

Filamin A regulates cardiovascular remodeling

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Filamin A (FLNA) is a large actin-binding cytoskeletal protein that is important for cell motility by stabilizing actin networks and integrating them with cell membranes. Interestingly, a C-terminal fragment of FLNA can be cleaved off by calpain to stimulate adaptive angiogenesis by transporting multiple transcription factors into the nucleus. Recently, increasing evidence suggests that FLNA participates in the pathogenesis of cardiovascular and respiratory diseases, in which the interaction of FLNA with transcription factors and/or cell signaling molecules dictate the function of vascular cells. Localized FLNA mutations associate with cardiovascular malformations in humans. A lack of FLNA in experimental animal models disrupts cell migration during embryogenesis and causes anomalies, including heart and vessels, similar to human malformations.

actin-binding

cell signaling

cytoskeleton

transcription

1. Introduction

Actin-binding cytoskeletal proteins are involved in the formation and maintenance of cell shape and morphology in response to external stimuli from surrounding connective tissue [1]. Filamins are one of the actin-binding proteins that mediate dynamic remodeling during cell movement. Recently, filamins have been shown to be involved in cell signaling and transcription. The filamin family has three members, i.e., filamin A (FLNA), filamin B (FLNB), and filamin C (FLNC). These isoforms exhibit 70% homology with their amino acids and 45% homology in hinge 1 (H1) and hinge 2 (H2) domains [2]. FLNA is the most studied isoform of filamins. Both the human *FLNA* and mouse *FLNA* gene are located on chromosome X. Human *FLNB* and *FLNC* genes are located on chromosomes 3 and 7, respectively, whereas mouse *FLNB* and *FLNC* genes are located on chromosomes 14 and 6, respectively [3]. Although FLNA and FLNB are very similar to one another, their human mutations at different genomic positions result in a wide variety of clinical phenotypes. Filamin isoforms are expressed strongly during embryogenesis. FLNA and FLNB are ubiquitously expressed throughout the body; however, the expression of FLNC is expressed by skeletal and cardiac muscles [3]. The long-elongated Y-shaped (240–280 kDa) polypeptide chain of FLNA (Figure 1) exists in either homo- or heterodimers and each chain consists of 24 immunoglobulin repeats, which are disrupted by two H1 and H2 domains, whereas H1 comprises between 15 and 16 Ig repeats and H2 between 23 and 24 Ig repeats (Figure 1) [4]. These hinge regions are proteolyzed by calpain and separate 24 Ig repeats into the rod 1 domain, which comprises 1–15 Ig repeats, rod 2 comprising 16–23 Ig repeats and a dimerization domain [4]. The hinge regions are proteolyzed by Ca²⁺-dependent calpains, and the cleavage of these sites produces a 90 kDa C-terminal fragment of FLNA (FLNA^{CT}) and a 200 kDa N-terminal fragment (FLNA^{NT}) [6].

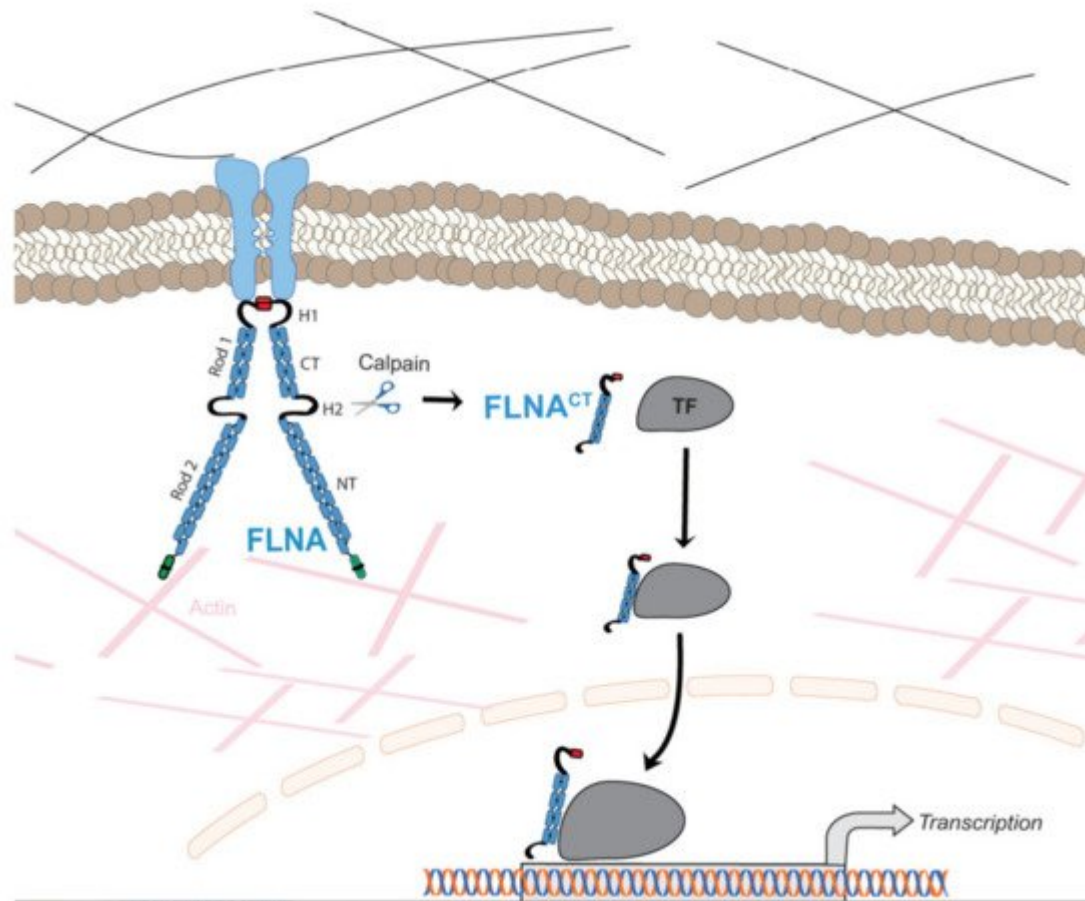


Figure 1. Schematic illustration of the regulation of function of transcriptional factors (TF) by FLNA. Membrane-bound FLNA mediates extracellular signals to the cytoskeleton. FLNA interacts with multiple TF in a cell-specific manner in the cytoplasm. Increased calpain protease activity cleaves membrane-bound FLNA releasing the FLNA^{CT}, which is translocated to the nucleus together with TF. As part of the transcriptional complex, FLNA^{CT} can also bind to promoter regions of target genes. Thus, FLNA^{CT} increases the transactivation function by facilitating translocation to the nucleus and/or nuclear retention and/or by working as a transcriptional coactivator.

1.1. Filamin A in Cellular Signaling and Migration

FLNA interacts with more than 90 partner proteins to execute multiple cellular functions [7], and mainly helps to provide scaffolding to its interacting partners. Due to the high level of similarity between the Ig repeats, multiple proteins bind at multiple sites of FLNA [8]. Other than providing a structural organization for cells, FLNA protein also mediates organ development, cell signaling, migration, proliferation, cell adhesion, phosphorylation, transcription, and the nuclear transportation of transcriptional factors [5].

Cell migration is a crucial step during embryogenesis, wound healing and also remodeling processes, such as myocardial infarction and atherosclerosis. These processes require the specific movement of particular cells to form different tissues. Cell migration is involved in cell polarization, protrusion in the direction of cell movement, retraction, and release from the rear. FLNA is present in most motile cells, at both the leading and rear end, and has been shown to be involved in the remodeling of the actin cytoskeleton, and in cell protrusion and retraction. In

addition, FLNA provides scaffolding for multiple cytoskeletal proteins by the integration of cell adhesion [2][4]. FLNA is more localized to lamellipodia, filopodia, stress fibers and focal adhesions [9]. The possible interactions could be due to the specific binding partners of FLNA, such as receptors and adhesion molecules that mostly reside in the cell region under migration [10]. Furthermore, it could also be through higher concentrations of FLNA in newly assembled actin sites in lamellipodia, due to greater avidity for the branched F-actin junctions [11][12]. Furthermore, endoplasmic reticulum stress inducer IRE1 α interacts with FLNA to induce the cytoskeletal remodeling and cell migration [13].

1.2. Therapeutic Use of Calpain Inhibitors

The H1 and H2 hinge regions of FLNA are proteolyzed by Ca²⁺-dependent calpains that participate in various cellular and physiologic activities, such as cytoskeletal remodeling [14], cell motility, [15], embryonic development [16], signal transduction pathways [17], apoptosis [18], the regulation of gene expression [19], and cell cycles [20]. Chemical calpain inhibitors are generally classified into two groups, i.e., non-peptide calpain inhibitors and peptidomimetic calpain inhibitors [21]. For example, calpain 1 regulates negatively erythrocyte deformability and filtration. As a result, inhibition of calpain 1 has been proposed as a therapeutic approach to treat sickle cell disease [21]. The inhibition of calpain 3 activity treats tibial muscular dystrophy. Either the overexpression or the blocking of calpain 3 activity inhibits disease progression [22]. Both calpain 1 and calpain 2 are extremely abundant in the heart [23]. Despite the enhanced activation of calpains detected during ischemic and reperfusion injury [24], calpain inhibitors in cardiovascular diseases have not been studied in detail. Interestingly, calpain inhibitors have the potential to block the phosphorylation of FAK1, Src, Cdk5 and MAPK kinases that are involved in angiogenesis [25].

2. Genetic Disorders Associated with Human Filamin A Mutations

Due to the wide range of function of FLNA in cell migration and cell signaling, mutations in the *FLNA* gene cause a varying spectrum of developmental malformations, which are also associated with cardiovascular malformations and diseases (Figure 2), whereas *FLNB* and *FLNC* mutations are mainly restricted to skeletal and cardiac muscle diseases, respectively. As these proteins have gained interest recently in the research field, their more predominant functions are yet to be explored. The first mutation to be identified in the *FLNA* gene was the null mutation, where coding amino acids are converted to a stop codon in exon 3, resulting in periventricular nodular heterotopia (PVNH), where a six-layer neocortex is not formed due to failed neuronal migration [26]. PVNH is mainly linked to females, while the majority of hemizygous males are confined to embryonic lethality, and live-born males display aortic dilation and die from a massive hemorrhage in the neonatal period [26]. Female PVNH patients run a high risk of strokes and are associated with other cardiovascular abnormalities, such as valvular abnormalities, persistent ductus arteriosus and aneurysms in the aorta [26]. The majority of PVNH patients with *FLNA* mutations also have thoracic aortic aneurysms [27]. Multiple missense mutations, resulting in substitutions in the actin-binding domain (ABD), have been identified, mainly in the Ig repeats of 9, 10, 14, 16, 22 and 23 of FLNA [28]. These mutations are found in otopalatodigital syndrome, Melnick-Needles syndrome and front metaphyseal dysplasia [28], and are also accompanied by cardiac, tracheobronchial and urological malformations, resulting in perinatal death [29]. However,

phenotypes caused by the otopalatodigital syndrome, Melnick-Needles syndrome and front metaphyseal dysplasia are very distinct from PVNH [30].

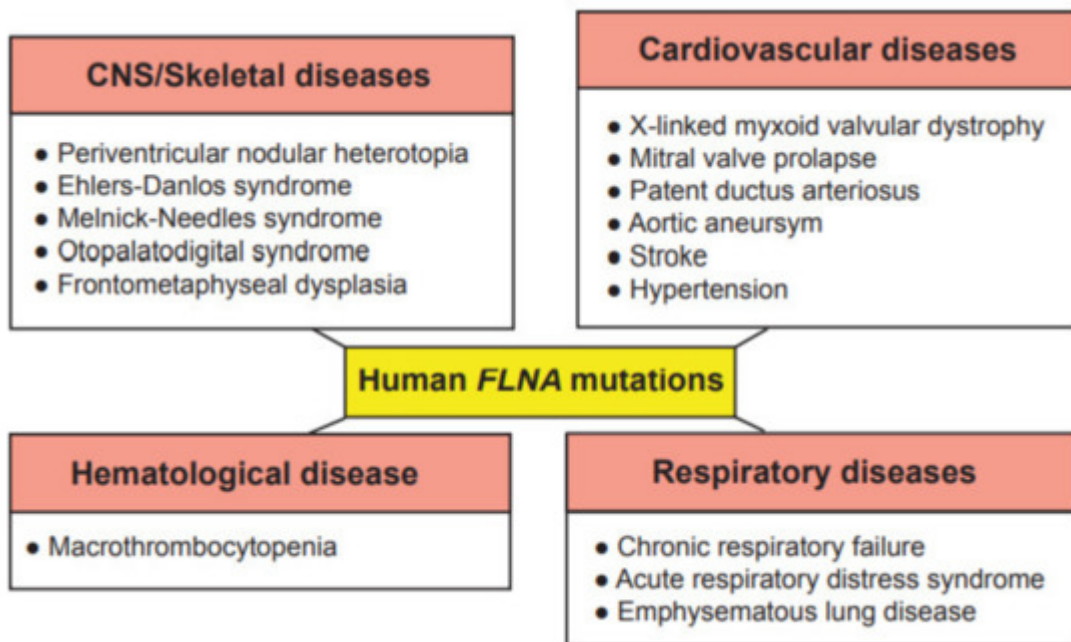


Figure 2. Genetic diseases associated with mutations in the human *FLNA* gene. These genetic diseases are grouped into central nervous system (CNS)/skeletal, cardiovascular, hematological and respiratory diseases.

Missense mutations in the *N*-terminal terminal region of the *FLNA* gene are associated with non-syndromic mitral valve dystrophy [31]. The genomic deletion of codons from exon 19 results in cardiac X-linked myxoid valvular dystrophy (XVMD), a specific cardiovascular malformation [32]. XVMD are frequently involved in vascular anomalies, which include mitral valve prolapse, as well as mitral and aortic regurgitation. Interestingly, it is known that a defective signaling cascade in TGF- β results in impaired mitral valve remodeling, and *FLNA* contributes to changes in cardiac valves by regulating the TGF- β signaling cascade via interaction with SMADs [33]. XVMD are not linked with PVNH and the other congenital disorders mentioned above. Interestingly, a human-induced pluripotent stem cell line from a 10-year-old male patient with cardiac valvular dysplasia, carrying a particular mutation in the *FLNA* gene (c. 84G \rightarrow A), using non-integrative Sendai virus reprogramming technology, has been established to the study molecular mechanisms of cardiac valvular dysplasia, as this cell line can differentiate into three germ layers in vivo [34].

Exon 21 nonsense mutations, frameshift and 4-shift mutations have been identified in *FLNA* to cause bilateral PVNH, along with Ehlers-Danlos syndrome, accompanied by minor cardiovascular malformations [35]. A variant of PVNH, associated with *FLNA* mutations and Ehlers-Danlos syndrome, leads to the development of aortic dilation during early childhood [36]. Novel pathogenic variants have been identified in the *FLNA* gene, causing respiratory failure in newborns [37]. In summary, these human *FLNA* mutations are classified as at least moderate pathogenic associations according to the guidelines provided by the American College of Medical Genetics and Genomics [38].

References

1. Zhou, X.; Borén, J.; Akyürek, L.M. Filamins in cardiovascular development. *Trends Cardiovasc. Med.* 2007, 17, 222–229.
2. Van der Flier, A.; Sonnenberg, A. Structural and functional aspects of filamins. *Biochim. Biophys. Acta* 2001, 1538, 99–117.
3. Lu, J.; Lian, G.; Lenkinski, R.; De Grand, A.; Vaid, R.R.; Bryce, T.; Stasenko, M.; Boskey, A.; Walsh, C.; Sheen, V. Filamin B mutations cause chondrocyte defects in skeletal development. *Hum. Mol. Genet.* 2007, 16, 1661–1675.
4. Stossel, T.P.; Condeelis, J.; Cooley, L.; Hartwig, J.H.; Noegel, A.; Schleicher, M.; Shapiro, S.S. Filamins as integrators of cell mechanics and signalling. *Nat. Rev. Mol. Cell Biol.* 2001, 2, 138–145.
5. Zhou, A.-X.; Hartwig, J.H.; Akyürek, L.M. Filamins in cell signaling, transcription and organ development. *Trends Cell Biol.* 2010, 20, 113–123.
6. Fox, J.E.; Goll, D.E.; Reynolds, C.C.; Phillips, D.R. Identification of two proteins (actin-binding protein and P235) that are hydrolyzed by endogenous Ca²⁺-dependent protease during platelet aggregation. *J. Biol. Chem.* 1985, 260, 1060–1066.
7. Zhou, J.; Kang, X.; An, H.; Lv, Y.; Liu, X. The function and pathogenic mechanism of filamin A. *Gene* 2021, 784, 145575.
8. Nakamura, F.; Stossel, T.P.; Hartwig, J.H. The filamins: Organizers of cell structure and function. *Cell Adh. Migr.* 2011, 5, 160–169.
9. Campbell, I.D. Studies of focal adhesion assembly. *Biochem. Soc. Trans.* 2008, 36, 263–266.
10. Bellanger, J.M.; Astier, C.; Sardet, C.; Ohta, Y.; Stossel, T.P.; Debant, A. The Rac1-and RhoG-specific GEF domain of Trio targets filamin to remodel cytoskeletal actin. *Nat. Cell Biol.* 2000, 2, 888–892.
11. Urban, E.; Jacob, S.; Nemethova, M.; Resch, G.P.; Small, J.V. Electron tomography reveals unbranched networks of actin filaments in lamellipodia. *Nat. Cell Biol.* 2010, 12, 429–435.
12. Nakamura, F.; Osborn, T.M.; Hartemink, C.A.; Hartwig, J.H.; Stossel, T.P. Structural basis of filamin A functions. *J. Cell Biol.* 2007, 179, 1011–1025.
13. Urra, H.; Henriquez, D.R.; Cánovas, J.; Villarroel-Campos, D.; Carreras-Sureda, A.; Pulgar, E.; Molina, E.; Hazari, Y.M.; Limia, C.M.; Alvarez-Rojas, S.; et al. IRE1 α governs cytoskeleton remodelling and cell migration through a direct interaction with filamin A. *Nat. Cell Biol.* 2018, 20, 942–953.

14. Davies, P.; Wallach, D.; Willingham, M.; Pastan, I.; Yamaguchi, M.; Robson, R. Filamin-actin interaction. Dissociation of binding from gelation by Ca²⁺-activated proteolysis. *J. Biol. Chem.* 1978, 253, 4036–4042.
15. Hemmings, L.; Rees, D.; Ohanian, V.; Bolton, S.; Gilmore, A.; Patel, B.; Priddle, H.; Trevithick, J.; Hynes, R.; Critchley, D. Talin contains three actin-binding sites each of which is adjacent to a vinculin-binding site. *J. Cell Sci.* 1996, 109, 2715–2726.
16. Arthur, J.S.C.; Elce, J.S.; Hegadorn, C.; Williams, K.; Greer, P.A. Disruption of the murine calpain small subunit gene, *Capn4*: calpain is essential for embryonic development but not for cell growth and division. *Mol. Cell. Biol.* 2000, 20, 4474–4481.
17. Kishimoto, A.; Mikawa, K.; Hashimoto, K.; Yasuda, I.; Tanaka, S.; Tominaga, M.; Kuroda, T.; Nishizuka, Y. Limited proteolysis of protein kinase C subspecies by calcium-dependent neutral protease (Calpain). *J. Biol. Chem.* 1989, 264, 4088–4092.
18. Kidd, V.J.; Lahti, J.M.; Teitz, T. Proteolytic regulation of apoptosis. *Semin. Cell Dev. Biol.* 2000, 11, 191–201.
19. Pariat, M.; Salvat, C.; Bebien, M.; Brockly, F.; Altieri, E.; Carillo, S.; Jariel-Encontre, I.; Piechaczyk, M. The sensitivity of c-Jun and c-Fos proteins to calpains depends on conformational determinants of the monomers and not on formation of dimers. *Biochem. J.* 2000, 345 Pt 1, 129–138.
20. Watanabe, N.; Woude, G.F.V.; Ikawa, Y.; Sagata, N. Specific proteolysis of the c-mos proto-oncogene product by calpain on fertilization of *Xenopus* eggs. *Nat. Cell Biol.* 1989, 342, 505–511.
21. Donkor, I. An updated patent review of calpain inhibitors (2012—2014). *Expert Opin. Ther. Pat.* 2014, 25, 17–31.
22. Baghdiguian, S.; Martin, M.; Richard, I.; Pons, F.; Astier, C.; Bourg, N.; Hay, R.; Chemaly, R.; Halaby, G.; Loiselet, J.; et al. Calpain 3 deficiency is associated with myonuclear apoptosis and profound perturbation of the I κ B α /NF- κ B pathway in limb-girdle muscular dystrophy type 2A. *Nat. Med.* 1999, 5, 503–511.
23. Ilian, M.A.; Gilmour, R.S.; Bickerstaffe, R. Quantification of ovine and bovine calpain I, calpain II, and calpastatin mRNA by ribonuclease protection assay. *J. Anim. Sci.* 1999, 77, 853–864.
24. Inserte, J.; Hernando, V.; Garcia-Dorado, D. Contribution of calpains to myocardial ischaemia/reperfusion injury. *Cardiovasc. Res.* 2012, 96, 23–31.
25. Leloup, L.; Wells, A. Calpains as potential anti-cancer targets. *Expert Opin. Ther. Targets* 2011, 15, 309–323.
26. Fox, J.W.; Lamperti, E.D.; Ekşioğlu, Y.Z.; E Hong, S.; Feng, Y.; A Graham, D.; Scheffer, I.; Dobyens, W.B.; A Hirsch, B.; A Radtke, R.; et al. Mutations in filamin 1 Prevent Migration of

- Cerebral Cortical Neurons in Human Periventricular Heterotopia. *Neuron* 1998, 21, 1315–1325.
27. Chen, M.H.; Choudhury, S.; Hirata, M.; Khalsa, S.K.; Chang, B.; Walsh, C.A. Thoracic aortic aneurysm in patients with loss of function Filamin A mutations: Clinical characterization, genetics, and recommendations. *Am. J. Med. Genet. Part A* 2018, 176, 337–350.
 28. Robertson, S.P.; Jenkins, Z.A.; Morgan, T.; Adès, L.; Aftimos, S.; Boute, O.; Fiskerstrand, T.; Garcia-Miñaur, S.; Grix, A.; Green, A.; et al. Frontometaphyseal dysplasia: Mutations in FLNA and phenotypic diversity. *Am J. Med. Genet.* 2006, 140A, 1726–1736.
 29. Robertson, S.P.; Twigg, S.R.F.; Sutherland-Smith, V.; Biancalana, V.; Gorlin, R.J.; Horn, D.; Kenwrick, S.J.; Kim, C.; Morava, E.; Newbury-Ecob, R.; et al. Localized mutations in the gene encoding the cytoskeletal protein filamin A cause diverse malformations in humans. *Nat. Genet.* 2003, 33, 487–491.
 30. Feng, Y.; Walsh, C.A. The many faces of filamin: A versatile molecular scaffold for cell motility and signalling. *Nat. Cell Biol.* 2004, 6, 1034–1038.
 31. Le Tourneau, T.; Le Scouarnec, S.; Cueff, C.; Bernstein, D.; Aalberts, J.J.; Lecointe, S.; Mérot, J.; Bernstein, J.A.; Oomen, T.; Dina, C.; et al. New insights into mitral valve dystrophy: A Filamin-A genotype–phenotype and outcome study. *Eur. Hear. J.* 2017, 39, 1269–1277.
 32. Kyndt, F.; Gueffet, J.-P.; Probst, V.; Jaafar, P.; Legendre, A.; Le Bouffant, F.; Toquet, C.; Roy, E.; McGregor, L.; Lynch, S.A.; et al. Mutations in the gene encoding filamin A as a cause for familial cardiac valvular dystrophy. *Circulation* 2007, 115, 40–49.
 33. Sasaki, A.; Masuda, Y.; Ohta, Y.; Ikeda, K.; Watanabe, K. Filamin associates with Smads and regulates transforming growth factor- β signaling. *J. Biol. Chem.* 2001, 276, 17871–17877.
 34. Li, X.; Lu, Y.; Wang, J.; Liu, M.; Wang, M.; Hu, L.; Du, W.; Wang, L.; Jiang, Z.; Gu, X.; et al. An integration-free iPSC line ZZUNEUi008-A derived from dermal fibroblasts of a child with cardiac valvular dysplasia carrying a mutation in FLNA gene. *Stem Cell Res.* 2020, 47, 101882.
 35. Parrini, E.; Ramazzotti, A.; Dobyns, W.B.; Mei, D.; Moro, F.; Veggiotti, P.; Marini, C.; Brilstra, E.H.; Bernardina, B.D.; Goodwin, L.; et al. Periventricular heterotopia: Phenotypic heterogeneity and correlation with filamin A mutations. *Brain* 2006, 129, 1892–1906.
 36. Sheen, V.L.; Jansen, A.; Chen, M.H.; Parrini, E.; Morgan, T.; Ravenscroft, R.; Ganesh, V.; Underwood, T.; Wiley, J.; Leventer, R.; et al. Filamin A mutations cause periventricular heterotopia with Ehlers-Danlos syndrome. *Neurology* 2005, 64, 254–262.
 37. Sasaki, E.; Byrne, A.T.; Phelan, E.; Cox, D.; Reardon, W. A review of filamin A mutations and associated interstitial lung disease. *Eur. J. Nucl. Med. Mol. Imaging* 2019, 178, 121–129.
 38. Richards, S.; Aziz, N.; Bale, S.; Bick, D.; Das, S.; Gastier-Foster, J.; Grody, W.W.; Hegde, M.; Lyon, E.; Committee ALQA; et al. Standards and guidelines for the interpretation of sequence

variants: A joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. *Genet. Med.* 2015, 17, 405–424.

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