

Therapeutic Potential of Intrabodies for Cancer Immunotherapy

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Tumor cells are characterized by overexpressed tumor-associated antigens or mutated neoantigens, which are expressed on the cell surface or intracellularly. One strategy of cancer immunotherapy is to target cell-surface-expressed tumor-associated antigens (TAAs) with therapeutic antibodies. Intrabodies are suitable to knockdown TAAs and neoantigens without off-target effects. Intrabodies can now be selected against virtually any protein inside the cell. RNA sequencing and proteome analysis of single tumor cells combined with computational methods is bringing forward the identification of new neoantigens for the selection of anti-cancer intrabodies, which can be easily performed using phage display antibody repertoires. Anti-cancer intrabodies demonstrated tumor growth inhibition in appropriate xenograft tumor mouse models. At the moment, the biggest challenge in translating TAA/neoantigen-directed intrabodies into the clinic is the specific targeting of the intrabodies to the tumor cells. The promising development of tumor-specific lipid nanoparticles which could be embedded with an mRNA transgene or new capsid-modified and tumor-specific recombinant AAVs should enable tumor-cell-specific intrabody transfection/transduction in cancer patients and may finally bring intrabodies into the clinic.

TAAs

neoantigens

intrabodies

cancer immunotherapy

1. Introduction

Cancer immunotherapy with monoclonal antibodies or antibody fragments comprises the targeting of antibodies to extracellular or intracellular tumor-associated antigens (TAAs) or mutated neoantigens ^{[1][2][3]}. In contrast to TAAs, neoantigens comprise tumor-specific mutations and are not expressed by normal cells. The binding of tumor-specific antibodies to extracellular TAAs or neoantigens activates natural killer cells, macrophages or the complement system, leading to the destruction of the tumor cells. In addition to naked monoclonal antibodies, bispecific antibodies, immunotoxins, immunocytokines and engineered CAR-T cells with a TAA or neoantigen-specific antibody fragment or TCR-like antibody fused to T-cell signal domains can be successfully applied ^{[4][5]}.

Intrabodies can now be selected against virtually any protein inside the cell and they have the potential to specifically inhibit the function of TAAs and even neoantigens in cancer patients. Two different kinds of intrabodies exist with different modes of action. One group of intrabodies comprises the ER intrabodies produced as scFvs inside the ER to inhibit the function of transitory proteins passing the secretory pathway ^[6]. Functional inhibition is performed through intrabody/antigen retention by the SE (KDEL) sequence fused to the C terminal end of the intrabody. Many ER intrabodies have been selected against overexpressed TAAs on the tumor cell surface ^{[6][7][8]}.

The other group are single domain antibodies (sdAbs) comprising only the variable domain of the heavy chain VHH from camels (nanobodies) or sharks or human VH and VLs and are stable in the cytoplasm or nucleus [9]. They inactivate their targets by altering their conformation or interfere with the binding of the target protein to its corresponding binding partner.

Anti-cancer intrabodies demonstrated tumor growth inhibition in appropriate xenograft tumor mouse models [10][11][12][13][14][15][16][17][18][19][20]. Furthermore, a scFv-Fc intrabody inhibited the function of the serin (727)-phosphorylated form of STAT3 (pSSTAT3) in vitro and in mice [21]. STAT3 is involved in proliferation and apoptosis processes. CAR T cells transduced with an anti-CD7 intrabody applied in T-cell acute lymphoblastic leukemia (T-ALL) have been described [22]. The resulting CAR T cells only eliminated CD7⁺ lymphoblastic leukemia T cells and not the CAR T cells also expressing CD7.

2. Intrabodies against Oncogenic Cell Surface Receptors

TAA passing the secretory pathway are mainly cell surface receptors. They are targeted by ER intrabodies. TAAs and neoantigens expressed in the cytosol or nucleus are targeted by single domain antibodies. ER intrabodies in the scFv format are very stable in the ER but not in the nucleus or cytoplasm whereas sdAbs are stable in both compartments [23]. ER intrabodies can be constructed from hybridoma clones as scFv or selected from human antibody phage display repertoires [8][24].

To construct intrabodies from hybridomas targeting cell surface receptors, the variable genes of the light and heavy chain of the monoclonal antibody are amplified with consensus primer, by adapter-ligated PCR, 5' RACE or by PCR with primers of the constant region using circularized cDNA [24]. Both variable genes are then assembled to a scFv fragment and cloned into an ER targeting vector, providing the ER retention signal fused to the start of the intrabody coding region after cloning. Most anti-cancer ER intrabodies targeting cell surface receptors (human IL-2 receptor, ErbB-2, TLR2) were constructed from hybridoma clones [25][26].

ER intrabodies were also generated using phage display antibody repertoires. A scFv intrabody targeting VEGFR2/KDR was selected from an immune phage display library of mice [27]. Recently, intrabodies targeting human papillomavirus 16 E6 and E7 oncoproteins for the treatment of established HPV-associated tumors were generated from a synthetic human antibody phage display library [28]. One scFv against 16 E7 was targeted to the ER and interferes with the binding of E7 to retinoblastoma tumor suppressor (pRb). The second intrabody fused with a nuclear localization sequence rescued p53 activity, leading to cell death.

Some ER intrabodies were not targeted to cell surface receptors but to membrane proteins in intracellular compartment. These include TLR9, which translocates into the endosome after activation with CpG DNA and is involved in pancreatic tumor development [29] and polysialyltransferases ST8SialII and ST8SialIV localized in the Golgi apparatus which are activated during the growth of several tumors [30]. All those intrabodies were generated from hybridoma clones [14][31].

3. Intrabodies against Cytoplasmic or Nucleus Located TAAS

Several intrabodies have been generated in recent years against intracellular antigens [32]. The sdAb against F-actin capping protein CapG inhibited breast tumor metastasis in a xenograft tumor mouse model [15]. Recently an anti-HIF-1 α nanobody was developed to decrease gemcitabine resistance in pancreatic cancer patients [33]. The intrabody competitively inhibited the binding of the transcription factor HIF-1 α heterodimer to the aryl hydrocarbon receptor nuclear translocator (ARNT), leading to the inhibition of the HIF-1/VEGF pathway in vitro. It is known that high HIF-1 α expression reduces sensitivity to gemcitabine, which is applied to treat pancreatic adenocarcinoma and other cancers [34]. Interestingly the nanobody reduced tumor growth in mice treated with the nanobody to 41.58%, treated with gemcitabine to 64.89% and treatment with both gemcitabine and nanobody to 80.44%, respectively.

In another approach, nanobodies against AKT2, an isoform of the three AKT isoforms (AKT1, AKT2 and AKT3) of serine/threonine protein kinase AKT (also known as protein kinase B), were developed. AKT regulates cell proliferation, survival, growth, migration and metabolism and is hyperactivated by phosphorylation in cancer [35]. One nanobody (Nb8) affects the phosphorylation and/or expression levels of a wide range of proteins downstream of AKT, resulting in a G0/G1 cell cycle arrest, the induction of autophagy, a reduction in focal adhesion count and loss of stress fibers.

Furthermore, a nanobody was recently reported to inhibit the interaction between Twist1 and p53 [36]. Twist1 facilitates the degradation of p53 by the negative regulator murine double minute 2-Protein (MDM2), which in wild-type cancer inhibits the transcriptional activity of p53 and induces its degradation. This study demonstrated that targeting Twist1/p53 interaction with nanobodies might be a therapeutic option.

Interestingly, the function of a new nanobody against a G protein-coupled receptor (GPCR) of the human cytomegalovirus (CMV), US28, which is involved in cancer progression in glioblastoma was demonstrated [37]. The nanobody (VUN103) inhibits constitutive US28 signaling by G protein displacement and inhibited US28-enhanced spheroid growth in glioblastoma tumor cells. Phosphorylation of STAT3 was completely inhibited to the level of uninfected cells in glioblastoma tumor cells infected with CMV and transduced with the VUN103 intrabody. The group developed another nanobody (Nb7) which binds ligand-bound US28 and inhibits signaling too. Both nanobodies recognize two different conformations of US28, the super activated US28 induced by ligand CX3CL1 and the constitutively active US28. In both cases, the binding of the nanobodies inhibited the recruitment of the G protein α q subunit (G α q).

Furthermore, a sdAb ER intrabody targeted an ER membrane receptor: the human immunoglobulin heavy chain (VH) targeted B-cell receptor-associated protein 31 (BAP31 and blocked cyclin kinase inhibitor p27kip1 degradation, leading to growth inhibition and apoptotic death in GC cells and GC tumors in mice [19]. Finally, a novel nucleus-located intracellular nanobody against HPV16 E6 oncoprotein was recently generated from an immune camelid single domain library, inhibiting tumor growth in HPV16 mouse models [19].

4. Intrabodies against Intracellular Neoantigens

Intracellular TAAs and neoantigens are targeted by single domain antibodies selected from camels, sharks or human VH or VL antibody repertoires by phage display or ribosomal display [32][38][39]. Recently a new technique was established for the selection of small molecules, peptide aptamers and cytoplasmic intrabodies: localization-based interaction screening (SOLIS) [40]. SOLIS employs two chimeric proteins in which a membrane localization motif (CaaX) is fused at the C-terminus of interest neoantigens while the catalytic domain of SOS is fused at the C-terminus of another protein of interest. Son of sevenless (SOS) is a guanine nucleotide exchange factor that activates Ras upon growth factor stimulation. Interaction between the two proteins of interest induces membrane localization of the SOS fusion protein and then cell proliferation due to the activation of the RasMAPK signaling pathway.

Intrabodies were selected against oncogenic Ras mutants using different approaches leading to several intrabodies in different formats: disulfide free scFv, human VH nanobody, complete IgG antibody, “RasIns” (fibronectins) and DARPins [10][11][12][13][41][42][43] (**Table 1**) (**Figure 1**). In its active form, Ras binds to GTP and can interact with multiple effector proteins such as Raf kinases, PI3K (phosphatidylinositol 3-kinase) and RalGDS (Ral guanine nucleotide dissociation stimulator) [44]. Ras mutants of the canonical *ras* gene family, *H-ras*, *N-ras* and *K-ras*, are frequently activated by point mutation in human cancers, predominantly at G12, G13 and Q61 residues, leading to impaired GTPase activity resulting in constitutively active mutants persistently binding GTP and promoting tumorigenesis and tumor malignancy [45]. KRAS G12V is present in 3.47% of AACR GENIE cases, with pancreatic adenocarcinoma, lung adenocarcinoma, colon adenocarcinoma, colorectal adenocarcinoma and rectal adenocarcinoma having the greatest prevalence. The proteins *H-ras*, *N-ras* and *K-ras* are also called *H-p21Ras*, *K-p21Ras* and *N-p21Ras*.

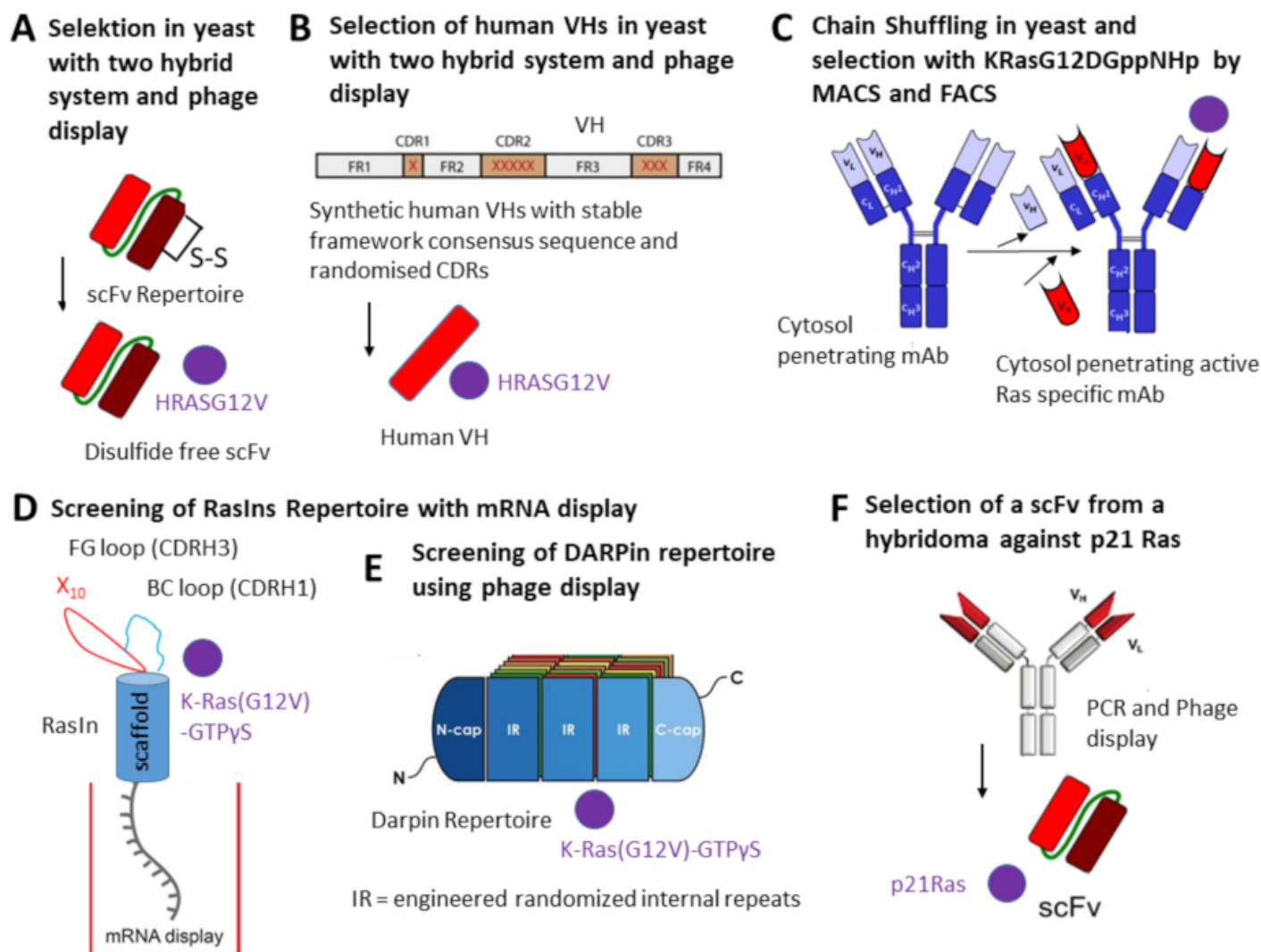


Figure 1. Strategies used to select intrabodies against Ras mutants.

Table 1. Reports of intrabody-mediated knockdown of the *ras* gene family.

Target	Selection of Intrabody	Physiological Knockdown Readout	Ref.
HRASG12V	A disulfide free scFv was selected with the intracellular antibody capture (IAC) technology [46].	When NIH 3T3 cells were co-transfected with the disulfide-free scFv and RAS mutant-dependent luciferase construct the number of transformed foci was reduced to 30%.	[41]
HRASG12V	A human VH domain was selected from two synthetic VH domain libraries with fully randomized complementarity determining regions (CDRs) introduced into a single stable intrabody framework. One human VH was selected in yeast.	The VH binds to activated GTP-bound wild-type HRAS and HRAS (G12V). Nude mice were injected subcutaneously with mouse (NIH3T3-EJ) or human (HT-1080 or DLD-1) tumor cells transduced with the anti-Ras intrabody. Tumors were not developed in mice when the sdAb was expressed in the	[10]

Target	Selection of Intrabody	Physiological Knockdown Readout	Ref.
		tumor cells compared to cells with an empty vector or an irrelevant intrabody.	
GTP-bound <i>K</i> - and <i>H</i> -Ras and the corresponding G12V mutants	Antibody-like ligands as intrabodies were developed (RasIn1 and RasIn2). They were selected by mRNA display using an antigen <i>K</i> -Ras(G12V)-GTPyS.	Detailed binding analysis demonstrated that RasIn1 and RasIn2 recognized the binding domain of the Raf kinase in activated <i>H</i> -RasG12V.	[42]
Ras mutants	This anti-Ras mutant antibody was engineered from a human antibody previously generated and is internalized through clathrin-mediated endocytosis using heparan sulfate proteoglycan (HSPG) as a receptor and escapes from early endosomes into the cytosol [47].	The intrabody (RT11-i) recognizes the GTP-bound active forms of wild-type (WT) KRas, NRas and HRas and their oncogenic mutants with mutations at positions 12, 13 or 61, such as KRasG12D, KRasG12V, KRasG13D, KRasQ61H, HRasG12V and NRasQ61R. RT11-i significantly inhibits the tumor growth of oncogenic Ras mutant tumor xenografts in mice.	[11]
Ras mutants	A new human IgG intrabody (inRas37) binding to activated GTP-bound Ras mutants with two-fold stronger activity was engineered from RT11-i.	Inhibition of tumor growth was seen in several xenograft tumor mice bearing different preestablished colorectal tumors.	[12]
p21 Ras	scFv antibody was generated from a hybridoma.	The scFv recognizes wild-type <i>H</i> -p21Ras, <i>K</i> -p21Ras and <i>N</i> -p21Ras [12] and their mutated variants. The intrabody significantly inhibited the tumor growth of nude mice with established tumors derived from human colon cancer cell line SW480 or human liver cancer cell line BEL-7402.	[13]
KRASG12V [41]	DARPins were selected from a phage display library by biopanning using biotinylated KRASG12V.	DARPins bound to an allosteric site of GDP or GTP-bound KRASWT and KRASG12V inhibiting KRAS nucleotide exchange and dimerization.	[43]

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ng the VH

and VL framework with a known stable consensus framework yielding high expression in bacteria and mammalian cell cytoplasm. When NIH 3T3 cells were co-transfected with the disulfide-free scFv and a RAS mutant-dependent luciferase construct the number of transformed foci was reduced to 30%. In a following approach, a human VH domain was selected from two synthetic VH domain libraries with fully randomized complementarity determining regions (CDRs) introduced into a single stable intrabody framework [10][41]. One human VH was selected in yeast. This VH binds to activated GTP-bound wild-type HRAS and HRAS (G12V). It was shown that the sdAb inhibited tumor growth and metastasis in a tumor xenograft mouse model. Nude mice were injected subcutaneously with mouse (NIH3T3-EJ) or human (HT-1080 or DLD-1) tumor cells transduced with the anti-Ras intrabody. Tumors were not developed in mice when the sdAb was expressed in the tumor cells compared to cells with an empty vector or an irrelevant intrabody. The crystal structure of intrabody and Ras showed that the intrabody covers the surface of RAS where most of the effectors (RAF, RALGDS and PI3K) interact.

Another approach utilizes antibody-like ligands exhibiting an immunoglobulin-like fold that are composed of the 10th fibronectin type III domain of human fibronectin, with two hypervariable loops that are structurally similar to antibody CDRH1 (BC loop) and CDRH3 (FG loop) regions. The RasIn1 and RasIn2 antibody-like ligands were developed as intrabodies that recognize active, GTP-bound *K*- and *H*-Ras and the corresponding G12V mutants [42]. The antibody-like ligands were selected by mRNA display using as antigen *K*-Ras (G12V)-GTPyS and a library comprising a modified 10FnIII scaffold containing a randomized biased CDR1 sequence derived from iDab#6 [10] and a naive randomized sequence of CDR3. Detailed binding analysis demonstrated that RasIn1 and RasIn2 recognized the binding domain of the Raf kinase in activated *H*-RasG12V

A very interesting study resulted in the human IgG intrabody RT11 against Ras mutants [11]. This anti-Ras mutant antibody was engineered from a human antibody previously generated and is internalized through clathrin-mediated endocytosis using heparan sulfate proteoglycan (HSPG) as a receptor and escapes from early endosomes into the cytosol [45]. The intrabody recognizes the GTP-bound active forms of wild-type (WT) KRas, NRas and HRas and their oncogenic mutants with mutations at positions 12, 13 or 61, such as KRasG12D, KRasG12V, KRasG13D, KRasQ61H, HRasG12V and NRasQ61R. Inactive GDP bound Ras WT and mutants were not recognized. To target the intrabody to tumor-associated integrins, the RGD10 cyclic peptide (targeting α_v -integrins) was genetically fused to the light chain of the intrabody (RT11i). It was shown that RT11-i significantly inhibits the tumor growth of oncogenic Ras mutant tumor xenografts in mice.

The work was continued by the same group by engineering a new human IgG intrabody (inRas37) binding to activated GTP-bound Ras mutants with two-fold stronger activity [12]. The endosomal escape was higher compared to the previously published intrabody RT11i and the cytosolic concentrations were two-fold higher in targeted cells. The inhibition of tumor growth was seen in several xenograft tumor mice bearing different preestablished colorectal tumors. Combination of an inhibitor of Yes-associated protein 1 (YAP1, a transcription factor which regulates cancer cell proliferation) and inRas37 showed a synergistic effect in Ras37-sensitive tumor cell lines.

An anti-p21 Ras scFv antibody was generated from a hybridoma recognizing wild-type *H*-p21Ras, *K*-p21Ras and *N*-p21Ras [13] and their mutated variants. The intrabody significantly inhibited the tumor growth of nude mice with established tumors derived from a human colon cancer cell line SW480 or a human liver cancer cell line BEL-7402. Intrabody was directly injected into the tumor by adenoviral gene transfer. Recently it was shown that the genetic fusion of the RGD4C peptide binding the integrin $\alpha_v\beta_3$ to the C-terminus of the anti-p21 Ras scFv, which could not penetrate the cell membrane alone, resulted in the penetration and growth inhibition of the human colon cancer cell line SW480 in vitro [48].

Recently, two other antibody-like ligands of the ankyrin repeat protein type (DARPin) [49] were selected that bind to an allosteric site of GDP- or GTP-bound KRASWT and KRASG12V, inhibiting KRAS nucleotide exchange and dimerization [43]. DARPins were isolated from a phage display library by biopanning using biotinylated KRASG12V.

Non-small cell lung cancer (NSCLC) is heterogeneous and KRAS G12C is the most prevalent of the KRAS mutations. Therapies targeting KRAS are beginning to show clinical potential, most notably with KRASG12C

inhibitors [50][51]. So far, other gene variants have not been targeted in advanced clinical studies. Therefore, using intrabodies that have been selected against intracellular neoantigens, for example against HRASG12V [10][41][42] KRASG12D, KRASG13D, KRASQ61H, KRASG12V, NRASQ61R [11], *H*- and *K*-Ras G12V [12], p21Ras [13] and KRASG12V [43], might be very important for developing advanced phase clinical studies.

Nevertheless, extensive intratumor heterogeneity (ITH) related to genetic diversity both within individual tumors and between primary and metastatic tumors for different cancer types has been newly discovered [52]. Peptide vaccines may only kill a small number of tumor cells if the neoantigens targeted are derived from mutated subclones. Similarly, intrabodies may also be ineffective if new driver neoantigens are elicited during treatment. In the future, it is essential to identify effective and common neoantigens containing tumor driver mutations. It might be possible to apply a mixture of intrabodies targeting different driver neoantigens or combine the intrabody with a small molecule inhibitor against another oncogenic neoantigen identified inside the tumor.

Mutation-derived neoantigens can be recognized by T cells and increased mutated neoantigens (neoantigen burden) influence the survival across diverse types of human cancers. Tumors with both a high clonal neoantigen burden and low neoantigen intratumoral heterogeneity encourage longer survival. Furthermore, a relationship between neoantigen burden and response to immune checkpoint inhibitors has been demonstrated too. Sensitivity to PD-1 and CTLA-4 blockade in patients with advanced NSCLC and melanoma was enhanced in tumors enriched for clonal neoantigens [53][54][55]. In this context, the treatment of anti-neoantigen intrabodies with immune checkpoint inhibitors may increase therapeutic efficiency.

5. Bringing Intrabodies into Cancer Patients: Delivery of Intrabodies with Nanoparticles or AAV

Many methods for intrabody delivery into target cells are available such as the delivery of intrabody genes by plasmids or viral transduction, or using polymeric and dendrimeric nanoparticles embedded with small hydrophobic and hydrophilic drugs, peptides, vaccines and antibodies [56]. The most promising and efficient methods for intrabody transduction and expression in cancer cells are the use of lipid nanoparticles with embedded mRNA [57][58][59][60] and transduction with a new generation of AAVs carrying the intrabody cDNA [61][62] (Figure 2).

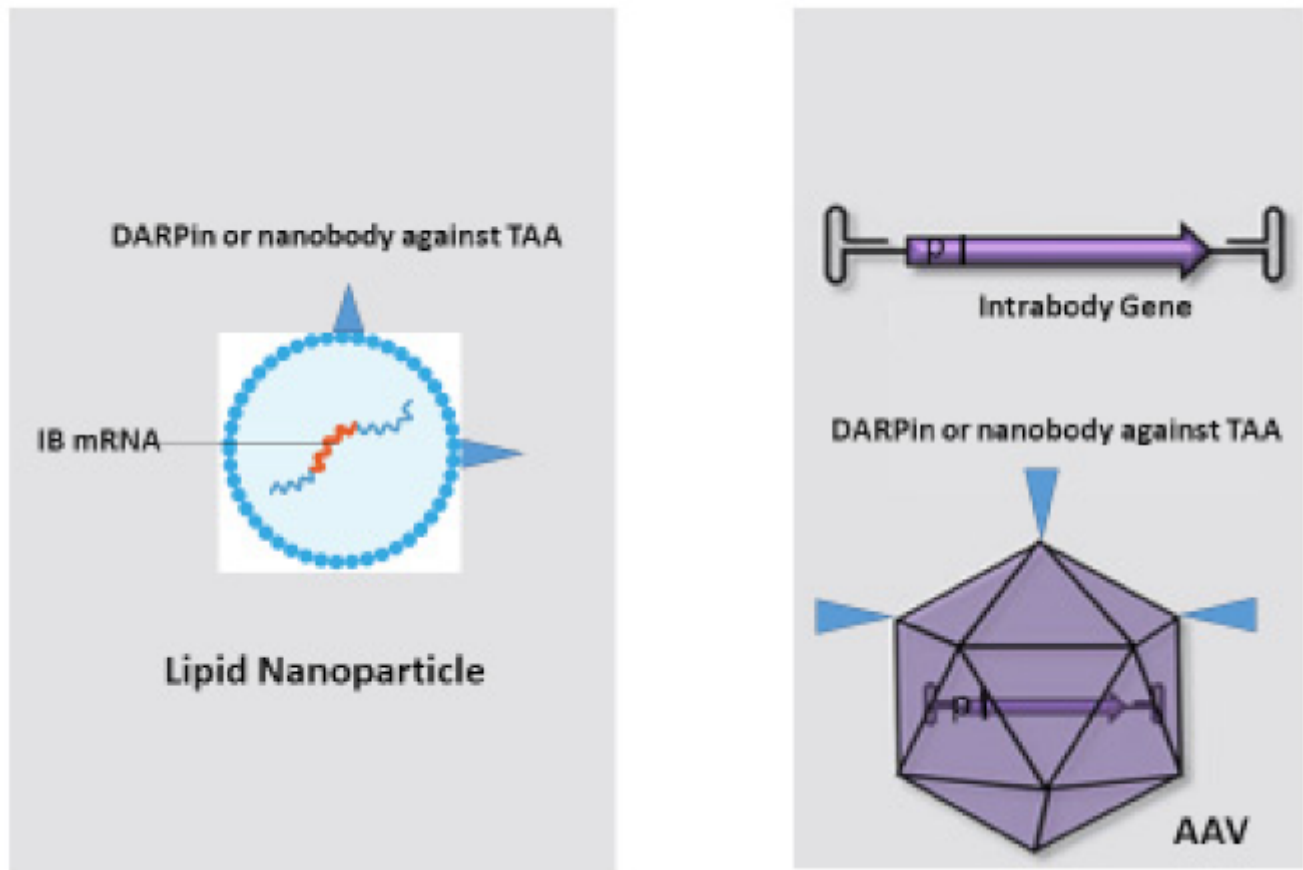


Figure 2. Tumor-cell-specific intrabody transfer with nanoparticle or AAV, *p* = promoter.

References

1. Gerber, H.-P.; Sibener, L.V.; Lee, L.J.; Gee, M. Intracellular targets as source for cleaner targets for the treatment of solid tumors. *Biochem. Pharmacol.* 2019, 168, 275–284.
2. Biernacki, M.A.; Bleakley, M. Neoantigens in Hematologic Malignancies. *Front. Immunol.* 2020, 11, 121.
3. Garcia-Garijo, A.; Fajardo, C.A.; Gros, A. Determinants for Neoantigen Identification. *Front. Immunol.* 2019, 10, 1392.
4. Naran, K.; Nundalall, T.; Chetty, S.; Barth, S. Principles of Immunotherapy: Implications for Treatment Strategies in Cancer and Infectious Diseases. *Front. Microbiol.* 2018, 9, 3158.
5. Yarmarkovich, M.; Marshall, Q.F.; Warrington, J.M.; Premaratne, R.; Farrel, A.; Groff, D.; Li, W.; di Marco, M.; Runbeck, E.; Truong, H.; et al. Cross-HLA targeting of intracellular oncoproteins with peptide-centric CARs. *Nature* 2021, 599, 477–484.
6. Marschall, A.L.; Dubel, S.; Boldicke, T. Recent Advances with ER Targeted Intrabodies. *Adv. Exp. Med. Biol.* 2016, 917, 77–93.

7. Zhang, C.; Ötjengerdes, R.M.; Roewe, J.; Mejias-Estevez, R.; Marschall, A.L.J. Applying Antibodies Inside Cells: Principles and Recent Advances in Neurobiology, Virology and Oncology. *BioDrugs* 2020, 34, 435–462.
8. Marschall, A.L.; Dubel, S.; Boldicke, T. Specific in vivo knockdown of protein function by intrabodies. *MAbs* 2015, 7, 1010–1035.
9. Böldicke, T. Single domain antibodies for the knockdown of cytosolic and nuclear proteins. *Protein Sci.* 2017, 26, 925–945.
10. Tanaka, T.; Williams, R.L.; Rabbitts, T.H. Tumour prevention by a single antibody domain targeting the interaction of signal transduction proteins with RAS. *EMBO J.* 2007, 26, 3250–3259.
11. Shin, S.-M.; Choi, D.-K.; Bae, J.; Kim, J.-S.; Park, S.-W.; Kim, Y.-S. Antibody targeting intracellular oncogenic Ras mutants exerts anti-tumour effects after systemic administration. *Nat. Commun.* 2017, 8, 15090.
12. Shin, S.-M.; Kim, J.-S.; Park, S.-W.; Jun, S.-Y.; Kweon, H.-J.; Choi, D.-K.; Lee, D.; Cho, Y.B.; Kim, Y.-S. Direct targeting of oncogenic RAS mutants with a tumor-specific cytosol-penetrating antibody inhibits RAS mutant-driven tumor growth. *Sci. Adv.* 2020, 6, eaay2174.
13. Yang, J.-L.; Pan, X.-Y.; Zhao, W.-X.; Hu, Q.-C.; Ding, F.; Feng, Q.; Li, G.-Y.; Luo, Y. The antitumor efficacy of a novel adenovirus-mediated anti-p21Ras single chain fragment variable antibody on human cancers in vitro and in vivo. *Int. J. Oncol.* 2016, 48, 1218–1228.
14. Somplatzki, S.; Mühlenhoff, M.; Kröger, A.; Gerardy-Schahn, R.; Böldicke, T. Intrabodies against the Polysialyltransferases ST8SialII and ST8SialIV inhibit Polysialylation of NCAM in rhabdomyosarcoma tumor cells. *BMC Biotechnol.* 2017, 17, 42.
15. Van Impe, K.; Bethuyne, J.; Cool, S.; Impens, F.; Ruano-Gallego, D.; De Wever, O.; Vanloo, B.; Van Troys, M.; Lambein, K.; Boucherie, C.; et al. A nanobody targeting the F-actin capping protein CapG restrains breast cancer metastasis. *Breast Cancer Res.* 2013, 15, R116.
16. Deshane, J.; Siegal, G.P.; Wang, M.; Wright, M.; Bucy, R.P.; Alvarez, R.D.; Curiel, D.T. Transductional efficacy and safety of an intraperitoneally delivered adenovirus encoding an anti-erbB-2 intracellular single-chain antibody for ovarian cancer gene therapy. *Gynecol. Oncol.* 1997, 64, 378–385.
17. Popkov, M.; Jendreyko, N.; McGavern, D.B.; Rader, C.; Barbas, C.F., 3rd. Targeting tumor angiogenesis with adenovirus-delivered anti-Tie-2 intrabody. *Cancer Res.* 2005, 65, 972–981.
18. Jendreyko, N.; Popkov, M.; Rader, C.; Barbas, C.F. Phenotypic knockout of VEGF-R2 and Tie-2 with an intradiabody reduces tumor growth and angiogenesis in vivo. *Proc. Natl. Acad. Sci. USA* 2005, 102, 8293–8298.

19. Chen, J.; Guo, H.; Jiang, H.; Namusamba, M.; Wang, C.; Lan, T.; Wang, T.; Wang, B. A BAP31 intrabody induces gastric cancer cell death by inhibiting p27(kip1) proteasome degradation. *Int. J. Cancer* 2019, 144, 2051–2062.
20. Zhang, W.; Shan, H.; Jiang, K.; Huang, W.; Li, S. A novel intracellular nanobody against HPV16 E6 oncoprotein. *Clin. Immunol.* 2021, 225, 108684.
21. Koo, M.Y.; Park, J.; Lim, J.M.; Joo, S.Y.; Shin, S.-P.; Shim, H.B.; Chung, J.; Kang, D.; Woo, H.A.; Rhee, S.G. Selective inhibition of the function of tyrosine-phosphorylated STAT3 with a phosphorylation site-specific intrabody. *Proc. Natl. Acad. Sci. USA* 2014, 111, 6269–6274.
22. Png, Y.T.; Vinanica, N.; Kamiya, T.; Shimasaki, N.; Coustan-Smith, E.; Campana, D. Blockade of CD7 expression in T cells for effective chimeric antigen receptor targeting of T-cell malignancies. *Blood Adv.* 2017, 1, 2348–2360.
23. Asaadi, Y.; Jouneghani, F.F.; Janani, S.; Rahbarizadeh, F. A comprehensive comparison between camelid nanobodies and single chain variable fragments. *Biomark. Res.* 2021, 9, 87.
24. Böldicke, T.; Somplatzki, S.; Sergeev, G.; Mueller, P.P. Functional inhibition of transitory proteins by intrabody-mediated retention in the endoplasmatic reticulum. *Methods* 2012, 56, 338–350.
25. Böldicke, T. Blocking translocation of cell surface molecules from the ER to the cell surface by intracellular antibodies targeted to the ER. *J. Cell Mol. Med.* 2007, 11, 54–70.
26. Kirschning, C.J.; Dreher, S.; Maaß, B.; Fichte, S.; Schade, J.; Köster, M.; Noack, A.; Lindenmaier, W.; Wagner, H.; Böldicke, T. Generation of anti-TLR2 intrabody mediating inhibition of macrophage surface TLR2 expression and TLR2-driven cell activation. *BMC Biotechnol.* 2010, 10, 31.
27. Böldicke, T.; Tesar, M.; Griesel, C.; Rohde, M.; Gröne, H.J.; Waltenberger, J.; Kollet, O.; Lapidot, T.; Yayon, A.; Weich, H. Anti-VEGFR-2 scFvs for cell isolation. Single-chain antibodies recognizing the human vascular endothelial growth factor receptor-2 (VEGFR-2/flk-1) on the surface of primary endothelial cells and preselected CD34+ cells from cord blood. *Stem Cells* 2001, 19, 24–36.
28. Paolini, F.; Amici, C.; Carosi, M.; Bonomo, C.; Di Bonito, P.; Venuti, A.; Accardi, L. Intrabodies targeting human papillomavirus 16 E6 and E7 oncoproteins for therapy of established HPV-associated tumors. *J. Exp. Clin. Cancer Res.* 2021, 40, 37.
29. Grimmig, T.; Moench, R.; Kreckel, J.; Haack, S.; Rueckert, F.; Rehder, R.; Tripathi, S.; Ribas, C.; Chandraker, A.; Germer, C.T.; et al. Toll Like Receptor 2, 4, and 9 Signaling Promotes Autoregulative Tumor Cell Growth and VEGF/PDGF Expression in Human Pancreatic Cancer. *Int. J. Mol. Sci.* 2016, 17, 2060.
30. Al-Saraireh, Y.M.J.; Sutherland, M.; Springett, B.R.; Freiburger, F.; Morais, G.R.; Loadman, P.M.; Errington, R.J.; Smith, P.J.; Fukuda, M.; Gerardy-Schahn, R.; et al. Pharmacological Inhibition of

- polysialyltransferase ST8Siall Modulates Tumour Cell Migration. PLoS ONE 2013, 8, e73366.
31. Reimer, E.; Somplatzki, S.; Zegenhagen, D.; Hänel, S.; Fels, A.; Bollhorst, T.; Hovest, L.G.; Bauer, S.; Kirschning, C.J.; Böldicke, T. Molecular cloning and characterization of a novel anti-TLR9 intrabody. *Cell. Mol. Biol. Lett.* 2013, 18, 433–446.
 32. Böldicke, T. Single domain antibodies for the knockdown of cytosolic and nuclear proteins. *Protein Sci.* 2017, 26, 925–945.
 33. Kang, E.A.G.; Hu, M.; Ren, H.; Wang, J.; Cheng, X.; Li, R.; Yuan, B.; Balan, Y.; Bai, Z.; Huang, H. VHH212 nanobody targeting the hypoxia-inducible factor 1 α suppresses angiogenesis and potentiates gemcitabine therapy in pancreatic cancer in vivo. *Cancer Biol. Med.* 2021, 18, 772–787.
 34. Shah, V.; Sheppard, B.; Sears, R.; Alani, A.W. Hypoxia: Friend or Foe for drug delivery in Pancreatic Cancer. *Cancer Lett.* 2020, 492, 63–70.
 35. Manning, B.D.; Toker, A. AKT/PKB Signaling: Navigating the Network. *Cell* 2017, 169, 381–405.
 36. D'Agostino, S.; Mazzega, E.; Praček, K.; Piccinin, S.; Pivetta, F.; Armellin, M.; Fortuna, S.; Maestro, R.; de Marco, A. Interference of p53:Twist1 interaction through competing nanobodies. *Int. J. Biol. Macromol.* 2022, 194, 24–31.
 37. De Groof, T.W.M.; Bergkamp, N.D.; Heukers, R.; Giap, T.; Bebelman, M.P.; Haas, R.G.-D.; Piersma, S.R.; Jimenez, C.R.; Garcia, K.C.; Ploegh, H.L.; et al. Selective targeting of ligand-dependent and -independent signaling by GPCR conformation-specific anti-US28 intrabodies. *Nat. Commun.* 2021, 12, 4357.
 38. Valdés-Tresanco, M.S.; Molina-Zapata, A.; Pose, A.G.; Moreno, E. Structural Insights into the Design of Synthetic Nanobody Libraries. *Molecules* 2022, 27, 2198.
 39. English, H.; Hong, J.; Ho, M. Ancient species offers contemporary therapeutics: An update on shark VNAR single domain antibody sequences, phage libraries and potential clinical applications. *Antib. Ther.* 2020, 3, 1–9.
 40. Kashima, D.; Kageoka, M.; Kimura, Y.; Horikawa, M.; Miura, M.; Nakakido, M.; Tsumoto, K.; Nagamune, T.; Kawahara, M. A Novel Cell-Based Intracellular Protein-Protein Interaction Detection Platform (SOLIS) for Multimo-dality Screening. *ACS Synth. Biol.* 2021, 10, 990–999.
 41. Tanaka, T.; Rabbitts, T.H. Intrabodies based on intracellular capture frameworks that bind the RAS protein with high af-finity and impair oncogenic transformation. *EMBO J.* 2003, 22, 1025–1035.
 42. Cetin, M.; Evenson, W.E.; Gross, G.G.; Jalali-Yazdi, F.; Krieger, D.; Arnold, D.; Takahashi, T.T.; Roberts, R.W. RasIns: Genetically Encoded Intrabodies of Activated Ras Proteins. *J. Mol. Biol.* 2017, 429, 562–573.

43. Bery, N.; Legg, S.; Debreczeni, J.; Breed, J.; Embrey, K.; Stubbs, C.; Kolasinska-Zwierz, P.; Barrett, N.; Marwood, R.; Watson, J.; et al. KRAS-specific inhibition using a DARPIn binding to a site in the allosteric lobe. *Nat. Commun.* 2019, 10, 2607.
44. Milburn, M.V.; Tong, L.; Devos, A.M.; Brünger, A.; Yamaizumi, Z.; Nishimura, S.; Kim, S.-H. Molecular Switch for Signal Transduction: Structural Differences Between Active and Inactive Forms of Protooncogenic ras Proteins. *Science* 1990, 247, 939–945.
45. Prior, I.A.; Lewis, P.D.; Mattos, C. A Comprehensive Survey of Ras Mutations in Cancer. *Cancer Res.* 2012, 72, 2457–2467.
46. Visintin, M.; Quondam, M.; Cattaneo, A. The intracellular antibody capture technology: Towards the high-throughput se-lection of functional intracellular antibodies for target validation. *Methods* 2004, 34, 200–214.
47. Choi, D.-K.; Bae, J.; Shin, S.-M.; Shin, J.-Y.; Kim, S.; Kim, Y.-S. A general strategy for generating intact, full-length IgG antibodies that penetrate into the cytosol of living cells. *mAbs* 2014, 6, 1402–1414.
48. Huang, C.C.; Liu, F.R.; Feng, Q.; Pan, X.Y.; Song, S.L.; Yang, J.L. RGD4C peptide mediates anti-p21Ras scFv entry into tumor cells and produces an inhibitory effect on the human colon cancer cell line SW480. *BMC Cancer* 2021, 21, 321.
49. Kawe, M.; Forrer, P.; Amstutz, P.; Plückthun, A. Isolation of Intracellular Proteinase Inhibitors Derived from Designed Ankyrin Repeat Proteins by Genetic Screening. *J. Biol. Chem.* 2006, 281, 40252–40263.
50. Burns, T.F.; Borghaei, H.; Ramalingam, S.S.; Mok, T.S.; Peters, S. Targeting KRAS-Mutant Non-Small-Cell Lung Cancer: One Mutation at a Time, with a Focus on KRAS G12C Mutations. *J. Clin. Oncol.* 2020, 38, 4208–4218.
51. Tang, D.; Kroemer, G.; Kang, R. Oncogenic KRAS blockade therapy: Renewed enthusiasm and persistent challenges. *Mol. Cancer* 2021, 20, 128.
52. Jamal-Hanjani, M.; Thanopoulou, E.; Peggs, K.S.; Quezada, S.A.; Swanton, C. Tumour heterogeneity and im-mune-modulation. *Curr. Opin. Pharmacol.* 2013, 13, 497–503.
53. Chan, T.; Yarchoan, M.; Jaffee, E.; Swanton, C.; Quezada, S.; Stenzinger, A.; Peters, S. Development of tumor mutation burden as an immunotherapy biomarker: Utility for the oncology clinic. *Ann. Oncol.* 2019, 30, 44–56.
54. McGranahan, N.; Furness, A.J.S.; Rosenthal, R.; Ramskov, S.; Lyngaa, R.; Saini, S.K.; Jamal-Hanjani, M.; Wilson, G.A.; Birkbak, N.J.; Hiley, C.T.; et al. Clonal neoantigens elicit T cell immunoreactivity and sensitivity to immune checkpoint blockade. *Science* 2016, 351, 1463–1469.

55. Samstein, R.M.; Lee, C.-H.; Shoushtari, A.N.; Hellmann, M.D.; Shen, R.; Janjigian, Y.Y.; Barron, D.A.; Zehir, A.; Jordan, E.J.; Omuro, A.; et al. Tumor mutational load predicts survival after immunotherapy across multiple cancer types. *Nat. Genet.* 2019, 51, 202–206.
56. Mitchell, M.J.; Billingsley, M.M.; Haley, R.M.; Wechsler, M.E.; Peppas, N.A.; Langer, R. Engineering precision nanoparticles for drug delivery. *Nat. Rev. Drug Discov.* 2021, 20, 101–124.
57. Guevara, M.L.; Persano, F.; Persano, S. Advances in Lipid Nanoparticles for mRNA-Based Cancer Immunotherapy. *Front. Chem.* 2020, 8, 589959.
58. Miao, L.; Zhang, Y.; Huang, L. mRNA vaccine for cancer immunotherapy. *Mol. Cancer* 2021, 20, 41.
59. Tenchov, R.; Bird, R.; Curtze, A.E.; Zhou, Q. Lipid Nanoparticles—From Liposomes to mRNA Vaccine Delivery, a Landscape of Research Diversity and Advancement. *ACS Nano* 2021, 15, 16982–17015.
60. Slastnikova, T.A.; Ulasov, A.V.; Rosenkranz, A.A.; Sobolev, A.S. Targeted Intracellular Delivery of Antibodies: The State of the Art. *Front. Pharmacol.* 2018, 9, 1208.
61. Hacker, U.T.; Bentler, M.; Kaniowska, D.; Morgan, M.; Büning, H. Towards Clinical Implementation of Adeno-Associated Virus (AAV) Vectors for Cancer Gene Therapy: Current Status and Future Perspectives. *Cancers* 2020, 12, 1889.
62. Colón-Thillet, R.; Jerome, K.R.; Stone, D. Optimization of AAV vectors to target persistent viral reservoirs. *Viol. J.* 2021, 18, 85.

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