

Heart Diet in Prevention of Heart Failure

Subjects: **Health Care Sciences & Services**

Contributor: Ram B. Singh , Jan Fedacko , Dominik Pella , Ghizal Fatima , Galal Elkilany , Mahmood Moshiri , Krasimira Hristova , Patrik Jakabcin , Natalia Vaňova

Antioxidants, such as polyphenolics and flavonoids, omega-3 fatty acids, and other micronutrients that are rich in Indo-Mediterranean-type diets, could be protective in sustaining the oxidative functions of the heart. The cardiomyocytes use glucose and fatty acids for the physiological functions depending upon the metabolic requirements of the heart. Apart from toxicity due to glucose, lipotoxicity also adversely affects the cardiomyocytes, which worsen in the presence of deficiency of endogenous antioxidants and deficiency of exogenous antioxidant nutrients in the diet. The high-sugar-and-high-fat-induced production of ceramide, advanced glycation end products (AGE) and triamino-methyl-N-oxide (TMAO) can predispose individuals to oxidative dysfunction and Ca-overloading. The alteration in the biology may start with normal cardiac cell remodeling to biological remodeling due to inflammation. It is proposed that a greater intake of high exogenous antioxidant restorative treatment (HEART) diet, polyphenolics and flavonoids, as well as cessation of red meat intake and egg, can cause improvement in the oxidative function of the heart, by inhibiting oxidative damage to lipids, proteins and DNA in the cell, resulting in beneficial effects in the early stage of the Six Stages of heart failure (HF).

Western diet

cardiomyocyte

oxidative stress

bioactive agents

dietary fat

cardiac failure

1. Oxidative Dysfunction in Heart Failure

It seems that behavioral risk factors such as Western diet, tobacco and alcohol intake, short sleep, and mental stress can cause an overproduction of free radicals, oxidative myocardial dysfunction and inflammation, which may alter the twist of the heart due to cardiomyocyte dysfunction and physiological remodeling initially ^[1]. The intracellular oxidative homeostasis in the cardiac cells is closely regulated by the production of ROS with limited intracellular defense mechanisms.

If the oxidative dysfunction continues, it may lead to pathological remodeling with cardiac damage in the form of increased high-sensitivity (hs) troponin T, in cardiac cells causing abnormalities in the global longitudinal strain ^[2]. In the cardiac cells, an overproduction of ROS may lead to the development and progression of maladaptive myocardial remodeling, which may be an early stage of heart failure (HF) ^{[3][4]}. Oxidative stress and ROS directly cause inflammation and impair the electrophysiology of the heart by targeting contractile machinery and cardiac components via the dysfunction of proteins that are crucial to excitation–contraction coupling, including sodium channels, L-type calcium channels, potassium channels, and the sodium–calcium exchanges ^{[1][2][3][4][5]}. Oxidative

stress may also cause alteration in the activity of the sarcoplasmic reticulum Ca^{2+} -adenosine triphosphatase (SERCA) as well as reduce myofilament calcium sensitivity [5]. In addition, oxidative stress can induce an energy deficit by influencing the protein function related to metabolism of energy [5]. Oxidative dysfunction may facilitate a pro-fibrotic function, as adaptation, by causing the proliferation of fibroblasts in the heart and matrix metallo-proteinases for extracellular remodeling, which may be the beginning of the hypertrophy of the heart [3][4].

It seems that the production of ROS in the heart is primarily completed by the mitochondria, xanthine oxidase, NADPH oxidases, and uncoupled nitric oxide synthase (NOS) [3]. The electron transport chain of the mitochondria may cause an overproduction of superoxide anion, contributing to cardiomyocyte damage with an increase in myocardial injury after an acute myocardial infarction [3]. There may be an increase in oxidative stress with an increased expression and activity of NADPH oxidase, due to multiple environmental and biological factors, such as angiotensin II, endothelin-1, mechanical stretch and tumor necrosis factor (TNF)- α [1][2][3][4][5]. The expression of xanthine oxidase and its activity is also increased due to damaging effects of behavioral risk factors such as tobacco intake and alcoholism in the heart exposed to these risk factors. It is proposed that oxidative dysfunction with increased oxidative stress may be the first stage of HF, which may be associated with cardiac damage and dysfunctional twist [1][2][5][6]. If there is a lower availability of endogenous antioxidants, super-oxide-dismutase (SOD), glutathione-peroxidase (GPS) and catalase or coenzyme Q10, it may cause the worsening of cardiac function, resulting in sub-endocardial damage, which may be the second stage of HF [6][7]. There may be an uncoupling of the NOS with structural instability, which further increases the generation of ROS, leading to left ventricular (LV) enlargement, dysfunction in the contraction [3], and remodeling of LV [3][4]. If the cardiac damage continues, it may lead to increased sympathetic activity with decline in parasympathetic activity causing neuro-hormonal dysfunction [1][2][3][4][5][6].

2. Left Ventricular Twist as Function of the Heart

Richard Lower FRCP (1631–1691) was the first to publish the twisting motion of the LV, in 1669, as “the wringing of a linen cloth to squeeze out the water”, which continues to intrigue the experts in their quest to understand cardiac function [8][9][10]. Apart from speckle tracking echocardiography (STE), magnetic resonance imaging (MRI) may be used to examine LV twist [10][11]. It appears to be crucial to examine twist function to understand the oxidative function of the heart, which would require quantification of the LV twist. The cardiac twist or torsion represents the mean longitudinal gradient of the net difference in the clockwise and counterclockwise rotation of the apex and base of the LV, as viewed from the apex of the left ventricle. The LV twist deforms the sub-endocardial fiber matrix, resulting in the storage of potential energy. A further deformation in the recoil of twist may cause the release of restoring forces, which contributes to diastolic relaxation of the LV with early diastolic filling [11]. Interestingly, systolic function may not be entirely normal, despite the normal ejection fraction (EF). There may be a decline in the left ventricular systolic long-axis at earlier stages, followed by evidence of more greater, subtle defects. On physical training, with decreased augmentation of function in the long-axis, impairment in systolic twist, decreased global strain, and electromechanical dys-synchrony will reduce the myocardial systolic reserve [11][12][13]. The twist function may alter during oxidative myocardial dysfunction, which may be an early marker of HF.

The physiology of twist mechanics indicate that LV twists in systole store optimal energy and, during the recoils (untwists) in diastole, cause energy release [12]. It seems that left ventricular ejection is aided by twist and untwist, which is helpful for the relaxation and filling of the ventricle. Thus, twist or torsion and rotation are crucial in cardiac contraction mechanics. Torsion or twist is accompanied by the wringing motion of the heart in its long axis produced by contraction of the myofibers in the wall of LV [8]. The apex and the base of the heart, during initial isovolumic contraction, both rotate in a counterclockwise method, if observed from apex to base. However, in the normal heart, the base of the heart has clockwise rotation during systole and the apex of the heart has counterclockwise rotation, causing a wringing movement. The cardiologists are not able to understand the utility of the twist function in clinical practice, which may be due to the problems in the measurement of cardiac rotation and torsion in the clinic [13][14]. It seems that three-dimensional STE may be an alternative method to assess the twist function, during plane motion. However, it seems that the measurement of the twist function would enhance the knowledge of physiological mechanics of the heart, such as the early diagnosis of abnormality in the rotation, indicating sub-endocardial dysfunction, the second stage of the six stages of HF, that may occur due to behavioral risk factors such as tobacco and Western diet. These risk factors may be also helpful in exploring the secrets of the diastole (a Rosetta stone), which could be a new concept in diastolic function and diastolic HF, via STE, in the light of neuro-humoral dysfunction [13][14][15][16][17]. It seems that the physician needs to have a closer look to understand the physio-pathogenesis of oxidative myocardial function and cardiac dysfunction, in particular, the LV twist and decline in myocardial strain [6]. There is an unmet need to use rotation and twist, as well as reversible sub-endocardial and diastole dysfunction in the diastole, via STE, as new markers of cardiac function, in the presence of oxidative dysfunction of the myocardium [15][16][17].

3. Oxidative Dysfunction and Inflammation as Targets for Therapeutic Antioxidants

Preclinical and clinical studies indicate that several therapeutic options are available to treat oxidative stress-associated cardiovascular diseases (CVDs) [1][3][4]. Many of the antioxidants, such as dietary content of phytochemicals, and novel polyphenols, have been examined for therapy, in view of the risk factors and inflammatory mediators of HF [4][18][19]. Apart from these, new therapeutic methods such as miRNA and nano-medicine are also available for the treatment of CVDs, in particular, HF, which may be tried, during the early stages of the Six Stages of HF. It seems that an increase in free fatty acids and oxidative dysfunction with reference to variability in biomarkers such as glucose levels, and levels of oxidative stress, predispose individuals to multifold greater inflammation and immune deficiency, leading to cardiac cell apoptosis and heart failure (HF) [20][21][22]. Decline in immunological responses may result in damage to other body systems contributing in diseases of associated body systems [20][21][22]. Free radicals are known to damage the cell membranes, causing the development of intracellular Ca^{2+} overload, activation of proteases and phospholipases, and alterations in mitochondrial gene expression in the cardiac cells, predisposing individuals to cardiomyocyte dysfunction [20][21][22][23][24]. Deficiency of protective antioxidants may predispose individuals to oxidative damage to proteins, enzymes, fatty acids and DNA [25][26][27]. It is possible that the cell damage may be reversed by the HEART diet. Experimental and epidemiological studies have also demonstrated that Western-type diets characterized by high sugar and

refined carbohydrates with a high glycemic index, as well as high-fat diet, red meat and preserved meat, may predispose individuals to increased risk of HF [25][26][27][28][29][30][31][32][33].

Apart from endogenous antioxidant defences, several exogenous antioxidants are available that may be administered for the treatment of HF. Since therapy with individual antioxidants in patients with CVDs has only had limited success, there is a need to determine the role of the Mediterranean diet, such as the HEART diet, in the management of HF, **Table 1**.

Table 1. Antioxidant defences and antioxidants available in the HEART diet.

| Indogenous Antioxidants | Exogenous Antioxidants from HEART Diet |
|-----------------------------|--|
| Enzymes | Vitamins |
| Superoxide dismutase (SOD) | Vitamin C, ascorbic acid, ascorbate |
| Glutathion peroxidase (GPS) | Vitaminss, E, tocopherol, tocotrienol |
| Glutathion reductase | Vitamin A, vitamin D |
| Glutathion-S-transferase | Polyphenolics and favonoids |
| Paraoxanase | Quercitin, resveratrol |
| Thioredoxin reductase | Catechins; Flavonols, Flavanols |
| Heme- oxygenase | Curcumin |
| Aldehyde dehydrogenase | Anthrocyanins |
| 8-Oxyguanine glycoselase | Phenolic acid |
| Catalase (Iron dependent) | Isoflavons/Genestein |
| Non-enzyme antioxidant | Carotinoids |
| Bilirubin | Alpha-carotine, beta-carotine |
| Coenzyme Q10 | Zeaxanthin |
| L-carnitine | Lutein |
| Alpha-lipoic acid | Lycopine |
| Melatonin | Beta-cryptixanthin |
| Uric acid, cholesterol | Minerals |
| Metal binding proteins | Magnesium |

| Indogenous Antioxidants | Exogenous Antioxidants from HEART Diet |
|------------------------------|--|
| Metallothioneine | Selenium, cromium |
| Lactoferrin | Zinc, copper |
| Transferrin | Fiber in the diet; oligosaccharides, polysaccharides |
| Ferritin | Fatty acids; Omega-3 and Monounsaturated |
| Ceruloplasmin (Cu dependent) | Amino acids; L-theanine, arginine, L-tryptophan |

1. Singh, R.B.; Komatsu, T.; Lee, W.C.; Watanabe, S.; Inoue, S.C.; Kiyoi, T.; Mogi, M.; Gaur, S.S.; Gautam, R. Effects of behavioral risk factors with reference to smoking on pathophysiology of cardiomyocyte dysfunction. *World Heart J.* 2020, 12, 9–14.

4. Effects of HEART Diet in Heart Failure

2. Singh, R.B.; Fedacko, J.; Goyal, R.; Rai, R.H.; Nandave, M.; Tonk, R.K.; Gaur, S.S.; Gautam, R.; Chakraborty, S. Pathophysiology and significance of antioxidants in heart failure, with reference to oxidative stress risk factors. *World Heart J.* 2020, 12, 15–22.

3. Van der Pol, A.; van Gilst, W.H.; Voors, A.A.; van der Meer, P. Treating oxidative stress in heart failure: Past, present and future. *Eur. J. Heart Fail.* 2019, 21, 425–435.

4. Najjar, R.S.; Feresin, R.G. Protective role of polyphenols in heart failure: Molecular targets and cellular mechanisms underlying their therapeutic potential. *Int. J. Mol. Sci.* 2021, 22, 1668.

5. Takimoto, E.; Kass, D.A. Role of oxidative stress in cardiac hypertrophy and remodeling. *Hypertension* 2007, 49, 241–248.

6. Singh, R.B.; Elkilany, G.; Fedacko, J.; Hristova, K.; Palmiero, P.; Singh, J.; Manal, M.A.; Badran, H.M. Evolution of the natural history of myocardial twist and diastolic dysfunction as cardiac dysfunction. In *Chronic Heart Failure, Pathophysiology and Management*; Singh, R.B., Fedacko, J., Elkilany, G., Hristova, K., Eds.; Elsevier: Cambridge, MA, USA, 2023; in press.

7. Mironczuk-Chodakowska, I.; Witkowska, A.M.; Zujew, M.E. Endogenous non-enzymatic antioxidants in the human body: A review. *Med. Sci.* 2018, 63, 68–78.

8. Mann, D.L.; Bristow, M.R. Mechanisms and models in heart failure: The biomechanical model and beyond. *Circulation* 2005, 111, 2837–2849.

9. Halabi, A.; Yang, H.; Wang, L.; Porter, E.; Huynh, Q.; Negishi, K.; Marwick, T.H. Evolution of myocardial dysfunction in asymptomatic patients at risk of heart failure. *JACC Cardiovasc. Imaging* 2021, 14, 350–361.

10. Lower, R. *Tractatus de Corde*; Oxford University Press: London, UK, 1669.

11. Sengupta, P.P.; Tajik, A.H.; ChandraSekaran, K.; Khandheria, B.K. Twist mechanisms of the left ventricle: Principles and application. *JACC Cardiovasc. Imaging* 2008, 1, 366–376.

12. Nakatani, S. Left ventricular rotation and twist: Why should we learn? *J. Cardiovasc. Ultrasound* 2011, 19, 1–6.

13. Sahay, S.K.; Hristova, K.; Nanda, M.C. Echocardiography [26] [27] On Toward deciphering the 'Rosetta stones' of left ventricular diastolic dysfunction. Echocardiography 2020, 37, 1886–1899. [CrossRef]

14. Singh, R.B.; Sozzi, F.B.; Fedacko, J.; Hristova, K.; Fatima, G.; Pella, D.; Cornelissen, G.; Isaza, A.; Pella, D.; Singh, J.; et al. Pre-heart failure at 2D- and 3D-speckle tracking echocardiography: A

5. Dietary Fat and Risk of Heart Failure

15. Singh, R.B.; Fedacko, J.; Elkilany, G.; Hristova, K.; Palmiero, P.; Pella, D.; Cornelissen, G.; Isaza, A.; Pella, D. 2020 Guidelines on Pre-Heart Failure in the Light of 2D and 3D Speckle Tracking Echocardiography. A Scientific Statement of the International College of Cardiology. World Heart J. 2020, 12, 50–70.

16. Khou, M.G.; Peshock, R.M.; Ayers, C.R.; de Lemos, J.A.; Drazner, M.H. A 4-tiered classification of left ventricular hypertrophy based on left ventricular geometry: The Dallas heart study. Circ. Cardiovasc. Imaging 2010, 3, 164–171.

17. Lacalzada, J.; de la Rosa, A.; Izquierdo, M.M.; Jiménez, J.J.; Iribarren, J.L.; García-González, M.J.; López, B.M.; Duque, M.A.; Barragán, A.; Hernández, C.; et al. Left ventricular global longitudinal systolic strain predicts adverse remodeling and subsequent cardiac events in patients with acute myocardial infarction treated with primary percutaneous coronary intervention. Int. J. Cardiovasc. Imaging 2015, 31, 575–584.

18. Hristova, K.; Singh, R.B.; Fedacko, J.; Toda, E.; Kumar, A.; Saxena, M.; Baby, A.; Takahashi, T.; HF [36]. De Meester, F.; Wilson, D.W. Causes and risk factors of congestive heart failure in India. World Heart J. 2013, 5, 13–20.

19. Fedacko, J.; Singh, R.B.; Gupta, A.; Hristova, K.; Toda, E.; Kumar, A.; Saxena, M.; Baby, A.; Singh, R.; Takahashi, T.; et al. Inflammatory mediators in chronic heart failure in North India. Acta Cardiol. 2014, 69, 391–394.

20. Simmonds, S.; Gijzen, H.; Heymans, S.; Van der Griend, A.; Van der Griend, A.; et al. Pathophysiology of atherosclerosis in HFpEF and HF. Eur. Heart J. 2019, 40, 100–108.

21. Elkilany, G.; Singh, R.B.; Hristova, K.; Milovanovic, B.; Chaves, H.; Wilson, D.W.; Saboo, B.; Mahashwari, A. Beyond drug therapy, nutritional perspectives in the management of chronic heart failure. World Heart J. 2015, 7, 83–88.

22. Singh, R.B.; Fedacko, J.; Pella, D. Coenzyme Q10 modulates remodeling possibly by decreasing angiotensin-converting enzyme in patients with acute coronary syndrome. Antioxidants 2018, 7, 99.

23. Fedacko, J.; Singh, R.B.; Niaz, M.A.; Bharadwaj, K.; Verma, N.; Gupta, A.K.; Singh, R.B. Association of coronary protective factors among patients with acute coronary syndromes. J. Cardiol. Ther. 2016, 4, 671–677.

24. Singh, R.B.; Cornelissen, G.; Takahashi, T.; Shastun, S.; Hristova, K.; El-Kilany, G.; Fatima, G.; Tyagi, G.; Mojto, V.; Suchday, S.; Alami, M.; Otsuka, K.; Saito, B. et al. Brain-heart interactions and glycating agents and peptides in the heart failure. *Worlds Heart J.* 2015, 7, 120–142.
25. Singh, R.B.; Hristova, K.; Fedacko, J.; El-Kilany, G.; Cornelissen, G. Chronic heart failure: A disease of the brain. *Heart Fail. Rev.* 2018, 24, 301–307.
26. Singh, R.B.; Wilczynska-Fedacko, J.; Takahashi, T.; Niaz, M.A.; Jain, S.; Fatima, G.; Manal, M.A.; Ahla, M.A.S. Association of Indo-Mediterranean neuroprotective dietary (MIND) pattern with loss of weight on two types of low-energy diet. It is possible that obesity has a microbial part, which might have important therapeutic potentials. *Int. J. Clin. Nutr.* 2021, 21, 11–20.
27. Wilczynska, A.; Fedacko, J.; Hristova, K.; Alkilany, G.; Fatima, G.; Tyagi, G.; Mojto, V.; Suchday, S. Association of dietary pattern and depression with risk of cardiovascular diseases. *Int. J. Clin. Nutr.* 2017, 17, 1–10.
28. Singh, R.B.; Gvozdiakova, A.; Singh, J.; Shastun, S.; Dhalla, N.S.; Pella, D.; Fedacko, J.; Cornelissen, G. Omega-3 PUFA, Omega-6 PUFA and mitochondrial dysfunction in relation to remodelling. In *Recent Advances in Mitochondrial Medicine and Coenzyme Q10*; Gvozdiakova, A., Cornelissen, G., Singh, R.B., Eds.; Nova Science Publishers: Hauppauge, NY, USA, 2018; Chapter 23, pp. 353–368.
29. Nettleton, J.A.; Steffen, L.M.; Loehr, L.R.; Rosamond, W.D.; Folsom, A.R. Incident heart failure is transition in mitochondrial permeability. It seems that the related mechanisms are complex, which could be on account of adaptation of the heart, in a situation, on saturated fat diets. There is an unmet need to have complete understanding of the effects of different types of dietary fats on phospholipids in the cell membrane of cardiac cells, metabolites of lipids and metabolic functions in the failing as well as in the normal heart. It is likely that existing nutrients such as ω-3 fatty acids, flavonoids, and coenzyme Q10 may inhibit the lipotoxicity caused by saturated fat and delay the progression of cardiac hypertrophy by improving twist function and sub-endothelial function, which may cause HFpEF in place of HFrEF. It seems that changes in fat consumption could be crucial in the management of HF. The influence of the HEART diet or Western type of diet on cardiac cells could be dependent on pro-inflammatory biomarkers that can damage cardiomyocytes. High-glucose or fast-food diets induce an increase in ceramides and high levels of TMAO on account of greater consumption of red meat (L Carnitine) and triglycerides. *Nutr. Metab. Cardiovasc. Dis.* 2011, 21, 941–946.
30. Ashaye, A.; Gaziano, J.; Dioussé, I. Red meat consumption and risk of heart failure in male physicians. *Nutr. Metab. Cardiovasc. Dis.* 2011, 21, 941–946.
31. Kaluza, J.; Akesson, A.; Wolk, A. Long-term processed and unprocessed red meat consumption and risk of heart failure: A prospective cohort study of women. *Int. J. Cardiol.* 2015, 193, 42–46.
32. Kaluza, J.; Akesson, A.; Wolk, A. Processed and unprocessed red meat consumption and risk of heart failure: Prospective study of men. *Circ. Heart Fail.* 2014, 7, 552–557.
33. Lechman, E.B.; Littleman, W.A.; Wolk, A. Dietary glycemic index, dietary glycemic load, and new-onset heart failure events: A prospective study of middle-aged and elderly women. *Am. J. Clin. Nutr.* 2010, 91, 65–71.
34. Ng, S.F.; Lin, R.C.; Laybutt, D.R.; Barres, R.; Owens, J.A.; Morris, M.J. Chronic high-fat diet in fathers' programs p-cell dysfunction in female rat offspring. *Nature* 2010, 467, 963–966.
35. Schiattarella, G.G.; Altamirano, E.; Tong, D.; French, K.M.; Villalobos, E.; Kim, S.Y.; Luo, X.; Jiang, N.; May, H.I.; Wang, Z.V. et al. Nitrosative stress drives heart failure with preserved ejection fraction. *Nature* 2019, 568, 351–356.

6. Mechanisms of Diet and Obesity in Heart Failure

There is evidence that the Western type of diet is a risk factor of obesity, whereas Mediterranean-style diets may have protective effects on obesity and HF [36,37,38]. It is possible to create diet-induced obesity in animal experiments, in selected strains, by use of a diet that is rich in fat (about 40% to 50% of energy, versus 10–15%) in conjunction with sugar (~20% to 30% sucrose). It appears that obesity may have complex influences on the

36. Wang, Z.; Klipfel, E.; Berne, A.; Koth, R.; Levin, S.; Dugan, B.; Feldstein, A.E.; Britton and changes, F.; Xie, G.; Hong, Y.-M. Gut flora, the metabolism and phosphatidylcholine promotes cardiovascular disease. *Nat. Rev. Clin. Endocrinol.* 2011, 7, 257–263.
37. Stanley, W.C.; Dabkowski, E.R.; Ribeiro, R.F., Jr.; O'Connell, K.A. Dietary fat and heart failure: Moving from lipotoxicity to lipoprotection. *Circ. Res.* 2012, 110, 764–776.
38. Leman, S.C.; Olendzki, B.; Wagner, R.; Li, W.; Gulver, A.L.; Ockene, J.; Goldberg, R.J. The dietary quality of persons with heart failure in NHANES 1999–2006. *J. Gen. Intern. Med.* 2010, 25, 135–140.
39. Lopaschuk, G.D.; Holmes, C.D.; Stanley, W.C. Cardiac energy metabolism in obesity. *Circ. Res.* 2007, 101, 335–347.

7. Protective Dietary Patterns in the Prevention of Heart Failure

40. Rajman, I.; Wolk, A.; Latsson, S.C. The relationship between sweetened beverages consumption and risk of heart failure in men. *Heart* 2015, 101, 1961–1965.
41. Tikellis, G.; Tobias, M.; Westcott, B.; Coughlan, M.; Prepetich, B.; Dabkowski, K.; Tager, A.; Bierhays, A.; Groppe, H.; Fergan, J. MacCardia inflammation associated with a Western diet is mediated via activation of RAGE by AGEs. *Am. J. Physiol. Endocrinol. Metab.* 2008, 295, E323–E330.
42. Dambrova, M.; Latkovskis, G.; Kuka, J.; Strele, I.; Konrade, I.; Grinberga, S.; Hartmane, D.; Pugovics, O.; Erglis, A.; Liepinsh, E. Diabetes is associated with higher trimethylamine N-oxide plasma levels. *Exp. Clin. Endocrinol. Diabetes* 2016, 124, 251–256.
43. Suzuki, T.; Heaney, L.M.; Bhandari, S.S.; Jones, D.J.L.; Ng, L.L. Trimethylamine N-oxide and the prognosis in acute heart failure. *Heart* 2016, 102, 841–848.
44. O'Shea, K.M.; Khairallah, R.J.; Sparagna, G.C.; Xu, W.; Hecker, P.A.; Robillard-Frayne, I.; Des Rosiers, C.; Kristian, T.; Murphy, R.C.; Fiskum, G.; et al. Dietary omega-3 fatty acids alter cardiac mitochondrial phospholipid composition and delay Ca²⁺-induced permeability transition. *J. Mol. Cell. Cardiol.* 2009, 47, 819–827.
45. Miatello, R.; Vazquez, M.; Penna, N.; Cruzado, M.; Zumino, A.P.; Bislari, N. Chronic administration of resveratrol prevents biochemical cardiovascular changes in fructose-fed rats. *Am. J. Hypertens.* 2005, 18, 864–870.
46. Ley, R.E.; Tambaugh, P.J.; Klein, S.; Gordon, J.I. Microbial ecology: Human gut microbes associated with obesity. *Nature* 2006, 444, 1022–1023.
47. Levitan, E.B.; Wolk, A.; Mittleman, M.A. Relation of consistency with the dietary approaches to stop hypertension diet and incidence of heart failure in men aged 45 to 79 years. *Am. J. Cardiol.* 2009, 104, 1416–1420.
48. There is growing evidence on the role of egg on risk

48. Cerdas, H.; Bernal, W.; Acuna-Miller, R.; Cava, M. A. Consistency with the DASH diet and incidence of heart failure. *Arch Intern Med*. 2009; 169, 851–857.
49. Hall, J.E.; da Silva, A.A.; do Carmo, J.M.; Dubinon, J.; Hamza, S.; Munusamy, S.; Smith, G.; Stec, D.E. Obesity-induced hypertension: Role of sympathetic nervous system, leptin, and melanocortins. *J. Biol. Chem.* 2010, 285, 17271–17276.
50. Okere, I.C.; Chandler, M.P.; McElfresh, T.A.; Rennison, J.H.; Sharov, V.; Sabbah, H.N.; Tserng, K.Y.; Hoit, B.D.; Ernsberger, P.; Young, M.E.; et al. Differential effects of saturated and unsaturated fatty acid diets on cardiomyocyte apoptosis, adipose distribution, and serum leptin. *Am. J. Physiol. Heart Circ. Physiol.* 2006, 291, H38–H44.
51. Nwozo, S.O.; Orojobi, F.; Adaramoye, O.A. Hypolipidemic and antioxidant potentials of *Xylopiia aethiopica* seed extract in hypercholesterolaemic rats. *J. Med. Foods* 2011, 14, 114–119.
52. Nwozo, S.O.; Lewis, Y.T.; Oyinloye, B.E. The effects of *Piper guineense* versus *Sesamum indicum* aqueous extracts on lipid metabolism and antioxidants in hypercholesterolemic rats. *Iran. J. Med. Sci. (IJMS)* 2017, 42, 449–456.
53. Singh, R.B.; Rastogi, S.S.; Verma, R.; Laxmi, B.; Singh Reema Ghosh, S.; Niaz, M.A. Randomized, controlled trial of cardioprotective diet in patients with recent acute myocardial infarction: Results of one year follow up. *BMJ* 2002, 304, 1115–1119.
54. Singh, R.B.; Dubnov, G.; Niaz, M.A.; Ghosh, S.; Singh, R.; Rastogi, S.S.; Manor, O.; Pella, D.; Berry, E.M. Effect of an Indo-Mediterranean diet on progression of coronary disease in high risk patients: A randomized single blind trial. *Lancet* 2002, 360, 1455–1461.
55. Fitzgerald, S.M.; Henegar, J.R.; Brands, M.W.; Henegar, L.K.; Hall, J.E. Cardiovascular and renal responses to a high-fat diet in Osborne-Mendel rats. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 2001, 281, R547–R552.
56. Pladevall, M.; Williams, K.; Guyer, H.; Sadurni, J.; Falces, C.; Ribes, A.; Pare, C.; Brotons, C.; Gabriel, R.; Serrano-Rios, M.; et al. The association between leptin and left ventricular hypertrophy: A population-based cross-sectional study. *J. Hypertens.* 2003, 21, 1467–1473.
57. Schaffer, J.E. Lipotoxicity: When tissues overeat. *Curr. Opin. Lipidol.* 2003, 14, 281–287.
58. Papandreou, C.; Hernández-Alonso, P.; Bulló, M.; Ruiz-Canela, M.; Li, J.; Guasch-Ferré, M.; Toledo, E.; Clish, C.; Corella, D.; Estruch, R.; et al. High plasma glutamate and a low glutamine-to-glutamate ratio are associated with increased risk of heart failure but not atrial fibrillation in the Prevención con Dieta Mediterránea (PREDIMED) Study. *J. Nutr.* 2020, 150, 2882–2889.
59. Wirth, J.; di Giuseppe, R.; Boeing, H.; Weikert, C. A Mediterranean-style diet, its components and the risk of heart failure: A prospective population-based study in a non-Mediterranean country. *Eur. J. Clin. Nutr.* 2016, 70, 1015–1021.

60. Djoussé, L.; Akinkuolie, A.O.; Wu, J.H.; Ding, E.L.; Gaziano, J.M. Fish consumption, omega-3 fatty acids and risk of heart failure: A meta-analysis. *Clin. Nutr.* 2012, 31, 846–853.
61. Djoussé, L.; Gaziano, J.M. Breakfast cereals and risk of heart failure in the Physicians' Health Study I. *Arch. Intern. Med.* 2007, 167, 2080–2085.
62. Mozaffarian, D.; Lemaitre, R.N.; King, I.B.; Song, X.; Spiegelman, D.; Sacks, F.M.; Rimm, E.B.; Siscovick, D.S. Circulating long-chain omega-3 fatty acids and incidence of congestive heart failure in older adults: The Cardiovascular Health Study. *Ann. Intern. Med.* 2011, 155, 160–170.
63. Murphy, M.P. How mitochondria produce reactive oxygen species. *Biochem. J.* 2009, 417, 1–13.
64. Nadal-Ginard, B.; Kajstura, J.; Leri, A.; Anversa, P. Myocyte death, growth, and regeneration in cardiac hypertrophy and failure. *Circ. Res.* 2003, 92, 139–150.
65. Abel, E.D.; Doenst, T. Mitochondrial adaptations to physiological vs. pathological cardiac hypertrophy. *Cardiovasc. Res.* 2011, 90, 234–242.
66. Paulus, W.J. Unfolding discoveries in heart failure. *N. Engl. J. Med.* 2020, 382, 679–682.
67. Amgalan, D.; Kitsis, R.N. A mouse model for the most common form of heart failure. *Nature* 2019, 568, 324–325.
68. Bogiatzi, C.; Gloor, G.; Allen-Vercoe, E.; Reid, G.; Wong, R.G.; Urquhart, B.L.; Dinculescu, V.; Ruetz, K.N.; Velenosi, T.J.; Pignanelli, M.; et al. Metabolic products of the intestinal microbiome and extremes of atherosclerosis. *Atherosclerosis* 2018, 273, 91–97.
69. Spence, J.D.; Srichaikul, K.K.; Jenkins, D.J.A. Cardiovascular Harm from Egg Yolk and Meat: More Than Just Cholesterol and Saturated Fat. *J. Am. Heart Assoc.* 2021, 10, e017066.
70. Magomedova, A.G. Characteristics of nutrition and health of pupils in the regions of Russia. *Munic. Educ. Innov. Exp.* 2021, 3, 56–64.
71. Wang, W.; Kang, P.M. Oxidative stress and antioxidant treatments in cardiovascular diseases. *Antioxidants* 2020, 9, 1292.

Retrieved from <https://encyclopedia.pub/entry/history/show/62859>