

# Medulloblastoma

Subjects: Oncology

Submitted by:  Terence Duarte

## Definition

Medulloblastoma is the most prevalent malignant brain tumor in children, while it accounts for only 1–2% of adult brain tumors. Recognized as a biologically heterogeneous disease, the World Health Organization (WHO) considers there to be four molecular subgroups: wingless-activated (WNT), sonic hedgehog-activated (SHH); Group 3; and Group 4. Recently, the picture became more complex when 12 different medulloblastoma subtypes were described, including two WNT subtypes, four SHH subtypes, three group 3 subtypes, and three group 4 subtypes, with each subgroup being characterized by specific mutations, copy number variations, transcriptomic/methylomic profiles, and clinical outcomes. For the SHH subgroup MB, germline or somatic mutations and a copy-number variation are the common drivers that affect critical genes involved in SHH signaling, including PTCH1 (patched 1 homologue), SUFU (suppressor of fused homologue), and SMO (smoothened), among others

---

## 1. Introduction

The most common genetic events, which occur in both pediatric and adult tumors, are loss-of-function, mutations, or deletions in PTCH1 and SUFU, which act as negative regulators of SHH signaling <sup>[1][2][3]</sup>. Activation of mutations and amplification of SMO or GLI2 (glioma associated oncogene homologue 2) also lead to constitutive activation of the SHH pathway <sup>[4][5]</sup>. Germline and somatic TP53 mutations predominantly coincident with GLI2 amplifications and are found exclusively in children between the ages of 8 and 17 years <sup>[6][7][8][9]</sup>. Somatic TERT (telomerase) promoter hotspot mutations are also associated with the SHH subgroup <sup>[10][11]</sup>. Mutation of PTEN (phosphatase and tensin homolog) is found in more than 5% of human SHH subgroup MB cases and is associated with decreased expression of PTEN mRNA and proteins in the cerebellum <sup>[9][12]</sup>. In addition, genetically engineered mouse models (GEMMs) carrying mutations/overexpression of those genes have also been developed to study this medulloblastoma subgroup <sup>[13][14][15]</sup>.

Medulloblastoma can also be viewed through the lens of the tumor microenvironment (TME), and its multiple roles in cancer offer an interesting way to identify the critical steps regulating medulloblastoma biology, disease progression, and overall survival <sup>[16][17][18][19][20][21][22][23][24][25][26][27][28][29]</sup>. In addition to tumor cells, the tumor microenvironment is characterized by diverse cell populations, including stem-like cells and tumor-associated components such as blood vessels <sup>[18]</sup>, immune cells <sup>[24][25]</sup>, neurons, endothelial cells, microglia <sup>[26]</sup>, macrophages <sup>[27][28]</sup>, and astrocytes <sup>[17][19][20][21][29]</sup>. The communication between these unique collections of cell types is implicated in therapy resistance <sup>[30][31][32]</sup>, immune infiltration, and inflammation <sup>[28]</sup>. Since tumor-associated cells could be the focus of therapeutic vulnerability, a comprehensive understanding of the interactions between the tumor cells and the tumor-associated components may provide new opportunities for targeted discoveries. In the SHH subgroup MB, recent studies have highlighted that the cellular diversity within tumors has a critical role in supporting the growth of tumor cells and the robustness of cancer <sup>[17][19][21][25][27][28][29][33][34]</sup>. In MB-prone mice with a SMO mutation, the TME contains tumor cell types that exist across a spectrum of differentiation states and tumor-derived cells that express markers for astrocytic and oligodendrocytic precursors <sup>[35]</sup>. This suggests that even in a tumor with a single pathway-activation mutation, diverse mechanisms may drive tumor growth, demonstrating the need to target multiple pathways simultaneously for therapeutic effectiveness.

## Astrocytes and the Medulloblastoma Microenvironment: The New Player within the Complex Ecosystem

Due to increasing evidence of an association between wound healing and the development of tumors, recent studies have investigated the complex functions of astrocytes involved in the support of medulloblastoma growth, as these specialized glial cells are involved in the functional recovery of the central nervous system (CNS) [17][20][21][36][37][38][39][40][41][42][43][44][45][46][47][48][49][50][51][52]. Astrocytes are specialized and heterogeneous cells that are essential modulators of local blood flow as well as being involved in the maintenance of homeostasis of extracellular fluids, ions, and transmitters [49][50]. In a healthy CNS, these glial cells participate in synaptic function and plasticity among other dynamic activities that are crucial for the neural circuit and neurological function and behavior [49][50]. In this context, recent studies have identified SHH signaling as an essential regulator of the molecular identity and functional properties of astrocytes [51][52]. Under normal conditions, astrocytes express the components of the SHH pathway, but do not secrete the SHH protein [53][54]. Recent in vivo studies have shown that the SHH pathway is active in astrocytes of the mature forebrain through the SHH transduction system, which includes the receptor PTCH1 as well as GLI transcription factors [54][55]. Other studies have also demonstrated that the SHH protein is mainly produced by neurons in several brain areas, including dopaminergic neurons [56], the Purkinje cells and mossy cells in the hippocampus, but not in astrocytes or oligodendrocytes [57]. In addition, under physiological stress or pathological conditions, it has been reported that astrocytes may be able to produce and become powerful sources of the SHH protein [58][59][60][61].

In the cerebellum, specialized, unipolar astrocytes called Bergmann glia (BG) have been shown to be capable of responding to the Purkinje-derived SHH protein from the postnatal stage through to adulthood [62]. Mice in which SMO is postnatally ablated in BG demonstrate reduced proliferation of granule cell precursors (GCP) and increased differentiation accompanied by a loss of SHH activity. In these animals, WNT signaling is ectopically elevated in GCP, suggesting that this pathway is involved in cross-talk with the SHH pathway, which helps to regulate GCP proliferation [62].

Astrocyte reactivity (AR) [38][39][40][41][42][43][44][45][49][51], an ubiquitous, complex, and multistage process, is known to be involved in different CNS pathologies, including trauma [39], inflammation [36], stem-cell repair [41], regeneration [42], peripheral metabolic disorders [43], neurodegenerative diseases [40][44], and tumor progression [34][45][47][63][64][65]. In the context of brain metastasis, reactive astrocytes have a dual role: they limit disease progression during the early stages and, later on, foster tumor growth [45]. During tumor progression, reactive astrocytes are key components of the microenvironment, and their function and crosstalk with other components of the TME have been targets of neuro-oncology research [17][21][33][63][64][65]. Astrocytes can act through paracrine secretion of degradative enzymes, cytokines, chemokines, and growth factors [36] and have multiple primary and branching endfeet that interact with tumor cells, facilitating growth, proliferation, survival, and invasion. Recent studies have demonstrated that, in brain tumors, astrocytes secrete cytokines and trophic factors and contribute to tumor growth, metastasis, and resistance to current therapy [41][43]. In primary gliomas and brain metastases, astrocytes establish gap junctions with tumor cells, and these functional connections are regulated by signaling molecules, such as connexin [43]. In response to these non-cell-autonomous stimuli, astrocytes can produce a multitude of molecular signals that can, in turn, influence many different neural and non-neural cell types, including cells involved in innate immune responses [63]. In parasite infections, astrocytes secrete the SHH protein which, in turn, induces the production of GRP78, an endoplasmic reticulum (ER) chaperone from the heat shock protein family [20]. Under ER stress, it is believed that the activation of GRP78 may increase cell survival through the unfolded protein response and may also protect cells from ER-stress-induced apoptosis by activating Bcl-2 and inhibiting Bak, Bax, Caspase, and CHOP [20]. In fact, astrocytes facilitate the formation of medulloblastoma tumoroids by secreting SHH proteins and generating the astrocyte-derived extracellular matrix [29].

The roles of astrocytes in the medulloblastoma microenvironment have been investigated, and studies have demonstrated that astrocytes secrete CD133, a key cancer stem cell marker that is involved in medulloblastoma tumorigenicity and alters gene expression, increasing invasion and adhesion by medulloblastoma cells [48]. Astrocytes can also have a direct influence on brain tumor stem cells that are

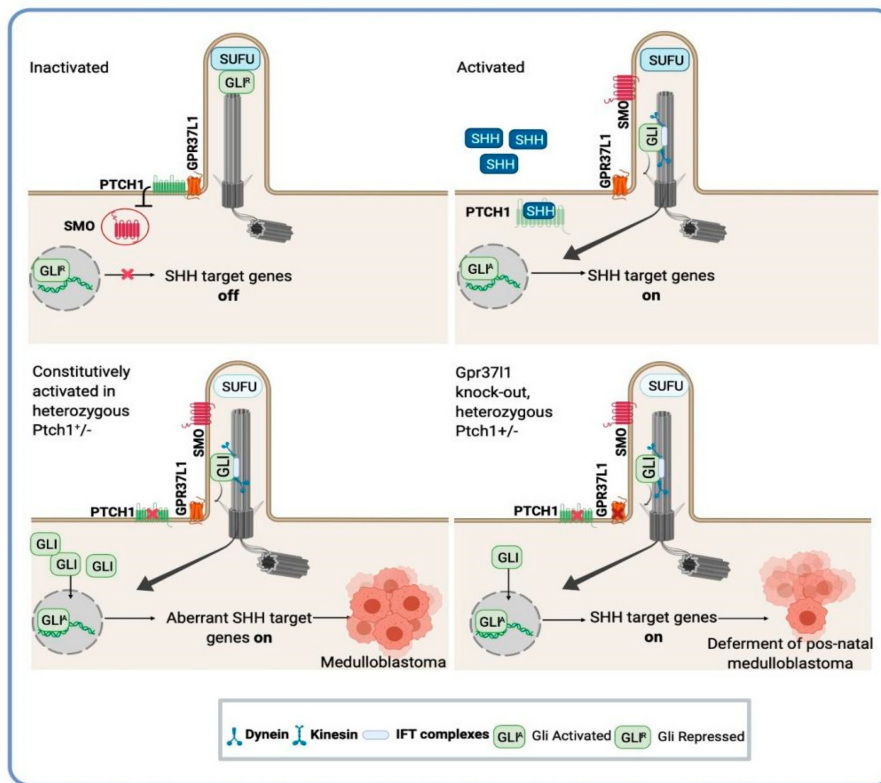
activated by several ligands, including SHH, which enriches the stem cell population [66]. These interactions are bi-directional, and tumor stem cells can provide signals that affect the surrounding astrocytes [65].

## **2. Novel Targets and Therapeutic Opportunities for Medulloblastoma: A Potential Application of Astrocytes-SHH Medulloblastoma Cross-Talk Research**

Current understating of the contribution of the TME to the growth of tumor cells has shifted the focus of neuro-oncology research, moving from exclusively targeting tumoral cells to targeting the tumor microenvironment or signals coming from it, as well as the interactions between them [67][68][21][29][33][66][69]. From multiple studies, it has become clear that the interplay between tumor cells and cells of the tumor microenvironment orchestrate events that are critical to tumor progression, and in this way, many cellular and molecular elements of the microenvironment are emerging as attractive targets for therapeutic strategies [17][23][24][29][31][70]. Although GPCs are the most studied cells regarding the origin of SHH MB and have been the focus of the search for targets in the medulloblastoma for some time, protein receptors and peptide factors from other cellular sources that impact SHH subgroup MB have attracted the attention of researchers more recently.

The G protein-coupled receptor (GPCR) family of proteins is widely dysregulated in cancer and yet is underexploited in oncology. Recent studies have shown that GPCRs can play multiple roles in cancer progression, including proliferation, survival, angiogenesis, metastasis, therapy resistance, and immune evasion upon activation by ligands produced by cancer cells or through the multiplicity of cells within the tumor stroma [69]. The mitogenic ciliary functions of G-protein coupled receptor 37-like 1 (GPR37L1) in SHH-SMO signaling are particularly attractive to target cancer via the tumor microenvironment. The GPR37L1, an orphan G-protein-coupled receptor is a selective marker of cerebellar BG astrocytes [70][71], and it specifically colocalizes and interacts with the PTCH1 protein in discrete areas of these Bergmann glia cell membranes in newborn mice [70]. The Bergmann glial cells possess the primary cilia (PC), which are antenna-like organelles required for sensing and transducing extracellular stimuli [72]. These PC are essential for the regulation of several signaling pathways, such as SHH and WNT, and they can promote tumorigenesis in medulloblastoma [72][73]. Primary cilia have been reliably detected in all cells of pre-neoplastic MB in PTCH1+/- mice [19]. Thus, the specific detection of primary cilia could be usefully applied for the study of early, pre-neoplastic MB lesions [73].

GPR37L1-/- mice present with precocious Bergmann glia, Purkinje neuron maturation, and increased levels of Purkinje secreted SHH protein, as well as SMO and the intracellular effectors of the SHH-SMO cascade, MYCN and GLI2 [70]. In cerebellar primary astrocyte cultures from GPR37L1-/- mouse pups, these astrocytes displayed striking increases in proliferative activity, PTCH1 protein expression and internalization, intracellular cholesterol content, and ciliary localization of SMO, as well as marked production of active SHH signaling [74]. Similar effects were reproduced by treating wild-type astrocytes with a putative prosaptide ligand of the GPR37L1 receptor [74]. Using GPR37L1-/-Ptch1+/- mice, Di Pietro et al. (2019) showed that genetic ablation of GPR37L1 in this medulloblastoma-prone mouse model can reduce the occurrence and severity of postnatal tumors [19] (**Figure 1**). These authors speculated that this receptor could be involved in the process of BG modulation of SHH production by Purkinje neurons and suggested the involvement of WNT3, a specific inhibitor of SHH-induced neuronal mitogenesis [19]. As GPCRs are the most “druggable” class of proteins currently known, the GPR37L1 receptors have become a highly valuable target for the development of novel therapies, and their use as a specific blocking agent may be an important target for medulloblastoma treatment.



**Figure 1.** In the absence of the sonic hedgehog ligand (SHH), the negative regulator PTCH1 is present on the ciliary membrane. In this state (Inactivated), suppressor of fused (SUFU) forms a complex with the GLI transcription factors in the periciliary region. SHH binding to PTCH1 (Activated) induces its translocation away from the cilium and promotes the entry of the activating receptor SMO. This process allows the migration of active GLI (GLI<sup>A</sup>) into the nucleus where the transcription of SHH target genes is activated. Mice heterozygous for loss-of-function PTCH1 mutations have a higher incidence of medulloblastoma [13][75][76]. In a GPR3711<sup>-/-</sup>Ptch1<sup>+/-</sup> mouse model, the lack of GPR3711 reduced the postnatal tumor occurrence of tumors and decreased the incidence of more aggressive tumor types [19].

Regarding the critical tumor-stroma interaction related to SHH subgroup MB, Snuderl et al. showed that stromal cells produce Placental growth factor (PIGF), a member of the vascular endothelial growth factor (VEGF) family, which is stimulated via paracrine SHH ligand secretion by the tumor cells [77]. In vitro results have demonstrated that PIGF, its receptor neuropilin 1 (Nrp1), and the MAPK signaling axis are critical for the survival of medulloblastoma cells, and in Smo/Smo transgenic mice, the blockade of PIGF with anti-PIGF antibodies was associated with significantly smaller tumors [77]. Together, these findings provide insight into the roles of SHH, PIGF, and Nrp1 in SHH subgroup MB, and as PIGF is dispensable during development, they support the idea that this SHH tumor and PIGF interaction may be used as a therapeutic approach for this pediatric tumor.

Added to the findings highlighted here, the effects of SHH protein secretion by tumor astrocytes on tumor progression and the adaptive transdifferentiation of the tumor and its reliance on astrocytic signals open new perspectives for discovering multiple potential therapeutic targets within the tumor-associated glial cells. With the large amount of genetic and epigenetic data from the subsets of human medulloblastoma, the design of future medulloblastoma therapies must be based on the identification of genes that are specifically expressed in these tumor-associated astrocytes.

## References

1. Raffel, C.; Jenkins, R.B.; Frederick, L.; Hebrink, D.; Alderete, B.; Fults, D.W.; James, C.D. Sporadic medulloblastomas contain PTCH mutations. *Cancer Res.* 1997, 57, 842–845.
2. Taylor, M.D.; Liu, L.; Raffel, C.; Hui, C.C.; Mainprize, T.G.; Zhang, X.; Agatep, R.; Chiappa, S.; Gao, L.; Lowrance, A.; et

- al. Mutations in *SUFU* predispose to medulloblastoma. *Nat. Genet.* 2002, 31, 306–310.
3. Brugières, L.; Pierron, G.; Chompret, A.; Paillerets, B.B.-D.; Di Rocco, F.; Varlet, P.; Pierre-Kahn, A.; Caron, O.; Grill, J.; Delattre, O. Incomplete penetrance of the predisposition to medulloblastoma associated with germ-line *SUFU* mutations. *J. Med. Genet.* 2009, 47, 142–144.
  4. Gibson, P.; Tong, Y.; Robinson, G.; Thompson, M.C.; Currle, D.S.; Eden, C.; Kranenburg, T.; Hogg, T.; Poppleton, H.; Martin, J.; et al. Subtypes of medulloblastoma have distinct developmental origins. *Nat. Cell Biol.* 2010, 12, 1095–1099.
  5. Buczkowicz, P.; Ma, J.; Hawkins, C. *GLI2* Is a Potential Therapeutic Target in Pediatric Medulloblastoma. *J. Neuropathol. Exp. Neurol.* 2011, 70, 430–437.
  6. Zhukova, N.; Ramaswamy, V.; Remke, M.; Pfaff, E.; Shih, D.J.H.; Martin, D.C.; Castelo-Branco, P.; Baskin, B.; Ray, P.N.; Bouffet, E.; et al. Subgroup-Specific Prognostic Implications of *TP53* Mutation in Medulloblastoma. *J. Clin. Oncol.* 2013, 31, 2927–2935.
  7. Louis, D.N.; Perry, A.; Reifenberger, G.; von Deimling, A.; Figarella-Branger, D.; Cavenee, W.K.; Ohgaki, H.; Wiestler, O.D.; Kleihues, P.; Ellison, D.W. The 2016 World Health Organization Classification of Tumors of the Central Nervous System: A summary. *Acta Neuropathol.* 2016, 131, 803–820.
  8. Ramaswamy, V.; Remke, M.; Bouffet, E.; Bailey, S.; Clifford, S.C.; Doz, F.; Kool, M.; Dufour, C.; Vassal, G.; Milde, T.; et al. Risk stratification of childhood medulloblastoma in the molecular era: The current consensus. *Acta Neuropathol.* 2016, 131, 821–831.
  9. Da Silva, L.S.; Mançano, B.M.; de Paula, F.E.; dos Reis, M.B.; de Almeida, G.C.; Matsushita, M.; Junior, C.A.; Evangelista, A.F.; Saggiaro, F.; Serafini, L.N.; et al. Expression of *GNAS*, *TP53*, and *PTEN* Improves the Patient Prognostication in Sonic Hedgehog (*SHH*) Medulloblastoma Subgroup. *J. Mol. Diagn.* 2020, 22, 957–966.
  10. Viana-Pereira, M.; Almeida, G.C.; Stavale, J.N.; Malheiro, S.; Clara, C.; Lobo, P.; Pimentel, J.; Reis, R. Study of *hTERT* and Histone 3 Mutations in Medulloblastoma. *Pathobiology* 2016, 84, 108–113.
  11. Remke, M.; Ramaswamy, V.; Peacock, J.; Shih, D.J.H.; Koelsche, C.; Northcott, P.A.; Hill, N.; Cavalli, F.M.G.; Kool, M.; Wang, X.; et al. *TERT* promoter mutations are highly recurrent in *SHH* subgroup medulloblastoma. *Acta Neuropathol.* 2013, 126, 917–929.
  12. Hartmann, W.; Dignon-Söntgerath, B.; Koch, A.; Waha, A.; Endl, E.; Dani, I.; Denkhaus, D.; Goodyer, C.G.; Sörensen, N.; Wiestler, O.D.; et al. Phosphatidylinositol 3'-Kinase/*AKT* Signaling Is Activated in Medulloblastoma Cell Proliferation and Is Associated with Reduced Expression of *PTEN*. *Clin. Cancer Res.* 2006, 12, 3019–3027.
  13. Goodrich, L.V.; Milenković, L.; Higgins, K.M.; Scott, M.P. Altered neural cell fates and medulloblastoma in mouse patched mutants. *Science* 1997, 277, 1109–1113.
  14. Kimura, H.; Stephen, D.; Joyner, A.; Curran, T. *Gli1* is important for medulloblastoma formation in *Ptc1*<sup>+/-</sup> mice. *Oncogene* 2005, 24, 4026–4036.
  15. Castellino, R.C.; Barwick, B.G.; Schniederjan, M.; Buss, M.C.; Becher, O.; Hambardzumyan, D.; Macdonald, T.J.; Brat, D.J.; Durden, D.L. Heterozygosity for *Pten* promotes tumorigenesis in a mouse model of medulloblastoma. *PLoS ONE* 2010, 5, e10849.
  16. Byrd, T.; Grossman, R.G.; Ahmed, N. Medulloblastoma-Biology and microenvironment: A review. *Pediatr. Hematol. Oncol.* 2012, 29, 495–506.
  17. Liu, Y.; Yuelling, L.W.; Wang, Y.; Du, F.; Gordon, R.E.; O'Brien, J.A. Astrocytes promote medulloblastoma progression through hedgehog secretion. *Cancer Res.* 2017, 77, 6692–6703.
  18. Hambardzumyan, D.; Becher, O.J.; Rosenblum, M.K.; Pandolfi, P.P.; Manova-Todorova, K.; Holland, E.C. *PI3K* pathway regulates survival of cancer stem cells residing in the perivascular niche following radiation in medulloblastoma in vivo. *Genes Dev.* 2008, 22, 436–448.
  19. Di Pietro, C.; La Sala, G.; Matteoni, R.; Marazziti, D.; Tocchini-Valentini, G.P. Genetic ablation of *Gpr3711* delays tumor occurrence in *Ptch1*<sup>+/-</sup> mouse models of medulloblastoma. *Exp. Neurol.* 2019, 312, 33–42.
  20. Chen, K.Y.; Chen, Y.J.; Cheng, C.J.; Jhan, K.Y.; Wang, L.C. Excretory/secretory products of *Angiostrongylus cantonensis* fifth-stage larvae induce endoplasmic reticulum stress via the Sonic hedgehog pathway in mouse astrocytes. *Parasit. Vectors* 2020, 13, 317.
  21. Yao, M.; Ventura, P.B.; Jiang, Y.; Rodriguez, F.J.; Wang, L.; Perry, J.S.A. Astrocytic trans-Differentiation Completes a Multicellular Paracrine Feedback Loop Required for Medulloblastoma Tumor Growth. *Cell* 2020, 180, 502–520.e19.
  22. Zhou, R.; Joshi, P.; Katsushima, K.; Liang, W.; Liu, W.; Goldenberg, N.A. The emerging field of noncoding RNAs and their importance in pediatric diseases. *J. Pediatr.* 2020, 221, S11–S19.
  23. Chung, A.S.; Ferrara, N. Targeting the tumor microenvironment with *Src* kinase inhibition. *Clin. Cancer Res.* 2010, 16, 775–777.
  24. Binnewies, M.; Roberts, E.W.; Kersten, K.; Chan, V.; Fearon, D.F.; Merad, M.; Coussens, L.M.; Gaborilovich, D.I.; Ostrand-Rosenberg, S.; Hedrick, C.C.; et al. Understanding the tumor immune microenvironment (TIME) for effective therapy. *Nat. Med.* 2018, 24, 541–550.
  25. Gajewski, T.F.; Schreiber, H.; Fu, Y.-X. Innate and adaptive immune cells in the tumor microenvironment. *Nat. Immunol.* 2013, 14, 1014–1022.

26. Amarante, M.K.; Vitiello, G.A.F.; Rosa, M.H.; Mancilla, I.A.; Watanabe, M.A.E. Potential use of CXCL12/CXCR4 and sonic hedgehog pathways as therapeutic targets in medulloblastoma. *Acta Oncol.* 2018, 57, 1134–1142.
27. Margol, A.S.; Robison, N.J.; Gnanachandran, J.; Hung, L.T.; Kennedy, R.J.; Vali, M.; Dhall, G.; Finlay, J.L.; Epstein, A.; Krieger, M.D.; et al. Tumor-Associated Macrophages in SHH Subgroup of Medulloblastomas. *Clin. Cancer Res.* 2015, 21, 1457–1465.
28. Pham, C.D.; Mitchell, D.A. Know your neighbors: Different tumor microenvironments have implications in immunotherapeutic targeting strategies across MB subgroups. *Oncoimmunology* 2016, 5, e1144002.
29. Cheng, Y.; Franco-Barraza, J.; Wang, Y.; Zheng, C.; Zhang, L.; Qu, Y.; Long, Y.; Cukierman, E.; Yang, Z.-J. Sustained hedgehog signaling in medulloblastoma tumoroids is attributed to stromal astrocytes and astrocyte-derived extracellular matrix. *Lab. Investig.* 2020, 100, 1208–1222.
30. Raviraj, R.; Nagaraja, S.S.; Selvakumar, I.; Mohan, S.; Nagarajan, D. The epigenetics of brain tumors and its modulation during radiation: A review. *Life Sci.* 2020, 256, 117974.
31. Hirata, E.; Sahai, E. Tumor microenvironment and differential responses to therapy. *Cold Spring Harb. Perspect. Med.* 2017, 7, a026781.
32. Hanahan, D.; Weinberg, R.A. Hallmarks of Cancer: The Next Generation. *Cell* 2011, 144, 646–674.
33. Tamayo-Orrego, L.; Charron, F. Recent advances in SHH medulloblastoma progression: Tumor suppressor mechanisms and the tumor microenvironment. *F1000Research* 2019, 8, 1823.
34. Raza, M.; Prasad, P.; Gupta, P.; Kumar, N.; Sharma, T.; Rana, M.; Goldman, A.; Sehrawat, S. Perspectives on the role of brain cellular players in cancer-associated brain metastasis: Translational approach to understand molecular mechanism of tumor progression. *Cancer Metastasis Rev.* 2018, 37, 791–804.
35. Ocasio, J.; Babcock, B.; Malawsky, D.; Weir, S.J.; Loo, L.; Simon, J.M.; Zylka, M.J.; Hwang, D.; Dismuke, T.; Sokolsky, M.; et al. scRNA-seq in medulloblastoma shows cellular heterogeneity and lineage expansion support resistance to SHH inhibitor therapy. *Nat. Commun.* 2019, 10, 5829.
36. Sofroniew, M.V. Multiple roles for astrocytes as effectors of cytokines and inflammatory mediators. *Neuroscientist* 2014, 20, 160–172.
37. Khakh, B.S.; Sofroniew, M.V. Diversity of astrocyte functions and phenotypes in neural circuits. *Nat. Neurosci.* 2015, 18, 942–952.
38. Anderson, M.A.; Ao, Y.; Sofroniew, M.V. Heterogeneity of reactive astrocytes. *Neurosci. Lett.* 2014, 565, 23–29.
39. Burda, J.E.; Bernstein, A.M.; Sofroniew, M.V. Astrocyte roles in traumatic brain injury. *Exp. Neurol.* 2016, 275, 305–315.
40. Tong, X.; Ao, Y.; Faas, G.C.; Nwaobi, S.E.; Xu, J.; Hausteiner, M.D.; Anderson, M.A.; Mody, I.; Olsen, M.; Sofroniew, M.V.; et al. Astrocyte Kir4.1 ion channel deficits contribute to neuronal dysfunction in Huntington’s disease model mice. *Nat. Neurosci.* 2014, 17, 694–703.
41. Robel, S.; Berninger, B.; Götz, M. The stem cell potential of glia: Lessons from reactive gliosis. *Nat. Rev. Neurosci.* 2011, 12, 88–104.
42. Silver, J.; Miller, J.H. Regeneration beyond the glial scar. *Nat. Rev. Neurosci.* 2004, 5, 146–156.
43. Zamanian, J.L.; Xu, L.; Foo, L.C.; Nouri, N.; Zhou, L.; Giffard, R.G.; Barres, B.A. Genomic analysis of reactive astrogliosis. *J. Neurosci.* 2012, 32, 6391–6410.
44. Pekny, M.; Pekna, M. Reactive gliosis in the pathogenesis of CNS diseases. *Biochim. Biophys Acta.* 2016, 1862, 483–491.
45. Wasilewski, D.; Priego, N.; Fustero-Torre, C.; Valiente, M. Reactive astrocytes in brain metastasis. *Front. Oncol.* 2017, 7, 298.
46. Placone, A.L.; Quiñones-Hinojosa, A.; Searson, P.C. The role of astrocytes in the progression of brain cancer: Complicating the picture of the tumor microenvironment. *Tumor Biol.* 2016, 37, 61–69.
47. Brandao, M.; Simon, T.; Critchley, G.; Giamas, G. Astrocytes, the rising stars of the glioblastoma microenvironment. *Glia* 2019, 67, 779–790.
48. Gronseth, E.; Gupta, A.; Koceja, C.; Kumar, S.; Kutty, R.G.; Rarick, K.; Wang, L.; Ramchandran, R. Astrocytes influence medulloblastoma phenotypes and CD133 surface expression. *PLoS ONE* 2020, 15, e0235852.
49. Nedergaard, M.; Ransom, B.; Goldman, S.A. New roles for astrocytes: Redefining the functional architecture of the brain. *Trends Neurosci.* 2003, 26, 523–530.
50. Barres, B.A. The Mystery and Magic of Glia: A perspective on their roles in health and disease. *Neuron* 2008, 60, 430–440.
51. Sofroniew, M.V. Molecular dissection of reactive astrogliosis and glial scar formation. *Trends Neurosci.* 2009, 32, 638–647.
52. Sofroniew, M.V. Astrocyte reactivity: Subtypes, states, and functions in CNS innate immunity. *Trends Immunol.* 2020, 41, 758–770.
53. Traiffort, E.; Charytoniuk, D.; Watroba, L.; Faure, H.; Sales, N.; Ruat, M. Discrete localizations of hedgehog signalling components in the developing and adult rat nervous system. *Eur. J. Neurosci.* 1999, 11, 3199–3214.
54. Garcia, A.D.; Petrova, R.; Eng, L.; Joyner, A.L. Sonic Hedgehog regulates discrete populations of astrocytes in the adult mouse forebrain. *J. Neurosci.* 2010, 30, 13597–13608.

55. Jiao, J.; Chen, D.F. Induction of neurogenesis in nonconventional neurogenic regions of the adult central nervous system by niche astrocyte-produced signals. *Stem Cells* 2008, 26, 1221–1230.
56. Gonzalez-Reyes, L.E.; Verbitsky, M.; Blesa, J.; Jackson-Lewis, V.; Paredes, D.; Tillack, K.; Phani, S.; Kramer, E.; Przedborski, S.; Kottmann, A. Sonic Hedgehog Maintains Cellular and Neurochemical Homeostasis in the Adult Nigrostriatal Circuit. *Neuron* 2012, 75, 306–319.
57. Gonzalez-Reyes, L.E.; Chiang, C.-C.; Zhang, M.; Johnson, J.; Arrillaga-Tamez, M.; Couturier, N.H.; Reddy, N.; Starikov, L.; Capadona, J.R.; Kottmann, A.H.; et al. Sonic Hedgehog is expressed by hilar mossy cells and regulates cellular survival and neurogenesis in the adult hippocampus. *Sci. Rep.* 2019, 9, 17402–17420.
58. Amankulor, N.M.; Hambardzumyan, D.; Pyonteck, S.M.; Becher, O.J.; Joyce, J.A.; Holland, E.C. Sonic hedgehog pathway activation is induced by acute brain injury and regulated by injury-related inflammation. *J. Neurosci.* 2009, 29, 10299–10308.
59. Alvarez, J.I.; Dodelet-Devillers, A.; Kebir, H.; Ifergan, I.; Fabre, P.J.; Terouz, S.; Sabbagh, M.; Wosik, K.; Bourbonnière, L.; Bernard, M.; et al. The Hedgehog Pathway Promotes Blood-Brain Barrier Integrity and CNS Immune Quiescence. *Science* 2011, 334, 1727–1731.
60. Sirko, S.; Behrendt, G.; Johansson, P.; Tripathi, P.; Costa, M.; Bek, S.; Heinrich, C.; Tiedt, S.; Colak, D.; Dichgans, M.; et al. Reactive Glia in the Injured Brain Acquire Stem Cell Properties in Response to Sonic Hedgehog. *Cell Stem Cell* 2013, 12, 426–439.
61. Pitter, K.; Tamagno, I.; Feng, X.; Ghosal, K.; Amankulor, N.; Holland, E.C.; Hambardzumyan, D. The SHH/Gli pathway is reactivated in reactive glia and drives proliferation in response to neurodegeneration-induced lesions. *Glia* 2014, 62, 1595–1607.
62. Cheng, F.Y.; Fleming, J.T.; Chiang, C. Bergmann glial Sonic hedgehog signaling activity is required for proper cerebellar cortical expansion and architecture. *Dev. Biol.* 2018, 440, 152–166.
63. Priego, N.; Valiente, M. The potential of astrocytes as immune modulators in brain tumors. *Front. Immunol.* 2019, 10, 1314.
64. Zhang, G.; Rich, J.N. Reprogramming the microenvironment: Tricks of tumor-derived astrocytes. *Cell Res.* 2020, 30, 633–634.
65. Gronseth, E.; Wang, L.; Harder, D.R.; Ramchandran, R. The Role of Astrocytes in Tumor Growth and Progression. In *Astrocyte-Physiology and Pathology*; IntechOpen: London, United Kingdom, 2018.
66. Liu, H.; Sun, Y.; O'Brien, J.; Franco-Barraza, J.; Qi, X.; Yuan, H.; Jin, W.; Zhang, J.; Gu, C.; Zhao, Z.; et al. Necroptotic astrocytes contribute to maintaining stemness of disseminated medulloblastoma through CCL2 secretion. *Neuro. Oncol.* 2020, 22, 625–638.
67. Cavalli, F.M.; Remke, M.; Rampasek, L.; Peacock, J.; Shih, D.J.H.; Luu, B.; Garzia, L.; Torchia, J.; Nor, C.; Morrissy, S.; et al. Intertumoral Heterogeneity within Medulloblastoma Subgroups. *Cancer Cell* 2017, 31, 737–754.e6.
68. Northcott, P.A.; Robinson, G.W.; Kratz, C.P.; Mabbott, D.J.; Pomeroy, S.L.; Clifford, S.C.; Rutkowski, S.; Ellison, D.W.; Malkin, D.; Taylor, M.; et al. Medulloblastoma. *Nat. Rev. Dis. Prim.* 2019, 5, 11.
69. Wu, V.; Yeerna, H.; Nohata, N.; Chiou, J.; Harismendy, O.; Raimondi, F.; Inoue, A.; Russell, R.B.; Tamayo, P.; Gutkind, J.S. Illuminating the Onco-GPCRome: Novel G protein-coupled receptor-driven oncocrine networks and targets for cancer immunotherapy. *J. Biol. Chem.* 2019, 294, 11062–11086.
70. Di Pietro, C.; Marazziti, D.; Lasala, G.; Abbaszadeh, Z.; Golini, E.; Matteoni, R.; Tocchini-Valentini, G.P. Primary Cilia in the Murine Cerebellum and in Mutant Models of Medulloblastoma. *Cell. Mol. Neurobiol.* 2017, 37, 145–154.
71. Marazziti, D.; Di Pietro, C.; Golini, E.; Mandillo, S.; Lasala, G.; Matteoni, R.; Tocchini-Valentini, G.P. Precocious cerebellum development and improved motor functions in mice lacking the astrocyte cilium-, patched 1-associated Gpr3711 receptor. *Proc. Natl. Acad. Sci. USA* 2013, 110, 16486–16491.
72. Han, Y.G.; Kim, H.J.; Dlugosz, A.A.; Ellison, D.W.; Gilbertson, R.J.; Alvarez-Buylla, A. Dual and opposing roles of primary cilia in medulloblastoma development. *Nat. Med.* 2009, 15, 1062–1065.
73. Han, Y.G.; Alvarez-Buylla, A. Role of primary cilia in brain development and cancer. *Curr. Opin. Neurobiol.* 2010, 20, 58–67.
74. La Sala, G.; Di Pietro, C.; Matteoni, R.; Bolasco, G.; Marazziti, D.; Tocchini-Valentini, G.P. Gpr3711/prosaposin receptor regulates Ptch1 trafficking, Shh production, and cell proliferation in cerebellar primary astrocytes. *J. Neurosci. Res.* 2021, 99, 1064–1083, Epub ahead of print.
75. Zurawel, R.H.; Allen, C.; Wechsler-Reya, R.; Scott, M.P.; Raffel, C. Evidence that haploinsufficiency of Ptch leads to medulloblastoma in mice. *Genes Chromosomes Cancer* 2000, 28, 77–81.
76. Mao, J.; Ligon, K.L.; Rakhlin, E.Y.; Thayer, S.P.; Bronson, R.T.; Rowitch, D.; McMahon, A.P. A novel somatic mouse model to survey tumorigenic potential applied to the Hedgehog pathway. *Cancer Res.* 2006, 66, 10171–10178.
77. Snuderl, M.; Batista, A.; Kirkpatrick, N.D.; de Almodovar, C.R.; Riedemann, L.; Walsh, E.C.; Anolik, R.; Huang, Y.; Martin, J.; Kamoun, W.; et al. Targeting placental growth factor/neuropilin 1 pathway inhibits growth and spread of medulloblastoma. *Cell* 2013, 152, 1065–1076.

**Keywords**

---

medulloblastoma;tumor progression;tumor microenvironment;tumor-associated astrocytes;hedgehog signaling;tumor-astrocytes cross talk

---

Retrieved from <https://encyclopedia.pub/15849>