

Omega-6 Fatty Acids

Subjects: Agriculture, Dairy & Animal Science

Contributor: Kelsey Harvey

Global beef production must increase in the next decades to meet the demands of a growing population, while promoting sustainable use of limited natural resources. Supplementing beef cattle with omega-6 fatty acids (FAs) is a nutritional approach shown to enhance production efficiency, with research conducted across different environments and sectors of the beef industry. Omega-6 FA from natural feed ingredients such as soybean oil are highly susceptible to ruminal biohydrogenation. Hence, our and other research groups have used soybean oil in the form of Ca soaps (CSSO) to lessen ruminal biohydrogenation, and maximize delivery of omega-6 FA to the duodenum for absorption. In cow–calf systems, omega-6 FA supplementation to beef cows improved pregnancy success by promoting the establishment of early pregnancy. Cows receiving omega-6 FA during late gestation gave birth to calves that were healthier and more efficient in the feedlot, suggesting the potential role of omega-6 FA on developmental programming. Supplementing omega-6 FA to young cattle also elicited programming effects toward improved adipogenesis and carcass quality, and improved calf immunocompetence upon a stress stimulus. Cattle supplemented with omega-6 FA during growing or finishing periods also experienced improved performance and carcass quality. All these research results were generated using cattle of different genetic composition (*Bos taurus* and *B. indicus* influenced), and in different environments (tropical, subtropical, and temperate region). Hence, supplementing omega-6 FA via CSSO is a sustainable approach to enhance the production efficiency of beef industries across different areas of the world.

Keywords: beef cattle ; growth ; health

1. Introduction

The United Nations estimates that beef production will need to increase by 120% in the next decades to feed a growing world population, ^[1]. The resources for beef production will become even more limited as the planet population increases and urban areas expand. Hence, management systems that promote sustainable beef production are warranted to meet production demands while fostering ecological stewardship and judicious use of limited natural resources.

Beef cattle operations across the world typically rely on forage as the primary nutrient source, which represents nearly 81% of the feed supplied to cattle during their productive lives ^[2]. The seasonal nature of forage production leads to variation in quantity and quality of forage, requiring supplementation strategies designed to correct nutrient deficiencies ^[3]. Fat supplementation has been extensively investigated in beef production systems, particularly as a means to provide energy to cattle ^{[4][5]}. However, supplemental fats can have nutraceutical benefits to cattle beyond their energy contribution ^{[6][7][8][9]}, particularly omega-6 fatty acids (FA) such as linoleic acid ^{[5][7]}. Research from our and other groups supplemented cattle with omega-6 FA using soybean oil as a source of linoleic acid in the form of Ca soaps to minimize ruminal biohydrogenation, and maximize delivery of omega-6 FA to the duodenum. Divalent cations such as Ca react with FA to form insoluble soaps that cannot be dissociated nor modified by the ruminal microbes. In turn, Ca soaps of FA are dissociated when exposed to the low pH of the abomasum, releasing the FA for duodenal absorption ^[10]. Therefore, the purpose of this review is to compile recent research on omega-6 FA supplementation via CSSO to beef cattle, and its potential to serve as a sustainable alternative to improve beef production efficiency.

2. Supplemental Omega-6 FA and Female Reproduction

Cow–calf systems are the foundation for global beef industries by determining the number of cattle available for harvest. Reproductive failure is a key factor limiting productivity in cow–calf operations, and pregnancy loss has been recognized as one of the main reproductive challenges in cattle ^[11]. Although ≥90% of fertile beef females effectively conceive after a single service, nearly 50% remain pregnant 30 days after service and even less females give birth to a live calf ^[12]. Management interventions to minimize pregnancy loss and promote embryonic survival are thus warranted, including supplementation with omega-6 FA. Linoleic acid and its omega-6 derivatives, however, serve as a precursor for prostaglandin (PG) F_{2α} synthesis ^[13], which triggers luteolysis and has embryotoxic effects during early gestation ^[14]. For

this reason, omega-6 FA supplementation was initially perceived as detrimental to the reproductive performance of beef cows [5].

Differing from this latter concept, our research group reported that supplementing omega-6 FA via CSSO to beef cows after artificial insemination (AI) increased pregnancy rates by 25% [15][16]. Across a series of trials, grazing *Bos indicus* beef cows supplemented with 100 g/day of CSSO for 28 days beginning after AI had greater pregnancy rates compared with cows supplemented with 100 g/day of Ca soaps of palm oil (iso-caloric and iso-lipidic control rich in palmitic acid) or unsupplemented (CON) cows (Figure 1). These results provide evidence that omega-6 FA supplementation improved, and did not impair [5], the reproductive performance of beef females. Moreover, increased pregnancy rates resulting from omega-6 FA were associated with pregnancy establishment because CSSO was offered during the early embryonic period [17], and independent of their contribution to energy intake as CSPALM resulted in similar pregnancy rates to CON.

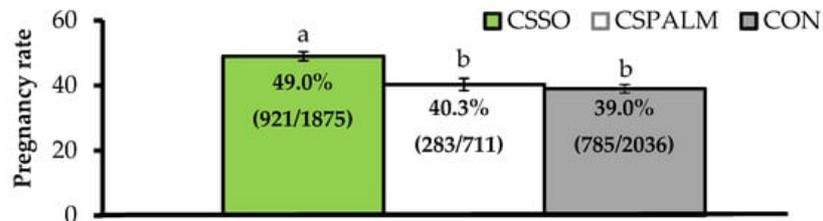


Figure 1. Pregnancy rates (as %) after artificial insemination in beef cows receiving Ca soaps of soybean oil (CSSO), Ca soaps of palm oil (CSPALM), or CON (unsupplemented) for 28 days after AI [15][16]. All values reported are least square means \pm standard error (represented as error bars). Means with different superscripts differ ($p \leq 0.05$). Information within parentheses indicate number of pregnant cows/total cows inseminated.

To provide biological support of the findings from Lopes et al. [15][16], Cooke et al. [18] investigated FA incorporation into reproductive tissues and physiological responses associated with pregnancy establishment. Grazing *B. indicus* beef cows were supplemented or not (CON) with 100 g/day of CSSO and slaughtered 19 days after AI. Cows receiving CSSO had greater incorporation of linoleic acid and its omega-6 derivatives into plasma, endometrium, corpus luteum, and conceptus. More specifically, CSSO supplementation increased intake and intestinal absorption of linoleic acid, which in turn was incorporated, elongated, desaturated, and accumulated into reproductive tissues, including as arachidonic acid in the conceptus (Table 1). These authors also evaluated factors associated with embryonic development and early pregnancy establishment on day 19 of gestation. These included conceptus size, mRNA expression of genes associated with pregnancy development in endometrial and luteal samples, and mRNA expression of interferon-tau (IFN- τ) by the conceptus; the conceptus-derived signal for maternal recognitions of pregnancy [17]. The increase in omega-6 FA accumulation, however, did not impact any of these variables, despite a tendency for increased IFN- τ concentration in uterine flushes collected from CSSO cows (10.9 vs. 7.3 ng/mL). Cows were slaughtered 19 days after AI to recover elongated conceptuses that still expressed IFN- τ mRNA [19] and provided enough tissue for both FA and mRNA expression analyses. The physiological processes responsible for pregnancy signaling to maternal tissues occur near days 15 to 17 of gestation [17]. Hence, Cooke et al. [18] evaluated maternal tissues and conceptuses after the critical period for pregnancy recognition, which prevented proper assessment of how omega-6 FA impacted expression of genes that mediate pregnancy establishment.

Table 1. Concentrations of fatty acids (FA) in samples collected on day 19 of gestation from *B. indicus* beef cows receiving or not (CON; n = 9) Ca soaps of soybean oil (CSSO; n = 9) after artificial insemination. Values reported are least square means \pm standard error. Adapted from Cooke et al. [18].

Item	CON	CSSO	p =
Plasma (mg of FA/g of plasma)			
Linoleic acid	0.275 \pm 0.022	0.540 \pm 0.021	<0.01
Arachidonic	0.023 \pm 0.005	0.025 \pm 0.004	0.74
Total omega-6 FA	0.296 \pm 0.023	0.565 \pm 0.022	<0.01
Endometrium (mg of FA/g of tissue)			
Linoleic acid	0.144 \pm 0.043	0.358 \pm 0.044	<0.01
Arachidonic	0.241 \pm 0.061	0.266 \pm 0.061	0.77

Item	CON	CSSO	p =
Total omega-6 FA	0.549 ± 0.136	0.938 ± 0.136	0.05
Corpus luteum (mg of FA/g of tissue)			
Linoleic acid	3.935 ± 0.543	7.035 ± 0.543	<0.01
Arachidonic	4.731 ± 0.349	4.942 ± 0.350	0.67
Total omega-6 FA	12.72 ± 1.19	17.78 ± 1.19	<0.01
Conceptus (mg of FA/g of tissue)			
Linoleic acid	0.022 ± 0.059	0.174 ± 0.062	0.08
Arachidonic	0.086 ± 0.043	0.312 ± 0.043	<0.01
Total omega-6 FA	0.384 ± 0.753	2.045 ± 0.755	0.13

To complement the results from Cooke et al. [18], Cipriano et al. [20] focused on conceptus- and endometrial-derived responses that mediate pregnancy signaling to maternal tissues on day 15 of gestation. Grazing *B. indicus* cows were supplemented or not (CON) with 100 g/day of CSSO beginning after AI. A subset of these cows were assigned to conceptus collection via transcervical flushing with saline followed by endometrial biopsy in the uterine horn ipsilateral to the corpus luteum 15 days after AI. The remaining cows were sampled for whole blood RNA extraction 20 days after AI, and pregnancy status was verified 28 days after AI. Supplementing omega-6 FA via CSSO increased conceptus length (2.58 vs. 1.15 cm) and mRNA expression of prostaglandin E synthase and IFN- τ by the conceptus, as well as mRNA expression of interferon-stimulated genes (ISG) in the whole blood (Figure 2). These results suggest that omega-6 FA supplementation enhanced conceptus development and IFN- τ synthesis during the pregnancy recognition period [17], corroborating the increased pregnancy rates to AI when CSSO was supplemented during early gestation [15][16]. The mRNA expression of ISGs have been used to gauge IFN- τ production and conceptus development from days 15 to 22 of gestation [21], given that IFN- τ synthesis upregulates mRNA expression of ISGs in circulating blood leukocytes [22]. Increased conceptus length and IFN- τ mRNA expression from supplemental omega-6 FA was associated with accumulation of arachidonic acid [18] and upregulation of prostaglandin E synthase mRNA in the conceptus. This enzyme converts PGH₂ to PGE₂ [23], which coordinates with IFN- τ endometrial functions that are critical for conceptus development and pregnancy signaling to maternal tissues [24]. In turn, CSSO supplementation did not impact the endometrial mRNA expression of *prostaglandin E synthase* and *cyclooxygenase-2* (Figure 2), suggesting that the effects of omega-6 FA on PG-related responses on day 15 of gestation may be specific to the conceptus due to heightened accumulation of arachidonic acid in this tissue and not in the endometrium [18].

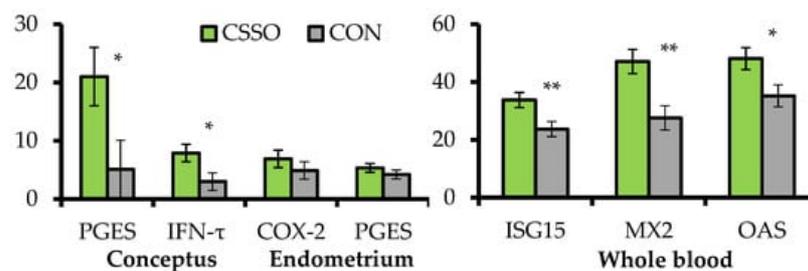


Figure 2. Expression of mRNA (relative fold change) in genes associated with pregnancy establishment in conceptus and endometrial samples collected on day 15 of gestation, and whole blood collected on day 20 of gestation from *B. indicus* cows receiving or not (CON; n = 10) Ca soaps of soybean oil (CSSO; n = 10) after artificial insemination. PGES = *prostaglandin E synthase*; IFN- τ = *interferon-tau*; COX-2 = *cyclooxygenase-2*; ISG15 = *interferon-stimulated gene 15*; MX2 = *myxovirus resistance 2*; OAS = *20,50-oligoadenylate synthetase*. All values reported are least square means \pm standard error (represented as error bars). Within variable, ** $p < 0.01$ and * $p \leq 0.05$. Adapted from Cipriano et al. [20].

Our initial efforts in characterizing the benefits of omega-6 FA to cattle reproduction were conducted with *B. indicus* cows reared in tropical conditions [15][16][18][20]. Pregnancy establishment and overall reproductive physiology differ between *B. indicus* and *B. taurus* females [25], and FA composition differs between tropical and temperate feed ingredients. Hence, Brandão et al. [26] conducted two trials evaluating omega-6 FA supplementation via CSSO to *B. taurus* cows in temperate conditions. In the first trial, grazing Angus cows were supplemented with 100 g/day of CSSO or prilled saturated fat (isocaloric and iso-lipidic control; CON+) for 21 days after AI. Similar to the findings from Lopes et al. [15][16], pregnancy rates following AI were increased by 17% in cows supplemented with omega-6 FA (Table 2). The companion trial focused on

conceptus- and endometrial-derived responses that mediate pregnancy signaling to maternal tissues with a design similar to Cipriano et al. [20], using Angus × Hereford cows that received 100 g/day of CSSO or CON+ beginning after AI. Supplementing omega-6 FA upregulated mRNA expression of IFN- τ by the conceptus and ISG in the whole blood, but did not increase conceptus length (11.3 vs. 11.4 cm for CSSO and CON, respectively) and mRNA expression of *prostaglandin E synthase*. Conceptus length across treatments was 11.4 ± 1.9 cm in Brandão et al. [26] and 2.4 ± 0.5 cm in Cipriano et al. [20], suggesting that *B. taurus* conceptus may be at an advanced stage of elongation on day 15 of gestation compared with *B. indicus* conceptus, and past the stage in which omega-6 FA impacts conceptus growth and expression of *prostaglandin E synthase*. Nevertheless, results from Brandão et al. [26] confirmed that omega-6 FA supplementation via CSSO to *B. taurus* cows also upregulated IFN- τ synthesis by the conceptus during the pregnancy recognition period, leading to increased pregnancy rates following fixed-time AI.

Table 2. Pregnancy rates and expression of mRNA (relative fold change) of genes associated with pregnancy establishment in *B. taurus* cows receiving Ca soaps of soybean oil (CSSO) or prilled saturated fat (CON+) after artificial insemination. Values reported are least square means ± standard error. Adapted from Brandão et al. [26] 1.

Item	CON+	CSSO	p =
Pregnancy rates to AI (n = 11/treatment), %	51.7 ± 4.1	60.2 ± 4.2	0.01
Physiological responses (n = 9/treatment)			
Endometrium, mRNA expression			
<i>Cyclooxygenase-2</i>	5.11 ± 1.32	4.88 ± 1.33	0.89
<i>Prostaglandin E synthase</i>	7.40 ± 1.05	5.76 ± 1.15	0.30
Conceptus, mRNA expression			
<i>Interferon-tau</i>	12.1 ± 3.6	21.3 ± 3.4	0.05
<i>Prostaglandin E synthase</i>	2.50 ± 0.49	2.22 ± 0.48	0.69
Whole blood, mRNA expression			
<i>Interferon-stimulated gene 15</i>	29.8 ± 4.9	43.1 ± 4.3	0.04
<i>Myxovirus resistance 2</i>	20.1 ± 2.8	20.2 ± 2.5	0.98
<i>20,50-oligoadenylate synthetase</i>	18.3 ± 2.9	26.8 ± 2.6	0.03

Collectively, supplementing omega-6 FA via CSSO increased incorporation of these FA into maternal and embryonic tissues and promoted IFN- τ synthesis by the conceptus during the maternal pregnancy recognition period, leading to increased pregnancy success in beef cows. These outcomes were generated across several research trials using nearly 6000 beef cows from different subspecies and managed in different environments, and were independent of the energy contribution of omega-6 FA given that iso-caloric and iso-lipidic control supplements were included. Hence, omega-6 FA supplementation is a nutritional alternative to enhance the reproductive efficiency of *B. taurus* and *B. indicus* beef cows reared in temperate and tropical environments.

3. Supplemental Omega-6 FA and Developmental Programming

The embryonic, fetal, and neonatal periods are the stages of life in which most developmental processes occur [27]. Nutrient supply during these periods exerts long-term consequences on the growth, development, and metabolic functioning of the offspring [28], leading to the concept of developmental programming [29]. Fetal developmental is sensitive to maternal nutrient status from oocyte maturation to parturition [30][31], and developmental plasticity remains susceptible to environmental stimuli during early postnatal life [32]. Dietary FAs provide a specific opportunity to nutritionally modulate developmental programming, as they differentially regulate expression of genes across metabolic tissues. For example, omega-3 FA limits adipose tissue accumulation by suppressing adipocyte differentiation [33][34], whereas omega-6 FA has been described as proadipogenic [35][36]. The fetal stage is critical for skeletal muscle and intramuscular adipocyte development [37][38]; hence, omega-6 FA supplementation during gestation can potentially enhance adipogenesis and thereby sites for marbling formation later in life [39].

4. Supplemental Omega-6 FA to Growing and Finishing Cattle

Weaning and feedlot receiving are two of the most stressful events in the beef production cycle, when cattle are exposed to a variety of physiological and physical stressors, including road transport, exposure to novel diets and environments, and comingling with new animals [40]. The combination of all of the stressors stimulates neuroendocrine and inflammatory reactions that directly impair cattle immunocompetence and productivity, leading to BRD incidence and reduced performance upon feedlot arrival [41]. Hence, strategies to increase the immunocompetence of cattle during the initial phases of the feedlot are warranted, including the use of omega-6 FA based on its immunomodulatory properties [42]. Research from our group demonstrated that omega-6 FA supplementation via CSSO to cattle upon feedlot arrival decreased plasma concentrations of inflammatory markers, but reduced feed intake and subsequent cattle ADG [8]. For this reason, our group evaluated omega-6 FA supplementation prior to feedlot arrival, by supplementing CSSO during a post-weaning preconditioning program [9]. Steers supplemented with omega-6 FA via CSSO during preconditioning had a greater feedlot-received ADG, which was attributed to reduced plasma concentrations of proinflammatory cytokines (Table 3). Moreover, CSSO steers had improved carcass marbling upon slaughter, which was associated with greater ADG upon feedlot arrival and potentially with metabolic imprinting effects, as omega-6 FA was supplemented when steers were 6 months old [9]. Hence, omega-6 FA supplementation prior to feedlot arrival should also be considered as a nutritional intervention to improve initial health and performance of feedlot cattle.

Table 3. Performance and health responses from steers supplemented or not (CON; n = 6) with Ca soaps of soybean oil (CSSO; n = 6) for 28 days prior to feedlot arrival (day 0). Values reported are least square means \pm standard error. Adapted from Cooke et al. [9].

Item	CON	CSSO	p =
Plasma tumor necrosis alpha, pg/mL (log)			
Day 0 (arrival)	1.74 \pm 0.21	1.91 \pm 0.21	0.58
Day 1	1.88 \pm 0.23	2.00 \pm 0.22	0.67
Day 3	2.23 \pm 0.20	1.55 \pm 0.20	0.03
Feedlot average daily gain, kg/d			
Initial phase (day 1 to 144)	1.17 \pm 0.02	1.25 \pm 0.02	0.02
Final phase (day 145 to slaughter)	2.10 \pm 0.05	2.09 \pm 0.05	0.86
Carcass traits			
Hot carcass weight, kg	394 \pm 6	402 \pm 6	0.31
<i>Longissimus</i> muscle area, cm ²	94.7 \pm 1.5	92.0 \pm 1.6	0.23
Yield grade	3.16 \pm 0.10	3.48 \pm 0.11	0.04
Marbling	444 \pm 18	515 \pm 19	0.01
Backfat, cm	1.55 \pm 0.06	1.63 \pm 0.06	0.38

Beef cattle are typically backgrounded on pasture after weaning in areas where forage is available for grazing [43], although supplemental nutrients are often required in this practice to meet the requirements of growing cattle [44]. Hess et al. [5] reviewed multiple studies in which omega-6 FA was supplemented to grazing cattle, but using grains and oilseeds highly susceptible to ruminal biohydrogenation [40]. To fill this gap in knowledge, Cappellozza et al. [45] evaluated performance and nutrient intake of grazing *B. indicus* bulls supplemented with omega-6 FA via CSSO. In this study, ADG was increased in bulls offered a grain-based supplement at 0.3% of their body weight fortified with omega-6 FA compared with bulls receiving an iso-caloric and iso-nitrogenous control supplement (0.92 vs. 0.81 kg/day, respectively). These authors also noted that bulls supplemented with omega-6 FA consumed less water (4.11 vs. 4.96% of body weight), and hypothesized that this outcome was due to reduced ruminal caloric increment from inclusion of CSSO into the supplement [45]. More specifically, CSSO partially replaced corn to maintain the supplement's iso-caloric and iso-nitrogenous status, whereas ruminal fermentation of starch resulted in greater heat production compared with rumen-inert fats [45][46].

Another area of limited research is the inclusion of omega-6 FA into feedlot diets, as these FA from natural sources can disrupt ruminal function, feed intake and efficiency, and overall cattle performance [5]. The use of CSSO may partially alleviate these concerns, as supplementing Ca soaps of cottonseed oil improved feed efficiency of feedlot *B. indicus* bulls

compared with cohorts receiving isocaloric and isonitrogenous diets [47]. Accordingly, Nascimento et al. [47] investigated the inclusion of omega-6 FA via CSSO, or a mixture of palm, soybean, and cottonseed oils fed as Ca soaps into feedlot diets (CSMIX). Supplemented CSSO or CSMIX increased energy intake, feed efficiency, ADG, and carcass merit of *B. indicus* finishing bulls compared with cohorts not receiving supplemental fat (Table 4). In turn, cattle performance and carcass traits were not improved by omega-6 FA supplementation via CSSO compared with the saturated + monounsaturated FA provided by the CSMIX (Table 4). Therefore, omega-6 FA inclusion via CSSO to feedlot diets improved cattle performance and efficiency by increasing the energy density of the diet, whereas a combination of saturated + monounsaturated FA appears to be more favorable for feedlot productivity and carcass quality [48][49].

Table 4. Performance and carcass traits of feedlot bulls supplemented or not (CON; n = 16) with Ca soaps of soybean oil (CSSO; n = 16) or a mixture of palm, soybean, and cottonseed oils (CSMIX; n = 15) until slaughter. Values reported are least square means ± standard error. Adapted from Nascimento et al. [47] 1.

Item	CON	CSSO	CSMIX	C1	C2
Performance					
Average daily gain, kg/d	1.14 ± 0.04	1.37 ± 0.05	1.48 ± 0.05	<0.01	0.11
Feed efficiency, g/kg	156 ± 3	168 ± 3	183 ± 3	<0.01	<0.01
Final body weight, kg	476 ± 6	508 ± 7	524 ± 7	<0.01	0.13
Carcass traits					
Hot carcass weight, kg	268 ± 4	284 ± 4	297 ± 4	<0.01	0.03
Longissimus muscle area, cm ²	67.8 ± 1.88	70.4 ± 1.94	75.4 ± 1.94	0.04	0.08
Backfat, cm	0.318 ± 0.035	0.439 ± 0.039	0.448 ± 0.040	0.01	0.87

5. Conclusions

This review compiled recent research on omega-6 FA supplementation via CSSO to beef cattle, and its benefits to production efficiency across different environments and sectors of the beef industry. Supplementing omega-6 FA increased the reproductive efficiency of beef cows by promoting the processes associated with early pregnancy establishment. Omega-6 FA also elicited positive effects during periods of developmental plasticity, such as gestation and early postnatal life. Supplementing omega-6 FA to beef cows during late gestation resulted in alterations in tissue differentiation and improved health and productivity of offspring. Similar effects on developmental programming were noted when omega-6 FA was supplemented to young calves. Lastly, supplementing omega-6 FA to growing cattle receiving forage-based diets resulted in enhanced immunocompetence, growth, and carcass merit, although such benefits were not evident when omega-6 FA was provided to feedlot cattle consuming high-concentrate diets. Collectively, this review provides research-based evidence that omega-6 FA supplementation via CSSO is a sustainable approach to improve beef production efficiency.

References

- 2050 High-Level Experts Forum: The Forum. Available online: (accessed on 11 February 2021).
- Watson, A.K.; MacDonald, J.C.; Erickson, G.E.; Kononoff, P.J.; Klopfenstein, T.J. FORAGES AND PASTURES SYMPOSIUM: Optimizing the use of fibrous residues in beef and dairy diets. *J. Anim. Sci.* 2015, 93, 2616–2625.
- Kunkle, W.E.; Johns, J.T.; Poore, M.H.; Herd, D.B. Designing supplementation programs for beef cattle fed forage-based diets. *J. Anim. Sci.* 2000, 77, 1–12.
- National Academies of Sciences, Engineering, and Medicine. Nutrient Requirements of Beef Cattle: Eighth Revised Edition; The National Academies Press: Washington, DC, USA, 2016; ISBN 978-0-309-31702-3.
- Hess, B.W.; Moss, G.E.; Rule, D.C. A decade of developments in the area of fat supplementation research with beef cattle and sheep. *J. Anim. Sci.* 2008, 86, E188–E204.
- Sumida, C.; Graber, R.; Nunez, E. Role of fatty acids in signal transduction: Modulators and messengers. *Prostaglandins Leukot. Essent. Fatty Acids.* 1993, 48, 117–122.
- Funston, R.N. Fat supplementation and reproduction in beef females. *J. Anim. Sci.* 2004, 82, E154–E161.

8. Araujo, D.B.; Cooke, R.F.; Hansen, G.R.; Staples, C.R.; Arthington, J.D. Effects of rumen-protected polyunsaturated fatty acid supplementation on performance and physiological responses of growing cattle after transportation and feedlot entry. *J. Anim. Sci.* 2010, 88, 4120–4132.
9. Cooke, R.F.; Bohnert, D.W.; Moriel, P.; Hess, B.W.; Mills, R.R. Effects of polyunsaturated fatty acid supplementation on ruminal in situ forage degradability, performance, and physiological responses of feeder cattle. *J. Anim. Sci.* 2011, 89, 3677–3689.
10. Sukhija, P.S.; Palmquist, D.L. Dissociation of calcium soaps of long-chain fatty acids in rumen fluid. *J. Dairy Sci.* 1990, 73, 1784–1787.
11. Bellows, D.S.; Ott, S.L.; Bellows, R.A. Review: Cost of reproductive diseases and conditions in cattle. *Prof. Anim. Sci.* 2002, 18, 26–32.
12. Reese, S.T.; Franco, G.A.; Poole, R.K.; Hood, R.; Fernandez Montero, L.; Oliveira Filho, R.V.; Cooke, R.F.; Pohler, K.G. Pregnancy loss in beef cattle: A meta-analysis. *Anim. Repro. Sci.* 2020, 106521.
13. Yaqoob, P.; Calder, P.C. Fatty acids and immune function: New insights into mechanisms. *Br. J. Nutr.* 2007, 98, S41–S45.
14. Inskip, E.K. Preovulatory, postovulatory, and postmaternal recognition effects of concentrations of progesterone on embryonic survival in the cow. *J. Anim. Sci.* 2004, 82 (Suppl. 13), E24–E39.
15. Lopes, C.N.; Scarpa, A.B.; Cappellozza, B.I.; Cooke, R.F.; Vasconcelos, J.L.M. effects of rumen-protected polyunsaturated fatty acid supplementation on reproductive performance of *Bos indicus* beef cows. *J. Anim. Sci.* 2009, 87, 3935–3943.
16. Lopes, C.N.; Cooke, R.F.; Reis, M.M.; Peres, R.F.G.; Vasconcelos, J.L.M. Strategic supplementation of calcium salts of polyunsaturated fatty acids to enhance reproductive performance of *Bos indicus* beef cows. *J. Anim. Sci.* 2011, 89, 3116–3124.
17. Spencer, T.E.; Bazer, F.W. Conceptus Signals for establishment and maintenance of pregnancy. *Reprod. Biol. Endocrinol.* 2004, 2, 49.
18. Cooke, R.F.; Cappellozza, B.I.; Guarnieri Filho, T.A.; Depner, C.M.; Lytle, K.A.; Jump, D.B.; Bohnert, D.W.; Cerri, R.L.A.; Vasconcelos, J.L.M. Effects of calcium salts of soybean oil on factors that influence pregnancy establishment in *bos indicus* beef cows. *J. Anim. Sci.* 2014, 92, 2239–2250.
19. Roberts, R.M.; Cross, J.C.; Leaman, D.W. Interferons as hormones of pregnancy. *Endocr. Rev.* 1992, 13, 432–452.
20. Cipriano, R.S.; Cooke, R.F.; Rodrigues, A.D.; Silva, L.G.T.; Bohnert, D.W.; Marques, R.S.; Vasconcelos, J.L.M.; Pires, A.V.; Cerri, R.L.A. Post-Artificial insemination supplementation with calcium salts of soybean oil influences pregnancy establishment factors in *Bos indicus* beef cows. *J. Anim. Sci.* 2016, 94, 4892–4902.
21. Fricke, P.M.; Carvalho, P.D.; Lucy, M.C.; Curran, F.; Herlihy, M.M.; Waters, S.M.; Larkin, J.A.; Crowe, M.A.; Butler, S.T. Effect of manipulating progesterone before timed artificial insemination on reproductive and endocrine parameters in seasonal-calving, pasture-based Holstein-Friesian cows. *J. Dairy Sci.* 2016, 99, 6780–6792.
22. Green, J.C.; Okamura, C.S.; Poock, S.E.; Lucy, M.C. Measurement of interferon-tau (IFN- τ) stimulated gene expression in blood leukocytes for pregnancy diagnosis within 18–20d after insemination in dairy cattle. *Anim. Reprod. Sci.* 2010, 121, 24–33.
23. Schmitz, G.; Ecker, J. The opposing effects of n-3 and n-6 fatty acids. *Prog. Lipid Res.* 2008, 47, 147–155.
24. Dorniak, P.; Bazer, F.W.; Spencer, T.E. Prostaglandins regulate conceptus elongation and mediate effects of interferon tau on the ovine uterine endometrium. *Biol. Reprod.* 2011, 84, 1119–1127.
25. Cooke, R.F.; Cardoso, R.C.; Cerri, R.L.A.; Lamb, G.C.; Pohler, K.G.; Riley, D.G.; Vasconcelos, J.L.M. Board Invited Review—Cattle adapted to tropical and subtropical environments (II): Genetic and reproductive considerations. *J. Anim. Sci.* 2020, 98, skaa015.
26. Brandão, A.P.; Cooke, R.F.; Schubach, K.M.; Marques, R.S.; Bohnert, D.W.; Carvalho, R.S.; Dias, N.W.; Timlin, C.L.; Clark-Deener, S.; Currin, J.F.; et al. Supplementing ca salts of soybean oil after artificial insemination increases pregnancy success in *Bos taurus* beef cows. *J. Anim. Sci.* 2018, 96, 2838–2850.
27. Koletzko, B.; Brands, B.; Poston, L.; Godfrey, K.; Demmelmair, H. Early nutrition programming of long-term health. *Proc. Nutr. Soc.* 2012, 71, 371–378.
28. Fall, C.H.D. Evidence for the intra-uterine programming of adiposity in later life. *Ann. Hum. Biol.* 2011, 38, 410–428.
29. Reynolds, L.P.; Borowicz, P.P.; Caton, J.S.; Vonnahme, K.A.; Luther, J.S.; Hammer, C.J.; Maddock Carlin, K.R.; Grazul-Bilska, A.T.; Redmer, D.A. Developmental programming: The concept, large animal models, and the key role of uteroplacental vascular development. *J. Anim. Sci.* 2010, 88, E61–E72.

30. Robinson, J.J.; Sinclair, K.D.; McEvoy, T.G. Nutritional effects on foetal growth. *Anim. Sci.* 1999, 68, 315–331.
31. Funston, R.N.; Larson, D.M.; Vonnahme, K.A. Effects of maternal nutrition on conceptus growth and offspring performance: Implications for beef cattle production. *J. Anim. Sci.* 2010, 88, E205–E215.
32. Hochberg, Z.; Feil, R.; Constancia, M.; Fraga, M.; Junien, C.; Carel, J.-C.; Boileau, P.; Le Bouc, Y.; Deal, C.L.; Lillycrop, K.; et al. Child health, developmental plasticity, and epigenetic programming. *Endocr. Rev.* 2011, 32, 159–224.
33. Raclot, T.; Groscolas, R.; Langin, D.; Ferré, P. Site-specific regulation of gene expression by n-3 polyunsaturated fatty acids in rat white adipose tissues. *J. Lipid Res.* 1997, 38, 1963–1972.
34. Okuno, M.; Kajiwara, K.; Imai, S.; Kobayashi, T.; Honma, N.; Maki, T.; Suruga, K.; Goda, T.; Takase, S.; Muto, Y.; et al. Perilla oil prevents the excessive growth of visceral adipose tissue in rats by down-regulating adipocyte differentiation. *J. Nutr.* 1997, 127, 1752–1757.
35. Cleary, M.P.; Phillips, F.C.; Morton, R.A. Genotype and Diet Effects in Lean and Obese Zucker Rats Fed Either Safflower or Coconut Oil Diets. *Exp. Biol. Med.* 1999, 220, 153–161.
36. Massiera, F.; Saint-Marc, P.; Seydoux, J.; Murata, T.; Kobayashi, T.; Narumiya, S.; Guesnet, P.; Amri, E.-Z.; Negrel, R.; Ailhaud, G. Arachidonic acid and prostacyclin signaling promote adipose tissue development: A human health concern? *J. Lipid. Res.* 2003, 44, 271–279.
37. Zhu, M.-J.; Ford, S.P.; Nathanielsz, P.W.; Du, M. Effect of maternal nutrient restriction in sheep on the development of fetal skeletal muscle. *Biol. Reprod.* 2004, 71, 1968–1973.
38. Yan, X.; Zhu, M.-J.; Dodson, M.V.; Du, M. Developmental programming of fetal skeletal muscle and adipose tissue development. *J. Genomics.* 2013, 1, 29–38.
39. Du, M.; Tong, J.; Zhao, J.; Underwood, K.R.; Zhu, M.; Ford, S.P.; Nathanielsz, P.W. Fetal programming of skeletal muscle development in ruminant animals. *J. Anim. Sci.* 2010, 88, E51–E60.
40. Duff, G.C.; Galyean, M.L. Recent advances in management of highly stressed, newly received feedlot cattle. *J. Anim. Sci.* 2007, 85, 823–840.
41. Cooke, R.F. Invited Paper: Nutritional and management considerations for beef cattle experiencing stress-induced inflammation. *Prof. Anim. Sci.* 2017, 33, 1–11.
42. Miles, E.A.; Calder, P.C. Modulation of immune function by dietary fatty acids. *Proc. Nutr. Soc.* 1998, 57, 277–292.
43. Thomson, D.U.; White, B.J. Backgrounding beef cattle. *Vet. Clin. North Am. Food Anim. Pract.* 2006, 22, 373–398.
44. National Academies of Sciences, Engineering, and Medicine (NASEM). *Nutrient Requirements of Beef Cattle Model*; The National Academies Press: Washington, DC, USA, 2000.
45. Cappelozza, B.I.; Velasco, A.C.; Tongu, C.; Moraes, G.; Dib, R.; Cervieri, R. Effects of supplement amount, with or without calcium salts of fatty acids, on growth performance and intake behavior of grazing *Bos indicus* bulls. *Trans. Anim. Sci.* 2020, 4, 799–808.
46. Finch, V.A. Body temperature in beef cattle: Its control and relevance to production in the tropics. *J. Anim. Sci.* 1986, 62, 531–542.
47. Carvalho, M.A.A.; Cappelozza, B.I.; Silva, B.; Castro, T.S.; Burim, M.R.; Cervieri, R.C. Supplementation with calcium salts of cottonseed oil improves performance of *Bos indicus* animals consuming finishing diets. *Trans. Anim. Sci.* 2020, 4, 967–973.
48. Nascimento, F.A.; Silva, N.C.; Prados, L.F.; Pacheco, R.D.L.; Johnson, B.J.; Cappelozza, B.I.; Resende, F.D.; Siqueira, G.R. Calcium salts of fatty acids with varying fatty acid profiles in diets of feedlot-finished *bos indicus* bulls: Impacts on intake, digestibility, performance, and carcass and meat characteristics. *J. Anim. Sci.* 2020, 98.
49. Choi, S.H.; Gang, G.O.; Sawyer, J.E.; Johnson, B.J.; Kim, K.H.; Choi, C.W.; Smith, S.B. Fatty acid biosynthesis and lipogenic enzyme activities in subcutaneous adipose tissue of feedlot steers fed supplementary palm oil or soybean oil. *J. Anim. Sci.* 2013, 91, 2091–2098.