Circulation 2009, 120, 427–436.
- 13. Rademacher, F.; Simanski, M.; Harder, J. RNase 7 in cutaneous defense. Int. J. Mol. Sci. 2016, 17, 560.
- Jacobsen, A.S.; Jenssen, H. Human cathelicidin LL-37 prevents bacterial biofilm formation. Future Med. Chem. 2012, 4, 1587–1599.
- 15. Fruitwala, S.; El-Naccache, D.W.; Chang, T.L. Multifaceted immune functions of human defensins and underlying mech anisms. Semin. Cell Dev. Biol. 2019, 88, 163–172.
- 16. Shafee, T.M.A.; Lay, F.T.; Phan, T.K.; Anderson, M.A.; Hulett, M.D. Convergent evolution of defensin sequence, structur e and function. Cell. Mol. Life Sci. 2017, 74, 663–682.
- 17. Howell, K.; de Leeuw, E. Cell adhesion properties of human defensins. Biochem. Biophys. Res. Commun. 2018, 502, 2 38–242.
- Ottolini, B.; Hornsby, M.J.; Abujaber, R.; MacArthur, J.A.L.; Badge, R.M.; Schwarzacher, T.; Albertson, D.G.; Bevins, C. L.; Solnick, J.V.; Hollox, E.J. Evidence of convergent evolution in humans and macaques supports an adaptive role for copy number variation of the β-defensin-2 gene. Genome Biol. Evol. 2014, 6, 3025–3038.
- Ericksen, B.; Wu, Z.; Lu, W.; Lehrer, R.I. Antibacterial Activity and Specificity of Six Human Alpha-Defensins. Antimicro b. Agents Chemother. 2005, 49, 269–275.
- Basso, V.; Garcia, A.; Tran, D.Q.; Schaal, J.B.; Tran, P.; Ngole, D.; Aqeel, Y.; Tongaonkar, P.; Ouellette, A.J.; Selsteda, M.E. Fungicidal Potency and Mechanisms of θ- Defensins against Multidrug-Resistant Candida Species. Antimicrob. A gents Chemother. 2018, 62, e00111-18.
- 21. Jenssen, H. Therapeutic approaches using host defence peptides to tackle herpes virus infections. Viruses 2009, 1, 93 9–964.
- 22. Xu, D.; Lu, W. Defensins: A Double-Edged Sword in Host Immunity. Front. Immunol. 2020, 11, 764.
- Semple, F.; Dorin, J.R. β-Defensins: Multifunctional modulators of infection, inflammation and more? J. Innate Immun. 2012, 4, 337–348.
- 24. Contreras, G.; Shirdel, I.; Braun, M.S.; Wink, M. Defensins: Transcriptional regulation and function beyond antimicrobial activity. Dev. Comp. Immunol. 2020, 104, 103556.
- 25. van Harten, R.M.; van Woudenbergh, E.; van Dijk, A.; Haagsman, H.P. Cathelicidins: Immunomodulatory antimicrobial s. Vaccines 2018, 6, 63.
- 26. Engelberg, Y.; Landau, M. The Human LL-37(17-29) antimicrobial peptide reveals a functional supramolecular structur e. Nat. Commun. 2020, 11, 3894.
- 27. Chen, J.; Zhai, Z.; Long, H.; Yang, G.; Deng, B.; Deng, J. Inducible expression of defensins and cathelicidins by nutrien ts and associated regulatory mechanisms. Peptides 2020, 123, 170177.
- Podaza, E.; Palacios, F.; Croci, D.O.; Risnik, D.; Yan, X.J.; Almejún, M.B.; Colado, A.; Elías, E.E.; Borge, M.; Morande, P.E.; et al. Expression and function of cathelicidin hCAP18/LL-37 in chronic lymphocytic leukemia. Haematologica 202 0, 105, e465–e469.
- 29. Khurshid, Z.; Naseem, M.; Asiri, F.Y.I.; Mali, M.; Khan, R.S.; Sahibzada, H.A.; Zafar, M.S.; Moin, S.F.; Khan, E. Significa nce and diagnostic role of antimicrobial cathelicidins (LI-37) peptides in oral health. Biomolecules 2017, 7, 80.
- Scheenstra, M.R.; van Harten, R.M.; Veldhuizen, E.J.A.; Haagsman, H.P.; Coorens, M. Cathelicidins Modulate TLR-Act ivation and Inflammation. Front. Immunol. 2020, 11, 1137.
- Coorens, M.; Scheenstra, M.R.; Veldhuizen, E.J.A.; Haagsman, H.P. Interspecies cathelicidin comparison reveals diver gence in antimicrobial activity, TLR modulation, chemokine induction and regulation of phagocytosis. Sci. Rep. 2017, 7, 40874.
- 32. Tripathi, S.; Wang, G.; White, M.; Qi, L.; Taubenberger, J.; Hartshorn, K.L. Antiviral activity of the human cathelicidin, LL -37, and derived peptides on seasonal and pandemic influenza A viruses. PLoS ONE 2015, 10, e0124706.
- Van Eijk, M.; Boerefijn, S.; Cen, L.; Rosa, M.; Morren, M.J.H.; Van Der Ent, C.K.; Kraak, B.; Dijksterhuis, J.; Valdes, I. D.; Haagsman, H.P.; et al. Cathelicidin-inspired antimicrobial peptides as novel antifungal compounds. Med. Mycol. 202 0, 58, 1073–1084.
- 34. Guo, S.; DiPietrio, L. Factors Affecting Wound Healing. J. Dent. Res. 2010, 89, 219–229.
- 35. Frykberg, R.G.; Banks, J. Challenges in the Treatment of Chronic Wounds. Adv. Wound Care 2015, 4, 560–582.

- Capparelli, R.; De Chiara, F.; Nocerino, N.; Montella, R.C.; Iannaccone, M.; Fulgione, A.; Romanelli, A.; Avitabile, C.; Bl aiotta, G.; Capuano, F. New perspectives for natural antimicrobial peptides: Application as antinflammatory drugs in a murine model. BMC Immunol. 2012, 13, 61.
- Scott, M.G.; Dullaghan, E.; Mookherjee, N.; Glavas, N.; Waldbrook, M.; Thompson, A.; Wang, A.; Lee, K.; Doria, S.; Ha mill, P.; et al. An anti-infective peptide that selectively modulates the innate immune response. Nat. Biotechnol. 2007, 2 5, 465–472.
- Wiegand, C.; Schönfelder, U.; Abel, M.; Ruth, P.; Kaatz, M.; Hipler, U.C. Protease and pro-inflammatory cytokine conce ntrations are elevated in chronic compared to acute wounds and can be modulated by collagen type i in vitro. Arch. Der matol. Res. 2010, 302, 419–428.
- 39. Koczulla, R.; Bals, R. Cathelicidin Antimicrobial Peptides Modulate Angiogenesis. In Therapeutic Neovascularization–Q uo Vadis? Deindl, E., Kupatt, C., Eds.; Springer: Dordrecht, The Netherlands, 2007; pp. 191–196.
- 40. Choi, K.Y.; Chow, L.N.Y.; Mookherjee, N. Cationic host defence peptides: Multifaceted role in immune modulation and i nflammation. J. Innate Immun. 2012, 4, 361–370.
- 41. Huang, H.J.; Ross, C.R.; Blecha, F. Chemoattractant properties of PR-39, a neutrophil antibacterial peptide. J. Leukoc. Biol. 1997, 61, 624–629.
- 42. Welkos, S.; Cote, C.K.; Hahn, U.; Shastak, O.; Jedermann, J.; Bozue, J.; Jung, G.; Ruchala, P.; Pratikhya, P.; Tang, T.; et al. Humanized θ-defensins (retrocyclins) enhance macrophage performance and protect mice from experimental ant hrax infections. Antimicrob. Agents Chemother. 2011, 55, 4238–4250.
- Tokumaru, S.; Sayama, K.; Shirakata, Y.; Komatsuzawa, H.; Ouhara, K.; Hanakawa, Y.; Yahata, Y.; Dai, X.; Tohyama, M.; Nagai, H.; et al. Induction of Keratinocyte Migration via Transactivation of the Epidermal Growth Factor Receptor by the Antimicrobial Peptide LL-37. J. Immunol. 2005, 175, 4662–4668.
- 44. Kim, S.-H.; Kim, Y.N.; Jang, Y.-S. Cutting Edge: LL-37–Mediated Formyl Peptide Receptor-2 Signaling in Follicular Den dritic Cells Contributes to B Cell Activation in Peyer's Patch Germinal Centers. J. Immunol. 2017, 206, 1600886.
- 45. Hemshekhar, M.; Choi, K.Y.G.; Mookherjee, N. Host defense peptide LL-37-mediated chemoattractant properties, but n ot anti-inflammatory cytokine IL-1RA production, is selectively controlled by Cdc42 Rho GTPase via G protein-coupled r eceptors and JNK mitogen-activated protein kinase. Front. Immunol. 2018, 9, 1871.
- Minns, D.; Smith, K.J.; Alessandrini, V.; Hardisty, G.; Melrose, L.; Jackson-Jones, L.; MacDonald, A.S.; Davidson, D.J.; Gwyer Findlay, E. The neutrophil antimicrobial peptide cathelicidin promotes Th17 differentiation. Nat. Commun. 2021, 12, 1285.
- A7. Niyonsaba, F.; Ushio, H.; Nagaoka, I.; Okumura, K.; Ogawa, H. The Human β-Defensins (-1, -2, -3, -4) and Cathelicidin LL-37 Induce IL-18 Secretion through p38 and ERK MAPK Activation in Primary Human Keratinocytes. J. Immunol. 200 5, 175, 1776–1784.
- Lande, R.; Chamilos, G.; Ganguly, D.; Demaria, O.; Frasca, L.; Durr, S.; Conrad, C.; Schröder, J.; Gilliet, M. Cationic an timicrobial peptides in psoriatic skin cooperate to break innate tolerance to self-DNA. Eur. J. Immunol. 2015, 45, 203–2 13.
- 49. Bhatt, T.; Bhosale, A.; Bajantri, B.; Mathapathi, M.S.; Rizvi, A.; Scita, G.; Majumdar, A.; Jamora, C. Sustained Secretion of the Antimicrobial Peptide S100A7 Is Dependent on the Downregulation of Caspase-8. Cell Rep. 2019, 29, 2546–255 5.e4.
- 50. Koczulla, R.; Von Degenfeld, G.; Kupatt, C.; Krötz, F.; Zahler, S.; Gloe, T.; Issbrücker, K.; Unterberger, P.; Zaiou, M.; Le bherz, C.; et al. An angiogenic role for the human peptide antibiotic LL-37/hCAP-18. J. Clin. Invest. 2003, 111, 1665–16 72.
- 51. Raziyeva, K.; Kim, Y.; Zharkinbekov, Z.; Kassymbek, K.; Jimi, S.; Saparov, A. Immunology of acute and chronic wound healing. Biomolecules 2021, 11, 700.
- 52. Chen, S.; Lu, Z.; Wang, F.; Wang, Y. Cathelicidin-WA polarizes E. coli K88-induced M1 macrophage to M2-like macroph age in RAW264.7 cells. Int. Immunopharmacol. 2018, 54, 52–59.
- 53. Gauglitz, G.G.; Bureik, D.; Zwicker, S.; Ruzicka, T.; Wolf, R. The antimicrobial peptides psoriasin (s100a7) and koebneri sin (S100A15) suppress extracellular matrix production and proliferation of human fibroblasts. Skin Pharmacol. Physiol. 2015, 28, 115–123.
- 54. Hilchie, A.L.; Wuerth, K.; Hancock, R.E. Immune modulation by multifaceted cationic host. Nat. Chem. Biol. 2013, 9, 76 1–768.
- 55. Kwiecien, K.; Zegar, A.; Jung, J.; Brzoza, P.; Kwitniewski, M.; Godlewska, U.; Grygier, B.; Kwiecinska, P.; Morytko, A.; C ichy, J. Architecture of antimicrobial skin defense. Cytokine Growth Factor Rev. 2019, 49, 70–84.

- 56. Marcinkiewicz, M.; Majewski, S. The role of antimicrobial peptides in chronic inflammatory skin diseases. Postep. Derm atol. Alergol. 2016, 33, 6–12.
- 57. Versey, Z.; da Cruz Nizer, W.S.; Russell, E.; Zigic, S.; DeZeeuw, K.G.; Marek, J.E.; Overhage, J.; Cassol, E. Biofilm-Inn ate Immune Interface: Contribution to Chronic Wound Formation. Front. Immunol. 2021, 12, 648554.
- 58. Pereira, L.; Neves, B.M.; Moura, L.; Cruz, M.T.; Carvalho, E. Neurotensin Decreases the Proinflammatory Status of Hu man Skin Fibroblasts and Increases Epidermal Growth Factor Expression. Int. J. Inflam. 2014, 2014, 248240.
- 59. Turner-Brannen, E.; Choi, K.Y.; Lippert, D.N.D.; Cortens, J.P.; Hancock, R.E.W.; El-Gabalawy, H.; Mookherjee, N. Mod ulation of interleukin-1β-induced inflammatory responses by a synthetic cationic innate defence regulator peptide, IDR-1002, in synovial fibroblasts. Arthritis Res. Ther. 2011, 13, R129.
- Mookherjee, N.; Hamill, P.; Gardy, J.; Blimkie, D.; Falsafi, R.; Chikatamarla, A.; Arenillas, D.J.; Doria, S.; Kollmann, T.R.; Hancock, R.E.W. Systems biology evaluation of immune responses induced by human host defence peptide LL-37 in mononuclear cells. Mol. Biosyst. 2009, 5, 483–496.
- Nijnik, A.; Madera, L.; Ma, S.; Waldbrook, M.; Elliott, M.R.; Easton, D.M.; Mayer, M.L.; Mullaly, S.C.; Kindrachuk, J.; Jen ssen, H.; et al. Synthetic cationic peptide IDR-1002 provides protection against bacterial infections through chemokine i nduction and enhanced leukocyte recruitment. J. Immunol. 2010, 184, 2539–2550.
- 62. Casqueiro, J.; Casqueiro, J.; Alves, C. Infections in patients with diabetes mellitus: A review of pathogenesis. Indian J. Endocrinol. Metab. 2012, 16 (Suppl. 1), S27–S36.
- 63. Prasad, S.V.; Fiedoruk, K.; Daniluk, T.; Piktel, E.; Bucki, R. Expression and function of host defense peptides at inflamm ation sites. Int. J. Mol. Sci. 2020, 21, 104.
- 64. Kolbinger, F.; Loesche, C.; Valentin, M.-A.; Jiang, X.; Cheng, Y.; Jarvis, P.; Peters, T.; Calonder, C.; Bruin, G.; Polus, F.; et al. β-Defensin 2 is a responsive biomarker of IL-17A–driven skin pathology in patients with psoriasis. J. Allergy Clin. I mmunol. 2017, 139, 923–932.e8.
- 65. Kim, B.E.; Leung, D.Y.M. Significance of skin barrier dysfunction in atopic dermatitis. Allergy Asthma Immunol. Res. 20 18, 10, 207–215.
- 66. Nguyen, H.L.T.; Trujillo-Paez, J.V.; Umehara, Y.; Yue, H.; Peng, G.; Kiatsurayanon, C.; Chieosilapatham, P.; Song, P.; O kumura, K.; Ogawa, H.; et al. Role of antimicrobial peptides in skin barrier repair in individuals with atopic dermatitis. In t. J. Mol. Sci. 2020, 21, 7607.
- 67. Mookherjee, N.; Hancock, R.E.W. Cationic host defence peptides: Innate immune regulatory peptides as a novel appro ach for treating infections. Cell. Mol. Life Sci. 2007, 64, 922–933.
- 68. Macleod, A.S.; Mansbridge, J.N. The Innate Immune System in Acute and Chronic Wounds. Adv. Wound Care 2015, 5, 65–78.
- 69. Lima, M.H.M.; Caricilli, A.M.; de Abreu, L.L.; Araújo, E.P.; Pelegrinelli, F.F.; Thirone, A.C.P.; Tsukumo, D.M.; Pessoa, A. F.M.; dos Santos, M.F.; de Moraes, M.A.; et al. Topical insulin accelerates wound healing in diabetes by enhancing the AKT and ERK pathways: A double-blind placebo-controlled clinical trial. PLoS ONE 2012, 7, e36974.
- 70. Gao, L.C.; Li, T.T.; Li, F.Y.; Tang, L.; Li, H.Y.; Wang, X.J.; Li, X.K. Role of ERK-1/2 signaling pathway in keratinacyte gro wth factor-2 stimulating human keratinocyte proliferation. Chin. Pharmacol. Bull. 2011, 27, 1405–1408.
- Yoon, S.; Seger, R. The extracellular signal-regulated kinase: Multiple substrates regulate diverse cellular functions. Gr owth Factors 2006, 24, 21–44.
- 72. Somanath, P.R.; Chen, J.; Byzova, T.V. Akt1 is necessary for the vascular maturation and angiogenesis during cutaneo us wound healing. Angiogenesis 2008, 11, 277–288.
- 73. Mouritzen, M.V.; Carvalho, E.; Abourayale, S.; Ejaz, R.; Ardon, C.B.; Roursgaard, M.; Dalgaard, L.T. Neurotensin, subst ance P, and insulin enhance cell migration. J. Pept. Sci. 2018, 24, e3093.
- 74. Hesketh, M.; Sahin, K.B.; West, Z.E.; Murray, R.Z. Macrophage phenotypes regulate scar formation and chronic wound healing. Int. J. Mol. Sci. 2017, 18, 1545.
- 75. Krzyszczyk, P.; Schloss, R.; Palmer, A.; Berthiaume, F. The role of macrophages in acute and chronic wound healing a nd interventions to promote pro-wound healing phenotypes. Front. Physiol. 2018, 9, 419.