

Noncoding RNAs in Macrophage Polarization

Subjects: Allergy

Contributor: Osamu Ishibashi, Stefan A. Muljo, Zohirul Islam

Allergy is a type 2 immune reaction triggered by antigens known as allergens, including food and environmental substances such as peanuts, plant pollen, fungal spores, and the feces and debris of mites and insects. Macrophages are myeloid immune cells with phagocytic abilities that process exogenous and endogenous antigens. Upon activation, they can produce effector molecules such as cytokines as well as anti-inflammatory molecules. The dysregulation of macrophage function can lead to excessive type 1 inflammation as well as type 2 inflammation, which includes allergic reactions. Thus, it is important to better understand how macrophages are regulated in the pathogenesis of allergies. Emerging evidence highlights the role of noncoding RNAs (ncRNAs) in macrophage polarization, which in turn can modify the pathogenesis of various immune-mediated diseases, including allergies.

Keywords: macrophage ; macrophage polarization ; allergy ; non-coding RNA

1. Introduction

Allergies affect millions of people worldwide and are characterized by an excessive type 2 immune response to normally harmless substances, generally known as antigens or allergens, specifically [1][2]. Consequently, this response leads to the development of various allergic symptoms, including asthma, allergic rhinitis, and atopic dermatitis. In the most severe cases, it can result in anaphylaxis and possibly death. According to the World Allergy Organization, the prevalence of allergic diseases has been continuously increasing in the industrialized world [3][4]. In addition, according to the World Health Organization, the number of asthma patients is expected to increase to 400 million by 2025 [4]. The process by which the immune system becomes sensitive to a particular allergen is called sensitization and is typically accompanied by the development of immunoglobulin E (IgE), a specific subclass of antibodies, against the allergen. Sensitization rates to one or more common allergens among schoolchildren are reported to be between 40% and 50% [5]. Since antigen E was isolated from the pollen of common ragweed (*Ambrosia artemisiifolia*) as the first antigen in 1962 [6], a variety of environmental and food allergens have been identified, including 106 allergens that have recently (between January 2019 and March 2021) been accepted by the Allergen Nomenclature Sub-Committee (<http://allergen.org/committee.php>, accessed on 5 July 2023) [7].

Antihistamines are widely used for symptomatic treatment of many allergic diseases with variable efficacy. Despite the identification of increasing varieties of antigens, there is no fundamental treatment to overcome allergic symptoms except for allergen immunotherapy or desensitization, whereby long-term remission is expected, against a few food and environmental allergens, e.g., cedar pollen [8]. Desensitization therapy is actively investigated because of its clinical potential; however, it harbors the intrinsic risk of inducing severe side effects such as anaphylaxis [9]. Thus, desensitization shots are co-administered with antihistamines. A drug that promotes immune tolerance by targeting macrophages, for instance, could make allergen desensitization safer and even more effective. In 2003, omalizumab, an IgE-blocking antibody received approval from the U.S. Food and Drug Administration (FDA), but it is not approved for all allergic conditions, and it is expensive [10]. Alternatively, blocking antibodies against specific allergens are being developed, but these will be even more expensive for patients that are allergic to multiple allergens. Therefore, an alternative remedy based on a new concept is desirable for this growing patient population.

Immune cells such as mast cells, basophils, dendritic cells, B cells, and specific T-cell subsets are well recognized as key players in allergic reactions. In contrast, to date, macrophages are not commonly associated with allergies. In the future, it would be key for the field to provide credible *in vivo* evidence that macrophages also play a role in modulating allergy first using mouse models but ultimately in human patients. However, several lines of evidence have recently revealed the crucial role of macrophages in developing and modulating these allergic responses [11][12][13][14][15][16]. For example, macrophages are the most abundant immune cells present in the lungs (approximately 70% of the immune cells) and play a crucial role in asthma caused by environmental-allergen-induced airway inflammation [17][18], suggesting that

macrophages, together with other immune cells could play a role in immune responses. Therefore, the role of macrophages in allergic diseases and the mechanism underlying their functional regulation deserve further study.

In one popular paradigm, macrophages can be divided into two major subclasses, i.e., M1 and M2, based on the inflammatory responses that they mediate, and the process by which macrophages differentiate in response to challenge is called macrophage polarization. Macrophage polarization is determined by the microenvironment (**Figure 1**). However, the mechanisms underlying *in vivo* macrophage polarization are complicated and remain largely unclarified, but various intracellular molecules, including signaling molecules and enzymes, and receptors have been shown to regulate macrophage polarization [19][20][21][22][23][24][25][26][27][28][29][30]. For example, it is not known whether dupilumab, a clinically approved biologic drug that blocks IL4R α signaling [31], leads to an *in vivo* reduction in M2 macrophages in patients receiving this drug. In the future, it would be important to investigate which immune cells are in fact being inhibited by dupilumab. Alternatively, it is possible that dupilumab switches macrophages into a tolerant state, for instance, by turning on the expression of anti-inflammatory molecules. Thus, it is crucial that the mechanism of action of dupilumab be investigated systematically at the cellular and molecular levels. Here, the researchers provide a consideration of macrophages and noncoding RNAs (ncRNAs). Accumulating evidence has revealed that ncRNAs, a class of functional RNAs not translated into proteins and associated with various pathological events, are associated with both macrophage polarization and allergies. ncRNAs are typically classified into two major types that have distinct functions, i.e., housekeeping ncRNAs and regulatory ncRNAs. The detailed classification of ncRNAs is discussed elsewhere [32][33]. Emerging evidence shows that these ncRNAs play roles in macrophage polarization related to allergies [34][35].

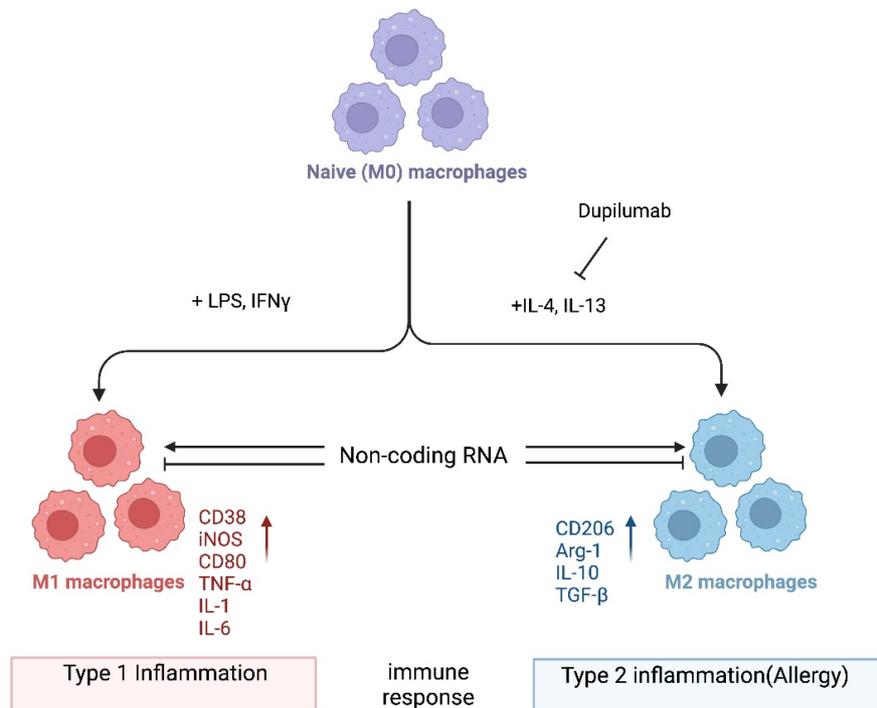


Figure 1. Macrophage polarization. Naïve (M0) macrophages in their inactive state can be polarized into either of two types of activated macrophages with distinct functions, M1 and M2 macrophages (also termed “classically activated” or “alternatively activated” macrophages, respectively), after exposure to certain stimuli. M1 and M2 macrophages are functionally associated with type 1 and type 2 immune reactions, respectively. Several mRNAs and proteins are used as markers to differentiate between these macrophages: i.e., arginase-1 (Arg-1) and CD206 for M2; CD38, CD80, and iNOS for M1 macrophages. However, the criteria for the subclassification of macrophages *in vivo* in different tissues still require further investigation. Noncoding RNAs might regulate the differentiation of macrophages and/or the function of M1 and M2 macrophages by modifying gene expression programs. Dupilumab is a currently available monoclonal antibody that blocks IL-4 and IL-13 signaling by targeting IL4R α [31]. This biologic drug is FDA-approved for allergic diseases such as eczema, asthma, and nasal polyps, which result in chronic sinusitis. Hypothetically, its mechanism of action is in part to inhibit M2 polarization. Tralokinumab, another FDA-approved monoclonal antibody, used for the treatment of atopic dermatitis, targets just the cytokine IL-13 (not depicted). Again, it is not well understood which cell types are being affected by this biologic drug. It would be interesting to directly compare dupilumab versus tralokinumab and assess the *in vivo* effects of each on macrophages and their noncoding transcriptome. Image created with [BioRender.com](https://www.biorender.com) (accessed on 28 November 2023).

As mentioned earlier, ncRNAs are RNA molecules that do not encode proteins and are essential in regulating gene expression at both transcriptional and post-transcriptional levels, which includes epigenetic regulation [36]. There are at

least three classes of ncRNAs that regulate gene expression: microRNAs (miRNAs), long ncRNAs (lncRNAs), and circular RNAs (circRNAs). Furthermore, ncRNAs that regulate protein activity have been described [37][38]. While miRNA-mediated regulation of gene expression occurs at the post-transcriptional level, lncRNAs and circRNAs may utilize diverse mechanisms of action. Emerging evidence has highlighted the critical role of ncRNAs in regulating macrophage polarization, which may lead to the development of allergies [39]. Although the mechanisms through which ncRNAs regulate macrophage polarization are diverse and complex, several studies have shown that ncRNAs potentially regulate M1 and M2 macrophage polarization by targeting the regulators of proinflammatory signaling pathways or regulating the expression of anti- or proinflammatory cytokines.

Although published studies have thus far highlighted the functional association between ncRNAs and macrophage polarization or that between ncRNAs and allergic diseases, few reports have described the ncRNA–macrophage polarization–allergy axis. Therefore, in the following sections, the researchers summarize these previous studies on how the individual ncRNA classes are involved in macrophage polarization and how that may relate to allergic diseases.

2. miRNA-Mediated Regulation of Macrophage Polarization

miRNAs are small (typically ~22 nt in length) ncRNAs that post-transcriptionally regulate gene and protein expression by binding to the 3'-untranslated region of the target mRNAs, which induces mRNA degradation and translational repression [35][40]. Several miRNAs have been demonstrated to regulate macrophage polarization related to allergic diseases.

The mannose receptor MRC1/CD206 is expressed in immune cells, and its expression level is pronouncedly elevated in M2 macrophages; therefore, it is generally accepted as an M2 macrophage marker [25]. MRC1/CD206 recognizes an extensive range of surface glycoproteins and plays a crucial role in a variety of immunological events, both physiologically and pathologically [41]. Interestingly, *miR-511-3p* is an miRNA that is transcribed from an intron of the *MRC1* gene. The expressions of *miR-511-3p* and MRC1/CD206 have been shown to be coregulated in macrophages [42][43]. In studies with the MRC1 knockout mouse model in which *miR-511-3p* expression is also deficient, Zhou et al. demonstrated that *miR-511-3p* downregulated M1 macrophage polarization, upregulated M2 macrophage polarization, and protected against cockroach allergen-induced lung inflammation [43]. In addition, it was reported by Do et al. that *miR-511-3p* promoted M2 macrophage polarization and attenuated cockroach-allergen-induced lung inflammation by targeting CCL2 [44]. Alternatively, Heinsbroek et al. demonstrated that *miR-511-3p* regulated intestinal inflammation by controlling macrophage-mediated microbial responses via the indirect upregulation of TLR-4 expression [45]. These findings suggest that *miR-511-3p* regulates macrophage functions and polarization by targeting multiple mRNAs.

Using an allergen-induced asthma knockout mouse model, Chung et al. reported that *miR-451a* negatively affects IL-4-induced M2 macrophage polarization by targeting and silencing the expression of Sirtuin 2 and promoting asthmatic inflammation [46]. Additionally, a few other studies were conducted using ovalbumin-induced allergic asthma mouse models to identify miRNAs in macrophage polarization. For example, Veremeyko et al. demonstrated that *miR-124* expression was upregulated in the lung alveolar macrophages of an ovalbumin-induced allergic lung inflammation mouse model and contributed to the development of M2, but not M1, macrophage polarization [47]. Another study by Shi et al. highlighted the involvement of *miR-142-5p* and *miR-130a-3p* in pulmonary macrophage polarization and asthma airway remodeling in ovalbumin-sensitized mice [48]. Additionally, Su et al. reported that *miR-142-5p* and *miR-130a-3p* functioned by targeting suppressor of cytokine signaling 1 (SOCS1) and peroxisome proliferator-activated receptor γ , respectively [49]. Notably, this study revealed that SOCS1 had a negative impact on the M2 macrophage polarization in mice [49], while the M2 polarization of human macrophages is enhanced by SOCS1 [50]. Again, such contradiction will need to be resolved before ncRNAs can be selected as allergy drug targets.

3. lncRNA-Mediated Regulation of Macrophage Polarization

lncRNAs are long (generally defined to be >200 nt in length) ncRNAs that regulate gene expression at various levels, which include chromatin remodeling, transcriptional regulation, and post-transcriptional regulation [38][39]. Several lncRNAs have been shown to regulate M2 macrophage polarization related to allergies. For example, the knockdown of receptor-type tyrosine protein phosphatase ϵ (PTPRE)-AS1, a lncRNA selectively expressed in IL-4-stimulated macrophages, was shown to promote M2 macrophage activation via the MAPK/ERK 1/2 pathway [51]. Wen et al. recently demonstrated that MIR222HG acts on the *miR146a-5p*/TRAF6/NF- κ B axis, leading to the attenuation of macrophage M2 polarization and allergic inflammation in allergic rhinitis [52]. The few studies investigating the lncRNA AK085865 have also highlighted its role in macrophage polarization [53][54]. In particular, the study conducted by Pei et al. showed that AK085865-deficient mice were protected from the allergic airway inflammation induced by Der f 1, a major mite allergen component of *Dermatophagoides farinae* [53]. They also found that AK085865 deletion suppressed M2 macrophage polarization, which

subsequently decreased their susceptibility to Der f 1-induced airway inflammation. In addition, Zhang et al. demonstrated that AK085865 specifically interacted with interleukin-enhancer-binding factor (ILF)-2 and functioned as a negative regulator of the ILF2–ILF3-complex-mediated biosynthesis of *miR-192*, which promotes M2 macrophage polarization through the direct targeting of interleukin-1 receptor-associated kinase (IRAK) 1 [54].

lnc-BAZ2B, a lncRNA dominantly expressed in monocytes and significantly upregulated in children with asthma, was also demonstrated to promote M2 macrophage polarization. Mechanistically, lnc-BAZ2B promotes the expression of BAZ2B mRNA by stabilizing its pre-mRNA, leading to enhanced IRF4 expression and M2 macrophage polarization [55]. Another lncRNA reported to regulate the pathological state of allergies is NKILA [56]. This lncRNA was demonstrated to limit the asthmatic airway inflammation, enhancing M2 macrophage polarization and inhibiting the NF-κB pathway in a mouse asthmatic model.

In contrast to many reports on the lncRNA-mediated regulation of M2 macrophage polarization in allergy, there are few reports on the lncRNA–M1 macrophage polarization-allergy axis. One of the few such studies, reported by Jiang et al., describes the contribution of lncRNA MEG8-sponging of *miR-181a-5p* to M1 macrophage polarization via regulating SHP2 expression in a rat model of IgA purpura, which is a type 3 allergic disease triggered by allergens such as drugs, food, or insect bites [57]. In another study, Zhu et al. demonstrated that lncRNA growth-arrest-specific transcript 5 (GAS5) is upregulated in exosomes isolated from the nasal mucus of allergic rhinitis patients and promotes M1 macrophage polarization by restraining autophagy and subsequently activating NF-κB signaling [58].

4. circRNA-Mediated Regulation of Macrophage Polarization

circRNAs are a recently discovered product of back splicing, and a subset of them do not encode for protein. Thus, they comprise a new category of ncRNAs that form covalently closed circular structures, which make them resistant to degradation by RNA exonucleases [59][60]. Since they are long-lived, a few circRNAs have been shown to act as molecular “sponges” that sequester miRNAs and/or RNA-binding proteins [61]. Although the function of most circRNAs remains poorly understood, a few circRNAs have been demonstrated to regulate the macrophage polarization associated with allergy. For example, Shang et al. reported that circ_0001359 was downregulated in ovalbumin-induced asthmatic mice compared with normal mice and circ_0001359-enriched exosomes secreted from adipose-derived stem-cells-attenuated airway remodeling via promoting polarization into M2-like macrophages [62]. Mechanistically, circ_0001359 was shown to regulate macrophage polarization by enhancing FoxO1 signaling via sponging *miR-183-5p* (Figure 2A) [62]. Recently, luteolin, a flavone reported to have a protective role in asthma, was shown to activate M2 and suppress M1 macrophage polarization via upregulating circ_0001326 in the human macrophage cell line THP-1 [63]. The same study also elucidated the underlying mechanism of how circ_0001326 regulates downstream gene expression, including *miR-136-5p* and USP4 (Figure 2B). Finally, it is also conceivable that synthetic circRNAs could be rationally designed to inhibit specific miRNAs to treat diseases such as allergy. Once miRNAs that promote allergy have been identified and validated, then one could simply multimerize the binding sites for miRNA(s) of interest into a synthetic circRNA that will serve to inhibit them and, in turn, allergy.

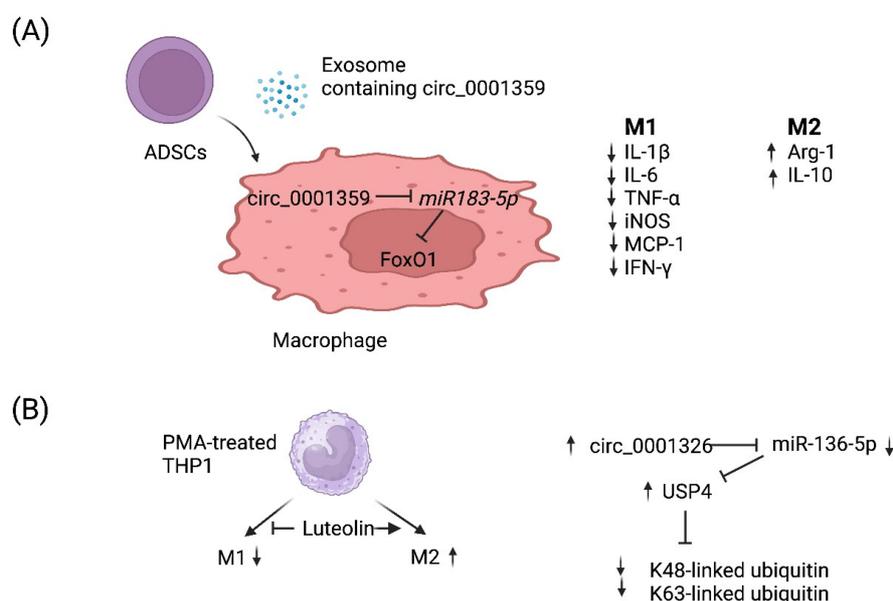


Figure 2. Working models of how two circRNAs regulate M1 vs. M2 macrophage polarization. (A) Adipose-derived stem cells (ADSCs) secrete exosomes that contain circ_0001359. Upon fusion with macrophages, circ_0001359 is released

into the cytoplasm and promotes M2-like macrophage polarization in an ovalbumin-induced asthma mouse model and lipopolysaccharide-induced RAW264.7 macrophages cells as evidenced by Arg-1 and IL-10 expression. In contrast, the expression of the following M1 effector molecules is suppressed by circ_0001359: IL-1 β , IL-6, TNF- α , iNOS, MCP-1, and IFN- γ . Mechanistically, circ_0001359 inhibits *miR-183-5p* via base pairing. Since *Foxo1* mRNA is directly repressed by *miR-183-5p*, FoxO1 activity is enhanced as a result and may be in part responsible for reprogramming macrophage cell fate. Image created with [BioRender.com](https://www.biorender.com) (accessed on 28 November 2023). (B) Luteolin (a naturally occurring flavonoid found in plants), known for its protective role in asthma, inhibits M1 macrophage polarization and promotes M2 activation in THP-1-derived macrophages. Luteolin-treated THP-1 macrophages induce expression of circ_0001326, inhibiting *miR-136-5p* via base pairing. Consequently, ubiquitin-specific protease 4 (USP4) is upregulated since it is directly repressed by *miR-136-5p*, and ultimately K48-linked and K63-linked ubiquitin is metabolized by USP4 since it is a deubiquitinase enzyme. Image created with [BioRender.com](https://www.biorender.com) (accessed on 28 November 2023).

References

1. Lee, T.H. Allergy: The unmet need. *Clin. Med.* 2003, 3, 303–305.
2. Pulendran, B.; Artis, D. New Paradigms in Type 2 Immunity. *Science* 2012, 337, 431–435.
3. Pawankar, R.; Canonica, G.W.; Holgate, S.T.; Lockey, R.F.; Blaiss, M.S. WAO White Book on Allergy: Update 2013 Executive Summary; World Allergy Organization: Milwaukee, WI, USA, 2013.
4. Pawankar, R.; Canonica, G.W.; Holgate, S.T.; Lockey, R.F. Allergic diseases and asthma: A major global health concern. *Curr. Opin. Allergy Clin. Immunol.* 2012, 12, 39–41.
5. Pawankar, R.; Mori, S.; Ozu, C.; Kimura, S. Overview on the pathomechanisms of allergic rhinitis. *Asia Pac. Allergy* 2011, 1, 157–167.
6. King, T.P.; Norman, P.S. Isolation studies of allergens from ragweed pollen. *Biochemistry* 1962, 1, 709–720.
7. Sudharson, S.; Kalic, T.; Hafner, C.; Breiteneder, H. Newly defined allergens in the WHO/IUIS Allergen Nomenclature Database during 01/2019-03/2021. *Allergy* 2021, 76, 3359–3373.
8. Thompson, C.P.; Silvers, S.; Shapiro, M.A. Intralymphatic immunotherapy for mountain cedar pollinosis: A randomized, double-blind, placebo-controlled trial. *Ann. Allergy Asthma Immunol.* 2020, 125, 311–318.e2.
9. James, C.; Bernstein, D.I. Allergen immunotherapy: An updated review of safety. *Curr. Opin. Allergy Clin. Immunol.* 2017, 17, 55–59.
10. Busse, W.; Corren, J.; Lanier, B.Q.; McAlary, M.; Fowler-Taylor, A.; Cioppa, G.D.; van As, A.; Gupta, N. Omalizumab, anti-IgE recombinant humanized monoclonal antibody, for the treatment of severe allergic asthma. *J. Allergy Clin. Immunol.* 2001, 108, 184–190.
11. Girodet, P.O.; Nguyen, D.; Mancini, J.D.; Hundal, M.; Zhou, X.; Israel, E.; Cernadas, M. Alternative Macrophage Activation Is Increased in Asthma. *Am. J. Respir. Cell Mol. Biol.* 2016, 55, 467–475.
12. Abdelaziz, M.H.; Abdelwahab, S.F.; Wan, J.; Cai, W.; Huixuan, W.; Jianjun, C.; Kumar, K.D.; Vasudevan, A.; Sadek, A.; Su, Z.; et al. Alternatively activated macrophages; a double-edged sword in allergic asthma. *J. Transl. Med.* 2020, 18, 58.
13. Balhara, J.; Gounni, A.S. The alveolar macrophages in asthma: A double-edged sword. *Mucosal Immunol.* 2012, 5, 605–609.
14. van der Veen, T.A.; de Groot, L.E.; Melgert, B.N. The different faces of the macrophage in asthma. *Curr. Opin. Pulm. Med.* 2020, 26, 62–68.
15. Hou, Y.; Wei, D.; Zhang, Z.; Guo, H.; Li, S.; Zhang, J.; Zhang, P.; Zhang, L.; Zhao, Y. FABP5 controls macrophage alternative activation and allergic asthma by selectively programming long-chain unsaturated fatty acid metabolism. *Cell Rep.* 2022, 41, 111668.
16. Mackaness, G.B. Cellular resistance to infection. *J. Exp. Med.* 1962, 116, 381–406.
17. Robbe, P.; Draijer, C.; Borg, T.R.; Luinge, M.; Timens, W.; Wouters, I.M.; Melgert, B.N.; Hylkema, M.N. Distinct macrophage phenotypes in allergic and nonallergic lung inflammation. *Am. J. Physiol. Cell. Mol. Physiol.* 2015, 308, L358–L367.
18. Draijer, C.; Robbe, P.; Boorsma, C.E.; Hylkema, M.N.; Melgert, B.N. Dual role of YM1+ M2 macrophages in allergic lung inflammation. *Sci. Rep.* 2018, 8, 5105.

19. Zhu, L.; Zhao, Q.; Yang, T.; Ding, W.; Zhao, Y. Cellular metabolism and macrophage functional polarization. *Int. Rev. Immunol.* 2015, 34, 82–100.
20. Bertani, F.R.; Mozetic, P.; Fioramonti, M.; Iuliani, M.; Ribelli, G.; Pantano, F.; Santini, D.; Tonini, G.; Trombetta, M.; Businaro, L.; et al. Classification of M1/M2-polarized human macrophages by label-free hyperspectral reflectance confocal microscopy and multivariate analysis. *Sci. Rep.* 2017, 7, 8965.
21. Mosser, D.M.; Edwards, J.P. Exploring the full spectrum of macrophage activation. *Nat. Rev. Immunol.* 2008, 8, 958–969.
22. Islam, Z.; Inui, T.; Ishibashi, O. Gpr137b is an orphan G-protein-coupled receptor associated with M2 macrophage polarization. *Biochem. Biophys. Res. Commun.* 2019, 509, 657–663.
23. Nathan, C.F.; Murray, H.W.; Wiebe, M.E.; Rubin, B.Y. Identification of interferon-gamma as the lymphokine that activates human macrophage oxidative metabolism and antimicrobial activity. *J. Exp. Med.* 1983, 158, 670–689.
24. Martinez, F.O.; Gordon, S. The M1 and M2 paradigm of macrophage activation: Time for reassessment. *F1000Prime Rep.* 2014, 6, 6–13.
25. Murray, P.J.; Allen, J.E.; Biswas, S.K.; Fisher, E.A.; Gilroy, D.W.; Goerdts, S.; Gordon, S.; Hamilton, J.A.; Ivashkiv, L.B.; Lawrence, T.; et al. Macrophage activation and polarization: Nomenclature and experimental guidelines. *Immunity* 2014, 41, 14–20.
26. Sica, A.; Mantovani, A. Macrophage plasticity and polarization: In vivo veritas. *J. Clin. Investig.* 2012, 122, 787–795.
27. Boutilier, A.J.; ElSawa, S.F. Macrophage Polarization States in the Tumor Microenvironment. *Int. J. Mol. Sci.* 2021, 22, 6995.
28. Li, J.; Kim, S.Y.; Lainez, N.M.; Coss, D.; Nair, M.G. Macrophage-Regulatory T Cell Interactions Promote Type 2 Immune Homeostasis Through Resistin-Like Molecule α . *Front. Immunol.* 2021, 12, 710406.
29. Wang, N.; Liang, H.; Zen, K. Molecular mechanisms that influence the macrophage m1-m2 polarization balance. *Front. Immunol.* 2014, 5, 614.
30. Liu, J.; Geng, X.; Hou, J.; Wu, G. New insights into M1/M2 macrophages: Key modulators in cancer progression. *Cancer Cell Int.* 2021, 21, 389.
31. Le Floch, A.; Allinne, J.; Nagashima, K.; Scott, G.; Birchard, D.; Asrat, S.; Bai, Y.; Lim, W.K.; Martin, J.; Huang, T.; et al. Dual blockade of IL-4 and IL-13 with dupilumab, an IL-4R α antibody, is required to broadly inhibit type 2 inflammation. *Allergy* 2020, 75, 1188–1204.
32. Dahariya, S.; Paddibhatla, I.; Kumar, S.; Raghuwanshi, S.; Palapati, A.; Gutti, R.K. Long non-coding RNA: Classification, biogenesis and functions in blood cells. *Mol. Immunol.* 2019, 112, 82–92.
33. Zhang, P.; Wu, W.; Chen, Q.; Chen, M. Non-Coding RNAs and their Integrated Networks. *J. Integr. Bioinform.* 2019, 16, 2019-0027.
34. Tian, C.; Gao, J.; Yang, L.; Yuan, X. Non-coding RNA regulation of macrophage function in asthma. *Cell. Signal.* 2023, 112, 110926.
35. Feketea, G.; Bocsan, C.I.; Popescu, C.; Gaman, M.; Stanciu, L.A.; Zdrenghea, M.T. A Review of Macrophage MicroRNAs' Role in Human Asthma. *Cells* 2019, 8, 420.
36. Peschansky, V.J.; Wahlestedt, C. Non-coding RNAs as direct and indirect modulators of epigenetic regulation. *Epigenetics* 2014, 9, 3–12.
37. Statello, L.; Guo, C.J.; Chen, L.L.; Huarte, M. Gene regulation by long non-coding RNAs and its biological functions. *Nat. Rev. Mol. Cell Biol.* 2021, 22, 159.
38. Mattick, J.S.; Amaral, P.P.; Carninci, P.; Carpenter, S.; Chang, H.Y.; Chen, L.L.; Chen, R.; Dean, C.; Dinger, M.E.; Fitzgerald, K.A. Long non-coding RNAs: Definitions, functions, challenges and recommendations. *Nat. Rev. Mol. Cell Biol.* 2023, 24, 430–447.
39. Goodarzi, V.; Nouri, S.; Nassaj, Z.S.; Bighash, M.; Abbasian, S.; Hagh, R.A. Long non coding RNAs reveal important pathways in childhood asthma: A future perspective. *Histochem. J.* 2023, 54, 257–269.
40. Squadrito, M.L.; Etzrodt, M.; De Palma, M.; Pittet, M.J. MicroRNA-mediated control of macrophages and its implications for cancer. *Trends Immunol.* 2013, 34, 350–359.
41. Martinez-Pomares, L. The mannose receptor. *J. Leukoc Biol.* 2012, 92, 1177–1186.
42. Squadrito, M.L.; Pucci, F.; Magri, L.; Moi, D.; Gilfillan, G.D.; Ranghetti, A.; Casazza, A.; Mazzone, M.; Lyle, R.; Naldini, L.; et al. miR-511-3p modulates genetic programs of tumor-associated macrophages. *Cell Rep.* 2012, 1, 141–154.

43. Zhou, Y.; Do, D.C.; Ishmael, F.T.; Squadrito, M.L.; Tang, H.M.; Tang, H.L.; Hsu, M.H.; Qiu, L.; Li, C.; Zhang, Y.; et al. Mannose receptor modulates macrophage polarization and allergic inflammation through miR-511-3p. *J. Allergy Clin. Immunol.* 2018, 141, 350–364.e8.
44. Do, D.C.; Mu, J.; Ke, X.; Sachdeva, K.; Qin, Z.; Wan, M.; Ishmael, F.T.; Gao, P. miR-511-3p protects against cockroach allergen-induced lung inflammation by antagonizing CCL2. *J. Clin. Investig.* 2019, 4, e126832.
45. Heinsbroek, S.E.; Squadrito, M.L.; Schilderink, R.; Hilbers, F.W.; Verseijden, C.; Hofmann, M.; Helmke, A.; Boon, L.; Wildenberg, M.E.; Roelofs, J.J.; et al. miR-511-3p, embedded in the macrophage mannose receptor gene, contributes to intestinal inflammation. *Mucosal Immunol.* 2016, 9, 960–973.
46. Chung, S.; Lee, Y.G.; Karpurapu, M.; Englert, J.A.; Ballinger, M.N.; Davis, I.C.; Park, G.Y.; Christman, J.W. Depletion of microRNA-451 in response to allergen exposure accentuates asthmatic inflammation by regulating Sirtuin2. *Am. J. Physiol. Cell. Mol. Physiol.* 2020, 318, L921–L930.
47. Veremeyko, T.; Siddiqui, S.; Sotnikov, I.; Yung, A.; Ponomarev, E.D. IL-4/IL-13-dependent and independent expression of miR-124 and its contribution to M2 phenotype of monocytic cells in normal conditions and during allergic inflammation. *PLoS ONE* 2013, 8, e81774.
48. Shi, J.; Chen, M.; Ouyang, L.; Wang, Q.; Guo, Y.; Huang, L.; Jiang, S. miR-142-5p and miR-130a-3p regulate pulmonary macrophage polarization and asthma airway remodeling. *Immunol. Cell Biol.* 2020, 98, 715–725.
49. Su, S.; Zhao, Q.; He, C.; Huang, D.; Liu, J.; Chen, F.; Chen, J.; Liao, J.Y.; Cui, X.; Zeng, Y.; et al. miR-142-5p and miR-130a-3p are regulated by IL-4 and IL-13 and control profibrogenic macrophage program. *Nat Commun.* 2015, 6, 8523.
50. Paoletti, A.; Rohmer, J.; Ly, B.; Pascaud, J.; Rivière, E.; Seror, R.; Le Goff, B.; Nocturne, G.; Mariette, X. Monocyte/Macrophage Abnormalities Specific to Rheumatoid Arthritis Are Linked to miR-155 and Are Differentially Modulated by Different TNF Inhibitors. *J. Immunol.* 2019, 203, 1766–1775.
51. Han, X.; Huang, S.; Xue, P.; Fu, J.; Liu, L.; Zhang, C.; Yang, L.; Xia, L.; Sun, L.; Huang, S.K.; et al. LncRNA PTPRE-AS1 modulates M2 macrophage activation and inflammatory diseases by epigenetic promotion of PTPRE. *Sci. Adv.* 2019, 5, eaax9230.
52. Wen, S.; Li, F.; Tang, Y.; Dong, L.; He, Y.; Deng, Y.; Tao, Z. MIR222HG attenuates macrophage M2 polarization and allergic inflammation in allergic rhinitis by targeting the miR146a-5p/TRAF6/NF- κ B axis. *Front. Immunol.* 2023, 14, 1168920.
53. Pei, W.; Zhang, Y.; Li, X.; Luo, M.; Chen, T.; Zhang, M.; Zhong, M.; Lv, K. LncRNA AK085865 depletion ameliorates asthmatic airway inflammation by modulating macrophage polarization. *Int. Immunopharmacol.* 2020, 83, 106450.
54. Zhang, Y.; Li, X.; Wang, C.; Zhang, M.; Yang, H.; Lv, K. LncRNA AK085865 Promotes Macrophage M2 Polarization in CVB3-Induced VM by Regulating ILF2-ILF3 Complex-Mediated miRNA-192 Biogenesis. *Mol. Ther. Nucleic Acids* 2020, 21, 441–451.
55. Xia, L.; Wang, X.; Liu, L.; Fu, J.; Xiao, W.; Liang, Q.; Han, X.; Huang, S.; Sun, L.; Gao, Y.; et al. Lnc-BAZ2B promotes M2 macrophage activation and inflammation in children with asthma through stabilizing BAZ2B pre-mRNA. *J. Allergy Clin. Immunol.* 2021, 147, 921–932.e9.
56. Li, Q.; Lu, L.; Li, X.; Lu, S. Long non-coding RNA NKILA alleviates airway inflammation in asthmatic mice by promoting M2 macrophage polarization and inhibiting the NF- κ B pathway. *Biochem. Biophys. Res. Commun.* 2021, 571, 46–52.
57. Jiang, M.; Dai, J.; Yin, M.; Jiang, C.; Ren, M.; Tian, L. LncRNA MEG8 sponging miR-181a-5p contributes to M1 macrophage polarization by regulating SHP2 expression in Henoch-Schonlein purpura rats. *Ann. Med.* 2021, 53, 1576–1588.
58. Zhu, X.; Sun, Y.; Yu, Q.; Wang, X.; Wang, Y.; Zhao, Y. Exosomal lncRNA GAS5 promotes M1 macrophage polarization in allergic rhinitis via restraining mTORC1/ULK1/ATG13-mediated autophagy and subsequently activating NF- κ B signaling. *Int. Immunopharmacol.* 2023, 121, 110450.
59. Yu, C.-Y.; Kuo, H.-C. The emerging roles and functions of circular RNAs and their generation. *J. Biomed. Sci.* 2019, 26, 29.
60. Memczak, S.; Jens, M.; Elefsinioti, A.; Torti, F.; Krueger, J.; Rybak, A.; Maier, L.; Mackowiak, S.D.; Gregersen, L.H.; Munschauer, M.; et al. Circular RNAs are a large class of animal RNAs with regulatory potency. *Nature* 2013, 495, 333–338.
61. Zang, J.; Lu, D.; Xu, A. The interaction of circRNAs and RNA binding proteins: An important part of circRNA maintenance and function. *J. Neurosci. Res.* 2020, 98, 87–97.
62. Shang, Y.; Sun, Y.; Xu, J.; Ge, X.; Hu, Z.; Xiao, J.; Ning, Y.; Dong, Y.; Bai, C. Exosomes from mmu_circ_0001359-Modified ADSCs Attenuate Airway Remodeling by Enhancing FoxO1 Signaling-Mediated M2-like Macrophage Activation. *Mol. Ther. Nucleic Acids* 2020, 19, 951–960.

63. Gong, B.; Zheng, Y.; Li, J.; Lei, H.; Liu, K.; Tang, J.; Peng, Y. Luteolin activates M2 macrophages and suppresses M1 macrophages by upregulation of hsa_circ_0001326 in THP-1 derived macrophages. *Bioengineered* 2022, 13, 5079–5090.

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