

Acetylcholine Receptors

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Acetylcholine is a widely distributed excitatory neurotransmitter. Within the human body, it is present in both branches of the autonomic nervous system: within the parasympathetic system in pre- and postganglionic cells, and within the sympathetic system in preganglionic cells. It is also a neurotransmitter at the periphery within the neuromuscular junction.

nAChR

α-conotoxins

three-finger α-neurotoxins

1. Introduction

The receptors are characterized by radial symmetry and have pentameric organization, with five subunits arranged radially around a central ion-conducting pore. There are 17 subunits described in vertebrates: $\alpha 1$ to $\alpha 10$, $\beta 1$ to $\beta 4$, γ , δ , and ϵ . $\alpha 5$ and $\beta 3$ subunits are considered auxiliary subunits, not contributing to the ACh binding site but influencing properties of the receptor [1][2]. $\alpha 1$, $\beta 1$, δ , ϵ , and γ are subunits within muscle-type of nAChR, with ϵ subunits present in adult receptor type, while γ subunit present in fetal receptor.

The orthosteric ligand-binding site is formed at the junction between two neighboring subunits, where loops A, B, and C, from principal subunits and loops D, E, and F, coalesce and form a hydrophobic cage from aromatic residues that provides a space for ACh binding. The aromatic residues protruding to the ligand-binding site are rather conserved; within the muscle $\alpha 1$ subunit, they include Tyr 93 from loop A, Trp 149 from loop B, and Tyr 190 and Tyr 198 from loop C. These aromatic residues within the principal subunit are assisted by aromatic residues of a complementary subunit, such as Trp 57 of the γ subunit [3]. The conserved aromatic residues interact with ligands via cation-π interaction.

The receptor structure was studied at an atomic level. The quest for receptor structure determination was facilitated through studies on acetylcholine-binding protein (AChBP), a soluble multimeric protein secreted by snail glial cells which buffers ACh concentration [4].

nAChRs are widely distributed within the central nervous system (CNS) and peripheral nervous system, as well as outside the nervous system. The most prevalent receptor subtype within the brain is the $\alpha 4\beta 2$ -containing receptor, the upregulation of which is observed following chronic nicotine consumption [5]. Existing in two stoichiometric forms (($\alpha 4$)₂($\beta 2$)₃ and ($\alpha 4$)₃($\beta 2$)₂), $\alpha 4\beta 2$ -containing receptors can vary in their sensitivities to ACh [6]. They play critical role within reward pathways, in particular, within nigrostriatal dopaminergic circuitry [7].

Disfunction in nAChRs within neuromuscular junctions can lead to myasthenic syndrome. Within the CNS, abnormal functioning of the receptors can manifest itself as Alzheimer's disease, schizophrenia, depression, and Parkinson's disease [8]. Outside the CNS, improper functionality of the receptors can lead to skin disorders such as pemphigus vulgaris and cancer.

Thus, knowledge about the functionality of the receptors is of paramount importance. Ligands targeting the receptors can help decipher physiological mechanisms, as well as prevent pathological processes where receptors are involved [9][10]. Natural products and, in particular, venoms of venomous animals, such as snails, snakes, spiders, and scorpions, contain biologically active compounds targeting nAChRs; thus, they are actively and thoroughly investigated [11].

2. Snail Venoms as a Source of Toxins Acting on Acetylcholine Receptors

Predatory snails within the *Conus* genus produce peptides—conotoxins that target different ion channels [12]. The peptides with a general structure CC-Xm-C-Xn-C, where C represent cysteines, Xm and Xn—variable number of residues between cysteines—constitute the family of α -conotoxins (α -Ctx). Structurally, α -conotoxins have very rigid three-dimensional structure due to the restraints imposed by disulfide bonds.

α 3/5 conotoxins predominantly target muscle subtypes of nAChRs [13]. α -Conotoxins MI, GI, and SIA have been shown to exhibit >10,000-fold selectivity for α/δ over the α/γ site in mammalian receptors. However, the reverse pattern is observed in Torpedo receptors. Conserved Pro 6 and Tyr 12 residues are critical for hydrophobic interaction between the δ subunit and MI [14].

Ac1.1a and Ac1.1b are almost identical α 3/5 conotoxins differing in only one residue at position 14 [15] (Figure 1). They exhibit selectivity towards α/δ over α/γ site. The first three residues before the Cys 4 have been shown to lack an effect on binding the target.

α3/5 conotoxins				α4/7 conotoxins			
Acl.1a	NGRCCHPACGKHFNC*	15		AnIB	--GGCCHSHPACAAANNQDψC*	17	
Acl.1b	NGRCCHPACGKHFSC*	15		ArIA	IRDECCSNPACRVNNNPHVCRRR	22	
CIA	NGRCCHPACGKHFSC*	15		ArIB	--DECCSNPACRVNNNPHVCRRR	20	
GI	--ECCNPACGRHYSC*	13		CIB	--GCCSNPVCHLEHPNAC*	16	
MI	-GRCCHPACGKKNYSC*	14		EI	-RDOCCYHPTCNMSNPQIC*	18	
SIA	--YCCHPACGKNFDC*	13		GIC	--GCCSHPACAGNNQHIC*	16	
SII	-GCCCNPACGPNYGCGTSCS	19		GID	IRDyCCSNPACRVNNNOHVC^	19	
α4/3 conotoxins				Lola	--EGCCSNPACRTNHPEVCD--	18	
ImI	GCCSDPRCAWRC*	12		LsIA	--SGCCSNPACRVNNNPNI*	17	
ImII	ACCSDRRCRWRC*	12		LtIA	--GCCARAACAGIHQEIC*	16	
RgIA	GCCSDPRCRYRCR	13		Lt1.3	--GCCSHPACSGNNPYFC*	16	
α4/4 conotoxins				LvIA	--GCCSHPACNVVDHPEIC*	16	
BuIA	---GCCSTPPCAVLYC*	13		MII	--GCCSNPVCHLEHSNLC*	16	
EIIA	QTPGCCWNPACVKNRC*	16		Mrl.7	--PECCTHPACHVSHPELC*	17	
PIB	ZSOGCCWNPACVKNRC*	16		PeIA	--GCCSHPACSVNHPELC*	16	
α4/6 conotoxins				PnIA	--GCCSLPPCAANNPDYψC*	16	
AuIB	-GCCSYPPCFATNPDC*	15		RegIIA	--GCCSHPACNVNNNPHIC*	16	
VIIA	RDCCSNPPCAHNNPDC*	16		TxIB	--GCCSDPPCRNKHPDLC*	16	
VnIB	GCCCSHPVCYTKNPNG*	17		Vcl.1	--GCCSDPRCNYDHPEIC*	16	
TxID	-GCCSHPVCSAMSPIC*	15					

Figure 1. The α-conotoxins with 3/5, 4/4, 4/6, and 4/7 spacing. The cysteines residues are shaded in gray and intrachain disulfide bonds are shown. The number of residues in each toxin is given to the right. * C—terminal carboxamide, ^ C—terminal carboxylate, ψ—sulfated tyrosine, O—hydroxyproline, Z—pyroglutamate.

α-Conotoxin GI blocks muscle-type nAChRs, with selectivity for the α/δ site in mouse muscle receptors [16], and the α/γ site in *Torpedo californica* electric organ receptors [17]. Pro 5, Gly 8, Arg 9, and Tyr 11 are critical residues for GI to target GI mutant demonstrated a three-fold increase in potency in mouse α1β1δε and a decreased potency in rat neuronal α9α10 nAChR, compared to wildtype GI [18].

NMR structures for both CIA and CIB alpha-conotoxins derived from *Conus catus* have been studied. α-Conotoxin CIA caused muscle paralysis in fish when tested in vivo [19], while 4/7 α-Conotoxin CIB blocked neuronal type nAChRs.

Overall, there are two conserved regions (residues 2 and 3 in the first cysteine loop, as well as residues 1, 2, and 4 in the second cysteine loop) inherent to α3/5 conotoxins (Figure 1), which include important residues for the binding to muscle-type nAChR. (1) it has Pro residue in place of expected Arg/Lys; (2) it has a long C-terminus. Structure–functional studies on SII are very scarce, except for one that characterized SII as possessing three disulfide bonds and no net positive charge [20].

Asp 5, Pro 6, Arg 7, and Trp 10 are critical residues of the ImI conotoxin, conferring its specificity for the α7 nAChRs [21].

Despite the fact that Iml and ImII conotoxins share a high sequence homology (9 of 12 amino acids are identical) (Figure 1), they were found to target different sites at the α 7 nAChR, since only Iml (and not ImII) showed competitive inhibition of the receptor when α -bungarotoxin (α -Bgtx) was co-added [22]. Mutational studies revealed that the amino acid residue at position 6 (Pro in α -Ctx Iml, Arg in α -Ctx ImII) determines the selectivity for a specific site, whereas the amino acid residue in position 9 (Ala in α -Ctx Iml, Arg in α -Ctx ImII) confers each of the toxins its optimal affinity to bind to their corresponding sites at the α 7 nAChR [22].

The selectivity and potency of α -conotoxin RgIA for the α 9 α 10 subtype of nAChR is conferred by Arg 9 and Arg 13 residues [23]. It is also interesting to note that [24][6]-dicarba RgIA is able to act specifically at α 9 α 10 nAChRs, losing its effect on N-type calcium channels [25]. This analogue could potentially be used in a clinical trial for eliciting an effect on a single receptor subtype.

α -conotoxin α 3 β 2 nAChRs with a significantly lower IC₅₀ than α 4 β 2 receptors [26]. In fact, there is a 40,000-fold difference between their IC₅₀ values [26]. The five residues of the α 6 subunit, Lys 85, Asp 187, Ile 188, Thr 198, and Tyr 205, were determined to be responsible for this effect [27].

It should be noted that α -conotoxin BuIA on nAChRs with β 4 subunits have slow off-time when compared to corresponding nAChRs with β 2 subunits [26]. This implies that BuIA can discriminate between different β subunits based on the time needed to unblock the receptor.

The α -conotoxin BuIA [T5A; P6O] was found to be selective for α 6 β 4 vs. α 6 β 2 nAChRs [28]. It was mainly achieved by P6O substitution, which resulted in a 2800-fold increase of IC₅₀ at α 6/ α 3 β 2 β 3 and only a 6-fold increase at α 6/ α 3 β 4 [28].

It is interesting to note that, though α -conotoxins EIa and PIB come from different *Conus* species, they are almost identical in sequence, except for one residue at the second position. Therefore, it is not surprising that they both have selectivity for muscle-type Pro 5, Ala 6, and Lys 9 residues located in the first and second loops, respectively, which were previously found to be critical for the muscle nAChR selectivity of α 3/5 conotoxins [29].

Alanine scanning mutagenesis revealed three residues in the AuIB conotoxin that affect the inhibition of α 3 β 4 nAChRs [30]. Hence, its substitution with alanine disrupted a favorable interaction of AuIB with the receptor. It was elucidated that Pro 6 caused inhibition due to its effect on 3D structure of the peptide, whereas Phe 9 was responsible for the interaction with a two-residue binding pocket (Trp 59 and Lys 61) of the β 4 subunit [30].

Structure–activity studies on the VilA conotoxin revealed that Arg 1 and His 11 are critical residues for binding to the target effectively. (a mutant with additional Leu residue at position 16) produced a 12-fold stronger interaction with α 3 β 2 nAChR when compared to the native peptide. This was explained by hydrophobicity Leu residue conferred to the overall structure of the VilA conotoxin [31].

The VnIB conotoxin selectively inhibits However, no structure–activity studies on this conopeptide have been carried out so far. It is hypothesized that all of the members of α 4/6 conotoxins, including VnIB, exhibit β 4 subunit

selectivity due to their common 4/6 substructure. The selectivity of VnIB toward $\alpha 6$ -containing nAChRs can potentially be explained by the five residues present in the second disulfide-loop, excluding Pro residue (YTKNPN) [32].

There are five residues (His 5, Pro 6, Val 7, Met 11, and Pro 13) of TxID critical for the inhibition of $\alpha 3\beta 4$ and $\alpha 6/\alpha 3\beta 4$ nAChRs [33]. [S9A] TxID mutant was found to discriminate between the two receptor subtypes, having a 46-fold higher affinity to $\alpha 3\beta 4$ than to $\alpha 6/\alpha 3\beta 4$ nAChRs [33].

Ala 11 and Gln 14 play a role in $\alpha 3\beta 2$ selectivity of AnIB, whereas the C-terminal amidation and sulfation of Tyr 16 are responsible for $\alpha 7$ nAChR binding [34].

ArIB [V11L; V16D] is considered to be the most selective $\alpha 7$ nAChR antagonist to date (IC₅₀ = 1.09 nM) [35].

Important residues conferring EI potency to bind mouse $\alpha 1\beta 1\delta\epsilon$ nAChR were found to be His 7. In addition, deletion of Arg1–Asn2–Hyp3 residues are also crucial, since their deletion causes a total loss of activity at muscle receptors [36]. [S13A] EI mutant exhibited increased potency and selectivity for $\alpha 1\beta 1\delta\epsilon$ nAChRs [36].

GIC is a very interesting conotoxin from a research standpoint, since it exhibits extreme potency and selectivity towards $\alpha 3\beta 2$ nAChRs (IC₅₀ = 1.1 nM) [37]. His 5 and Gln 13 were found to be the most important residues that confer GIC its potency and selectivity, respectively. In addition, Ala 7, Asn 11, and Asn 12 residues of the peptide were involved in binding to the principal site of the receptor [37].

Ala 10, Val 13 and Val 18 are important residues of the GID conotoxin, conferring it selectivity for $\alpha 4\beta 2$ over $\alpha 3\beta 2$ nAChRs [38]. In particular, GID[V18N] analog exhibits the highest selectivity for $\alpha 4\beta 2$, with no inhibitory effect on the $\alpha 3\beta 2$ subtype [38].

The structure–functional study conducted to the Lo1a conotoxin found that its Asp-18 residue is critical for the selectivity of the peptide for neuronal vs. muscle subtype nAChRs [39]. This was evident from the observation that Lo1a- Δ D and Lo1a-RRR analogues, which either lacked Asp 18 or had it replaced with 3 Arg residues, respectively, acquired affinity for the adult muscle subtype $\alpha 1\beta 1\delta\epsilon$, aside from $\alpha 7$ receptors [39].

Ser-1 and Gly 2 are important residues for the affinity of LsIA to $\alpha 3\beta 2$ and $\alpha 7$ nAChRs [40]. In addition, carboxylation of the C-terminus of LsIA resulted in its selectivity for $\alpha 3\beta 2$ over $\alpha 7$ to increase 9-fold [40].

Selectivity of the Lt1.3 conotoxin for $\alpha 3\beta 2$ receptors is attributed to a small hydrophobic Gly 10 residue, whereas Asn 11, Asn 12, Pro 13, Tyr 14, and Phe 15 residues contribute to the receptor binding [41].

His 5, Pro 6, Ala 7, and His 12 of LvIA are important residues that interact with $\alpha 3$ -containing nAChRs [42]. LvIA exhibits unusual selectivity toward $\alpha 3\beta 2$ vs. $\alpha 6/\alpha 3\beta 2\beta 3$ nAChRs due to Asn 9 and Asp 11 residues [43].

Asp 5, Pro 6, and His 12 are the main residues of the MII conotoxin, which confers its potency for binding to the rat $\alpha 3\beta 2$ nAChR subtype [44]. The MII [H9A; L15A] analog of α -conotoxin MII is selective towards $\alpha 6$ -containing receptors (IC₅₀ on rat $\alpha 6/\alpha 3\beta 2\beta 3$ = 2.4 nM), and can discriminate between $\alpha 6$ and $\alpha 3$ subunits [45].

Mr1.7 [E2A] is potent and selective for the $\alpha 3\beta 2$ nAChR subtype, with no inhibitory effect on other subtypes [46]. However, the substitution of Ser 12 with Ala results in the loss of selectivity to $\alpha 3\beta 2$, with new binding ability to other subtypes ($\alpha 3\beta 4$, $\alpha 2\beta 4$, and $\alpha 7$) [46].

The PeIA conotoxin can be made >15,000-fold more potent on $\alpha 6/\alpha 3\beta 2\beta 3$ vs. $\alpha 3\beta 2$ nAChRs via substituting Asn 11 with either Lys or Arg [47]. On the contrary, PeIA-4566, which is a synthetic peptide composed of non-natural amino acids, targets $\alpha 3\beta 2$ over $\alpha 6/\alpha 3\beta 2\beta 3$, with a 300-fold difference in potency [48]. V10A; E14N] analog blocks $\alpha 6/\alpha 3\beta 2\beta 3$ and $\alpha 6/\alpha 3\beta 4$ nicotinic receptors with IC₅₀ values of 223 pM and 65 nM, respectively [49].

Residues from Leu 5 to Pro 13 and Tyr 15 are important residues of PnIA, conferring its inhibitory effect on $\alpha 7$ nAChRs [50]. In addition, the PnIA [A10L] mutant is able to exclusively bind $\alpha 7$ nAChRs [51].

The RegIIA [N11A; N12A] analog selectively blocks $\alpha 3\beta 4$ vs. $\alpha 3\beta 2$ nAChRs, with a 1000-fold difference in potency [52].

TxIB is unique due to its ability to selectively bind $\alpha 6/\alpha 3\beta 2\beta 3$ nAChRs, while lacking any capability to bind to other receptor subtypes [53]. However, structure-functional studies accounting for this have not been conducted yet.

It is generally accepted that α -Ctxs are antagonists of nAChRs, but MrIC (PECCTHPACHVSNPELC), which is a peptide variant of Mr1.7, plays a full agonist role of endogenous human $\alpha 7$ nAChRs in the presence of PNU (positive allosteric modulator of $\alpha 7$ nAChR subtype) [54]. It is suggested that the Pro residue in the N-terminus of the peptide has a role in binding $\alpha 7$ nAChR [54].

It was found that Asp 5 to Arg 7, and Asp 11 to Ile 15, are critical residues for the inhibitory effect of Vc1.1 at the $\alpha 9\alpha 10$ nAChR subtype [55]. In addition, mutations at positions 4 and 9 (substitution of Ser 4 with either Lys or Arg and substitution of Asn 9 with either Ala, Leu, or Ile) increased the potency of Vc1.1 at $\alpha 9\alpha 10$ nAChRs [55].

Due to the fact that nAChRs are implicated in a range of pathological conditions, α -conotoxins targeting nAChRs pose themselves as attractive candidates for drug development. For example, α -conotoxins RgIA and Vc1.1 demonstrate the analgesic effect in animal models of neuropathic pain [56][57][58][59][60][61][62][63]. Such analgesic effect works through targeting $\alpha 9\alpha 10$. The mechanism of analgesia mediated by RgIA and Vc1.1 conotoxins is not currently clear, but there are indications that it involves immune-mediated pathways.

RgIA was shown to be potent in reducing mechanical allodynia and oxaliplatin-induced hypersensitivity to thermal and mechanical stimuli [64], as well as accelerating the recovery of nerve damage following chronic constriction injury (CCI) [57]. Other than alleviating neuropathic pain, RgIA was also shown to be able to reduce the severity of dextran sodium sulfate-induced colitis in an animal model [65].

Vc1.1 was shown to reduce mechanical allodynia in rats with diabetic neuropathy induced by streptozotocin, and in rats with CCI [56][61]. Vc1.1 reduced mechanical hyperalgesia in rats with peripheral nerve ligation when the toxin was injected intramuscularly [62].

Another α -conotoxin with a potential therapeutic application due to its antitumor activity is TxID. The toxin inhibited the growth of lung cancer cells [66].

The results from in vitro and animal studies serve as an encouragement for drug development. There are, however, hindrances along the way [67], which stem from the peptidic nature of the toxins, the potential off-target effects, and different potencies on human receptors vs. rodent ones. For example, Vc1.1 was discontinued from phase II human clinical trials due to its low affinity on human $\alpha 9\beta 10$ nAChRs [68].

The α -conotoxins mentioned in the main text are shown, together with their targets and potencies.

References

1. Scholze, P.; Huck, S. The A5 Nicotinic Acetylcholine Receptor Subunit Differentially Modulates A4 β 2* and A3 β 4* Receptors. *Front. Synaptic Neurosci.* 2020, 12, 54.
2. Boorman, J.P.; Beato, M.; Groot-Kormelink, P.J.; Broadbent, S.D.; Sivilotti, L.G. The Effects of B3 Subunit Incorporation on the Pharmacology and Single Channel Properties of Oocyte-Expressed Human A3 β 4 Neuronal Nicotinic Receptors. *J. Biol. Chem.* 2003, 278, 44033–44040.
3. Gharpure, A.; Noviello, C.M.; Hibbs, R.E. Progress in Nicotinic Receptor Structural Biology. *Neuropharmacology* 2020, 171, 108086.
4. Karlin, A. A Touching Picture of Nicotinic Binding. *Neuron* 2004, 41, 841–842.
5. Govind, A.P.; Vezina, P.; Green, W.N. Nicotine-Induced Upregulation of Nicotinic Receptors: Underlying Mechanisms and Relevance to Nicotine Addiction. *Biochem. Pharmacol.* 2009, 78, 756–765.
6. Carbone, A.-L.; Moroni, M.; Groot-Kormelink, P.-J.; Bermudez, I. Pentameric Concatenated (A4)2(B2)3 and (A4)3(B2)2 Nicotinic Acetylcholine Receptors: Subunit Arrangement Determines Functional Expression. *Br. J. Pharmacol.* 2009, 156, 970–981.
7. McGranahan, T.M.; Patzlaff, N.E.; Grady, S.R.; Heinemann, S.F.; Booker, T.K. A4 β 2 Nicotinic Acetylcholine Receptors on Dopaminergic Neurons Mediate Nicotine Reward and Anxiety Relief. *J. Neurosci.* 2011, 31, 10891.
8. Winek, K.; Soreq, H.; Meisel, A. Regulators of Cholinergic Signaling in Disorders of the Central Nervous System. *J. Neurochem.* 2021.

9. Dineley, K.T.; Pandya, A.A.; Yakel, J.L. Nicotinic ACh Receptors as Therapeutic Targets in CNS Disorders. *Trends Pharmacol. Sci.* 2015, 36, 96–108.
10. Quik, M.; Zhang, D.; McGregor, M.; Bordia, T. Alpha7 Nicotinic Receptors as Therapeutic Targets for Parkinson's Disease. *Biochem. Pharmacol.* 2015, 97, 399–407.
11. Pennington, M.W.; Czerwinski, A.; Norton, R.S. Peptide Therapeutics from Venom: Current Status and Potential. *Bioorgan. Med. Chem.* 2018, 26, 2738–2758.
12. Jin, A.-H.; Muttenthaler, M.; Dutertre, S.; Himaya, S.W.A.; Kaas, Q.; Craik, D.J.; Lewis, R.J.; Alewood, P.F. Conotoxins: Chemistry and Biology. *Chem. Rev.* 2019, 119, 11510–11549.
13. McIntosh, J.M.; Santos, A.D.; Olivera, B.M. Conus Peptides Targeted to Specific Nicotinic Acetylcholine Receptor Subtypes. *Annu. Rev. Biochem.* 1999, 68, 59–88.
14. Jacobsen, R.B.; DelaCruz, R.G.; Grose, J.H.; McIntosh, J.M.; Yoshikami, D.; Olivera, B.M. Critical Residues Influence the Affinity and Selectivity of α -Conotoxin MI for Nicotinic Acetylcholine Receptors. *Biochemistry* 1999, 38, 13310–13315.
15. Liu, L.; Chew, G.; Hawrot, E.; Chi, C.; Wang, C. Two Potent A3/5 Conotoxins from Piscivorous *Conus Achatinus*. *Acta Biochim. Biophys. Sin.* 2007, 39, 438–444.
16. Groebe, D.R.; Dumm, J.M.; Levitan, E.S.; Abramson, S.N. Alpha-Conotoxins Selectively Inhibit One of the Two Acetylcholine Binding Sites of Nicotinic Receptors. *Mol. Pharmacol.* 1995, 48, 105.
17. Groebe, D.R.; Gray, W.R.; Abramson, S.N. Determinants Involved in the Affinity of α -Conotoxins GI and SI for the Muscle Subtype of Nicotinic Acetylcholine Receptors. *Biochemistry* 1997, 36, 6469–6474.
18. Ning, J.; Li, R.; Ren, J.; Zhangsun, D.; Zhu, X.; Wu, Y.; Luo, S. Alanine-Scanning Mutagenesis of α -Conotoxin GI Reveals the Residues Crucial for Activity at the Muscle Acetylcholine Receptor. *Mar. Drugs* 2018, 16, 507.
19. Giribaldi, J.; Wilson, D.; Nicke, A.; El Hamdaoui, Y.; Laconde, G.; Faucherre, A.; Moha Ou Maati, H.; Daly, N.L.; Enjalbal, C.; Dutertre, S. Synthesis, Structure and Biological Activity of CIA and CIB, Two α -Conotoxins from the Predation-Evoked Venom of *Conus Catus*. *Toxins* 2018, 10, 222.
20. Ramilo, C.A.; Zafaralla, G.C.; Nadasdi, L.; Hammerland, L.G.; Yoshikami, D.; Gray, W.R.; Kristipati, R.; Ramachandran, J.; Miljanich, G. Novel .Alpha.- and .Omega.-Conotoxins and *Conus Striatus* Venom. *Biochemistry* 1992, 31, 9919–9926.
21. Quiram, P.A.; Sine, S.M. Structural Elements in α -Conotoxin Iml Essential for Binding to Neuronal A7 Receptors. *J. Biol. Chem.* 1998, 273, 11007–11011.
22. Ellison, M.; McIntosh, J.M.; Olivera, B.M. α -Conotoxins Iml and ImII: Similar A7 nicotinic receptor antagonists act at different sites. *J. Biol. Chem.* 2003, 278, 757–764.

23. Ellison, M.; Feng, Z.-P.; Park, A.J.; Zhang, X.; Olivera, B.M.; McIntosh, J.M.; Norton, R.S. α -RgIA, a Novel Conotoxin That Blocks the A9 α 10 NACHR: Structure and Identification of Key Receptor-Binding Residues. *J. Mol. Biol.* 2008, 377, 1216–1227.

24. Millar, N.S.; Gotti, C. Diversity of Vertebrate Nicotinic Acetylcholine Receptors. *Neuropharmacology* 2009, 56, 237–246.

25. Chhabra, S.; Belgi, A.; Bartels, P.; van Lierop, B.J.; Robinson, S.D.; Kompella, S.N.; Hung, A.; Callaghan, B.P.; Adams, D.J.; Robinson, A.J.; et al. Dicarba Analogues of α -Conotoxin RgIA. Structure, Stability, and Activity at Potential Pain Targets. *J. Med. Chem.* 2014, 57, 9933–9944.

26. Azam, L.; Dowell, C.; Watkins, M.; Stitzel, J.A.; Olivera, B.M.; McIntosh, J.M. α -Conotoxin BuIA, a Novel Peptide from Conus Bullatus, Distinguishes among Neuronal Nicotinic Acetylcholine Receptors. *J. Biol. Chem.* 2005, 280, 80–87.

27. Kim, H.-W.; McIntosh, J.M. A6 NACHR Subunit Residues That Confer α -Conotoxin BuIA Selectivity. *FASEB J.* 2012, 26, 4102–4110.

28. Azam, L.; Maskos, U.; Changeux, J.-P.; Dowell, C.D.; Christensen, S.; De Biasi, M.; McIntosh, J.M. α -Conotoxin BuIA[T5A;P6O]: A Novel Ligand That Discriminates between A6 β 4 and A6 β 2 Nicotinic Acetylcholine Receptors and Blocks Nicotine-Stimulated Norepinephrine Release. *FASEB J.* 2010, 24, 5113–5123.

29. Quinton, L.; Servent, D.; Girard, E.; Molgó, J.; Le Caer, J.-P.; Malosse, C.; Haidar, E.A.; Lecoq, A.; Gilles, N.; Chamot-Rooke, J. Identification and Functional Characterization of a Novel α -Conotoxin (EIIA) from Conus Ermineus. *Anal. Bioanal. Chem.* 2013, 405, 5341–5351.

30. Grishin, A.A.; Cuny, H.; Hung, A.; Clark, R.J.; Brust, A.; Akondi, K.; Alewood, P.F.; Craik, D.J.; Adams, D.J. Identifying Key Amino Acid Residues That Affect α -Conotoxin AulB Inhibition of A3 β 4 Nicotinic Acetylcholine Receptors. *J. Biol. Chem.* 2013, 288, 34428–34442.

31. Li, L.; Liu, N.; Ding, R.; Wang, S.; Liu, Z.; Li, H.; Zheng, X.; Dai, Q. A Novel 4/6-Type Alpha-Conotoxin VlIA Selectively Inhibits NACHR A3 β 2 Subtype. *Acta Biochim. Biophys. Sin.* 2015, 47, 1023–1028.

32. van Hout, M.; Valdes, A.; Christensen, S.B.; Tran, P.T.; Watkins, M.; Gajewiak, J.; Jensen, A.A.; Olivera, B.M.; McIntosh, J.M. α -Conotoxin VnIB from Conus Ventricosus Is a Potent and Selective Antagonist of A6 β 4* Nicotinic Acetylcholine Receptors. *Neuropharmacology* 2019, 157, 107691.

33. Wu, Y.; Zhangsun, D.; Zhu, X.; Kaas, Q.; Zhangsun, M.; Harvey, P.J.; Craik, D.J.; McIntosh, J.M.; Luo, S. α -Conotoxin [S9A]TxID Potently Discriminates between A3 β 4 and A6/A3 β 4 Nicotinic Acetylcholine Receptors. *J. Med. Chem.* 2017, 60, 5826–5833.

34. Loughnan, M.L.; Nicke, A.; Jones, A.; Adams, D.J.; Alewood, P.F.; Lewis, R.J. Chemical and Functional Identification and Characterization of Novel Sulfated α -Conotoxins from the Cone Snail *Conus Anemone*. *J. Med. Chem.* 2004, 47, 1234–1241.

35. Whiteaker, P.; Christensen, S.; Yoshikami, D.; Dowell, C.; Watkins, M.; Gulyas, J.; Rivier, J.; Olivera, B.M.; McIntosh, J.M. Discovery, Synthesis, and Structure Activity of a Highly Selective A7 Nicotinic Acetylcholine Receptor Antagonist. *Biochemistry* 2007, 46, 6628–6638.

36. Ning, J.; Ren, J.; Xiong, Y.; Wu, Y.; Zhangsun, M.; Zhangsun, D.; Zhu, X.; Luo, S. Identification of Crucial Residues in α -Conotoxin EI Inhibiting Muscle Nicotinic Acetylcholine Receptor. *Toxins* 2019, 11, 603.

37. Lin, B.; Xu, M.; Zhu, X.; Wu, Y.; Liu, X.; Zhangsun, D.; Hu, Y.; Xiang, S.-H.; Kasheverov, I.E.; Tsetlin, V.I.; et al. From Crystal Structure of α -Conotoxin GIC in Complex with Ac-AChBP to Molecular Determinants of Its High Selectivity for A3 β 2 NACHR. *Sci. Rep.* 2016, 6, 22349.

38. Banerjee, J.; Yongye, A.B.; Chang, Y.-P.; Gyanda, R.; Medina-Franco, J.L.; Armishaw, C.J. Design and Synthesis of α -Conotoxin GID Analogue as Selective A4 β 2 Nicotinic Acetylcholine Receptor Antagonists. *Biopolymers* 2014, 102, 78–87.

39. Lebbe, E.K.M.; Peigneur, S.; Maiti, M.; Devi, P.; Ravichandran, S.; Lescrinier, E.; Ulens, C.; Waelkens, E.; D’Souza, L.; Herdewijn, P.; et al. Structure-Function Elucidation of a New α -Conotoxin, Lo1a, from Conus Longurionis. *J. Biol. Chem.* 2014, 289, 9573–9583.

40. Inserra, M.C.; Kompella, S.N.; Vetter, I.; Brust, A.; Daly, N.L.; Cuny, H.; Craik, D.J.; Alewood, P.F.; Adams, D.J.; Lewis, R.J. Isolation and Characterization of α -Conotoxin LsIA with Potent Activity at Nicotinic Acetylcholine Receptors. *Biochem. Pharmacol.* 2013, 86, 791–799.

41. Chen, J.; Liang, L.; Ning, H.; Cai, F.; Liu, Z.; Zhang, L.; Zhou, L.; Dai, Q. Cloning, Synthesis and Functional Characterization of a Novel α -Conotoxin Lt1.3. *Mar. Drugs* 2018, 16, 112.

42. Luo, S.; Zhangsun, D.; Schroeder, C.I.; Zhu, X.; Hu, Y.; Wu, Y.; Weltzin, M.M.; Eberhard, S.; Kaas, Q.; Craik, D.J.; et al. A Novel A4/7-Conotoxin LvIA from Conus Lividus That Selectively Blocks A3 β 2 vs. A6/A3 β 2 β 3 Nicotinic Acetylcholine Receptors. *FASEB J.* 2014, 28, 1842–1853.

43. Xu, M.; Zhu, X.; Yu, J.; Yu, J.; Luo, S.; Wang, X. The Crystal Structure of Ac-AChBP in Complex with α -Conotoxin LvIA Reveals the Mechanism of Its Selectivity towards Different NACHR Subtypes. *Protein Cell* 2017, 8, 675–685.

44. Everhart, D.; Cartier, G.E.; Malhotra, A.; Gomes, A.V.; McIntosh, J.M.; Luetje, C.W. Determinants of Potency on Alpha-Conotoxin MII, a Peptide Antagonist of Neuronal Nicotinic Receptors. *Biochemistry* 2004, 43, 2732–2737.

45. McIntosh, J.M.; Azam, L.; Staheli, S.; Dowell, C.; Lindstrom, J.M.; Kuryatov, A.; Garrett, J.E.; Marks, M.J.; Whiteaker, P. Analogs of Alpha-Conotoxin MII Are Selective for Alpha6-Containing Nicotinic Acetylcholine Receptors. *Mol. Pharmacol.* 2004, 65, 944–952.

46. Wang, S.; Zhao, C.; Liu, Z.; Wang, X.; Liu, N.; Du, W.; Dai, Q. Structural and Functional Characterization of a Novel α -Conotoxin Mr1.7 from Conus Marmoreus Targeting Neuronal NACHR A3 β 2, A9 α 10 and A6/A3 β 2 β 3 Subtypes. *Mar. Drugs* 2015, 13, 3259–3275.

47. Hone, A.J.; Ruiz, M.; Scadden, M.; Christensen, S.; Gajewiak, J.; Azam, L.; McIntosh, J.M. Positional Scanning Mutagenesis of α -Conotoxin PeIA Identifies Critical Residues That Confer Potency and Selectivity for A6/A3 β 2 β 3 and A3 β 2 Nicotinic Acetylcholine Receptors. *J. Biol. Chem.* 2013, 288, 25428–25439.

48. Hone, A.J.; Fisher, F.; Christensen, S.; Gajewiak, J.; Larkin, D.; Whiteaker, P.; McIntosh, J.M. PeIA-5466: A Novel Peptide Antagonist Containing Non-Natural Amino Acids That Selectively Targets A3 β 2 Nicotinic Acetylcholine Receptors. *J. Med. Chem.* 2019, 62, 6262–6275.

49. Hone, A.J.; Scadden, M.; Gajewiak, J.; Christensen, S.; Lindstrom, J.; McIntosh, J.M. α -Conotoxin PeIA[S9H,V10A,E14N] Potently and Selectively Blocks A6 β 2 β 3 versus A6 β 4 Nicotinic Acetylcholine Receptors. *Mol. Pharmacol.* 2012, 82, 972–982.

50. Luo, S.; Nguyen, T.A.; Cartier, G.E.; Olivera, B.M.; Yoshikami, D.; McIntosh, J.M. Single-Residue Alteration in α -Conotoxin PnIA Switches Its NACHR Subtype Selectivity. *Biochemistry* 1999, 38, 14542–14548.

51. Hogg, R.C.; Hopping, G.; Alewood, P.F.; Adams, D.J.; Bertrand, D. Alpha-Conotoxins PnIA and [A10L]PnIA Stabilize Different States of the Alpha7-L247T Nicotinic Acetylcholine Receptor. *J. Biol. Chem.* 2003, 278, 26908–26914.

52. Kompella, S.N.; Hung, A.; Clark, R.J.; Marí, F.; Adams, D.J. Alanine Scan of α -Conotoxin RegIIA Reveals a Selective A3 β 4 Nicotinic Acetylcholine Receptor Antagonist. *J. Biol. Chem.* 2015, 290, 1039–1048.

53. Luo, S.; Zhangsun, D.; Zhu, X.; Wu, Y.; Hu, Y.; Christensen, S.; Harvey, P.J.; Akcan, M.; Craik, D.J.; McIntosh, J.M. Characterization of a Novel α -Conotoxin TxID from Conus Textile That Potently Blocks Rat A3 β 4 Nicotinic Acetylcholine Receptors. *J. Med. Chem.* 2013, 56, 9655–9663.

54. Jin, A.-H.; Vetter, I.; Dutertre, S.; Abraham, N.; Emidio, N.B.; Inserra, M.; Murali, S.S.; Christie, M.J.; Alewood, P.F.; Lewis, R.J. MrIC, a Novel α -Conotoxin Agonist in the Presence of PNU at Endogenous A7 Nicotinic Acetylcholine Receptors. *Biochemistry* 2014, 53, 1–3.

55. Halai, R.; Clark, R.J.; Nevin, S.T.; Jensen, J.E.; Adams, D.J.; Craik, D.J. Scanning Mutagenesis of Alpha-Conotoxin Vc1.1 Reveals Residues Crucial for Activity at the Alpha9alpha10 Nicotinic Acetylcholine Receptor. *J. Biol. Chem.* 2009, 284, 20275–20284.

56. McIntosh, J.M.; Absalom, N.; Chebib, M.; Elgoyhen, A.B.; Vincler, M. Alpha9 Nicotinic Acetylcholine Receptors and the Treatment of Pain. *Biochem. Pharmacol.* 2009, 78, 693–702.

57. Di Cesare Mannelli, L.; Cinci, L.; Micheli, L.; Zanardelli, M.; Pacini, A.; McIntosh, J.M.; Ghelardini, C. α -Conotoxin RgIA Protects against the Development of Nerve Injury-Induced Chronic Pain and Prevents Both Neuronal and Glial Derangement. *Pain* 2014, 155, 1986–1995.

58. Hone, A.J.; Servent, D.; McIntosh, J.M. A9-Containing Nicotinic Acetylcholine Receptors and the Modulation of Pain. *Br. J. Pharmacol.* 2018, 175, 1915–1927.

59. Mohammadi, S.; Christie, M.J. A9-Nicotinic Acetylcholine Receptors Contribute to the Maintenance of Chronic Mechanical Hyperalgesia, but Not Thermal or Mechanical Allodynia. *Mol. Pain* 2014, 10, 1744–8069.

60. Mohammadi, S.A.; Christie, M.J. Conotoxin Interactions with A9 α 10-NAChRs: Is the A9 α 10-Nicotinic Acetylcholine Receptor an Important Therapeutic Target for Pain Management? *Toxins* 2015, 7, 3916–3932.

61. Vincler, M.; McIntosh, J.M. Targeting the A9 α 10 Nicotinic Acetylcholine Receptor to Treat Severe Pain. *Expert Opin. Ther. Targets* 2007, 11, 891–897.

62. Satkunanathan, N.; Livett, B.; Gayler, K.; Sandall, D.; Down, J.; Khalil, Z. Alpha-Conotoxin Vc1.1 Alleviates Neuropathic Pain and Accelerates Functional Recovery of Injured Neurones. *Brain Res.* 2005, 1059, 149–158.

63. Romero, H.K.; Christensen, S.B.; Di Cesare Mannelli, L.; Gajewiak, J.; Ramachandra, R.; Elmslie, K.S.; Vetter, D.E.; Ghelardini, C.; Iadonato, S.P.; Mercado, J.L.; et al. Inhibition of A9 α 10 Nicotinic Acetylcholine Receptors Prevents Chemotherapy-Induced Neuropathic Pain. *Proc. Natl. Acad. Sci. USA* 2017, 114, E1825–E1832.

64. Christensen, S.B.; Hone, A.J.; Roux, I.; Kniazeff, J.; Pin, J.-P.; Upert, G.; Servent, D.; Glowatzki, E.; McIntosh, J.M. RgIA4 Potently Blocks Mouse A9 α 10 NAChRs and Provides Long Lasting Protection against Oxaliplatin-Induced Cold Allodynia. *Front. Cell. Neurosci.* 2017, 11, 219.

65. AlSharari, S.D.; Toma, W.; Mahmood, H.M.; Michael McIntosh, J.; Imad Damaj, M. The A9 α 10 Nicotinic Acetylcholine Receptors Antagonist α -Conotoxin RgIA Reverses Colitis Signs in Murine Dextran Sodium Sulfate Model. *Eur. J. Pharmacol.* 2020, 883, 173320.

66. Qian, J.; Liu, Y.-Q.; Sun, Z.-H.; Zhangsun, D.-T.; Luo, S.-L. Identification of Nicotinic Acetylcholine Receptor Subunits in Different Lung Cancer Cell Lines and the Inhibitory Effect of Alpha-Conotoxin TxID on Lung Cancer Cell Growth. *Eur. J. Pharmacol.* 2019, 865, 172674.

67. Bertrand, D.; Terry, A.V.J. The Wonderland of Neuronal Nicotinic Acetylcholine Receptors. *Biochem. Pharmacol.* 2018, 151, 214–225.

68. Del Bufalo, A.; Cesario, A.; Salinaro, G.; Fini, M.; Russo, P. Alpha9 Alpha10 Nicotinic Acetylcholine Receptors as Target for the Treatment of Chronic Pain. *Curr. Pharm. Des.* 2014, 20, 6042–6047.

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