

# Porcine Stroke Models

Subjects: Zoology

Contributor: TERESA GASULL

In the search of animal stroke models providing translational advantages for biomedical research, pigs are large mammals with interesting brain characteristics and wide social acceptance. Compared to rodents, pigs have human-like highly gyrencephalic brains. In addition, increasingly through phylogeny, animals have more sophisticated white matter connectivity; thus, ratios of white-to-gray matter in humans and pigs are higher than in rodents. Swine models provide the opportunity to study the effect of stroke with emphasis on white matter damage and neuroanatomical changes in connectivity, and their pathophysiological correlate.

Keywords: stroke, swine, gyrencephalic brain, white matter

---

## 1. Introduction

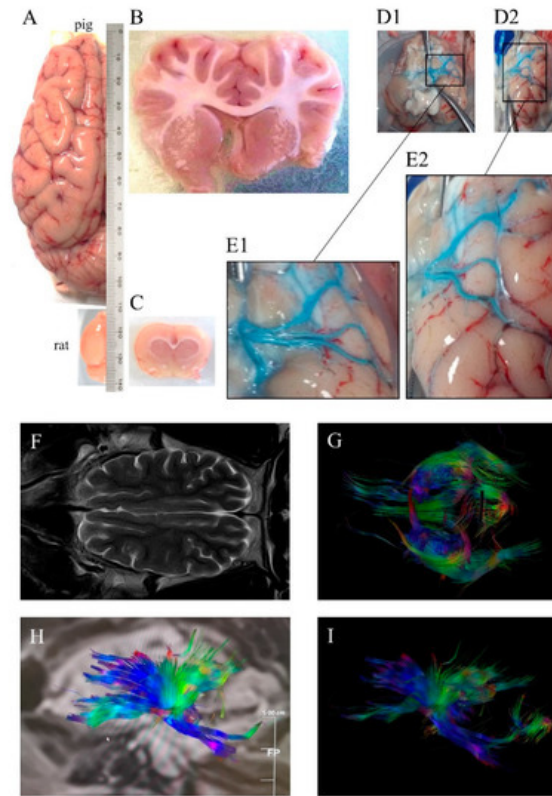
Stroke is a life-threatening disease that causes neuronal loss and subsequent high rates of mortality or permanent disability. Around 17 million stroke cases occur each year worldwide, causing 6 million fatalities and leaving around 6 million patients with serious disabilities. There are two major types of strokes: The ischemic type, which is associated with the occlusion of a cerebral artery and accounts for 85% of all strokes, and the hemorrhagic stroke, which results from blood spill because of an arterial wall rupture. Clinically, stroke treatment is limited to interventions that restore blood flow in the ischemic stroke type, either pharmacologically or via mechanical thrombectomy, and only a small 15% of all stroke patients might benefit from these therapies. Many of the pathophysiological mechanisms of stroke (e.g., excitotoxicity, the core and penumbra concepts, cortical spreading depolarization, excess of free radical production, or inflammation) have been primarily identified as drivers of neuronal death in rodent models with gray matter (GM)-rich brains before being confirmed in human stroke. In contrast, the pathways that contribute to the complex pathology of neuronal soma-devoid areas of white matter (WM) following a stroke event are relatively understudied, and mostly use rodent models or perinatal hypoxic models of cerebral palsy. However, damage to the WM areas is increasingly recognized as a cause of long-term cognitive and motor disabilities in most stroke survivors. This is why research, development, and characterization of swine models of stroke, which combine both the mechanistic knowledge gained in rodent research and a greater degree of neuroanatomical and connectivity similarities with humans, may play a key role to bridge the gap from pre-clinical findings to clinical implementation.

## 2. Pig Brain to Model Human Stroke Pathophysiology

Pigs are big mammals with interesting characteristics for their use in biomedical translational research as compared to dogs and non-human primates (NHPs) <sup>[1]</sup>. There are ethical challenges associated with the fact that NHPs are highly close to humans, and canine models are not as socially accepted as other models due to their setting as companion animals in western cultures <sup>[1][2]</sup>. Conversely, due to their generalized use in the meat industry, pigs are higher-order animals that do not arise the same ethical issues as other animals do in a vast majority of the population <sup>[3]</sup>.

In contrast with lissencephalic brains of rodents—and some commonly used NHPs like the marmoset <sup>[1][4]</sup>—pigs have human-like highly gyrencephalic brains (Figure 1), resembling lobes, gyri, and sulci/fissures of the human brain anatomy <sup>[5][6][7]</sup> with a comparable organization of motor and somatosensory areas to other mammals <sup>[8]</sup>. This resemblance with humans has also been found in several brain structures such as the limbic system, the brainstem, as well as other subcortical and diencephalic nuclei <sup>[5]</sup>. Swine brain mass is comparable to or greater than that of commonly used NHP models <sup>[5][9]</sup> with a well-developed prefrontal cortex in terms of cytoarchitecture and connectivity <sup>[10]</sup>. The development of the brain is remarkably similar to that in humans, including the myelination process <sup>[6][10][11][12]</sup>, showing remarkable similarities in the resting-state networks and connectivity <sup>[13]</sup>. Due to fibrous dura mater, the increase in intracranial pressure (ICP) in large animal models of stroke is similar to that observed in humans suffering stroke <sup>[14]</sup>. Importantly, swine models allow a trustworthy tracking of brain damage evolution using the same non-invasive multimodal imaging sequences and instruments used in clinical practice <sup>[15][16]</sup> (Figure 1). Pigs also display complex individual behaviors and

social interactions that can be analyzed using a number of validated tests [17][18]. Additionally, pigs have some neurovascular characteristics [19][20], including the diameter of their cerebral vessels, which make them suitable for pre-first-in-human validation of endovascular devices or new neurosurgical techniques [15].



**Figure 1.** (A) Dorsal view of pig and rat brain hemispheres showing differences in size and gyrencephaly. Coronal (B) pig and (C) rat sections showing differences in white matter. (D1) Ventral view of pig brain (without cerebellum) injected unilaterally i.a. ex vivo a blue dye, (D2) ventrolateral view of the same brain, (E1) and (E2) magnifications of the insets in (D1) and (D2), respectively, showing three main middle cerebral arteries. (F) Horizontal pig brain section obtained by magnetic resonance T2 sequence, (G) tractography using diffusion tensor imaging (DTI) and (H,I) sagittal pig brain tractography, with colors denoting directionality. Images were obtained at the Comparative Medicine and Bioimage Centre of Catalonia (CMCiB).

As a general rule, larger brains require longer fibers to connect distant cerebral areas. Across species, brain connectivity through white matter increases more rapidly than brain size [1][21]. The ratios of white-to-gray matter in humans and pigs are similar and much higher than those of mice and rats: Rodents have a WM brain composition of 10% compared to the >60% of both humans and swine [1][22][23] (Figure 1). The failure in stroke clinical trials of many neuroprotectants that are beneficial in rodents is thought to be in part due to the fact that the study of treatments with potential protection against ischemic axonal damage in the preclinical phase are seldom considered [24][25]. Hence, using larger gyrencephalic animals with a WM volume similar to that in humans could be pivotal in the preclinical study of stroke pathophysiology and of new clinically effective treatments [26][27].

Another aspect of the swine that should not be disregarded in stroke research is its size (Figure 1), as large animal species offer some advantages compared to smaller species. To begin with, depending on the strain and at sexual maturity, pigs can have a weight similar to that of humans, allowing the use of equipment and procedures specialized for humans in the clinical arena [2][3][28]. Additionally, their size allows repeated sampling of body fluids with minimal physiological disturbance, which eases performing longitudinal studies [3][29]. Moreover, pigs are much closer genetically to humans than mice [30][31], and have the potential to offer a better modeling of human gene regulation and function [32]; also, several models of pathologies and comorbidities usually found in stroke patients, such as obesity or atherosclerosis, have been established in swine [33]. The shorter the phylogenetic distance between the model and the modeled species, the better inference of the results obtained is anticipated [2]; moreover, animal models with comorbidities could potentially improve that inference. In sum, all these features make the pig an excellent research animal to obtain results with more translation to the clinic.

### 3. Conclusions and Future Perspectives

The pig brain resemblance to the human brain provides a unique opportunity to study structural brain damage by non-invasive imaging techniques used in humans and to compare with acute histological and metabolic measurements not available in human studies unless in fatal cases. In addition, new gene-manipulation procedures and comorbidity models in pigs can provide in the years to come the opportunity to experimentally assess the roles of specific molecules and pathways in the pathophysiology of stroke.

The main limitations of swine brain to model human stroke are: (1) The presence of the rete mirabile, that handicaps endovascular access to intracranial arteries to produce focal brain ischemia; (2) the fact that pig plasminogen has resistance to tissue plasminogen activator (tPA)-mediated activation; (3) the significant contribution of both posterior and anterior cerebral arteries to the circulation through the circle of Willis; and that (4) pigs take longer than rodents to reach sexual maturity, adult animal models are interesting due to the relationship between ageing and the disease, but mature pigs of conventional strains can be difficult to manage considering their size. The size issue can be solved by using adult miniature pigs, which in turn are more stable strains adapted for their use in biomedical research. However, studies using aged animals are still lacking. Stroke incidence is notably affected by age and aged brains are in an enhanced inflammatory and oxidative state that renders them more vulnerable to ischemia. Concerning the rete mirabile, the study of Cui et al. mentioned above reports achieving focal cerebral ischemia by an endovascular embolization of the rete unilaterally. The replication of these results would translate into models in which a much less severe surgery is needed to induce the pathology, thus leading to a refinement of the current techniques according to the 3 Rs principle. Even so, new techniques and/or new stroke-inducing paradigms in swine (adapted from rodent stroke models, for instance clot induction by ferrous chloride or by thrombin or innovative approaches) will foster the role of porcine stroke models to bridge the gap from pre-clinical findings to clinical implementation.

---

### References

1. Sorby-adams, A.J.; Vink, R.; Turner, R.J. Large animal models of stroke and traumatic brain injury as translational tools. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 2019.
2. Gieling, E.T.; Schuurman, T.; Nordquist, R.E.; Van der Staay, F.J. The pig as a model animal for studying cognition and neurobehavioral disorders. In *Molecular and Functional Models in Neuropsychiatry*; Springer Science & Business Media: Berlin, Germany, 2011; Volume 7, pp. 359–383. ISBN 9783642287190.
3. Schomberg, D.T.; Tellez, A.; Meudt, J.J.; Brady, D.A.; Dillon, K.N.; Arowolo, F.K.; Wicks, J.; Rousselle, S.D.; Shanmuganayagam, D. Miniature swine for preclinical modeling of complexities of human disease for translational scientific discovery and accelerated development of therapies and medical devices. *Toxicol. Pathol.* 2016, 44, 299–314.
4. Walker, J.; MacLean, J.; Hatsopoulos, N.G. The marmoset as a model system for studying voluntary motor control. *Dev. Neurobiol.* 2017, 77, 273–285.
5. Lind, N.M.; Moustgaard, A.; Jelsing, J.; Vajta, G.; Cumming, P.; Hansen, A.K. The use of pigs in neuroscience: Modeling brain disorders. *Neurosci. Biobehav. Rev.* 2007, 31, 728–751.
6. Sauleau, P.; Lapouble, E.; Val-Laillet, D.; Malbert, C.H. The pig model in brain imaging and neurosurgery. *Animal* 2009, 3, 1138–1151.
7. Kentar, M.; Mann, M.; Sahm, F.; Olivares-Rivera, A.; Sanchez-Porras, R.; Zerelles, R.; Sakowitz, O.W.; Unterberg, A.W.; Santos, E. Detection of spreading depolarizations in a middle cerebral artery occlusion model in swine. *Acta Neurochir. (Wien)* 2020, 162, 581–592.
8. Craner, S.L.; Ray, R.H. Somatosensory cortex of the neonatal pig: I. Topographic organization of the primary somatosensory cortex (SI). *J. Comp. Neurol.* 1991, 306, 24–38.
9. Roth, G.; Dicke, U. Evolution of the brain and intelligence. *Trends Cogn. Sci.* 2005, 9, 250–257.
10. Jelsing, J.; Hay-Schmidt, A.; Dyrby, T.; Hemmingsen, R.; Uylings, H.B.M.; Pakkenberg, B. The prefrontal cortex in the Göttingen minipig brain defined by neural projection criteria and cytoarchitecture. *Brain Res. Bull.* 2006, 70, 322–336.
11. Conrad, M.S.; Dilger, R.N.; Johnson, R.W. Brain growth of the domestic pig (*Sus scrofa*) from 2 to 24 weeks of age: A longitudinal MRI study. *Dev. Neurosci.* 2012, 34, 291–298.
12. Ryan, M.C.; Sherman, P.; Rowland, L.M.; Wijtenburg, S.A.; Acheson, A.; Fieremans, E.; Veraart, J.; Novikov, D.S.; Hong, L.E.; Sladky, J.; et al. Miniature pig model of human adolescent brain white matter development. *J. Neurosci. Methods* 2018, 296, 99–108.

13. Simchick, G.; Shen, A.; Campbell, B.; Park, H.J.; West, F.D.; Zhao, Q. Pig brains have homologous resting-state networks with human brains. *Brain Connect.* 2019, 9, 566–579.
14. Gabrielian, L.; Willshire, L.W.; Helps, S.C.; Van Den Heuvel, C.; Mathias, J.; Vink, R. Intracranial pressure changes following traumatic brain injury in rats: Lack of significant change in the absence of mass lesions or hypoxia. *J. Neurotrauma* 2011, 28, 2103–2111.
15. Cai, B.; Wang, N. Large animal stroke models vs. Rodent stroke models, pros and cons, and combination? In *Brain Edema XVI*; Springer: Cham, Switzerland, 2016; Volume 121, pp. 77–81.
16. Kaiser, E.E.; Waters, E.S.; Fagan, M.M.; Scheulin, K.M.; Platt, S.R.; Jeon, J.H.; Fang, X.; Kinder, H.A.; Shin, S.K.; Duberstein, K.J.; et al. Characterization of tissue and functional deficits in a clinically translational pig model of acute ischemic stroke. *Brain Res.* 2020, 1736.
17. Mendl, M.; Held, S.; Byrne, R.W. Pig cognition. *Curr. Biol.* 2010, 20, 796–798.
18. Kornum, B.R.; Knudsen, G.M. Cognitive testing of pigs (*Sus scrofa*) in translational biobehavioral research. *Neurosci. Biobehav. Rev.* 2011, 35, 437–451.
19. Swindle, M.M.; Smith, A.C. Comparative anatomy and physiology of the pig. *Scand. J. Lab. Anim. Sci.* 1998, 25, 11–21.
20. Ashwini, C.; Shubha, R.; Jayanthi, K. Comparative anatomy of the circle of Willis in man, cow, sheep, goat, and pig. *Neuroanatomy* 2008, 7, 54–65.
21. Zhang, K.; Sejnowski, T.J. A universal scaling law between gray matter and white matter of cerebral cortex. *Proc. Natl. Acad. Sci. USA* 2000, 97, 5621–5626.
22. Baltan, S.; Besancon, E.F.; Mbow, B.; Ye, Z.C.; Hamner, M.A.; Ransom, B.R. White matter vulnerability to ischemic injury increases with age because of enhanced excitotoxicity. *J. Neurosci.* 2008, 28, 1479–1489.
23. Sommer, C.J. Ischemic stroke: Experimental models and reality. *Acta Neuropathol.* 2017, 133, 245–261.
24. Gladstone, D.J.; Black, S.E.; Hakim, A.M. Toward wisdom from failure: Lessons from neuroprotective stroke trials and new therapeutic directions. *Stroke* 2002, 33, 2123–2136.
25. Matute, C.; Ransom, B.R. Roles of white matter in central nervous system pathophysiologies. *ASN Neuro* 2012, 4, 89–101.
26. Fisher, M.; Feuerstein, G.; Howells, D.W.; Hurn, P.D.; Kent, T.A.; Savitz, S.I.; Lo, E.H. Update of the stroke therapy academic industry roundtable preclinical recommendations. *Stroke* 2009, 40, 2244–2250.
27. Hainsworth, A.H.; Allan, S.M.; Boltze, J.; Cunningham, C.; Farris, C.; Head, E.; Ihara, M.; Isaacs, J.D.; Kalaria, R.N.; Lesnik Oberstein, S.A.M.J.; et al. Translational models for vascular cognitive impairment: A review including larger species. *BMC Med.* 2017, 15.
28. Swindle, M.M.; Makin, A.; Herron, A.J.; Clubb, F.J.; Frazier, K.S. Swine as models in biomedical research and toxicology testing. *Vet. Pathol.* 2012, 49, 344–356.
29. Bassols, A.; Costa, C.; Eckersall, P.D.; Osada, J.; Sabrià, J.; Tibau, J. The pig as an animal model for human pathologies: A proteomics perspective. *Proteom. Clin. Appl.* 2014, 8, 715–731.
30. Wernersson, R.; Schierup, M.H.; Jørgensen, F.G.; Gorodkin, J.; Panitz, F.; Stærfeldt, H.H.; Christensen, O.F.; Mailund, T.; Hornshøj, H.; Klein, A.; et al. Pigs in sequence space: A 0.66X coverage pig genome survey based on shotgun sequencing. *BMC Genom.* 2005, 6, 1–7.
31. Jørgensen, F.G.; Hobolth, A.; Hornshøj, H.; Bendixen, C.; Fredholm, M.; Schierup, M.H. Comparative analysis of protein coding sequences from human, mouse and the domesticated pig. *BMC Biol.* 2005, 3.
32. Robert, C.; Kapetanovic, R.; Beraldi, D.; Watson, M.; Archibald, A.L.; Hume, D.A. Identification and annotation of conserved promoters and macrophage-expressed genes in the pig genome. *BMC Genom.* 2015, 16, 1–17.
33. Renner, S.; Blutke, A.; Clauss, S.; Deeg, C.A.; Kemter, E.; Merkus, D.; Wanke, R.; Wolf, E. Porcine models for studying complications and organ crosstalk in diabetes mellitus. *Cell Tissue Res.* 2020.