

Single-Cell Analysis of CTCs and Biomarker Detections

Subjects: **Oncology**

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The field of single-cell analysis has advanced rapidly in the last decade and is providing new insights into the characterization of intercellular genetic heterogeneity and complexity, especially in human cancer. Circulating and disseminated tumor cells (CTCs and DTCs) are cancer cells that dissociate from primary and metastatic cancer sites and enter the circulation with potential to seed distant metastases. CTCs can be enriched or isolated from a simple blood liquid biopsy. Analysis of multiple single CTCs has the potential to allow the identification and characterization of cancer heterogeneity to guide best therapy and predict therapeutic response.

whole genome amplification

circulating tumor cell (CTC)

single-cell analysis

biomarkers

1. Breast Cancer

Breast cancer (BC) is the most common female cancer and CTC is a predictive marker of poor survival and metastatic relapse ^[1]. The detection rate of CTCs correlates with the number of metastatic sites, and BC patients with brain metastasis may have the highest CTC counts ^[2].

The hormone status of BC, such as expression of the estrogen receptor (ER) or progesterone receptor (PR), indicates the feasibility of ER-targeted endocrine therapy ^[3]. However, no correlation was found between total CTC number and/or ER expression status as determined by immunocyto staining and the intensity of ER staining in primary tumors ^[4]. Only 81.3% of patients were positive for ER expression in CTCs, while ER-negative CTCs were also found in ER-positive patients, delineating the genetic inconsistencies between CTC counts. ER status in CTCs might have predictive power with regard to response and resistance to endocrine therapy and may thus help in the choice of better treatment options ^[4]. One study performed Sanger sequencing on CTC WGAs (MALBAC), which resulted in the identification of the *ESR1*-Y537S variant known to produce a constitutively active receptor and *ESR1*-T570I (a novel mutation) in exon 8 ^[5]. This study found *ESR1*-Y537S heterozygously and homozygously in single CTCs and confirmed mutations in matched cell-free DNA (cfDNA) in one patient. Interestingly, in another patient, heterozygote *ESR1*-T570I and homozygote *ESR1*-Y537S were found in a single CTC, but *ESR1*-T570I could not be detected in matched cfDNA ^[5]. Thus, using two entities extractable from a blood biopsy, CTCs and cfDNA biomarkers may complement each other and enhance the chance of finding disease-related variants. However, in another study that screened for exon 4, 6 and 8 *ESR1* mutations after WGA (Picoplex, MALBAC), none was found in individual CTCs ^[4].

The PI3K/AKT/mTOR pathway (Phosphoinositide 3-kinase/protein kinase B/mammalian target of rapamycin) regulates cell growth, survival, and angiogenesis. Upregulated activity has been linked to oncogenesis and is a major therapeutic target [6]. In BC, mutations in PIK3CA are found in about 40% of ER-positive cancers and have been implicated in resistance to HER2-based therapies [7]. Pharmacologic targeting of PIK3Ca in HR (hormone receptor) +/HER2-metastatic BC offers significant benefits to patients with endocrine therapy resistance [8]. Several single CTC-based studies [8][9][10][11][12] were conducted to study mutations in the PIK3CA gene. Heterogenous expression of PIK3CA mutations among CTCs and matched primary tumors, and even among CTCs from the same patient, was observed. Individual PIK3CA mutations found in Ampli1-amplified CTCs included E542K and H1047R [8], as well as E542K, E545K and H1047R, as was determined in a second study [10]. Another study found PIK3CA mutations (E542K, E545K, H1047R, H1047L and M1043V) in exon 9 and 20 in at least one CTC in 36.4% of the patients tested [13]; similar data were reported in other studies [11][12] (Table 1).

Table 1. The application of WGA and biomarker detection of single CTCs in various cancer types.

Studies (Author, Year)	CTC Isolation	CTC Recovery	WGA Kits	Downstream Molecular Analysis	CTCs+ Patients Analyzed	CTC Nr Analyzed for WGA	Main Findings in Genetic Mutations and Alterations
<i>mBC or HER2- mBC</i>							
Babayan, A. et al., 2013 [4]	Density gradient	Micromanipulator TransferMan NK2	PicoPlex	Multiplex PCR	4	8 single CTCs	<i>ESR1</i> mutations in exons 4, 6 and 8 were not found
De Luca, F. et al., 2016 [14]	CellSearch	DEPArray	Ampli1	NGS (Ion AmpliSeq Cancer Hotspot panel v2)	4	3–5 single CTCs per patient	51 sequence variants in 25 genes were found, including somatic mutations in <i>TP53</i> (8 mutations) and <i>PDGFRA</i> (3 mutations). High intra- and inter-patient heterogeneity, discordance in mutational status between CTCs and primary tissue
Gasch, C. et al., 2016 [13]	CellSearch	Micromanipulator TransferMan NK2	GenomiPhi, Ampli1	Sanger sequencing, PCR	33	114 single CTCs	<i>PIK3CA</i> mutations in exon 9 and 20
Kaur, P. et al., 2020 [15]	Microfluidic ANGLE Parsortix	NA	REPLI-g	WES (SNVs, CNAs and SVs)	5	5 CTCs and 5 WBCs	Elevated C>T mutational signature in patient samples. Low VAFs for somatic variants in CTCs compared to metastasis, complex rearrangement patterns were observed, high discordance between paired samples, marked heterogeneity of somatic landscape
Li, S. et al., 2020 [2]	CellCollector	CellCollector	REPLI-g	NGS (HiSeq X-Ten Illumina)	17	0–15 CTCs	Different metastatic sites have their own corresponding high-frequency mutation genes
Neumann, M. H. et al., 2016 [16]	CellSearch	CellCelector	Ampli1	For library preparation, the multiplex PCR-based Ion Torrent AmpliSeqTM	2	7 single CTCs	Functional <i>PIK3CA</i> SNP (G to A, E545K) was detected in CTCs of patient 1 but not in CTCs of patient 2

Studies (Author, Year)	CTC Isolation	CTC Recovery	WGA Kits	Downstream Molecular Analysis	CTCs+ Patients Analyzed	CTC Nr Analyzed for WGA	Main Findings in Genetic Mutations and Alterations
				technology with Ampli1 CHPCustom Beta panel			
Neves, R. P. et al., 2014 [12]	CellSearch	FACS	Ampli1	aCGH (CNAs), qPCR	30	192 single CTCs	72.9% WGA success rate, 46.2% of WGA products show <i>CCND1</i> amplification, mutations in <i>PIK3CA</i> exon 20 in c.3140 were found in CTCs (2/12 analyzed patients), <i>TP53</i> mutations in exons 5, 7 and 8 were not found
Paolillo, C. et al., 2017 [5]	CellSearch	DEPArray	MALBAC	Sanger sequencing	3	40 single CTCs and 12 WBCs	<i>ESR1</i> mutations (Y537S and T570I) were identified
Pestrin, M. et al., 2014 [10]	CellSearch	DEPArray	Ampli1	Sanger sequencing (hotspot regions in <i>PIK3CA</i> exon 9, 20)	18	115 single CTCs	33% of patients had an identified <i>PI3KCA</i> mutation. Six different mutations in the <i>PI3KCA</i> gene, such as c.3140A>G, c.1633G>A, c.1624G>A, c.1624G>A, etc., were identified
Polzer, B. et al., 2014 [11]	CellSearch	DEPArray	Ampli1	ERBB2 qPCR (CNV), <i>PIK3CA</i> Sequencing, aCGH	66	510 single CTCs and 189 leukocytes	<i>PIK3CA</i> mutations in exon 9 and 20. Analysis of ERBB2 alterations
Schneck, H. et al., 2013 [8]	CellSearch	NA	Ampli1	Multiplex PCR, SNaPshot	44	NA	<i>PIK3CA</i> mutations in exon 9 and 20, such as E545K and H1047R, were detected, but E542K, E545G and E545A were not found
Wang, Y. et al., 2018 [17]	FACS combined with oHSV1-hTERT-GFP viral infection	FACS	MALBAC	WGS for CTC, WGS and WES for matched primary and metastatic tissue	8	11 single CTCs	SNVs accumulated sporadically among CTCs and matched primary tumors, at least 2 CTCs shared 394 SNVs, SNV mutations in <i>APC</i> and <i>LRP1B</i> genes co-occurred in CTC-shared and bulk tissue, CTC behaviour-related SNVs were verified
Zou, L. et al., 2020 [18]	CellSearch	Micropipetting	MALBAC	WGS (CNV and gene set enrichment analysis)	2	Single CTCs, but number is unknown	Different frequencies of CNVs between newly diagnosed and recurrent liver metastasis; similar CNV patterns among isolated CTCs of recurrent BCLM and recurrent liver metastasis; 25 genes were identified as CNV signatures of BCLM, including β -defensins and defensins
<i>PC or mCRPC</i>							
Faugeroux, V. et al.,	ISSET filtration, CellSearch,	Self-seeding microwell chips,	Ampli1	WES (10x depth	11	179 WGA samples or	Shared <i>GRM8</i> , <i>TP53</i> and <i>PTEN</i> mutations in epithelial CTC samples and other CTC-exclusive variants

Studies (Author, Year)	CTC Isolation	CTC Recovery	WGA Kits	Downstream Molecular Analysis	CTCs+ Patients Analyzed	CTC Nr Analyzed for WGA	Main Findings in Genetic Mutations and Alterations
2018 [19]	RosetteSep	FACS, laser microdissection		coverage)		34 WES	
Greene, S. B. et al., 2016 [20]	Epic Sciences	Eppendorf TransferMan NK4 micromanipulator	SeqPlex Enhanced	Sequencing with Illumina NextSeq500 using a High Output kit in a Paired-End 2x150 format (PE 2x150) (CNV)	7	67 single CTCs	AR amplification and PTEN loss
Gupta, S. et al., 2016 [21]	CellSearch, RBC lysis and CD45 depletion	IE/FACS	RepliGene, WGA4	aCGH (CNV)	16	16 CTCs and matched leukocytes	AR amplification in 50% of CTC samples, ERG genomic amplification in 40% of patients, PTEN loss, genomic alteration in chromatin reading and proliferative pathways
Magbanua, M. J. et al., 2012 [22]	CellSearch, IE/FACS	IE/FACS	WGA4	aCGH	12	9 patient bulk CTCs	Gains in 8q and loss in 8p; gains in the AR region of chr X of CTCs, including AR gains in 78% of cases
Rangel-Pozzo, A. et al., 2020 [23]	ScreenCell filtration	Laser microdissection	Ampli1	WES	9	21 single CTCs and 4 lymphocytes	Genetic variations in nine telomere maintenance pathways, including telomeric repeat-binding factor 2 (TRF2), SNVs and indels associated with telomere maintenance genes and known cancer drug response; presence of CNAs in 11 different pathways, including the DNA damage repair (DDR) pathway
Wu, Y. et al., 2016 [24]	Density gradient, negative and positive selection with magnetic beads	Laser microdissection	PicoPLEX (<40 cells), WGA2 kit (GenomePlex for microdissected tissues)	SNP array profiling (CytoSNP-12 and omni1-Quad bead chips, Nspl 250k, SNP6.0, and CytoScanHD arrays), Nanostring (nCounter Cancer CN panel)	8	8 disseminated tumor cells (bulk cells)	Gain of Ch 7 and 8q, loss in 8p, 12q23, 10q26, 13q and 16q21. AR gain, TMPRSS2/ERG alterations and MYC and other gained regions, FOXO1 gene deletion
Lung Cancer							
He, Y. et al., 2017 [25]	CellCollector	CellCollector	REPLI-g	NGS (hotspot panel v2)	5	6 CTCs	44 cancer-related genes existed in mutations in the analyzed CTCs and some cancer-related mutations were identified in KIT, SMARCB1 and TP53 genes

2. Prostate Cancer

Prostate cancer (PC) is the most common cancer type diagnosed in men; eventually, it develops into castrate-resistant prostate cancer (CRPC) following standard of care androgen deprivation therapy (ADT). Commonly altered genes during CRPC progression include *AR* (androgen receptor), *ERG* (ETS-related gene), *c-MET* (tyrosine-protein kinase MET), *PTEN* (phosphatase and tension homology deleted on chromosome 10)

Studies (Author, Year)	CTC Isolation	CTC Recovery	WGA Kits	Downstream Molecular Analysis	CTCs+ Patients Analyzed	CTC Nr Analyzed for WGA	Main Findings in Genetic Mutations and Alterations
Lu, S. et al., 2020 [26]	CellSearch	DEPArray	MALBAC, REPLI-g, WGA4, Ampli1	Targeted sequencing, WES, WGS	4	80 single CTCs and 11 WBCs	Comparative study, MALBAC WGA coupled with LP-WGS is a robust workflow [24] for CNV profiling, but none of the WGA methods achieve sufficient sensitivity and specificity by WES
Mariscal, J. et al., 2016 [27]	CELLection Epithelial Enrich Dynabeads	NA	WTA2	Gene expression profiling (Agilent 4x44k gene expression arrays), qPCR	42 NSCLC patients and 16 controls	NA	CTC-specific expression profile associates with the PI3K/AKT, ERK1/2 and NF-kB pathways. NOTCH1, PTP4A3, LGALS3 and ITGB3 were further validated by RT-qPCR in an independent cohort of NSCLC patients
Nakamura, I. T. et al., 2021 [28]	AutoMACS	DEPArray	SMARTer PicoPLEX	NGS (Today OncoPanel, AmpliSeq for Illumina comprehensive cancer panel, WGS) and Sanger sequencing	2	40 single floating tumor cells in pleural effusion	EGFR exon 19 deletion was confirmed in 63.2% of samples from case 1, detection of 85% EML4-ALK fusion in case 2, alectinib-resistant mutation of ALK (p.G1202R) in case 2. A BRCA1 truncating mutation and an RAF1 oncogenic mutation were identified
Ni, X. et al., 2013 [29]	CellSearch	Micropipetting	MALBAC	WGS at ~0.1x sequencing depth and WES for SNV/indel	11	72 single CTCs (including 4 leucocytes)	EGFR mutations (such as one INDEL p.K746_A750del), PIK3CA (such as p.E545K), RB1 (p.R320*) and TP53 mutations (such as p.T155I) were only shared between the liver metastatic tumor and CTCs; gain region in chromosome 8q contains the c-Myc gene; gain in chromosome 5p, which contains the telomerase reverse transcriptase (TERT) gene; chromosomal regions, including 3q29, 17q22, 17q25.3 and 20p13, had significant gain in all 19 CTCs of patients
Colorectal Cancer							
Fabbri, F. et al., 2013 [30]	OncoQuick	DEPArray	Ampli1	Sequencing and pyrosequencing	21	16 samples or cases	KRAS gene mutations in 50% of cases. G12C, G12D and G13D-KRAS mutations in one patient in three different groups of CTCs
Gasch, C. et al., 2013 [9]	CellSearch	Micromanipulator TransferMan NK2	GenomePlex, GenomiPhi	Targeted sequencing for KRAS, BRAF and PIK3CA gene, qPCR for EGFR	5	69 single CTCs	EGFR amplification in 7/26 CTCs, KRAS mutations (G12V) in 33% of CTCs, PIK3CA mutations (E545A and E542K) in 39% of CTCs, no BRAF locus change detected
Li, R. et al., 2019 [31]	Microfluidic chip (SCIGA-chip)	Microfluidic chip (SCIGA-chip)	MDA	Illumina sequencing	1	2 single CTCs and 1	A novel method involving all processing steps from blood collection to WGA preparation, 11 shared somatic

3. Lung Cancer

The detection of certain driver mutations, such as in *EGFR* and *ALK* fusion, is associated with the early stages of lung cancer, its development and drug resistance [25]. Genetic analysis of CTCs from the same patient can give overall information about deletions, fusions, insertions and SNVs in the metastatic tumor and such changes can be monitored during treatment, even in the presence of cell-to-cell heterogeneity; however, a large number of CTCs needs to be sequenced [29].

Ni. et al. observed number of mutations in different genes, such as *EGFR*, *PIK3CA*, *RB1* and *TP53*, after exome sequencing of single-CTC WGA products. Amongst these alterations, one INDEL in the *EGFR* gene (K746_A750del), which is a target for tyrosine kinase inhibitors (TKIs), was found in CTCs as well as in the primary and metastatic tumors of the patients, while other mutations in *PIK3CA* (E545K), *TP53* (T155I) and *RB1* (R320*) genes were only observed in CTCs and metastatic tumors in the liver. This study also found some common CNV regions that have important roles in cancer development, such as cell proliferation, differentiation and protecting chromosomal ends from degradation. These regions include regions of gain in chromosome 8q, the *c-Myc* gene

Studies (Author, Year)	CTC Isolation	CTC Recovery	WGA Kits	Downstream Molecular Analysis (SNPs/SVs)	CTCs+ Patients Analyzed	CTC Nr Analyzed for WGA	Main Findings in Genetic Mutations and Alterations
[29]							
Pancreatic Cancer							
Court, C.M et al., 2016 [32]	Density gradient and NanoVelcro/LCM microchip	Laser microdissection	REPLI-g	Sanger sequencing	12	119 single CTCs and 103 WBCs	<i>KRAS</i> mutations in 92% of patients and 33 out of 119 single CTCs sequenced (resulting in a 27.7% detection rate in single CTCs). No <i>KRAS</i> mutants were found in any WBCs
Melanoma							
Reid, A. L. et al., 2014 [33]	RBC lysis, immune-magnetic beads	NA	REPLI-g	ddPCR and castPCR	15	30 CTCs	Comparative study of ddPCR and castPCR. <i>BRAF</i> -V600E/K mutations were detected
Ruiz, C. et al., 2016 [34]	RBC lysis	Micromanipulator	GenomePlex	CNV analysis	40	Single CTCs and WBCs	Deletions of <i>CDKN2A</i> and <i>PTEN</i> ; amplifications of <i>BRAF</i> , <i>TERT</i> , <i>MDM2</i> and <i>KRAS</i> ; chromosomal amplifications in chr12, 17 and 19
[28]							
Mixed patient cohort							
Aljohani, H.M. et al., 2018 [35]	RBC lysis, CD45 depletion and EpCam positive selection	FACS	REPLI-g	Sanger sequencing, ddPCR	10	NA	Mutations (R34G, E79Q, E82G) in <i>Nrf2</i> in isolated CTCs, some mutations in the Keap/Nrf2/ARE pathway
Ferrarini, A. et al., 2018 [36]	CellSearch	DEPArray	Ampli1	WGS (CNAs), aCGH	3	15 single CTCs and 7 WBCs	A large amplification (100 Mbp) on chr 8, including the <i>c-MYC</i> gene, copy number loss was detected in the <i>BRCA2</i> locus
Gao, Y. et al., 2017 [37]	CellSearch	Micropipetting	MALBAC	WGS and WES for SNV/indels, SVs, CNs	23	97 single CTCs	Homozygous deletion of <i>PTEN</i> ; amplification of the <i>MYC</i> gene; 11 focal regions were identified, including well-known tumor suppressor genes or oncogenes, which were deleted or amplified

main therapeutic target; however, responses to EGFR inhibition are variable [9]. The key mutations found in single-cell analysis of CRC CTCs so far are *KRAS*, *PIK3CA* and *EGFR* mutations. Significant heterogeneous expression of *KRAS*, *PIK3CA* and *EGFR* was found among CTCs within the same patient and between different individuals [9][30]. A mutational discordance between primary tumor tissue and CTC WGAs was observed for *KRAS*, and remarkably different *KRAS* mutations in different single-CTC WGAs from the same individual patients have been observed [9][30]. CTCs were observed with increased EGFR expression in some patients, and *EGFR* gene amplification was identified in 7 out of 26 CTC WGAs for three patients [9].

5. Other Cancer Types

Pancreatic cancer is a lethal cancer with a less than 10% 5-year survival rate. *KRAS* is the predominant mutated gene in pancreatic cancer, and targeting *KRAS* may be an attractive therapy, despite many trial failures for anti-*KRAS* therapies [41]. *KRAS* mutations have been detected in 92% of patients, with a detection rate of 27.7% in total Note: aCGH: array comparative genomic hybridization; chr: chromosome; CNA: copy number alteration; CNV: copy number variant; mCRPC: metastatic castration resistant prostate cancer; ddPCR: droplet digital PCR; FACS: fluorescence activated cell sorting; IE: immunomagnetic enrichment; ddPCR: droplet digital polymerase chain reaction; RBC: red blood cell; SNV: single nucleotide variant; SNP: single nucleotide polymorphism; SV: structural variant; WBC: white blood cell; WES: whole exome sequencing; WGA4 and WGA2: different versions of GenomePlex; WGS: whole genome sequencing; WTA: whole transcriptome amplification; WTS: whole transcriptome sequencing; NA: not available.

References

1. Bidard, F.-C.; Proudhon, C.; Pierga, J.-Y. Circulating tumor cells in breast cancer. *Mol. Oncol.* 2016, 10, 418–430.
2. Li, S.; Yang, S.; Shi, J.; Ding, Y.; Gao, W.; Cheng, M.; Sun, Y.; Xie, Y.; Sang, M.; Yang, H.; et al. Recognition of the organ-specific mutations in metastatic breast cancer by circulating tumor cells isolated in vivo. *Neoplasma* 2021, 68, 31–39.
3. Allison, K.H.; Hammond, M.E.H.; Dowsett, M.; McKernin, S.E.; Carey, L.A.; Fitzgibbons, P.L.; Hayes, D.F.; Lakhani, S.R.; Chavez-MacGregor, M.; Perlmutter, J.; et al. Estrogen and Progesterone Receptor Testing in Breast Cancer: ASCO/CAP Guideline Update. *J. Clin. Oncol.* 2020, 38, 1346–1366.
4. Babayan, A.; Hannemann, J.; Spötter, J.; Müller, V.; Pantel, K.; Joosse, S.A. Heterogeneity of estrogen receptor expression in circulating tumor cells from metastatic breast cancer patients. *PLoS ONE* 2013, 8, e75038.
5. Paolillo, C.; Mu, Z.; Rossi, G.; Schiewer, M.J.; Nguyen, T.; Austin, L.; Capoluongo, E.; Knudsen, K.; Cristofanilli, M.; Fortina, P. Detection of Activating Estrogen Receptor Gene (ESR1) Mutations in Single Circulating Tumor Cells. *Clin. Cancer Res.* 2017, 23, 6086–6093.
6. Peng, Y.; Wang, Y.; Zhou, C.; Mei, W.; Zeng, C.J.F.i.O. PI3K/Akt/mTOR Pathway and Its Role in Cancer Therapeutics: Are We Making Headway? *Front. Oncol.* 2022, 12, 819128.
7. Fusco, N.; Malapelle, U.; Fassan, M.; Marchiò, C.; Buglioni, S.; Zupo, S.; Criscitiello, C.; Vigneri, P.; Dei Tos, A.P.; Maiorano, E.; et al. PIK3CA Mutations as a Molecular Target for Hormone Receptor-Positive, HER2-Negative Metastatic Breast Cancer. *Front. Oncol.* 2021, 11, 644737.
8. Schneck, H.; Blassl, C.; Meier-Stiegen, F.; Neves, R.P.; Janni, W.; Fehm, T.; Neubauer, H. Analysing the mutational status of PIK3CA in circulating tumor cells from metastatic breast cancer patients. *Mol. Oncol.* 2013, 7, 976–986.
9. Gasch, C.; Bauernhofer, T.; Pichler, M.; Langer-Freitag, S.; Reeh, M.; Seifert, A.M.; Mauermann, O.; Izbic, J.R.; Pantel, K.; Riethdorf, S. Heterogeneity of epidermal growth factor receptor status and mutations of KRAS/PIK3CA in circulating tumor cells of patients with colorectal cancer. *Clin. Chem.* 2013, 59, 252–260.
10. Pestrin, M.; Salvianti, F.; Galardi, F.; De Luca, F.; Turner, N.; Malorni, L.; Pazzagli, M.; Di Leo, A.; Pinzani, P. Heterogeneity of PIK3CA mutational status at the single cell level in circulating tumor cells from metastatic breast cancer patients. *Mol. Oncol.* 2015, 9, 749–757.
11. Polzer, B.; Medoro, G.; Pasch, S.; Fontana, F.; Zorzino, L.; Pestka, A.; Andergassen, U.; Meier-Stiegen, F.; Czyz, Z.T.; Alberter, B.; et al. Molecular profiling of single circulating tumor cells with diagnostic intention. *EMBO Mol. Med.* 2014, 6, 1371–1386.
12. Neves, R.P.; Raba, K.; Schmidt, O.; Honisch, E.; Meier-Stiegen, F.; Behrens, B.; Möhlendick, B.; Fehm, T.; Neubauer, H.; Klein, C.A.; et al. Genomic high-resolution profiling of single

- CKpos/CD45neg flow-sorting purified circulating tumor cells from patients with metastatic breast cancer. *Clin. Chem.* 2014, 60, 1290–1297.
13. Gasch, C.; Oldopp, T.; Mauermann, O.; Gorges, T.M.; Andreas, A.; Coith, C.; Müller, V.; Fehm, T.; Janni, W.; Pantel, K.; et al. Frequent detection of PIK3CA mutations in single circulating tumor cells of patients suffering from HER2-negative metastatic breast cancer. *Mol. Oncol.* 2016, 10, 1330–1343.
 14. De Luca, F.; Rotunno, G.; Salvianti, F.; Galardi, F.; Pestrin, M.; Gabellini, S.; Simi, L.; Mancini, I.; Vannucchi, A.M.; Pazzagli, M.; et al. Mutational analysis of single circulating tumor cells by next generation sequencing in metastatic breast cancer. *Oncotarget* 2016, 7, 26107–26119.
 15. Kaur, P.; Campo, D.; Porras, T.B.; Ring, A.; Lu, J.; Chairez, Y.; Su, Y.; Kang, I.; Lang, J.E. A Pilot Study for the Feasibility of Exome-Sequencing in Circulating Tumor Cells Versus Single Metastatic Biopsies in Breast Cancer. *Int. J. Mol. Sci.* 2020, 21, 4826.
 16. Neumann, M.H.; Schneck, H.; Decker, Y.; Schömer, S.; Franken, A.; Endris, V.; Pfarr, N.; Weichert, W.; Niederacher, D.; Fehm, T.; et al. Isolation and characterization of circulating tumor cells using a novel workflow combining the CellSearch(®) system and the CellCelector(™). *Biotechnol. Prog.* 2016, 33, 125–132.
 17. Wang, Y.; Guo, L.; Feng, L.; Zhang, W.; Xiao, T.; Di, X.; Chen, G.; Zhang, K. Single nucleotide variant profiles of viable single circulating tumour cells reveal CTC behaviours in breast cancer. *Oncol. Rep.* 2018, 39, 2147–2159.
 18. Zou, L.; Imani, S.; Maghsoudloo, M.; Shasaltaneh, M.D.; Gao, L.; Zhou, J.; Wen, Q.; Liu, S.; Zhang, L.; Chen, G. Genome-wide copy number analysis of circulating tumor cells in breast cancer patients with liver metastasis. *Oncol. Rep.* 2020, 44, 1075–1093.
 19. Faugeroux, V.; Lefebvre, C.; Paillet, E.; Pierron, V.; Marcaillou, C.; Tourlet, S.; Billiot, F.; Dogan, S.; Oulhen, M.; Vielh, P.; et al. An Accessible and Unique Insight into Metastasis Mutational Content through Whole-Exome Sequencing of Circulating Tumor Cells in Metastatic Prostate Cancer. *Eur. Urol. Oncol.* 2020, 3, 498–508.
 20. Greene, S.B.; Dago, A.E.; Leitz, L.J.; Wang, Y.; Lee, J.; Werner, S.L.; Gendreau, S.; Patel, P.; Jia, S.; Zhang, L.; et al. Chromosomal Instability Estimation Based on Next Generation Sequencing and Single Cell Genome Wide Copy Number Variation Analysis. *PLoS ONE* 2016, 11, e0165089.
 21. Gupta, S.; Li, J.; Kemeny, G.; Bitting, R.L.; Beaver, J.; Somarelli, J.A.; Ware, K.E.; Gregory, S.; Armstrong, A.J. Whole Genomic Copy Number Alterations in Circulating Tumor Cells from Men with Abiraterone or Enzalutamide-Resistant Metastatic Castration-Resistant Prostate Cancer. *Clin. Cancer Res.* 2017, 23, 1346–1357.
 22. Magbanua, M.J.; Sosa, E.V.; Scott, J.H.; Simko, J.; Collins, C.; Pinkel, D.; Ryan, C.J.; Park, J.W. Isolation and genomic analysis of circulating tumor cells from castration resistant metastatic

- prostate cancer. *BMC Cancer* 2012, 12, 78.
23. Rangel-Pozzo, A.; Liu, S.; Wajnberg, G.; Wang, X.; Ouellette, R.J.; Hicks, G.G.; Drachenberg, D.; Mai, S. Genomic Analysis of Localized High-Risk Prostate Cancer Circulating Tumor Cells at the Single-Cell Level. *Cells* 2020, 9, 1863.
 24. Wu, Y.; Schoenborn, J.R.; Morrissey, C.; Xia, J.; Larson, S.; Brown, L.G.; Qu, X.; Lange, P.H.; Nelson, P.S.; Vessella, R.L.; et al. High-Resolution Genomic Profiling of Disseminated Tumor Cells in Prostate Cancer. *J. Mol. Diagn.* 2016, 18, 131–143.
 25. He, Y.; Shi, J.; Shi, G.; Xu, X.; Liu, Q.; Liu, C.; Gao, Z.; Bai, J.; Shan, B. Using the New CellCollector to Capture Circulating Tumor Cells from Blood in Different Groups of Pulmonary Disease: A Cohort Study. *Sci. Rep.* 2017, 7, 9542.
 26. Lu, S.; Chang, C.J.; Guan, Y.; Szafer-Glusman, E.; Punnoose, E.; Do, A.; Suttman, B.; Gagnon, R.; Rodriguez, A.; Ers, M.; et al. Genomic Analysis of Circulating Tumor Cells at the Single-Cell Level. *J. Mol. Diagn.* 2020, 22, 770–781.
 27. Mariscal, J.; Alonso-Nocelo, M.; Muinelo-Romay, L.; Barbazan, J.; Vieito, M.; Abalo, A.; Gomez-Tato, A.; Maria de Los Angeles, C.C.; Garcia-Caballero, T.; Rodriguez, C.; et al. Molecular Profiling of Circulating Tumour Cells Identifies Notch1 as a Principal Regulator in Advanced Non-Small Cell Lung Cancer. *Sci. Rep.* 2016, 6, 37820.
 28. Nakamura, I.T.; Ikegami, M.; Hasegawa, N.; Hayashi, T.; Ueno, T.; Kawazu, M.; Yagishita, S.; Goto, Y.; Shinno, Y.; Kojima, Y.; et al. Development of an optimal protocol for molecular profiling of tumor cells in pleural effusions at single-cell level. *Cancer Sci.* 2021, 112, 2006–2019.
 29. Ni, X.; Zhuo, M.; Su, Z.; Duan, J.; Gao, Y.; Wang, Z.; Zong, C.; Bai, H.; Chapman, A.R.; Zhao, J.; et al. Reproducible copy number variation patterns among single circulating tumor cells of lung cancer patients. *Proc. Natl. Acad. Sci. USA* 2013, 110, 21083–21088.
 30. Fabbri, F.; Carloni, S.; Zoli, W.; Ulivi, P.; Gallerani, G.; Fici, P.; Chiadini, E.; Passardi, A.; Frassineti, G.L.; Ragazzini, A.; et al. Detection and recovery of circulating colon cancer cells using a dielectrophoresis-based device: KRAS mutation status in pure CTCs. *Cancer Lett.* 2013, 335, 225–231.
 31. Li, R.; Jia, F.; Zhang, W.; Shi, F.; Fang, Z.; Zhao, H.; Hu, Z.; Wei, Z. Device for whole genome sequencing single circulating tumor cells from whole blood. *Lab Chip* 2019, 19, 3168–3178.
 32. Court, C.M.; Ankeny, J.S.; Sho, S.; Hou, S.; Li, Q.; Hsieh, C.; Song, M.; Liao, X.; Rochefort, M.M.; Wainberg, Z.A.; et al. Reality of Single Circulating Tumor Cell Sequencing for Molecular Diagnostics in Pancreatic Cancer. *J. Mol. Diagn.* 2016, 18, 688–696.
 33. Reid, A.L.; Freeman, J.B.; Millward, M.; Ziman, M.; Gray, E.S. Detection of BRAF-V600E and V600K in melanoma circulating tumour cells by droplet digital PCR. *Clin. Biochem.* 2014, 48, 999–1002.

34. Ruiz, C.; Li, J.; Luttgen, M.S.; Kolatkar, A.; Kendall, J.T.; Flores, E.; Topp, Z.; Samlowski, W.E.; McClay, E.; Bethel, K.; et al. Limited genomic heterogeneity of circulating melanoma cells in advanced stage patients. *Phys. Biol.* 2016, 12, 016008.
35. Aljohani, H.M.; Aittaleb, M.; Furgason, J.M.; Amaya, P.; Deeb, A.; Chalmers, J.J.; Bahassi, E.M. Genetic mutations associated with lung cancer metastasis to the brain. *Mutagenesis* 2018, 33, 137–145.
36. Ferrarini, A.; Forcato, C.; Buson, G.; Tononi, P.; Del Monaco, V.; Terracciano, M.; Bolognesi, C.; Fontana, F.; Medoro, G.; Neves, R.; et al. A streamlined workflow for single-cells genome-wide copy-number profiling by low-pass sequencing of LM-PCR whole-genome amplification products. *PLoS ONE* 2018, 13, e0193689.
37. Gao, Y.; Ni, X.; Guo, H.; Su, Z.; Ba, Y.; Tong, Z.; Guo, Z.; Yao, X.; Chen, X.; Yin, J.; et al. Single-cell sequencing deciphers a convergent evolution of copy number alterations from primary to circulating tumor cells. *Genome Res.* 2017, 27, 1312–1322.
38. Antonarakis, E.S.; Lu, C.; Wang, H.; Lubber, B.; Nakazawa, M.; Roeser, J.C.; Chen, Y.; Mohammad, T.A.; Chen, Y.; Fedor, H.L. AR-V7 and resistance to enzalutamide and abiraterone in prostate cancer. *N. Engl. J. Med.* 2014, 371, 1028–1038.
39. Khan, T.; Becker, T.M.; Scott, K.F.; Descallar, J.; de Souza, P.; Chua, W.; Ma, Y. Prognostic and Predictive Value of Liquid Biopsy-Derived Androgen Receptor Variant 7 (AR-V7) in Prostate Cancer: A Systematic Review and Meta-Analysis. *Front. Oncol.* 2022, 12, 868031.
40. Ciombor, K.; Strickler, J.; Bekaii-Saab, T.; Yaeger, R. BRAF-Mutated Advanced Colorectal Cancer: A Rapidly Changing Therapeutic Landscape. *J. Clin. Oncol.* 2022.
41. Yu, J.; Gemenetzi, G.; Kinny-Köster, B.; Habib, J.R.; Groot, V.P.; Teinor, J.; Yin, L.; Pu, N.; Hasanain, A.; van Oosten, F.; et al. Pancreatic circulating tumor cell detection by targeted single-cell next-generation sequencing. *Cancer Lett.* 2020, 493, 245–253.

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