

# Targeting Autophagy for Cancer Treatment

Subjects: **Oncology**

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Autophagy is a tightly regulated catabolic process that facilitates nutrient recycling from damaged organelles and other cellular components through lysosomal degradation. Deregulation of this process has been associated with the development of several pathophysiological processes, such as cancer and neurodegenerative diseases. In cancer, autophagy has opposing roles, being either cytoprotective or cytotoxic. Thus, deciphering the role of autophagy in each tumor context is crucial. Moreover, autophagy has been shown to contribute to chemoresistance in some patients. In this regard, autophagy modulation has recently emerged as a promising therapeutic strategy for the treatment and chemosensitization of tumors, and has already demonstrated positive clinical results in patients.

autophagy

anticancer therapy

autophagy inhibitors

autophagic cell death

chemoresistance

chemosensitization

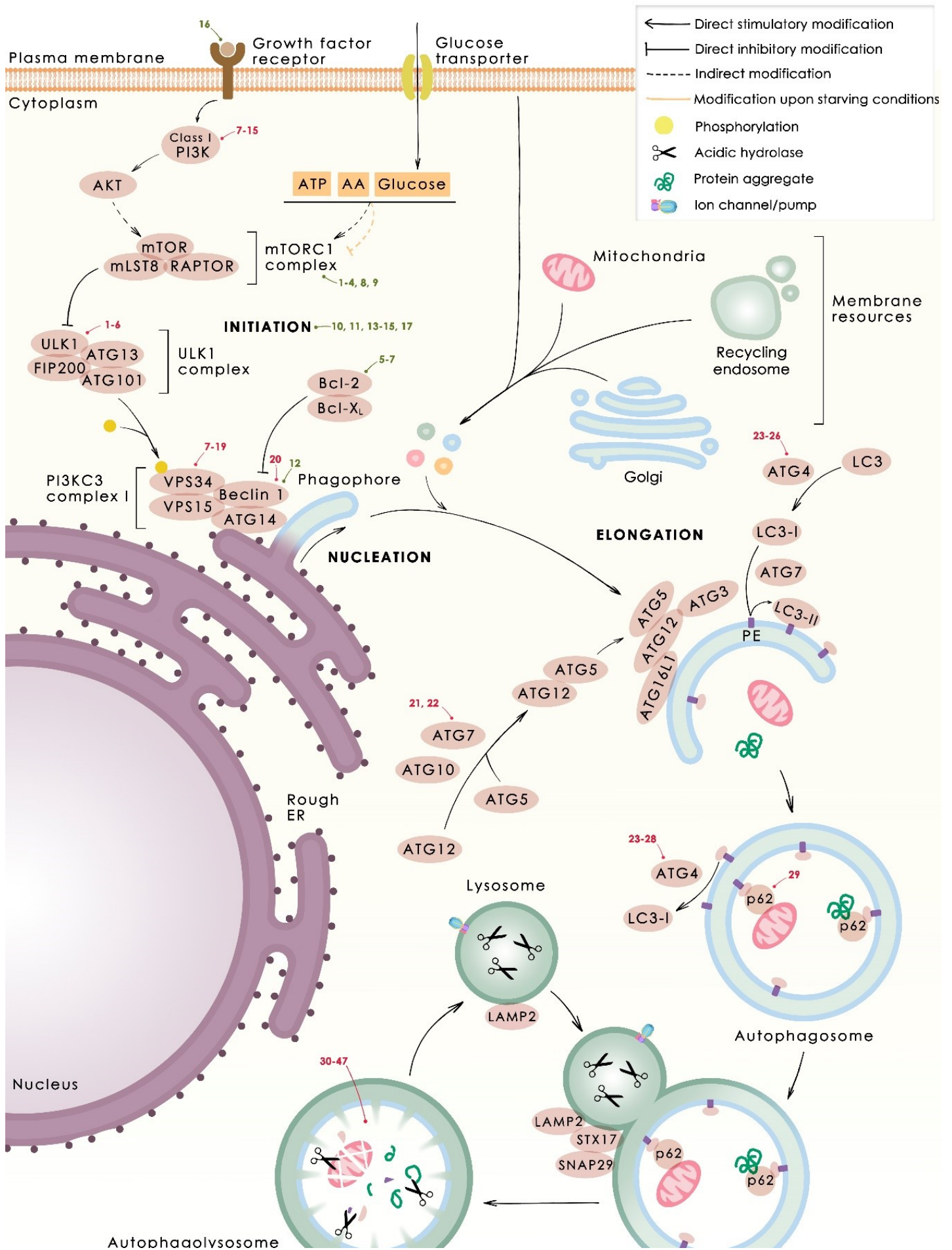
## 1. Introduction

Cellular homeostasis is crucial for cell survival and refers to all processes involved in the maintenance of an internal steady state at the level of the cell. Autophagy is one of the main catabolic mechanisms that contributes to cellular homeostasis, through the degradation and recycling of cytoplasmic components and organelles in the lysosomes <sup>[1][2]</sup>. This process confers the ability to adapt to environmental stresses, preventing cellular damage, and promoting cell survival, even in starving conditions, thus having a main physiologic cytoprotective role. It is a process tightly regulated and its dysfunction has been related to several pathologies, such as neurodegeneration, cancer, or aging <sup>[3]</sup>. Hence, autophagy modulation is emerging as a promising new therapeutic strategy to treat these malignancies <sup>[4]</sup>. Indeed, more than 120 clinical trials related to the process of autophagy were initiated to date. The majority of those target autophagy for cancer treatment, already showing promising results, for instance, using chloroquine or hydroxychloroquine as single agents or in combination therapies <sup>[5][6]</sup>. Nevertheless, the role of autophagy in cancer is somewhat controversial. Cytotoxic or cytoprotective roles have been reported depending on the cellular context <sup>[7]</sup>. Therefore, the deep understanding of autophagy regulation and the identification of its role in each cellular context is crucial for the selection of an appropriate therapeutic intervention involving autophagy modulation in cancer.

## 2. Therapeutic Strategies Targeting Autophagy

Modulation of autophagy has emerged as a promising therapeutic option for cancer treatment. Due to the dual role of autophagy in cancer cells, activators as well as inhibitors have been described as feasible chemotherapeutic agents.

In this section, we compiled different therapeutic interventions targeting autophagy, either for its stimulation or for its inhibition (**Figure 1**).



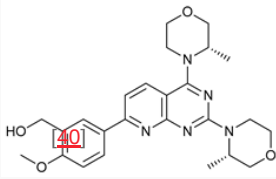
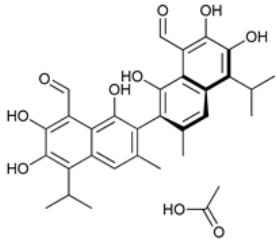
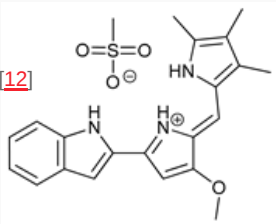
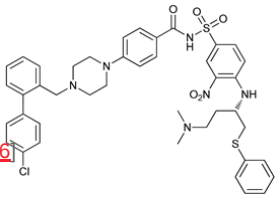
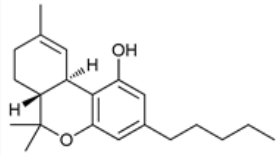
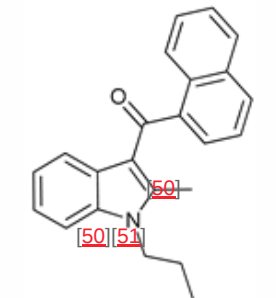
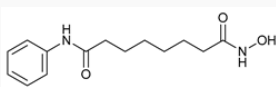
**Figure 1. Mechanism of autophagy.** The phases of the process (nucleation, elongation, maturation, degradation), with the main proteins that participate in each one, are depicted. Autophagy activators (green) and inhibitors (red) are marked where they interfere with the autophagy process. Numbers correspond to those compounds listed in table 1 and 2, respectively.

2.1. Autophagy Stimulation for Cancer Treatment

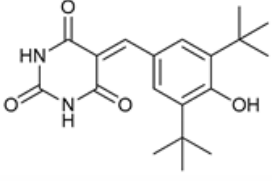
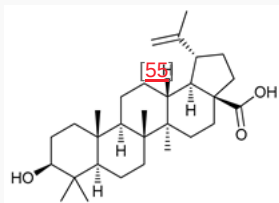
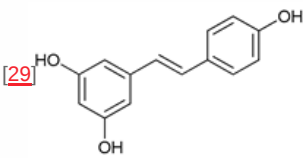
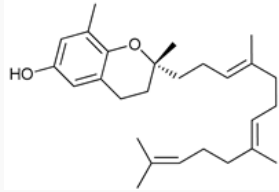
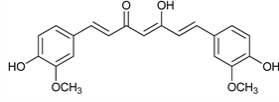
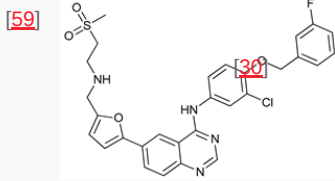
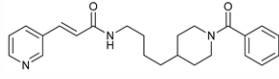
Induction of ACD has become an interesting alternative to overcome resistance to apoptosis and to exploit a caspase independent cell death for cancer treatment. In the following sections, compounds for which the mechanism of action is based on stimulating autophagy are described (Table 1).

Table 1. Autophagy activators.

Mechanism of Action/Type	Name	Structure	Number in Figure 1	Refs.
mTOR Inhibitors	Rapamycin		1	[8][9][10][11]
	Temsirolimus (CCI779)		2	[12][13]
	Everolimus (RAD001)		3	[14][15]

Mechanism of Action/Type	Name	Structure	Number in Figure 1	Refs.
autophagy [39], inhibition of autophagy isolated from <i>Streptococcus</i> [41][42]	AZD8055		4	[16][17]
	(-)-gossypol (AT-101)		5	[43][44] [18][19] [20][21]
	Obatoclax (GX15-070)		6	[22][23] [24]
BH3 Mimetics	ABT-737		7	[25] [17]
	Δ9-Tetrahydrocannabinol (THC)		8	[47] [26][27] [28]
Cannabinoids	JWH-015		9	[29]
Histone Deacetylase Inhibitors	Suberoylanilide hydroxamic acid (SAHA, Vorinostat)		10	[30] [21]

na [19], and colon cancer cells [20]. Obatoclax (GX15-070) is another BH3 mimetic that has shown autophagic-mediated necroptosis in oral squamous cell carcinoma [22], rhabdomyosarcoma cells [23], and acute lymphoblastic leukemia cells [24]. Moreover, obatoclax induced autophagy in adenoid cystic carcinoma [52]

Mechanism of Action/Type	Name	Structure <sup>[53]</sup>	Number in Figure 1 <sup>[25]</sup>	Refs.	effectivity
Cannabinoids are a group of chemical compounds present in the plant cannabis ( <i>Cannabis sativa</i> ) [54].  [27][28]	MHY2256		11	[31]	CB <sub>1</sub> and CB <sub>2</sub> receptors
	Betulinic acid		12	[32]	has been shown to induce canonical apoptosis or growth inhibition through
	Resveratrol		13	[33]	Wnt/b-catenin signaling pathway
	δ-Tocotrienol		14	[34]	induced by the inhibition of autophagy
	Curcumin		15	[35]	class III phosphatase
Others	Lapatinib		16	[36][37]	in both in vitro and in vivo
	APO866		17	[38]	

Some natural compounds have shown promising anticancer activities based on autophagy stimulation. Betulinic acid is a pentacyclic triterpenoid derived from widespread plants that has shown to induce ACD in multiple myeloma cells with high levels of Bcl-2 expression. This derivative acts as an attenuator for mitochondrial-mediated apoptosis, promoting ACD by inducing Beclin-1 phosphorylation [32]. Resveratrol, a polyphenol compound widely found in plants, has been shown to inhibit cell proliferation in breast cancer stem-like cells via suppressing the Wnt/b-catenin signaling pathway [33]. This pathway, which regulates critical genes in tissue development and homeostasis, is aberrantly activated in many cancers and its inhibition has been reported to be related with autophagy processes [33][60]. δ-Tocotrienol is one of the four isomers that comprises vitamin E that has shown cytotoxic effects against prostate cancer cells in vitro through autophagy activation via ER stress [34]. Curcumin is a

major constituent of *Curcuma longa* (turmeric) that induces autophagy, which has been shown to elicit a dual role protecting or leading to cell death depending on the duration of the treatment and concentration used [35].

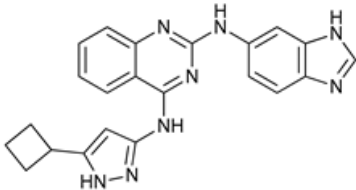
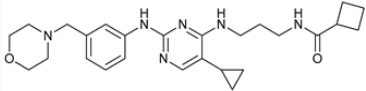
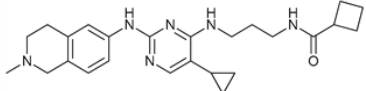
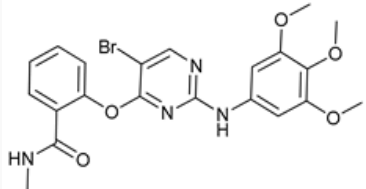
2.1.6. Others

Other compounds have been reported to induce ACD in cancer. For example, lapatinib is a small molecule tyrosine kinase inhibitor, targeting epidermal growth factor receptors that is capable of inducing ACD in hepatocellular carcinoma [36] and in acute leukemia cell lines [37]. APO866 is an inhibitor of nicotinamide adenine dinucleotide (NAD) biosynthesis that has shown anticancer activity through induction of ACD in cells from hematological malignancies [38].

2.2. Autophagy Inhibition for Cancer Treatment

In several tumors, autophagy has a protective role; therefore, its inhibition could be an interesting approach for tumor treatment. There are several autophagy inhibitors that block the process of autophagy at different steps, which we detail below (Table 2).

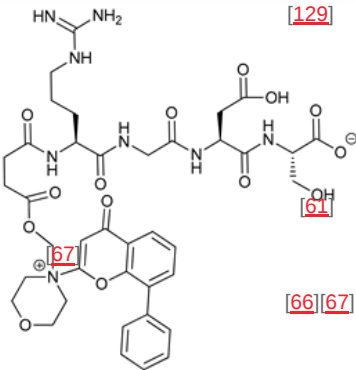
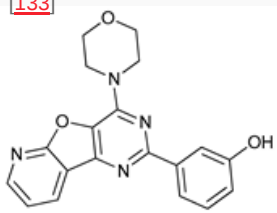
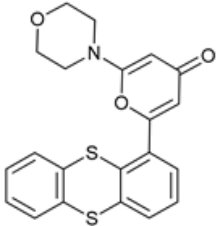
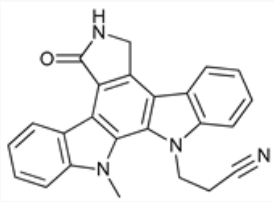
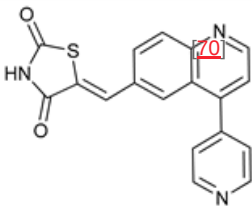
Table 2. Autophagy inhibitors.

Mechanism of Action	Name	Structure	Number in Figure 1	Refs.
ULK Inhibitors	Compound 6		1	[61]
	MRT68921		2	[62][63]
	MRT67307		3	[62][63]
	SBI-0206965		4	[64][65][66][67]

Mechanism of Action	Name	Structure	Number in Figure 1	Refs.
Pan PI3k Inhibitors	ULK-100		5	[68]
	ULK-101		6	[68]
	3MA		7	[69][70][71]
	3 MA derivatives		8	[72]
	Wortmannin		9	[73][74]
	LY294002		10	[75]

2.2.1. ULK Inhibitors

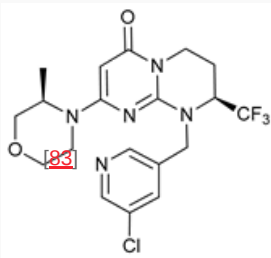
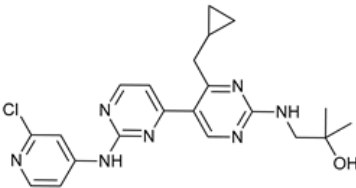
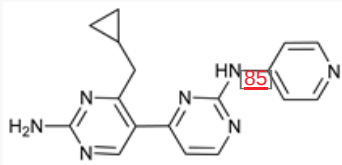
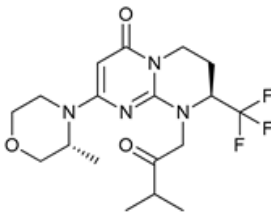
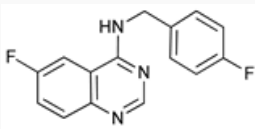
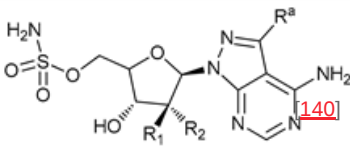


Mechanism of Action	Name	Structure	Number in Figure 1	Refs.
<a href="#">[128]</a> <a href="#">[129]</a> <a href="#">[64]</a> <a href="#">[130]</a> <a href="#">[131]</a> <a href="#">[132]</a> <a href="#">[65]</a> <a href="#">[66]</a> <a href="#">[65]</a> <a href="#">[64]</a>	SF1126		11	<a href="#">[76]</a> <a href="#">[77]</a>
<a href="#">[68]</a>	PI103		12	<a href="#">[78]</a>
<a href="#">[134]</a>	KU55933		13	<a href="#">[79]</a>
<a href="#">[135]</a>	Gö6976		14	<a href="#">[79]</a> <a href="#">[69]</a>
autophagy. Under starving presence of nutrients it pro reduces the expression of effective at high concentra derivatives have been syntl	GSK1059615		15	<a href="#">[80]</a> <a href="#">[81]</a> <a href="#">[71]</a>

of PI3Ks [\[73\]](#)[\[74\]](#). LY294002 is a synthetic small molecule [\[75\]](#) with poor solubility and short half-life. A conjugate analog of LY294002, named SF1126, was designed to accumulate in integrin expressing tissues, improving LY294002 solubility and pharmacokinetic, favoring its accumulation in the tumor site and showing antitumor and antiangiogenic properties in mouse models [\[76\]](#)[\[77\]](#). Other non-selective Pan PI3K inhibitors are PI103 [\[78\]](#), KU55933, Gö6976 [\[79\]](#), and GSK1059615 [\[80\]](#)[\[81\]](#)[\[136\]](#).

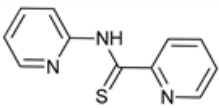
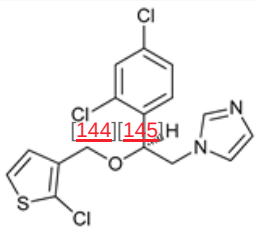
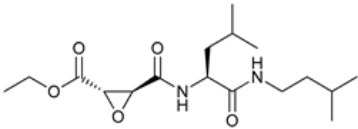
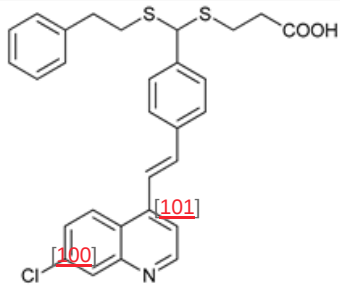
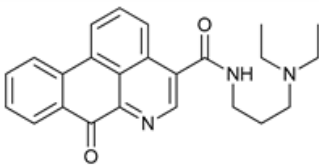
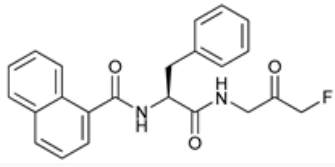
2.2.3. VPS34 (PI3KC3) Complex Inhibitors

VPS34 is a PI3KC3 that transforms PI to PI3P. VPS34 forms a complex with several subunits needed for its activation, such as VPS15 (also known as p150), ATG14, and Beclin-1. Autophagy can be blocked by inhibition of

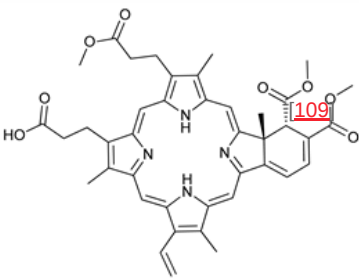
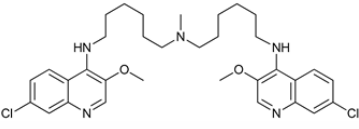
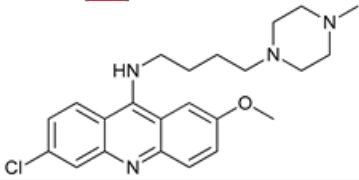
Mechanism of Action	Name	Structure	Number in Figure 1	Refs.	
VPS34 (PI3KC3) Inhibitors	<a href="#">[78]</a> <a href="#">[82]</a> SAR405 <a href="#">[84]</a>		16	<a href="#">[78]</a>	inhibitor I and II, vation or ore than et unique e against oted that function, dosomal may also
	<a href="#">[137]</a> <a href="#">[138]</a> VPS34-IN1		17	<a href="#">[78]</a> <a href="#">[82]</a>	
	PIK-III		18	<a href="#">[83]</a>	ndirectly through
	Compound 31		19	<a href="#">[84]</a>	I factors, atophagy
ATG Inhibitors	Spautin-1		20 <a href="#">[86]</a>	<a href="#">[85]</a>	ABARAP. of micro
	ATG7 inhibitor		21 <a href="#">[139]</a>	WO2018/089786 <a href="#">[140]</a>	ansion of ortant for e a good screened
<a href="#">[141]</a>	ATG7 inhibitor, miR154	UAGGUUAUCCGUGUUGCCUUCG	22	<a href="#">[86]</a>	not only

ume of the autophagosomes, which is accompanied by suppression of tumor growth in  
eous mouse model [\[87\]](#). Tioconazole is an antifungal drug that binds to the active site of  
ATG4 blocking autophagy flux reducing cell viability and sensitizing tumor cells to doxorubicin in a xenograft mouse  
model [\[88\]](#). Other ATG4B inhibitors that suppress autophagy in cell lines and in vivo inhibiting cell proliferation are  
UAMC-2526, a derivative of benzotropolones stable in plasma [\[89\]](#), and LV-320, a styrylquinoline [\[90\]](#).

It should be noticed that the roles of ATG4B in cancer are not well understood and some of the ATG4 inhibitors  
showed only inhibition in LC3-PE delipidation, but not in the autophagosome formation such as S130 [\[91\]](#) and FMK-  
9a [\[92\]](#)[\[93\]](#)[\[94\]](#). Additionally, some studies are focused on the evaluation of different markers that may predict the

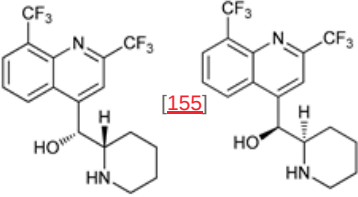
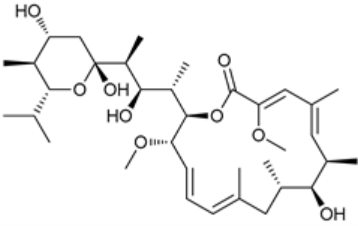
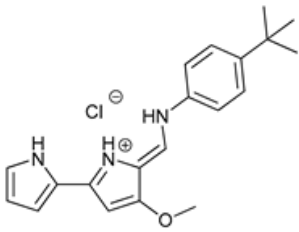
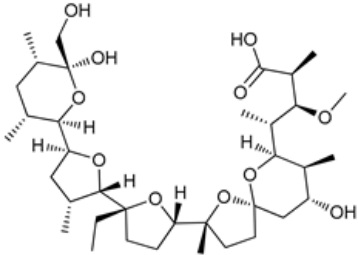
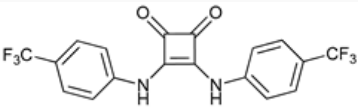
Mechanism of Action [143]	[142] Name	Structure	Number in Figure 1	Refs.
[96][98] [146]	NSC185058		23	[87]
	Tioconazol		24	[88] [144][145]
	[97]UAMC-2526		25	[89]
	LV320		26	[90] [100][101]
	+ S130		27	[99] [91]
	FMK-9a [149]		28	[92][93][94]

has been considered safe, 7.5% of patients [148] and c [150]. This toxicity limitation, along with inconsistencies in the results obtained in the clinic, have led to the study of new and more potent autophagy inhibitors [151]. Thus, CQ analogs that exert more potent autophagy inhibitory activity have been synthesized. Lys05 is a dimeric analog of CQ that accumulates within acidic organelles, including lysosomes, more potently than HCQ [102]. DQ661, a dimeric quinacrine (DQ), not only inhibits lysosomal catabolism, including autophagy, but also targets palmitoyl-protein thioesterase-1, resulting in the inhibition of mTORC1 signaling. DQ661 has shown effects on tumor mouse models alone and it also overcame resistance to gemcitabine [104]. Another antimalaria compound found to inhibit autophagy with antitumoral properties is VATG-027 [105]. On the other hand, mefloquine is also accumulated in lysosomes disrupting autophagy, it induces apoptosis and inhibits multidrug resistance protein1 (MDR1) being effective in multidrug-resistant tumor cells [107]. Mefloquine sensitizes chronic myeloid leukemia (CML) cells derived from patients in chronic phase to TK inhibitors showing selectivity for stem/progenitor tumoral cells to normal cells [106].

Mechanism of Action	Name	Structure	Number in Figure 1	Refs.
+ Autophagy Formation  +  2+	[110]  Verteporfin		29	[109]  [95][96][97][98] [111][112]
	Lysosome Inhibitors Lysosomotropic Agents	Chloroquine	30	[99][100] [114]
		Hydroxychloroquine	31	[116] [101]
	[117]  [118]	Lys05	32	[102][103]
[119]	DQ661		33	[104] [152]
	VATG-027		34	[120] [105] [122]

to inhibit autophagy by disruption of lysosomal function, which confers tumors to [124].

gested that the anti-tumor effects of lysosomal inhibitors may be independent of autophagy inhibition since they also interfere in other cellular mechanisms producing non-autophagy related effects [153][154][155][156][157][158][159][160]. Remarkably, disruption of the lysosomes not only blocks autophagy, but lysosomal permeabilization releases proteases such as cathepsins that are active at cytosolic pH and participate in apoptosis and apoptosis-like and necrosis-like cell death [161][162][163]. Additionally, lysosomes also participate in tumor invasion, hence, these inhibitors have shown to be effective against metastasis [103][164][165][166], targeting cancer stem cells [167], and inducing tumor vessel normalization [168].

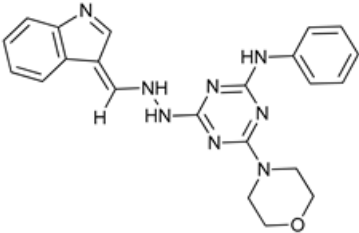
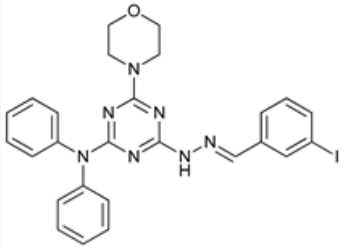
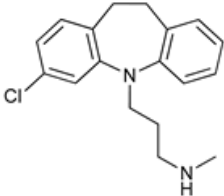
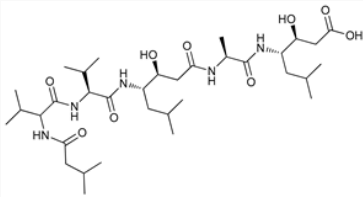
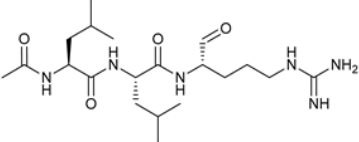
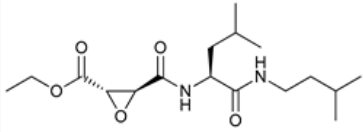
Mechanism of Action	Name	Structure	Number in Figure 1	Refs.
Drug resistance in sarcoma virus	[164]  Mefloquine		35	[106][107] [169]
	Ganoderma lucidum polysaccharide (GLP)		36	[108][109][110]
Vacuolar H <sup>+</sup> ATPase Inhibitors	Bafilomycin A1		37	[111][112][113]
Ionophores	Tambjamines		38	[114]
	Monensin		39	[115]
	Squaramides		40	[116]

; Han, L.; Weng, J.; Wang, K.; Chen, T. Rapamycin inhibits proliferation and induces autophagy in human neuroblastoma cells. Biosci. Rep. 2018, 38, 1–8.

10. R.Y.; Pei, H.L.; Gu, W.D.; Huang, J.; Wang, Z.G. Autophagic inhibitor attenuates rapamycin-induced inhibition of proliferation in cultured A549 lung cancer cells. Eur. Rev. Med. Pharmacol. Sci. 2014, 18, 806–810.

11. Xie, Z.G.; Xie, Y.; Dong, Q.R. Inhibition of the mammalian target of rapamycin leads to autophagy activation and cell death of MG63 osteosarcoma cells. Oncol. Lett. 2013, 6, 1465–1469.

12. Liu, W.; Huang, S.; Chen, Z.; Wang, H.; Wu, H.; Zhang, D. Temsirolimus, the mTOR inhibitor, induces autophagy in adenoid cystic carcinoma: In vitro and in vivo. Pathol. Res. Pract. 2014, 210, 764–769.

Mechanism of Action	Name	Structure	Number in Figure 1	Refs.
Inhibition of Autophagosome-Lysosome Fusion	WX8 family		41	[117]
	Vacuolin-1		42	[118]
	Desmethyldomipramine		43	[119]
Acid Protease Inhibitors	Pepstatin A		44	[120]
	Leupeptin		45	[120]
	E64d		46	[121]
Others	Nanoparticles		47	[122][123][124]

22. Sulkshane, P.; Teni, T. BH3 mimetic Obatoclax (GX15-070) mediates mitochondrial stress predominantly via MCL-1 inhibition and induces autophagy-dependent necroptosis in human oral cancer cells. *Oncotarget* 2017, 8, 60060–60079.

23. Basit, F.; Cristofanon, S.; Fulda, S. Obatoclax (GX15-070) triggers necroptosis by promoting the assembly of the necrosome on autophagosomal membranes. *Cell Death Differ.* 2013, 20, 1161–1173.

24. Bonapace, L.; Bornhauser, B.C.; Schmitz, M.; Cario, G.; Ziegler, U.; Niggli, F.K.; Schäfer, B.W.; Schrappe, M.; Stanulla, M.; Bourquin, J.P. Induction of autophagy-dependent necroptosis is required for childhood acute lymphoblastic leukemia cells to overcome glucocorticoid resistance. *J. Clin. Investig.* 2010, 120, 1310–1323.
25. Yao, X.; Li, X.; Zhang, D.; Xie, Y.; Sun, B.; Li, H.; Sun, L.; Zhang, X. B-cell lymphoma 2 inhibitor ABT-737 induces Beclin1- and reactive oxygen species-dependent autophagy in Adriamycin-resistant human hepatocellular carcinoma cells. *Tumor Biol.* 2017, 39, 1–12.
26. Armstrong, J.L.; Hill, D.S.; McKee, C.S.; Hernandez-Tiedra, S.; Lorente, M.; Lopez-Valero, I.; Anagnostou, M.E.; Babatunde, F.; Corazzari, M.; Redfern, C.P.F.; et al. Exploiting cannabinoid-induced cytotoxic autophagy to drive melanoma cell death. *J. Investig. Dermatol.* 2015, 135, 1629–1637.
27. Hernández-Tiedra, S.; Fabriàs, G.; Dávila, D.; Salanueva, Í.J.; Casas, J.; Montes, L.R.; Antón, Z.; García-Taboada, E.; Salazar-Roa, M.; Lorente, M.; et al. Dihydroceramide accumulation mediates cytotoxic autophagy of cancer cells via autolysosome destabilization. *Autophagy* 2016, 12, 2213–2229.
28. Salazar, M.; Carracedo, A.; Salanueva, Í.J.; Hernández-Tiedra, S.; Lorente, M.; Egia, A.; Vázquez, P.; Blázquez, C.; Torres, S.; García, S.; et al. Cannabinoid action induces autophagy-mediated cell death through stimulation of ER stress in human glioma cells. *J. Clin. Investig.* 2009, 119, 1359–1372.
29. Vara, D.; Salazar, M.; Olea-Herrero, N.; Guzmán, M.; Velasco, G.; Díaz-Laviada, I. Anti-tumoral action of cannabinoids on hepatocellular carcinoma: Role of AMPK-dependent activation of autophagy. *Cell Death Differ.* 2011, 18, 1099–1111.
30. Han, H.; Li, J.; Feng, X.; Zhou, H.; Guo, S.; Zhou, W. Autophagy-related genes are induced by histone deacetylase inhibitor suberoylanilide hydroxamic acid via the activation of cathepsin B in human breast cancer cells. *Oncotarget* 2017, 8, 53352–53365.
31. De, U.; Son, J.Y.; Sachan, R.; Park, Y.J.; Kang, D.; Yoon, K.; Lee, B.M.; Kim, I.S.; Moon, H.R.; Kim, H.S. A new synthetic histone deacetylase inhibitor, MHY2256, induces apoptosis and autophagy cell death in endometrial cancer cells via p53 acetylation. *Int. J. Mol. Sci.* 2018, 19, 2743.
32. Zhou, H.; Luo, W.; Zeng, C.; Zhang, Y.; Wang, L.; Yao, W.; Nie, C. PP2A mediates apoptosis or autophagic cell death in multiple myeloma cell lines. *Oncotarget* 2017, 8, 80770–80789.
33. Fu, Y.; Chang, H.; Peng, X.; Bai, Q.; Yi, L.; Zhou, Y.; Zhu, J.; Mi, M. Resveratrol inhibits breast cancer stem-like cells and induces autophagy via suppressing Wnt/ $\beta$ -catenin signaling pathway. *PLoS ONE* 2014, 9, e102535.

34. Fontana, F.; Moretti, R.M.; Raimondi, M.; Marzagalli, M.; Beretta, G.; Procacci, P.; Sartori, P.; Montagnani Marelli, M.; Limonta, P.  $\delta$ -Tocotrienol induces apoptosis, involving endoplasmic reticulum stress and autophagy, and paraptosis in prostate cancer cells. *Cell Prolif.* 2019, 52, 1–15.
35. Deng, Q.; Liang, L.; Liu, Q.; Duan, W.; Jiang, Y.; Zhang, L. Autophagy is a major mechanism for the dual effects of curcumin on renal cell carcinoma cells. *Eur. J. Pharmacol.* 2018, 826, 24–30.
36. Chen, Y.J.; Chi, C.W.; Su, W.C.; Huang, H.L. Lapatinib induces autophagic cell death and inhibits growth of human hepatocellular carcinoma. *Oncotarget* 2014, 5, 4845–4854.
37. Chen, Y.J.; Fang, L.W.; Su, W.C.; Hsu, W.Y.; Yang, K.C.; Huang, H.L. Lapatinib induces autophagic cell death and differentiation in acute myeloblastic leukemia. *Onco. Targets. Ther.* 2016, 9, 4453–4464.
38. Ginet, V.; Puyal, J.; Rummel, C.; Aubry, D.; Breton, C.; Cloux, A.J.; Majjigapu, S.R.; Sordat, B.; Vogel, P.; Bruzzone, S.; et al. A critical role of autophagy in antileukemia/lymphoma effects of APO866, an inhibitor of NAD biosynthesis. *Autophagy* 2014, 10, 603–617.
39. Hua, H.; Kong, Q.; Zhang, H.; Wang, J.; Luo, T.; Jiang, Y. Targeting mTOR for cancer therapy. *J. Hematol. Oncol.* 2019, 12, 71.
40. Noda, T.; Ohsumi, Y. Tor, a phosphatidylinositol kinase homologue, controls autophagy in yeast. *J. Biol. Chem.* 1998, 273, 3963–3966.
41. Sehgal, S.N.; Baker, H.; Vézina, C. Rapamycin (Ay-22,989), a New Antifungal Antibiotic. II. Fermentation, Isolation and Characterization. *J. Antibiot. (Tokyo).* 1975, 28, 727–732.
42. Vézina, C.; Kudelski, A.; Sehgal, S.N. Rapamycin (AY 22,989) A NEW ANTIFUNGAL ANTIBIOTIC. *J. Antibiot. (Tokyo).* 1975, 28, 721–726.
43. Pattingre, S.; Espert, L.; Biard-Piechaczyk, M.; Codogno, P. Regulation of macroautophagy by mTOR and Beclin 1 complexes. *Biochimie* 2008, 90, 313–323.
44. Benjamin, D.; Colombi, M.; Moroni, C.; Hall, M.N. Rapamycin passes the torch: A new generation of mTOR inhibitors. *Nat. Rev. Drug Discov.* 2011, 10, 868–880.
45. Chiarini, F.; Evangelisti, C.; Lattanzi, G.; McCubrey, J.A.; Martelli, A.M. Advances in understanding the mechanisms of evasive and innate resistance to mTOR inhibition in cancer cells. *Biochim. Biophys. Acta. Mol. Cell Res.* 2019, 1866, 1322–1337.
46. Mao, B.; Gao, S.; Weng, Y.; Zhang, L.; Zhang, L. Design, synthesis, and biological evaluation of imidazo[1,2-b]pyridazine derivatives as mTOR inhibitors. *Eur. J. Med. Chem.* 2017, 129, 135–150.
47. Chen, Y.; Lee, C.H.; Tseng, B.Y.; Tsai, Y.H.; Tsai, H.W.; Yao, C.L.; Tseng, S.H. AZD8055 exerts antitumor effects on colon cancer cells by inhibiting mTOR and cell-cycle progression. *Anticancer Res.* 2018, 38, 1445–1454.



48. Carew, J.S.; Kelly, K.R.; Nawrocki, S.T. Mechanisms of mTOR inhibitor resistance in cancer therapy. *Target. Oncol.* 2011, 6, 17–27.
49. Merino, D.; Kelly, G.L.; Lessene, G.; Wei, A.H.; Roberts, A.W.; Strasser, A. BH3-Mimetic Drugs: Blazing the Trail for New Cancer Medicines. *Cancer Cell* 2018, 34, 879–891.
50. Opydo-Chanek, M.; Gonzalo, O.; Marzo, I. Multifaceted anticancer activity of BH3 mimetics: Current evidence and future prospects. *Biochem. Pharmacol.* 2017, 136, 12–23.
51. Czabotar, P.E.; Lessene, G.; Strasser, A.; Adams, J.M. Control of apoptosis by the BCL-2 protein family: Implications for physiology and therapy. *Nat. Rev. Mol. Cell Biol.* 2014, 15, 49–63.
52. Liang, L.Z.; Ma, B.; Liang, Y.J.; Liu, H.C.; Zhang, T.H.; Zheng, G.S.; Su, Y.X.; Liao, G.Q. Obatoclox induces Beclin 1- and ATG5-dependent apoptosis and autophagy in adenoid cystic carcinoma cells. *Oral Dis.* 2015, 21, 470–477.
53. Koehler, B.C.; Jassowicz, A.; Scherr, A.L.; Lorenz, S.; Radhakrishnan, P.; Kautz, N.; Elssner, C.; Weiss, J.; Jaeger, D.; Schneider, M.; et al. Pan-Bcl-2 inhibitor Obatoclox is a potent late stage autophagy inhibitor in colorectal cancer cells independent of canonical autophagy signaling. *BMC Cancer* 2015, 15, 1–11.
54. Śledziński, P.; Zeyland, J.; Słomski, R.; Nowak, A. The current state and future perspectives of cannabinoids in cancer biology. *Cancer Med.* 2018, 7, 765–775.
55. Costa, L.; Amaral, C.; Teixeira, N.; Correia-Da-Silva, G.; Fonseca, B.M. Cannabinoid-induced autophagy: Protective or death role? *Prostaglandins Other Lipid Mediat.* 2016, 122, 54–63.
56. Mrakovcic, M.; Kleinheinz, J.; Fröhlich, L.F. Histone deacetylase inhibitor-induced autophagy in tumor cells: Implications for p53. *Int. J. Mol. Sci.* 2017, 18, 1883.
57. Newbold, A.; Falkenberg, K.J.; Prince, H.M.; Johnstone, R.W. How do tumor cells respond to HDAC inhibition? *FEBS J.* 2016, 283, 4032–4046.
58. Mrakovcic, M.; Bohner, L.; Hanisch, M.; Fröhlich, L.F. Epigenetic targeting of autophagy via HDAC inhibition in tumor cells: Role of p53. *Int. J. Mol. Sci.* 2018, 19, 3952.
59. Mann, B.S.; Johnson, J.R.; Cohen, M.H.; Justice, R.; Pazdur, R. FDA Approval Summary: Vorinostat for Treatment of Advanced Primary Cutaneous T-Cell Lymphoma. *Oncologist* 2007, 12, 1247–1252.
60. Wang, J.; Ren, X.R.; Piao, H.; Zhao, S.; Osada, T.; Premont, R.T.; Mook, R.A.; Morse, M.A.; Lysterly, H.K.; Chen, W. Niclosamide-induced Wnt signaling inhibition in colorectal cancer is mediated by autophagy. *Biochem. J.* 2019, 476, 535–546.
61. Lazarus, M.B.; Novotny, C.J.; Shokat, K.M. Structure of the human autophagy initiating kinase ULK1 in complex with potent inhibitors. *ACS Chem. Biol.* 2015, 10, 257–261.

62. Skah, S.; Richartz, N.; Duthil, E.; Gilljam, K.M.; Bindesbøll, C.; Naderi, E.H.; Eriksen, A.B.; Ruud, E.; Dirdal, M.M.; Simonsen, A.; et al. cAMP-mediated autophagy inhibits DNA damage-induced death of leukemia cells independent of p53. *Oncotarget* 2018, 9, 30434–30449.
63. Petherick, K.J.; Conway, O.J.L.; Mpamhanga, C.; Osborne, S.A.; Kamal, A.; Saxty, B.; Ganley, I.G. Pharmacological inhibition of ULK1 kinase blocks mammalian target of rapamycin (mTOR)-dependent autophagy. *J. Biol. Chem.* 2015, 290, 11376–11383.
64. Lu, J.; Zhu, L.; Zheng, L.P.; Cui, Q.; Zhu, H.H.; Zhao, H.; Shen, Z.J.; Dong, H.Y.; Chen, S.S.; Wu, W.Z.; et al. Overexpression of ULK1 Represents a Potential Diagnostic Marker for Clear Cell Renal Carcinoma and the Antitumor Effects of SBI-0206965. *EBioMedicine* 2018, 34, 85–93.
65. Dower, C.M.; Bhat, N.; Gebru, M.T.; Chen, L.; Wills, C.A.; Miller, B.A.; Wang, H.-G. Targeted Inhibition of ULK1 Promotes Apoptosis and Suppresses Tumor Growth and Metastasis in Neuroblastoma. *Mol. Cancer Ther.* 2018, 17, 2365–2376.
66. Tang, F.; Hu, P.; Yang, Z.; Xue, C.; Gong, J.; Sun, S.; Shi, L.; Zhang, S.; Li, Z.; Yang, C.; et al. SBI0206965, a novel inhibitor of Ulk1, suppresses non-small cell lung cancer cell growth by modulating both autophagy and apoptosis pathways. *Oncol. Rep.* 2017, 37, 3449–3458.
67. Egan, D.F.; Chun, M.G.H.; Vamos, M.; Zou, H.; Rong, J.; Miller, C.J.; Lou, H.J.; Raveendra-Panickar, D.; Yang, C.-C.; Sheffler, D.J.; et al. Small molecule inhibition of the autophagy kinase ULK1 and identification of ULK1 substrates. *Mol. Cell* 2015, 59, 285–297.
68. Martin, K.R.; Celano, S.L.; Solitro, A.R.; Gunaydin, H.; Scott, M.; O'Hagan, R.C.; Shumway, S.D.; Fuller, P.; MacKeigan, J.P. A Potent and Selective ULK1 Inhibitor Suppresses Autophagy and Sensitizes Cancer Cells to Nutrient Stress. *iScience* 2018, 8, 74–84.
69. Seglen, P.O.; Gordon, P.B. 3-Methyladenine: Specific inhibitor of autophagic/lysosomal protein degradation in isolated rat hepatocytes. *Proc. Natl. Acad. Sci. USA* 1982, 79, 1889–1892.
70. Wu, Y.T.; Tan, H.L.; Shui, G.; Bauvy, C.; Huang, Q.; Wenk, M.R.; Ong, C.N.; Codogno, P.; Shen, H.M. Dual role of 3-methyladenine in modulation of autophagy via different temporal patterns of inhibition on class I and III phosphoinositide 3-kinase. *J. Biol. Chem.* 2010, 285, 10850–10861.
71. Zou, Z.; Zhang, J.; Zhang, H.; Liu, H.; Li, Z.; Cheng, D.; Chen, J.; Liu, L.; Ni, M.; Zhang, Y.; et al. 3-Methyladenine can depress drug efflux transporters via blocking the PI3K-AKT-mTOR pathway thus sensitizing MDR cancer to chemotherapy. *J. Drug Target.* 2014, 22, 839–848.
72. Wu, Y.; Wang, X.; Guo, H.; Zhang, B.; Zhang, X.B.; Shi, Z.J.; Yu, L. Synthesis and screening of 3-MA derivatives for autophagy inhibitors. *Autophagy* 2013, 9, 595–603.
73. Powis, G.; Bonjouklian, R.; Berggren, M.M.; Gallegos, A.; Abraham, R.; Ashendel, C.; Zalkow, L.; Matter, W.F.; Dodge, J.; Grindey, G.; et al. Wortmannin, a Potent and Selective Inhibitor of Phosphatidylinositol-3-kinase. *Cancer Res.* 1994, 54, 2419–2423.

74. Thelen, M.; Wymann, M.P.; Langen, H. Wortmannin binds specifically to 1-phosphatidylinositol 3-kinase while inhibiting guanine nucleotide-binding protein-coupled receptor signaling in neutrophil leukocytes. *Proc. Natl. Acad. Sci. USA* 1994, 91, 4960–4964.
75. Vlahos, C.J.; Matter, W.F.; Hui, K.Y.; Brown, R.F. A Specific Inhibitor of Phosphatidylinositol 3 Kinase, 2-(4-Morpholinyl)-8-phenyl-4H-1-benzopyran-4-one (LY294002). *J. Biol. Chem.* 1994, 269, 5241–5248.
76. Garlich, J.R.; De, P.; Dey, N.; Jing, D.S.; Peng, X.; Miller, A.; Murali, R.; Lu, Y.; Mills, G.B.; Kundra, V.; et al. A vascular targeted pan phosphoinositide 3-kinase inhibitor prodrug, SF1126, with antitumor and antiangiogenic activity. *Cancer Res.* 2008, 68, 206–215.
77. Qin, A.; Li, Y.; Zhou, L.; Xing, C.; Lu, X. Dual PI3K-BRD4 Inhibitor SF1126 Inhibits Colorectal Cancer Cell Growth in Vitro and in Vivo. *Cell. Physiol. Biochem.* 2019, 52, 758–768.
78. Ronan, B.; Flamand, O.; Vescovi, L.; Dureuil, C.; Durand, L.; Fassy, F.; Bachelot, M.F.; Lamberton, A.; Mathieu, M.; Bertrand, T.; et al. A highly potent and selective Vps34 inhibitor alters vesicle trafficking and autophagy. *Nat. Chem. Biol.* 2014, 10, 1013–1019.
79. Farkas, T.; Daugaard, M.; Jäättelä, M. Identification of small molecule inhibitors of phosphatidylinositol 3-kinase and autophagy. *J. Biol. Chem.* 2011, 286, 38904–38912.
80. Xie, J.; Li, Q.; Ding, X.; Gao, Y. GSK1059615 kills head and neck squamous cell carcinoma cells possibly via activating mitochondrial programmed necrosis pathway. *Oncotarget* 2017, 8, 50814–50823.
81. Bei, S.; Li, F.; Li, H.; Li, J.; Zhang, X.; Sun, Q.; Feng, L. Inhibition of gastric cancer cell growth by a PI3K-mTOR dual inhibitor GSK1059615. *Biochem. Biophys. Res. Commun.* 2019, 511, 13–20.
82. Bago, R.; Malik, N.; Munson, M.J.; Prescott, A.R.; Davies, P.; Sommer, E.; Shpiro, N.; Ward, R.; Cross, D.; Ganley, I.G.; et al. Characterization of VPS34-IN1, a selective inhibitor of Vps34, reveals that the phosphatidylinositol 3-phosphate-binding SGK3 protein kinase is a downstream target of class III phosphoinositide 3-kinase. *Biochem. J.* 2014, 463, 413–427.
83. Dowdle, W.E.; Nyfeler, B.; Nagel, J.; Elling, R.A.; Liu, S.; Triantafellow, E.; Menon, S.; Wang, Z.; Honda, A.; Pardee, G.; et al. Selective VPS34 inhibitor blocks autophagy and uncovers a role for NCOA4 in ferritin degradation and iron homeostasis in vivo. *Nat. Cell Biol.* 2014, 16, 1069–1079.
84. Pasquier, B.; El-Ahmad, Y.; Filoche-Rommé, B.; Dureuil, C.; Fassy, F.; Abecassis, P.Y.; Mathieu, M.; Bertrand, T.; Benard, T.; Barrière, C.; et al. Discovery of (2 S)-8-[(3 R)-3-Methylmorpholin-4-yl]-1-(3-methyl-2-oxobutyl)-2-(trifluoromethyl)-3,4-dihydro-2 H -pyrimido[1,2- a ]pyrimidin-6-one: A novel potent and selective inhibitor of Vps34 for the treatment of solid tumors. *J. Med. Chem.* 2015, 58, 376–400.
85. Liu, J.; Xia, H.; Kim, M.; Xu, L.; Li, Y.; Zhang, L.; Cai, Y.; Norberg, H.V.; Zhang, T.; Furuya, T.; et al. Beclin1 Controls the Levels of p53 by Regulating the Deubiquitination Activity of USP10 and

- USP13. *Cell* 2011, 147, 223–234.
86. Zhang, J.; Mao, S.; Wang, L.; Zhang, W.; Zhang, Z.; Guo, Y.; Wu, Y.; Yi, F.; Yao, X. MicroRNA-154 functions as a tumor suppressor in bladder cancer by directly targeting ATG7. *Oncol. Rep.* 2019, 41, 819–828.
  87. Akin, D.; Wang, S.K.; Habibzadegah-Tari, P.; Law, B.; Ostrov, D.; Li, M.; Yin, X.M.; Kim, J.S.; Horenstein, N.; Dunn, W.A. A novel ATG4B antagonist inhibits autophagy and has a negative impact on osteosarcoma tumors. *Autophagy* 2014, 10, 2021–2035.
  88. Liu, P.-F.; Hsu, C.-J.; Tseng, H.-H.; Wu, C.-H.; Shu, C.-W.; Tsai, K.-L.; Yang, L.-W.; Tsai, W.-L.; Cheng, J.-S.; Chang, H.-W.; et al. Drug repurposing screening identifies tioconazole as an ATG4 inhibitor that suppresses autophagy and sensitizes cancer cells to chemotherapy. *Theranostics* 2018, 8, 830–845.
  89. Kurdi, A.; Cleenewerck, M.; Vangestel, C.; Lyssens, S.; Declercq, W.; Timmermans, J.P.; Stroobants, S.; Augustyns, K.; De Meyer, G.R.Y.; Van Der Veken, P.; et al. ATG4B inhibitors with a benzotropolone core structure block autophagy and augment efficiency of chemotherapy in mice. *Biochem. Pharmacol.* 2017, 138, 150–162.
  90. Bosc, D.; Vezenkova, L.; Bortnik, S.; An, J.; Xu, J.; Choutka, C.; Hannigan, A.M.; Kovacic, S.; Loo, S.; Clark, P.G.K.; et al. A new quinoline-based chemical probe inhibits the autophagy-related cysteine protease ATG4B OPEN. *Sci. Rep.* 2018, 8, 11653.
  91. Fu, Y.; Hong, L.; Xu, J.; Zhong, G.; Gu, Q.; Gu, Q.; Guan, Y.; Zheng, X.; Dai, Q.; Luo, X.; et al. Discovery of a small molecule targeting autophagy via ATG4B inhibition and cell death of colorectal cancer cells in vitro and in vivo. *Autophagy* 2019, 15, 295–311.
  92. Qiu, Z.; Kuhn, B.; Aebi, J.; Lin, X.; Ding, H.; Zhou, Z.; Xu, Z.; Xu, D.; Han, L.; Liu, C.; et al. Discovery of Fluoromethylketone-Based Peptidomimetics as Covalent ATG4B (Autophagin-1) Inhibitors. *ACS Med. Chem. Lett.* 2016, 7, 802–806.
  93. Xu, D.; Xu, Z.; Han, L.; Liu, C.; Zhou, Z.; Qiu, Z.; Lin, X.; Tang, G.; Shen, H.; Aebi, J.; et al. Identification of new ATG4B inhibitors based on a novel high-throughput screening platform. *SLAS Discov.* 2017, 22, 338–347.
  94. Chu, J.; Fu, Y.; Xu, J.; Zheng, X.; Gu, Q.; Luo, X.; Dai, Q.; Zhang, S.; Liu, P.; Hong, L.; et al. ATG4B inhibitor FMK-9a induces autophagy independent on its enzyme inhibition. *Arch. Biochem. Biophys.* 2018, 644, 29–36.
  95. Donohue, E.; Tovey, A.; Vogl, A.W.; Arns, S.; Sternberg, E.; Young, R.N.; Roberge, M. Inhibition of autophagosome formation by the benzoporphyrin derivative verteporfin. *J. Biol. Chem.* 2011, 286, 7290–7300.
  96. Liu-Chittenden, Y.; Huang, B.; Shim, J.S.; Chen, Q.; Lee, S.-J.; Anders, R.A.; Liu, J.O.; Pan, D. Genetic and pharmacological disruption of the TEAD-YAP complex suppresses the oncogenic

- activity of YAP. *Genes Dev.* 2012, 26, 1300–1305.
97. Donohue, E.; Thomas, A.; Maurer, N.; Manisali, I.; Zeisser-Labouebe, M.; Zisman, N.; Anderson, H.J.; Ng, S.S.W.; Webb, M.; Bally, M.; et al. The Autophagy Inhibitor Verteporfin Moderately Enhances the Antitumor Activity of Gemcitabine in a Pancreatic Ductal Adenocarcinoma Model. *J. Cancer* 2013, 4, 585–596.
  98. Lui, J.W.; Xiao, S.; Ogomori, K.; Hammarstedt, J.E.; Little, E.C.; Lang, D. The efficiency of verteporfin as a therapeutic option in pre-clinical models of melanoma. *J. Cancer* 2019, 10, 1–10.
  99. Poole, B.; Ohkuma, S. Effect of weak bases on the intralysosomal pH in mouse peritoneal macrophages. *J. Cell Biol.* 1981, 90, 665–669.
  100. Solomon, V.R.; Lee, H. Chloroquine and its analogs: A new promise of an old drug for effective and safe cancer therapies. *Eur. J. Pharmacol.* 2009, 625, 220–233.
  101. Shi, T.T.; Yu, X.X.; Yan, L.J.; Xiao, H.T. Research progress of hydroxychloroquine and autophagy inhibitors on cancer. *Cancer Chemother. Pharmacol.* 2017, 79, 287–294.
  102. McAfee, Q.; Zhang, Z.; Samanta, A.; Levi, S.M.; Ma, X.-H.; Piao, S.; Lynch, J.P.; Uehara, T.; Sepulveda, A.R.; Davis, L.E.; et al. Autophagy inhibitor Lys05 has single-agent antitumor activity and reproduces the phenotype of a genetic autophagy deficiency. *Proc. Natl. Acad. Sci. USA* 2012, 109, 8253–8258.
  103. Baquero, P.; Dawson, A.; Mukhopadhyay, A.; Kuntz, E.M.; Mitchell, R.; Olivares, O.; Ianniciello, A.; Scott, M.T.; Dunn, K.; Nicastri, M.C.; et al. Targeting quiescent leukemic stem cells using second generation autophagy inhibitors. *Leukemia* 2019, 33, 981–994.
  104. Rebecca, V.W.; Nicastri, M.C.; Mclaughlin, N.; Fennelly, C.; Ronghe, A.; Nofal, M.; Lim, C.; Witze, E.; Chude, C.I.; Zhang, G.; et al. A unified approach to targeting the lysosome's degradative and growth signaling roles. *Cancer Discov.* 2017, 7, 1266–1283.
  105. Goodall, M.L.; Wang, T.; Martin, K.R.; Kortus, M.G.; Kauffman, A.L.; Trent, J.M.; Gately, S.; MacKeigan, J.P. Development of potent autophagy inhibitors that sensitize oncogenic BRAF V600E mutant melanoma tumor cells to vemurafenib. *Autophagy* 2014, 10, 1120–1136.
  106. Lam Yi, H.; Than, H.; Sng, C.; Cheong, M.A.; Chuah, C.; Xiang, W. Lysosome Inhibition by Mefloquine Preferentially Enhances the Cytotoxic Effects of Tyrosine Kinase Inhibitors in Blast Phase Chronic Myeloid Leukemia. *Transl. Oncol.* 2019, 12, 1221–1228.
  107. Sharma, N.; Thomas, S.; Golden, E.B.; Hofman, F.M.; Chen, T.C.; Petasis, N.A.; Schönthal, A.H.; Louie, S.G. Inhibition of autophagy and induction of breast cancer cell death by mefloquine, an antimalarial agent. *Cancer Lett.* 2012, 326, 143–154.
  108. Cheng, S.; Sliva, D. *Ganoderma lucidum* for cancer treatment: We are close but still not there. *Integr. Cancer Ther.* 2015, 14, 249–257.

109. Pan, H.; Wang, Y.; Na, K.; Wang, Y.; Wang, L.; Li, Z.; Guo, C.; Guo, D.; Wang, X. Autophagic flux disruption contributes to *Ganoderma lucidum* polysaccharide-induced apoptosis in human colorectal cancer cells via MAPK/ERK activation. *Cell Death Dis.* 2019, 10.
110. Wu, K.; Na, K.; Chen, D.; Wang, Y.; Pan, H.; Wang, X. Effects of non-steroidal anti-inflammatory drug-activated gene-1 on *Ganoderma lucidum* polysaccharides-induced apoptosis of human prostate cancer PC-3 cells. *Int. J. Oncol.* 2018, 53, 2356–2368.
111. Yamamoto, A.; Tagawa, Y.; Yoshimori, T.; Moriyama, Y.; Masaki, R.; Tashiro, Y. Bafilomycin A1 Prevents Maturation of Autophagic Vacuoles by Inhibiting Fusion between Autophagosomes and Lysosomes in Rat Hepatoma Cell Line. *Cell Struct. Funct.* 1998, 23, 33–42.
112. Bowman, E.J.; Siebers, A.; Altendorf, K. Bafilomycins; A class of inhibitors of membrane ATPases from microorganisms, animal cells, and plant cells. *Proc. Natl. Acad. Sci. USA* 1988, 85, 7972–7976.
113. Mauvezin, C.; Neufeld, T.P. Bafilomycin A1 disrupts autophagic flux by inhibiting both V-ATPase-dependent acidification and Ca-P60A/SERCA-dependent autophagosome-lysosome fusion. *Autophagy* 2015, 11, 1437–1438.
114. Rodilla, A.M.; Korrodi-Gregório, L.; Hernando, E.; Manuel-Manresa, P.; Quesada, R.; Pérez-Tomás, R.; Soto-Cerrato, V. Synthetic tamjbamine analogues induce mitochondrial swelling and lysosomal dysfunction leading to autophagy blockade and necrotic cell death in lung cancer. *Biochem. Pharmacol.* 2017, 126, 23–33.
115. Grinde, B. Effect of carboxylic ionophores on lysosomal protein degradation in rat hepatocytes. *Exp. Cell Res.* 1983, 149, 27–35.
116. Busschaert, N.; Park, S.H.; Baek, K.H.; Choi, Y.P.; Park, J.; Howe, E.N.W.; Hiscock, J.R.; Karagiannidis, L.E.; Marques, I.; Félix, V.; et al. A synthetic ion transporter that disrupts autophagy and induces apoptosis by perturbing cellular chloride concentrations. *Nat. Chem.* 2017, 9, 667–675.
117. Sharma, G.; Guardia, C.M.; Roy, A.; Vassilev, A.; Saric, A.; Griner, L.N.; Marugan, J.; Ferrer, M.; Bonifacino, J.S.; DePamphilis, M.L. A family of PIKFYVE inhibitors with therapeutic potential against autophagy-dependent cancer cells disrupt multiple events in lysosome homeostasis. *Autophagy* 2019, 15, 1694–1718.
118. Lu, Y.; Dong, S.; Hao, B.; Li, C.; Zhu, K.; Guo, W.; Wang, Q.; Cheung, K.-H.; Wong, C.W.M.; Wu, W.-T.; et al. Vacuolin-1 potently and reversibly inhibits autophagosome-lysosome fusion by activating RAB5A. *Autophagy* 2014, 10, 1895–1905.
119. Rossi, M.; Munarriz, E.R.; Bartesaghi, S.; Milanese, M.; Dinsdale, D.; Guerra-Martin, M.A.; Bampton, E.T.W.; Glynn, P.; Bonanno, G.; Knight, R.A.; et al. Desmethylclomipramine induces the

- accumulation of autophagy markers by blocking autophagic flux. *J. Cell Sci.* 2009, 122, 3330–3339.
120. Libby, P.; Goldberg, A.L. Leupeptin, a protease inhibitor, decreases protein degradation in normal and diseased muscles. *Science* (80-. ). 1978, 199, 534–536.
  121. Tanida, I.; Minematsu-Ikeguchi, N.; Ueno, T.; Kominami, E. Lysosomal turnover, but not a cellular level, of endogenous LC3 is a marker for autophagy. *Autophagy* 2005, 1, 84–91.
  122. Stern, S.T.; Adiseshaiah, P.P.; Crist, R.M. Autophagy and lysosomal dysfunction as emerging mechanisms of nanomaterial toxicity. *Part. Fibre Toxicol.* 2012, 9, 20.
  123. Ma, X.; Wu, Y.; Jin, S.; Tian, Y.; Zhang, X.; Zhao, Y.; Yu, L.; Liang, X.J. Gold nanoparticles induce autophagosome accumulation through size-dependent nanoparticle uptake and lysosome impairment. *ACS Nano* 2011, 5, 8629–8639.
  124. Cui, Z.; Zhang, Y.; Xia, K.; Yan, Q.; Kong, H.; Zhang, J.; Zuo, X.; Shi, J.; Wang, L.; Zhu, Y.; et al. Nanodiamond autophagy inhibitor allosterically improves the arsenical-based therapy of solid tumors. *Nat. Commun.* 2018, 9, 4347.
  125. Hosokawa, N.; Hara, T.; Kaizuka, T.; Kishi, C.; Takamura, A.; Miura, Y.; Iemura, S.; Natsume, T.; Takehana, K.; Yamada, N.; et al. Nutrient-dependent mTORC1 Association with the ULK1–Atg13–FIP200 Complex Required for Autophagy. *Mol. Biol. Cell* 2009, 20, 1981–1991.
  126. Chen, S.; Wang, C.; Yeo, S.; Liang, C.C.; Okamoto, T.; Sun, S.; Wen, J.; Guan, J.L. Distinct roles of autophagy-dependent and -independent functions of FIP200 revealed by generation and analysis of a mutant knock-in mouse model. *Genes Dev.* 2016, 30, 856–869.
  127. Mercer, C.A.; Kaliappan, A.; Dennis, P.B. A novel, human Atg13 binding protein, Atg101, interacts with ULK1 and is essential for macroautophagy. *Autophagy* 2009, 5, 649–662.
  128. Lee, E.J.; Tournier, C. The requirement of uncoordinated 51-like kinase 1 (ULK1) and ULK2 in the regulation of autophagy. *Autophagy* 2011, 7, 689–695.
  129. Chaikuad, A.; Koschade, S.E.; Stolz, A.; Zivkovic, K.; Pohl, C.; Shaid, S.; Ren, H.; Lambert, L.J.; Cosford, N.D.P.; Brandts, C.H.; et al. Conservation of structure, function and inhibitor binding in UNC-51-like kinase 1 and 2 (ULK1/2). *Biochem. J.* 2019, 2, BCJ20190038.
  130. Yun, M.; Bai, H.Y.; Zhang, J.X.; Rong, J.; Weng, H.W.; Zheng, Z.S.; Xu, Y.; Tong, Z.T.; Huang, X.X.; Liao, Y.J.; et al. ULK1: A promising biomarker in predicting poor prognosis and therapeutic response in human nasopharyngeal carcinoma. *PLoS ONE* 2015, 10, e0117375.
  131. Zou, Y.; Chen, Z.; He, X.; He, X.; Wu, X.; Chen, Y.; Wu, X.; Wang, J.; Lan, P. High expression levels of unc-51-like kinase 1 as a predictor of poor prognosis in colorectal cancer. *Oncol. Lett.* 2015, 10, 1583–1588.

132. Jiang, S.; Li, Y.; Zhu, Y.H.; Wu, X.Q.; Tang, J.; Li, Z.; Feng, G.K.; Deng, R.; Li, D.D.; Luo, R.Z.; et al. Intensive expression of UNC-51-like kinase 1 is a novel biomarker of poor prognosis in patients with esophageal squamous cell carcinoma. *Cancer Sci.* 2011, 102, 1568–1575.
133. Dite, T.A.; Langendorf, C.G.; Hoque, A.; Galic, S.; Rebello, R.J.; Ovens, A.J.; Lindqvist, L.M.; Ngoei, K.R.W.; Ling, N.X.Y.; Furic, L.; et al. AMP-activated protein kinase selectively inhibited by the type II inhibitor SBI-0206965. *J. Biol. Chem.* 2018, 293, 8874–8885.
134. Lindmo, K. Regulation of membrane traffic by phosphoinositide 3-kinases. *J. Cell Sci.* 2006, 119, 605–614.
135. Liu, P.; Cheng, H.; Roberts, T.M.; Zhao, J.J. Targeting the phosphoinositide 3-kinase (PI3K) pathway in cancer. *Nat Rev Drug Discov* 2009, 8, 627–644.
136. Leontieva, O.V.; Blagosklonny, M. V Gerosuppression by pan-mTOR inhibitors. *Aging (Albany, NY).* 2016, 8, 3535–3551.
137. Funderburk, S.F.; Wang, Q.J.; Yue, Z. Public Access NIH Public Access. *Trends Cell Biol.* 2010, 20, 355–362.
138. Valet, C.; Levade, M.; Chicanne, G.; Bilanges, B.; Cabou, C.; Viaud, J.; Gratacap, M.-P.; Gaits-Iacovoni, F.; Vanhaesebroeck, B.; Payrastre, B.; et al. A dual role for the class III PI3K, Vps34, in platelet production and thrombus growth. *Blood* 2017, 130, 2032–2042.
139. Tanida, I.; Sou, Y.S.; Ezaki, J.; Minematsu-Ikeguchi, N.; Ueno, T.; Kominami, E. HsAtg4B/HsApg4B/autophagin-1 cleaves the carboxyl termini of three human Atg8 homologues and delipidates microtubule-associated protein light chain 3- and GABAA receptor-associated protein-phospholipid conjugates. *J. Biol. Chem.* 2004, 279, 36268–36276.
140. Yu, Z.Q.; Ni, T.; Hong, B.; Wang, H.Y.; Jiang, F.J.; Zou, S.; Chen, Y.; Zheng, X.L.; Klionsky, D.J.; Liang, Y.; et al. Dual roles of Atg8—PE deconjugation by Atg4 in autophagy. *Autophagy* 2012, 8, 883–892.
141. Fu, Y.; Huang, Z.; Hong, L.; Lu, J.H.; Feng, D.; Yin, X.M.; Li, M. Targeting ATG4 in cancer therapy. *Cancers* 2019, 11, 649.
142. Tran, E.; Chow, A.; Goda, T.; Wong, A.; Blakely, K.; Rocha, M.; Taeb, S.; Hoang, V.C.; Liu, S.K.; Emmenegger, U. Context-dependent role of ATG4B as target for autophagy inhibition in prostate cancer therapy. *Biochem. Biophys. Res. Commun.* 2013, 441, 726–731.
143. Bortnik, S.; Choutka, C.; Horlings, H.M.; Leung, S.; Baker, J.H.E.; Lebovitz, C.; Dragowska, W.H.; Go, N.E.; Bally, M.B.; Minchinton, A.I.; et al. Identification of breast cancer cell subtypes sensitive to ATG4B inhibition. *Oncotarget* 2016, 7.
144. Donohue, E.; Balgi, A.D.; Komatsu, M.; Roberge, M. Induction of Covalently Crosslinked p62 Oligomers with Reduced Binding to Polyubiquitinated Proteins by the Autophagy Inhibitor



Verteporfin. PLoS ONE 2014, 9, e114964.

145. Zhang, H.; Ramakrishnan, S.K.; Triner, D.; Centofanti, B.; Maitra, D.; Győrffy, B.; Sebolt-Leopold, J.S.; Dame, M.K.; Varani, J.; Brenner, D.E.; et al. Tumor-selective proteotoxicity of verteporfin inhibits colon cancer progression independently of YAP1. *Sci. Signal.* 2015, 8, ra98.
146. Ota, M.; Sasaki, H. Mammalian Tead proteins regulate cell proliferation and contact inhibition as transcriptional mediators of Hippo signaling. *Development* 2008, 135, 4059–4069.
147. Wei, H.; Wang, F.; Wang, Y.; Li, T.; Xiu, P.; Zhong, J.; Sun, X.; Li, J. Verteporfin suppresses cell survival, angiogenesis and vasculogenic mimicry of pancreatic ductal adenocarcinoma via disrupting the YAP-TEAD complex. *Cancer Sci.* 2017, 108, 478–487.
148. Melles, R.B.; Marmor, M.F. The risk of toxic retinopathy in patients on long-term hydroxychloroquine therapy. *JAMA Ophthalmol.* 2014, 132, 1453–1460.
149. Costedoat-Chalumeau, N.; Hulot, J.-S.; Amoura, Z.; Delcourt, A.; Maisonobe, T.; Dorent, R.; Bonnet, N.; Sablé, R.; Lechat, P.; Wechsler, B.; et al. Cardiomyopathy Related to Antimalarial Therapy with Illustrative Case Report. *Cardiology* 2007, 107, 73–80.
150. Marmor, M.F.; Kellner, U.; Lai, T.Y.Y.; Melles, R.B.; Mieler, W.F.; Lum, F. Recommendations on Screening for Chloroquine and Hydroxychloroquine Retinopathy (2016 Revision). *Ophthalmology* 2016, 123, 1386–1394.
151. Bristol, M.L.; Emery, S.M.; Maycotte, P.; Thorburn, A.; Chakradeo, S.; Gewirtz, D.A. Autophagy inhibition for chemosensitization and radiosensitization in cancer: Do the preclinical data support this therapeutic strategy? *J. Pharmacol. Exp. Ther.* 2013, 344, 544–552.
152. Bongiorno-Borbone, L.; Giacobbe, A.; Compagnone, M.; Eramo, A.; De Maria, R.; Peschiaroli, A.; Melino, G. Anti-tumoral effect of desmethylclomipramine in lung cancer stem cells. *Oncotarget* 2015, 6, 16926–16938.
153. Boya, P.; Casares, N.; Perfettini, J.; Dessen, P.; Larochette, N.; Métivier, D.; Meley, D.; Souquere, S.; Pierron, G.; Codogno, P.; et al. Inhibition of Macroautophagy Triggers Apoptosis. *Mol. Cell. Biol.* 2005, 25, 1025–1040.
154. Verbaanderd, C.; Maes, H.; Schaaf, M.B.; Sukhatme, V.P.; Pantziarka, P.; Sukhatme, V.; Agostinis, P.; Bouche, G. Repurposing drugs in oncology (ReDO)—Chloroquine and hydroxychloroquine as anti-cancer agents. *Ecancermedicalscience* 2017, 11, 1–35.
155. Piao, S.; Ojha, R.; Rebecca, V.W.; Samanta, A.; Ma, X.H.; McAfee, Q.; Nicastri, M.C.; Buckley, M.; Brown, E.; Winkler, J.D.; et al. ALDH1A1 and HLTF modulate the activity of lysosomal autophagy inhibitors in cancer cells. *Autophagy* 2017, 13, 2056–2071.
156. Maycotte, P.; Aryal, S.; Ct, C.; Thorburn, J. Chloroquine sensitizes breast cancer cells to chemotherapy independent of autophagy. *Autophagy* 2012, 8, 200–212.

157. Eng, C.H.; Wang, Z.; Tkach, D.; Toral-Barza, L.; Ugwonali, S.; Liu, S.; Fitzgerald, S.L.; George, E.; Frias, E.; Cochran, N.; et al. Macroautophagy is dispensable for growth of KRAS mutant tumors and chloroquine efficacy. *Proc. Natl. Acad. Sci. USA* 2016, 113, 182–187.
158. Degtyarev, M.; De Mazière, A.; Orr, C.; Lin, J.; Lee, B.B.; Tien, J.Y.; Prior, W.W.; Van Dijk, S.; Wu, H.; Gray, D.C.; et al. Akt inhibition promotes autophagy and sensitizes PTEN-null tumors to lysosomotropic agents. *J. Cell Biol.* 2008, 183, 101–116.
159. Kwakye-Berko, F.; Meshnick, S.R. Binding of chloroquine to DNA. *Mol. Biochem. Parasitol.* 1989, 35, 51–55.
160. Loehberg, C.R.; Strissel, P.L.; Dittrich, R.; Strick, R.; Dittmer, J.; Dittmer, A.; Fabry, B.; Kalender, W.A.; Koch, T.; Wachter, D.L.; et al. Akt and p53 are potential mediators of reduced mammary tumor growth by Chloroquine and the mTOR inhibitor RAD001. *Biochem. Pharmacol.* 2012, 83, 480–488.
161. Enzenmüller, S.; Gonzalez, P.; Debatin, K.M.; Fulda, S. Chloroquine overcomes resistance of lung carcinoma cells to the dual PI3K/mTOR inhibitor PI103 by lysosome-mediated apoptosis. *Anticancer. Drugs* 2013, 24, 14–19.
162. Kroemer, G.; Jäätelä, M. Lysosomes and autophagy in cell death control. *Nat. Rev. Cancer* 2005, 5, 886–897.
163. Carew, J.S.; Nawrocki, S.T.; Kahue, C.N.; Zhang, H.; Yang, C.; Chung, L.; Houghton, J.A.; Huang, P.; Giles, F.J.; Cleveland, J.L. Targeting autophagy augments the anticancer activity of the histone deacetylase inhibitor SAHA to overcome Bcr-Abl-mediated drug resistance. *Blood* 2007, 110, 313–322.
164. Morgan, M.J.; Fitzwalter, B.E.; Owens, C.R.; Powers, R.K.; Sottnik, J.L.; Gamez, G.; Costello, J.C.; Theodorescu, D.; Thorburn, A. Metastatic cells are preferentially vulnerable to lysosomal inhibition. *Proc. Natl. Acad. Sci. USA* 2018, 115, E8479–E8488.
165. Fennelly, C.; Amaravadi, R.K. Lysosomal Biology in Cancer. *Methods Mol Biol* 2017, 1594, 293–308.
166. Uhlman, A.; Folkers, K.; Liston, J.; Pancholi, H.; Hinton, A. Effects of Vacuolar H(+)-ATPase Inhibition on Activation of Cathepsin B and Cathepsin L Secreted from MDA-MB231 Breast Cancer Cells. *Cancer Microenviron.* 2017, 10, 49–56.
167. Balic, A.; Sørensen, M.D.; Trabulo, S.M.; Sainz, B.; Cioffi, M.; Vieira, C.R.; Miranda-Lorenzo, I.; Hidalgo, M.; Kleeff, J.; Erkan, M.; et al. Chloroquine Targets Pancreatic Cancer Stem Cells via Inhibition of CXCR4 and Hedgehog Signaling. *Mol. Cancer Ther.* 2014, 13, 1758–1771.
168. Maes, H.; Kuchnio, A.; Peric, A.; Moens, S.; Nys, K.; DeBock, K.; Quaegebeur, A.; Schoors, S.; Georgiadou, M.; Wouters, J.; et al. Tumor vessel normalization by chloroquine independent of autophagy. *Cancer Cell* 2014, 26, 190–206.

169. Mulcahy Levy, J.M.; Thompson, J.C.; Griesinger, A.M.; Amani, V.; Donson, A.M.; Birks, D.K.; Morgan, M.J.; Mirsky, D.M.; Handler, M.H.; Foreman, N.K.; et al. Autophagy Inhibition Improves Chemosensitivity in BRAFV600E Brain Tumors. *Cancer Discov* 2014, 4, 773–780.
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