# **Quinone-Based Redox Mediators**

Subjects: Green & Sustainable Science & Technology

Contributor: Haikun Zhang

Quinone compounds are a class of substances that contain cyclic unsaturated diketones, especially carbocyclic compounds and their derivatives with quinone groups. These compounds contain either q-quinone or o-quinone groups and can be divided into benzoquinone, naphthoquinone, anthraquinone, and phenanthraquinone according to their structural characteristics. Structurally, all compounds containing quinone groups have the potential to act as redox mediators to accelerate electron transfer.

quinoid compounds

redox mediators

natural substances

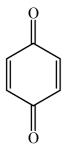
artificially synthesized quinone compounds

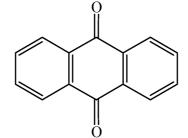
small molecules

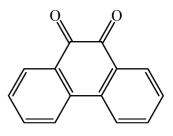
QRMs

## 1. Introduction

Quinone compounds are a class of substances that contain cyclic unsaturated diketones, especially carbocyclic compounds and their derivatives with quinone groups. These compounds contain either q-quinone or o-quinone groups and can be divided into benzoquinone, naphthoquinone, anthraquinone, and phenanthraquinone according to their structural characteristics (Figure 1). More broadly, quinoid compounds also include a variety of non-carbocyclic quinone-like substances, such as riboflavin, flavin mononucleotide, flavin adenine dinucleotide, and humic substances. They can be found in the pigments of plants (mainly Polygonaceae, Rubiaceae, Rheaceae, Liliaceae, and Leguminaceae), a few animals (Cochineal), and bacteria, as well as in the metabolites of some lower plant lichens and fungi. When these animals, plants, and bacteria die, the biological debris is subsequently decomposed and mineralized by microbes. During this process, numerous nonbiodegradable quinone compounds remain, leading to their wide distribution in natural environments (including almost all soils, waters and sediments). In fact, these quinoid compounds are not only widely found in nature but are also widely involved in environmental processes.







Benzoquinone

Naphthoquinone

Anthraquinone

Phenanthrenequinone

**Figure 1.** Structures of benzoquinone, naphthoquinone, anthraguinone, and phenanthrenequinone.

Since the 20th century (1970s), the environmental importance of these quinoid compounds has received increasing attention. In 1967, Roxon et al. [1] reported that riboflavin, a quinone-like compound, can accelerate the transformation of some organic contaminants. Subsequently, an increasing number of researchers began to focus on the application of quinone compounds (e.g., anthraquinone-2,6-disulfonate, anthraquinone-2-sulfonate, humus, carbon materials, etc.) for the transformation of environmental contaminants [2]. A mechanistic study found that the quinone group can be reversibly oxidized and reduced between quinone and hydroquinone due to its electron affinity, which enables it to serve as an electron shuttle in multiple redox reactions. In addition, the presence of quinoid compounds can accelerate reactions by lowering the activation energy of the total reaction and may even be a prerequisite for the reaction to take place under some situations. To date, many kinds of quinoid compounds have been proven to be capable of facilitating electron transfer in redox reactions between diverse inorganic or organic compounds, including organic matter, metals, and minerals [1][2].

Over the past twenty years, quinoid compounds have attracted more attention than before because of their significant interactions with microbes, which may further increase their environmental importance. Since the 1990s, extensive studies have been conducted in this field [2][3][4][5][6]. Researchers have found that the reduction and reoxidation of quinone groups can be coupled to the anaerobic respiration of electrochemically active bacteria in the presence of various electron donors, such as saccharides, short-chain fatty acids, alcohol,  $H_2$ , and even some organic pollutants. During this process, quinone is first reduced to hydroquinone; subsequently, hydroquinone can either directly reduce various electron acceptors, such as azo compounds [7][8], nitroaromatics [9], polyhalogenated pollutants [10][11], Fe(III) [12], Cr(VI) [13], Se(IV), and Te(IV) [14], or serve as electron donors to support the bioreduction of  $NO_3^-/NO_2^-$  [15], As(V) [16], and perchlorate [17] with their reoxidation. Overall, the presence of quinoid compounds considerably enhances the electron transfer in many anaerobic biotransformation processes.

Structurally, all compounds containing quinone groups have the potential to act as redox mediators to accelerate electron transfer. However, things do not work this way in real reactions. In addition to the functional groups, the redox potential ( $E'_0$ ) of effective quinoid redox mediators (QRMs) must also be between those of the terminal electron acceptor and electron donor to reduce the activation energy of reactions. In general,  $E'_0$  is an indication of the catalytic capacity of QRMs and should be in the range of -440 mV to -50 mV [18][19]. Vitamin B12 (-530 mV) was reported to be a very poor redox mediator due to its low  $E'_0$  [20]. In this research, taking the source and availability into consideration, the common QRMs are classified into four primary categories, including natural substances, artificially synthesized quinone compounds, microbiologically excreted small molecules, and immobilized QRMs.

## 2. Natural Substances

Humic substances, which widely exist in various environments, can be defined as some of the most important natural quinone mediators. Humic substances primarily originate from the degradation of plants and mainly include three types: humic acid, fulvic acid, and humin. In 1996, Lovley et al. [3] first reported that humic substances can

participate in some microbial respiration processes. Subsequently, an increasing number of studies have confirmed that humic substances are important redox mediators in nature, especially for some microbe-driven redox reactions (for instance, dissimilatory metal reduction bacteria, sulfate reducing bacteria, and methanogens) [5][21][22]. Humic substances have complex structures; however, quinone groups are considered to be the most important functional groups that are directly involved in electron transport since they can flexibly interconvert between hydroquinone and semiquinone via a reversible electron transfer reaction [23]. It has been reported that some non-quinone groups in humic substances are also able to transfer electrons, but the capacity of some specific non-quinone groups is much lower than that of quinone groups [23][24]. Accordingly, considering their ubiquity in the environment and efficient conductive capacity, humic substances may play a significant role in the biogeochemical cycles of matter and elements on a global scale.

# 3. Artificially Synthesized Quinone Compounds

In addition to natural quinone mediators, there are also many artificially synthesized quinones, including 9,10anthraguinone-2-carboxylic acid, anthraguinone-2,6-disulfonate (AQDS), anthraguinone-2-sulfonate (AQS), etc. Therein, AQDS and AQS are the most studied artificially synthesized quinone compounds. Both AQDS and AQS can be prepared via a chemical synthesis method: reacting anthraquinone (a natural raw material) first with sulfuric acid in the presence of mercury and then with sodium hydroxide (Reaction diagram 1). They were reported to play similar roles; however, their catalytic performance and mechanism during the electron transfer process vary greatly depending on their molecular structures and polarities, as well as the presence of different microorganisms and pollutants [7][18][25]. AQS is suggested to be a more powerful mediator than AQDS. This is because the redox potential of AQS is lower than that of AQDS. Moreover, the additional sulfonate group of AQDS diminishes the ease with which the compound approaches the functional enzyme. In addition, the roles of these artificially synthesized quinones change as reaction conditions change. For example, Zhou et al. [26] demonstrated that AQDS could first act as an electron acceptor during the anammox denitrification process in the presence of iron-reducing bacteria; then, the reduced AQDS can chemically react with Fe(III) (Figure 2a). In addition, for microbes, AQDS can simultaneously be used as an electron donor and an electron acceptor (Figure 2b). Wang et al. [27] reported that AQDS can enhance the coupled biotransformation of phenol and nitrate: AQDS is first reduced to AQH2DS (electron acceptor) by Shewanella sp. XB in the presence of phenol, and then AQH2DS (electron donor) provides electrons to strain XB for its further bioreduction of nitrate. On the whole, artificially synthesized quinone compounds are more targeted and efficient than humic substances.

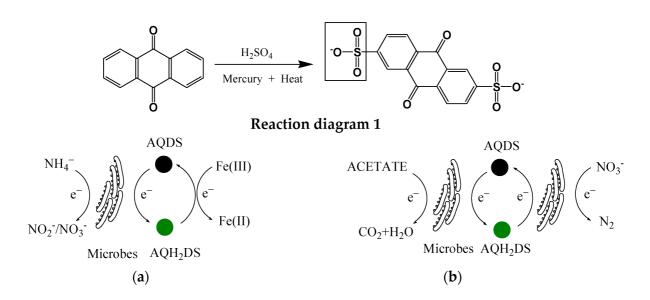


Figure 2. The role of AQDS in different reactions. (a) Anammox reaction; (b) denitrification reaction.

# 4. Microbial Secretions That Function as QRMs

In addition to natural quinone mediators and artificially synthesized quinone compounds, the soluble small molecules, such as riboflavin (RF), flavin mononucleotide (FMN), flavin adenine dinucleotide, menadione, and phenazines, excreted by various bacteria (Shewanella, Klebsiella, Pseudomonas, Enterococcus, Methylosinus, Bacillus, etc.) can also function as ORMs [28][29][30][31][32]. These small molecules can be used not only extracellularly as electron shuttles in the biotransformation of solid metal oxides and electrodes but also intracellularly as coenzyme factors of some flavin proteins to participate in the anaerobic biotransformation of organic pollutants and heavy metals [21][29]. Taking the most studied bacteria, Shewanella sp., as examples, they were reported to have the ability to secrete RF and FMN, which can help bacterial cells transport electrons for reducing iron oxides, such as hematite, goethite, ferrihydrite, and lepidocrocite [30][32]. Similarly, Tian et al. reported that the RF and FMN secreted by two Gram-positive bacteria, Bacillus cereus strain DIF1 and Rhodococcus ruber strain DIF2, can improve the electron transfer of microbial fuel cells [33]. In addition, researchers also found that FMN/FMNH<sub>2</sub> acts as a redox pair that may have multiple roles in the azo or chromium reductase-catalyzed reduction of azo dyes, Cr(VI), and nitroaromatic compounds. During this process, the FMN cofactor is first completely reduced to FMNH<sub>2</sub>, and second, these electrons are transferred from FMNH<sub>2</sub> to the electron acceptor, resulting in the formation of the corresponding azo or chromium reductase FMNH $_2$  [34][35][36][37][38][39][40]. The results of these studies also demonstrated that FMN/FMNH2 may play a similar role for the pairs of AQDS/AQH2DS in the process of electron transfer (Figure 3).

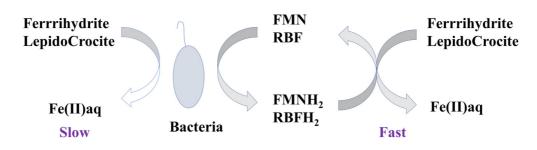


Figure 3. The role of FMN/RF during the bioreduction of iron(III) oxides [30].

## 5. Immobilized QRMs

Immobilized QRMs are designed for the practical application of known QRMs. Previous studies have demonstrated that quinone compounds can enhance the bioreduction of various contaminants [7][41][42][43]. In practical applications, however, these soluble quinone compounds, such as AQS and AQDS, need to be continuously added into reaction systems to increase the removal rates of contaminants, which results in an increase in running costs and secondary contamination. The immobilization of QRMs via physical or chemical methods not only maintains the catalytic performance of quinone groups but also realizes their effective recovery and reuse. This strategy is beneficial for their practical application. To date, many carriers, including activated carbon felt, anion exchange resins, and calcium alginate, have been developed for the immobilization of quinone compounds [44][45][46]. The previous studies confirmed that soluble quinone compounds can be covalently immobilized on the surface of polyurethane foam, graphene oxide and Fe<sub>3</sub>O<sub>4</sub>, which can be used to enhance the bioreduction of azo dyes, nitrobenzene, and chloronitrobenzenes [47][48][49][50]. These immobilized QRMs can have better catalytic performance than soluble QRMs during some biocatalytic reactions, further indicating that immobilized QRMs have potential application prospects, especially in the field of water pollution treatment. When QRMs are immobilized, they act as solid redox mediators to facilitate electron transfer by extending the direct electron transfer distance or as equivalents of functional enzymes (e.g., MtrC) as substrates for enzymatic reactions

#### References

- 1. Roxon, J.J.; Ryan, A.J.; Wright, S.E. Enzymatic reduction of tartrazine by Proteus vulgaris from rats. Food Cosmet. Toxicol. 1967, 5, 645–656.
- 2. Husain, M.; Husain, Q. Applications of Redox Mediators in the Treatment of Organic Pollutants by Using Oxidoreductive Enzymes: A Review. Crit. Rev. Environ. Sci. Technol. 2008, 38, 1–42.
- 3. Lovley, D.R.; Bluntharris, E.L.; Ejp, P.; Woodward, J.C.; Coates, J.D. Humic substances as electron acceptors for microbial respiration. Nature 1996, 382, 445–448.
- 4. Lovley, D.R.; Fraga, J.L.; Coates, J.D.; Blunt-Harris, E.L. Humics as an electron donor for anaerobic respiration. Environ. Microbiol. 1999, 1, 89–98.
- 5. Van der Zee, F.P.; Cervantes, F.J. Impact and application of electron shuttles on the redox (bio)transformation of contaminants: A review. Biotechnol. Adv. 2009, 27, 256–277.
- 6. Zhang, Y.; Zhang, Z.; Liu, W.; Chen, Y. New applications of quinone redox mediators: Modifying nature-derived materials for anaerobic biotransformation process. Sci. Total Environ. 2020, 744, 140652.

- 7. Rau, J.; Knackmuss, H.J.; Stolz, A. Effects of different quinoid redox mediators on the anaerobic reduction of azo dyes by bacteria. Environ. Sci. Technol. 2002, 36, 1497–1504.
- 8. van der Zee, F.P.; Bouwman, R.H.; Strik, D.P.; Lettinga, G.; Field, J.A. Application of redox mediators to accelerate the transformation of reactive azo dyes in anaerobic bioreactors. Biotechnol. Bioeng. 2001, 75, 691–701.
- 9. Li, L.; Wang, J.; Zhou, J.; Yang, F.; Jin, C.; Qu, Y.; Li, A.; Zhang, L. Enhancement of nitroaromatic compounds anaerobic biotransformation using a novel immobilized redox mediator prepared by electropolymerization. Bioresour. Technol. 2008, 99, 6908–6916.
- Fu, Q.S.; Barkovskii, A.L.; Adriaens, P. Reductive transformation of dioxins: An assessment of the contribution of dissolved organic matter to dechlorination reactions. Environ. Sci. Technol. 1999, 33, 3837–3842.
- 11. Perlinger, J.A.; Angst, W.; Schwarzenbach, R.P. Kinetics of the reduction of hexachloroethane by juglone in solutions containing hydrogen sulfide. Environ. Sci. Technol. 1996, 30, 3408–3417.
- 12. Wolf, M.; Kappler, A.; Jiang, J.; Meckenstock, R.U. Effects of humic substances and quinones at low concentrations on ferrihydrite reduction by Geobacter metallireducens. Environ. Sci. Technol. 2009, 43, 5679–5685.
- 13. Liu, G.; Yang, H.; Wang, J.; Jin, R.; Zhou, J.; Lv, H. Enhanced chromate reduction by resting Escherichia coli cells in the presence of quinone redox mediators. Bioresour. Technol. 2010, 101, 8127–8131.
- 14. Wang, X.; Liu, G.; Zhou, J.; Wang, J.; Jin, R.; Lv, H. Quinone-mediated reduction of selenite and tellurite by Escherichia coli. Bioresour. Technol. 2011, 102, 3268–3271.
- 15. Aranda-Tamaura, C.; Estrada-Alvarado, M.I.; Texier, A.C.; Cuervo, F.; Gómez, J.; Cervantes, F.J. Effects of different quinoid redox mediators on the removal of sulphide and nitrate via denitrification. Chemosphere 2007, 69, 1722–1727.
- 16. Field, J.A.; Sierra-Alvarez, R.; Cortinas, I.; Feijoo, G.; Moreira, M.T.; Kopplin, M.; Gandolfi, A.J. Facile reduction of arsenate in methanogenic sludge. Biodegradation 2004, 15, 185–196.
- 17. Thrash, J.C.; Van Trump, J.I.; Weber, K.A.; Miller, E.; Achenbach, L.A.; Coates, J.D. Electrochemical stimulation of microbial perchlorate reduction. Environ. Sci. Technol. 2007, 41, 1740–1746.
- 18. Brown, J.P. Reduction of polymeric azo and nitro dyes by intestinal bacteria. Appl. Environ. Microbiol. 1981, 41, 1283–1286.
- 19. Rau, J.; Stolz, A. Oxygen-insensitive nitroreductases NfsA and NfsB of Escherichia coli function under anaerobic conditions as lawsone-dependent azo reductases. Appl. Environ. Microbiol. 2003, 69, 3448–3455.

- 20. Santos, A.B.D.; Bisschops, I.A.; Cervantes, F.J.; van Lier, J.B. Effect of different redox mediators during thermophilic azo dye reduction by anaerobic granular sludge and comparative study between mesophilic (30 °C) and thermophilic (55 °C) treatments for decolourisation of textile wastewaters. Chemosphere 2004, 55, 1149–1157.
- 21. Liu, X.; Shi, L.; Gu, J.-D. Microbial electrocatalysis: Redox mediators responsible for extracellular electron transfer. Biotechnol. Adv. 2018, 36, 1815–1827.
- 22. Smith, J.A.; Nevin, K.P.; Lovley, D.R. Syntrophic growth via quinone-mediated interspecies electron transfer. Front. Microbiol. 2015, 6, 121.
- 23. Ratasuk, N.; Nanny, M.A. Characterization and quantification of reversible redox sites in humic substances. Environ. Sci. Technol. 2007, 41, 7844–7850.
- 24. Kaden, J.; Galushko, A.S.; Schink, B. Cysteine-mediated electron transfer in syntrophic acetate oxidation by cocultures of Geobacter sulfurreducens and Wolinella succinogenes. Arch. Microbiol. 2002, 178, 53–58.
- 25. Hong, Y.G.; Guo, J.; Xu, Z.-C.; Xu, M.-Y.; Sun, G.-P. Humic substances act as electron acceptor and redox mediator for microbial dissimilatory azoreduction by Shewanella decolorationis S12. J. Microbiol. Biotechnol. 2007, 17, 428–437.
- 26. Zhou, G.W.; Yang, X.-R.; Li, H.; Marshall, C.W.; Zheng, B.-X.; Yan, Y.; Su, J.-Q.; Zhu, Y.G. Electron shuttles enhance anaerobic ammonium oxidation coupled to iron(III) reduction. Environ. Sci. Technol. 2016, 50, 9298–9307.
- 27. Wang, J.; Zhou, Y.; Li, P.; Lu, H.; Jin, R.; Liu, G. Effects of redox mediators on anaerobic degradation of phenol by Shewanella sp. XB. Appl. Biochem. Biotechnol. 2015, 175, 3162–3172.
- 28. Bosire, E.M.; Rosenbaum, M.A. Electrochemical potential influences phenazine production, electron transfer and consequently electric current generation by Pseudomonas aeruginosa. Front. Microbiol. 2017, 8, 892.
- 29. Light, S.H.; Su, L.; Rivera-Lugo, R.; Cornejo, J.A.; Louie, A.; Iavarone, A.T.; Ajo-Franklin, C.M.; Portnoy, D.A. A flavin-based extracellular electron transfer mechanism in diverse Gram-positive bacteria. Nature 2018, 562, 140–144.
- 30. Shi, Z.; Zachara, J.M.; Shi, L.; Wang, Z.; Moore, D.A.; Kennedy, D.W.; Fredrickson, J.K. Redox reactions of reduced flavin mononucleotide (FMN), riboflavin (RBF), and anthraquinone-2, 6-disulfonate (AQDS) with ferrihydrite and lepidocrocite. Environ. Sci. Technol. 2012, 46, 11644–11652.
- 31. Wang, Y.; Newman, D.K. Redox reactions of phenazine antibiotics with ferric (hydr)oxides and molecular oxygen. Environ. Sci. Technol. 2008, 42, 2380–2386.

- 32. Wang, Z.; Shi, Z.; Shi, L.; White, G.F.; Richardson, D.J.; Clarke, T.A.; Fredrickson, J.K.; Zachara, J.M. Effects of soluble flavin on heterogeneous electron transfer between surface-exposed bacterial cytochromes and iron oxides. Geochim. Cosmochim. Acta 2015, 163, 299–310.
- 33. Tian, T.; Fan, X.; Feng, M.; Su, L.; Zhang, W.; Chi, H.; Fu, D. Flavin-mediated extracellular electron transfer in Gram-positive bacteria Bacillus cereus DIF1 and Rhodococcus ruber DIF2. RSC Adv. 2019, 9, 40903–40909.
- 34. Ackerley, D.; Gonzalez, C.; Keyhan, M.; Blake, R.; Matin, A. Mechanism of chromate reduction by the Escherichia coli protein, NfsA, and the role of different chromate reductases in minimizing oxidative stress during chromate reduction. Environ. Microbiol. 2004, 6, 851–860.
- 35. Liu, G.; Zhou, J.; Jin, R.; Zhou, M.; Wang, J.; Lu, H.; Qu, Y. Enhancing survival of Escherichia coli by expression of azoreductase AZR possessing quinone reductase activity. Appl. Microbiol. Biotechnol. 2008, 80, 409–416.
- 36. Liu, G.; Zhou, J.; Lv, H.; Xiang, X.; Wang, J.; Zhou, M.; Qv, Y. Azoreductase from Rhodobacter sphaeroides AS1. 1737 is a flavodoxin that also functions as nitroreductase and flavin mononucleotide reductase. Appl. Microbiol. Biotechnol. 2007, 76, 1271–1279.
- 37. Morokutti, A.; Lyskowski, A.; Sollner, S.; Pointner, E.; Fitzpatrick, T.B.; Kratky, C.; Gruber, K.; Macheroux, P. Structure and function of YcnD from Bacillus subtilis, a flavin-containing oxidoreductase. Biochemistry 2005, 44, 13724–13733.
- 38. Park, C.; Keyhan, M.; Wielinga, B.; Fendorf, S.; Matin, A. Purification to homogeneity and characterization of a novel Pseudomonas putida chromate reductase. Appl. Environ. Microbiol. 2000, 66, 1788–1795.
- 39. Thatoi, H.; Das, S.; Mishra, J.; Rath, B.P.; Das, N. Bacterial chromate reductase, a potential enzyme for bioremediation of hexavalent chromium: A review. J. Environ. Manag. 2014, 146, 383–399.
- 40. Vorontsov, I.I.; Minasov, G.; Brunzelle, J.S.; Shuvalova, L.; Kiryukhina, O.; Collart, F.R.; Anderson, W.F. Crystal structure of an apo form of Shigella flexneri ArsH protein with an NADPH-dependent FMN reductase activity. Protein Sci. 2007, 16, 2483–2490.
- 41. Gu, B.; Yan, H.; Zhou, P.; Watson, D.B.; Park, M.; Istok, J. Natural humics impact uranium bioreduction and oxidation. Environ. Sci. Technol. 2005, 39, 5268–5275.
- 42. Luan, F.; Burgos, W.D.; Xie, L.; Zhou, Q. Bioreduction of nitrobenzene; natural organic matter, and hematite by Shewanella putrefaciens CN32. Environ. Sci. Technol. 2010, 44, 184–190.
- 43. O'Loughlin, E.J. Effects of electron transfer mediators on the bioreduction of lepidocrocite (y-FeOOH) by Shewanella putrefaciens CN32. Environ. Sci. Technol. 2008, 42, 6876–6882.

- 44. Alvarez, L.; Perez-Cruz, M.; Rangel-Mendez, J.; Cervantes, F. Immobilized redox mediator on metal-oxides nanoparticles and its catalytic effect in a reductive decolorization process. J. Hazard. Mater. 2010, 184, 268–272.
- 45. Cervantes, F.J.; Garcia-Espinosa, A.; Moreno-Reynosa, M.A.; Rangel-Mendez, J.R. Immobilized redox mediators on anion exchange resins and their role on the reductive decolorization of azo dyes. Environ. Sci. Technol. 2010, 44, 1747–1753.
- 46. Li, L.; Zhou, J.; Wang, J.; Yang, F.; Jin, C.; Zhang, G. Anaerobic biotransformation of azo dye using polypyrrole/anthraquinonedisulphonate modified active carbon felt as a novel immobilized redox mediator. Sep. Purif. Technol. 2009, 66, 375–382.
- 47. Lu, H.; Zhang, H.; Wang, J.; Zhou, Y. A novel quinone/reduced graphene oxide composite as a solid-phase redox mediator for chemical and biological Acid Yellow 36 reduction. RSC Adv. 2014, 4, 47297–47303.
- 48. Zhang, H.K.; Hu, X. Catalytic reduction of NACs by nano Fe3O4/quinone composites in the presence of a novel marine exoelectrogenic bacterium under hypersaline conditions. RSC Adv. 2017, 7, 11852–11861.
- 49. Zhang, H.K.; Lu, H.; Zhang, S.; Liu, G.; Li, G.; Zhou, J.; Wang, J. A novel modification of poly (ethylene terephthalate) fiber using anthraquinone-2-sulfonate for accelerating azo dyes and nitroaromatics removal. Sep. Purif. Technol. 2014, 132, 323–329.
- 50. Zhang, H.K.; Lu, H.; Wang, J.; Zhou, J.T.; Sui, M. Cr(VI) reduction and Cr(III) immobilization by Acinetobacter sp. HK-1 with the assistance of a novel quinone/graphene oxide composite. Environ. Sci. Technol. 2014, 48, 12876–12885.

Retrieved from https://encyclopedia.pub/entry/history/show/118085