

Molecular Characterization of XX Maleness

Subjects: [Endocrinology & Metabolism](#)

Contributor: Rodolfo Rey

Androgens and anti-Müllerian hormone (AMH), secreted by the foetal testis, are responsible for the development of male reproductive organs and the regression of female anlagen. Virilization of the reproductive tract in association with the absence of Müllerian derivatives in the XX foetus implies the existence of testicular tissue, which can occur in the presence or absence of SRY. Recent advancement in the knowledge of the opposing gene cascades driving to the differentiation of the gonadal ridge into testes or ovaries during early foetal development has provided insight into the molecular explanation of XX maleness.

Sex differentiation

XX male

Disorders of Sex Development

Testis

Ovary

Ovotestis

1. Introduction

Ovarian differentiation and female internal and external genitalia are the expected pathway in the mammalian XX foetus (Figure 1). Only rarely, may XX gonads follow the testicular differentiation pathway, and consequently internal and external genitalia are virilised by testicular hormones. This “sex-reversal” condition was initially characterised in humans and named “XX male” [1], since no hint of a disorder of sex development was present until adulthood, when these males sought medical attention for infertility (see below). Alternatively, testicular tissue was also observed together with ovarian tissue in the same individual carrying an XX karyotype. The condition was known previously as “hermaphroditism”, and most frequently internal and external genitalia are ambiguous since the testicular moiety is functionally insufficient to induce full virilisation during embryonic and foetal development. Finally, virilisation of an XX foetus with ovarian development and absence of testicular tissue can be the consequence of an androgen excess of extragonadal origin. Indeed, disorders of adrenal steroidogenesis—like congenital adrenal hyperplasia [2], androgen-secreting adrenal or ovarian tumours or maternal use of anabolic steroids [3], and placental aromatase deficiency [4] can lead to foetal virilisation in the absence of testicular tissue. In this review, we will address only those conditions in which there is testicular differentiation despite an XX karyotype, with special emphasis in conditions affecting human individuals.

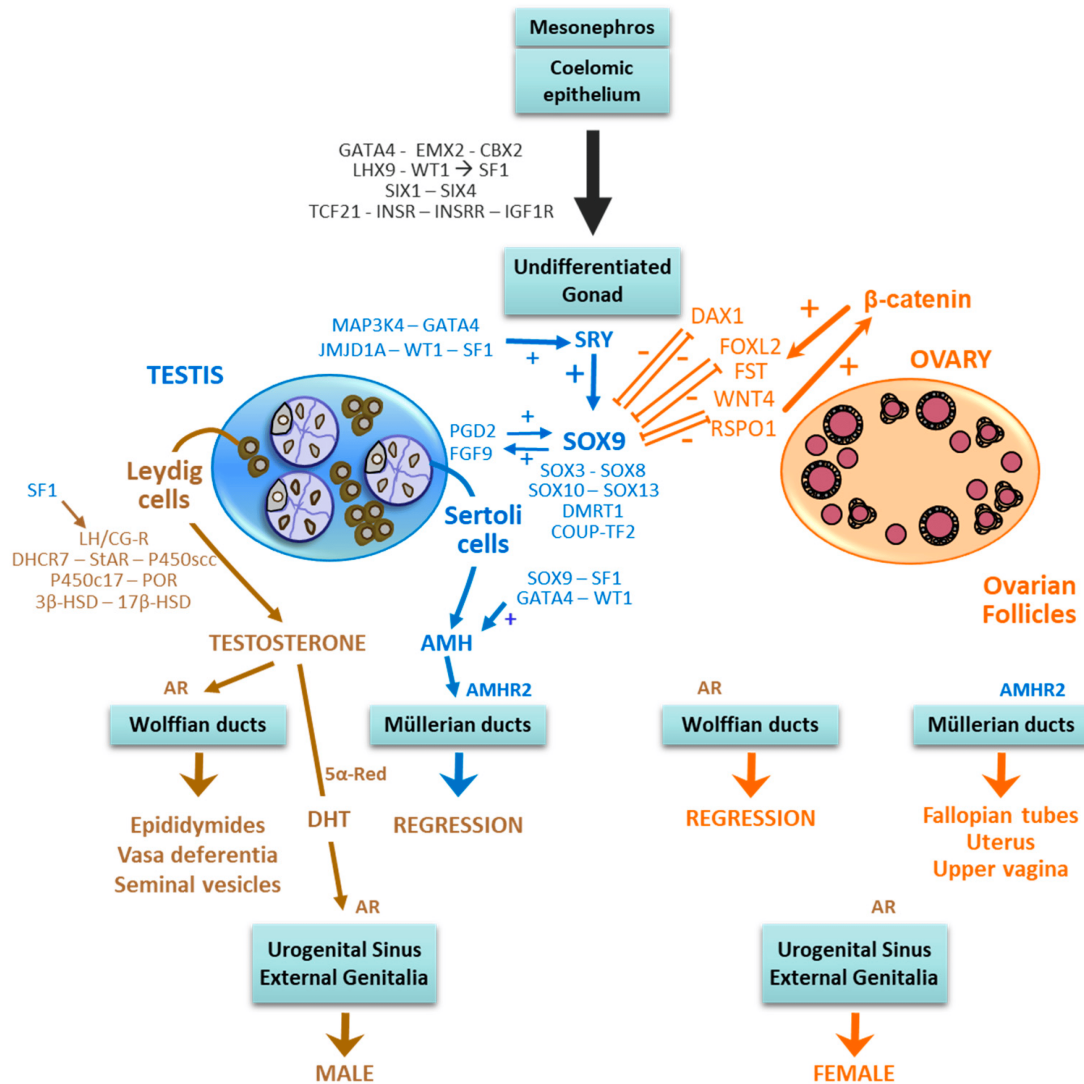


Figure 1. Sex differentiation during embryonic and foetal life in mammals: the mesonephros and the coelomic epithelium are stabilized and their cells proliferate, in response to the action of GATA4, EMX2, CBX2, LHX9 and WT1 -which upregulate SF1-, SIX1 and SIX4, TCF21 and members of the IGF family, to form the undifferentiated gonad. In XY foetuses, SRY expression is triggered by MAP3K4, GATA4, JMJD1A, WT1 and SF1; SRY upregulates SOX9 and other SOX family members. SOX9, PGD2 and FGF9 establish a positive feed-forward loop, which increases SOX9 expression, which prevails over DAX1, FOXL2, WNT4 and RSPO1, thus promoting testicular differentiation. The developing testis secretes testosterone and anti-Müllerian hormone (AMH), responsible for male differentiation of internal and external genitalia; testosterone can be transformed into dihydrotestosterone (DHT), a more potent androgen, through the action of the enzyme 5α-reductase (5α-Red); both testosterone and DHT act on the same androgen receptor (AR). In the XX foetuses, due to the absence of SRY, SOX9 expression remains low and is overcome by DAX1, FOXL2, WNT4 and RSPO1, which upregulate β-catenin; a feed-forward loop between these pro-ovarian factors is established, resulting in the differentiation of the female gonad. Since the ovary does not secrete androgens and AMH, internal and external genitalia develop through the female pathway. Modified with permission from Rey and Grinspon, 2011 [5]. Copyright 2010 Elsevier Ltd.

2. Molecular Mechanisms Underlying Foetal Sex Differentiation

2.1. The Sexually Undifferentiated Stage

Irrespective of their sex-chromosome constitution, all mammalian embryos are anatomically, histologically and functionally undifferentiated from a sexual standpoint during the first stages of development, approximately six weeks in humans and 10 days in mice. The gonadal ridges are bipotential, i.e., they can form ovaries or testes. Concomitantly, XX and XY embryos develop the anlagen for both male and female internal reproductive ducts; each pair of ducts is unipotential: the Wolffian (mesonephric) ducts may give rise only to male functional reproductive derivatives, whereas, the Müllerian (paramesonephric) ducts may give rise only to female functional structures. The primordia of the external genitalia are bipotential, like the gonads.

During the bipotential period, the XX and the XY gonadal ridges show identical expression profiles for purportedly pro-testicular factors, like SOX9 and FGF9, and pro-ovarian factors, like WNT4, RSPO1, and DAX1 [6]. Cell proliferation in the somatic component of the gonadal ridges is an essential process, which is regulated at this stage by LHX9 interaction with WT1 resulting in the modulating SF1 expression [7,8]. Other factors responsible for somatic cell proliferation are TCF21, INSR, IGF1R, and INSR (Figure 1). During early gonadal differentiation, cell proliferation is more critical in the male testis than in the ovary [9]. WT1 and SF1 play also key roles in the whole process of urogenital organogenesis. WT1, named after Wilms' tumour (nephroblastoma), is a transcriptional and post-transcriptional regulator [10] that is expressed early in the urogenital ridge and plays an essential role in the development of the kidneys and gonads. SF1 is encoded by the *NR5A1* gene. Initially described as a regulator of steroid hydroxylases in the adrenal cortex, SF1 is an orphan nuclear receptor also expressed in the hypothalamus, the pituitary and the gonads [11]. In the early embryo, SF1 is essential for the development of the gonadal primordium [12].

2.2. Testicular Differentiation

The differentiation of the gonadal ridges into ovaries or testes requires a delicate dosage balance in the timing and levels of expression of several genes [13]. In most mammalian embryos, the transient expression of *SRY*, which maps to the Y chromosome, triggers a cascade of gene interactions ultimately leading to the formation of a testis from the indifferent gonadal ridge [14]. *SRY* expression initiates in the middle of the gonad and expands toward the poles [15]. The timing and level of *SRY* expression are critical for proper testis differentiation: delayed or decreased expression results in dysgenetic testicular or ovotesticular differentiation in the mouse [16,17,18]. In most mammals, the *SRY*-box gene *SOX9* is the earliest upregulated gene in the testis pathway downstream of *SRY* (Figure 1), followed by *CITED4* and other members of the SOX family, including *SOX3*, *SOX8*, *SOX10* and *SOX13*. Many other genes are also critical for testicular differentiation [13,19,20]. *FGF9* has a role in Sertoli cell differentiation: *Fgf9*-knockout mice have dysgenetic gonads [21].

In human foetuses, Sertoli cell differentiation, characterised by the expression of SOX9 [22], AMH [23,24] and DHH [25,26,27], and cord formation begin in the central part of the gonad [28] by the end of the seventh week. Differentiating Sertoli cells also express growth factors, like nerve growth factors (NGFs), which can induce cell migration from the mesonephros acting through their receptors TRKA (NTRK1) and TRKC (NTRK3) [29,30]. In the interstitial compartment, Leydig cells differentiate amongst connective tissue and blood vessels. The formation of the coelomic vessel is a particular feature of early testicular differentiation [31,32].

The origin of Leydig cells is less clear: the mesonephros, the coelomic epithelium, the neural crest and resident SF1-expressing cells in the adreno-gonadal primordium have been proposed as their precursors [33,34,35,36,37]. Leydig cells differentiate a few days later than Sertoli cells initially depending, at least partially, on Sertoli cell-secreted PDGFs binding to PDGFR α [38] independently of gonadotropin action [39,40]. Nonetheless, in humans further Leydig cell differentiation requires placental chorionic gonadotropin (hCG) in the first and second trimesters of foetal life and foetal pituitary luteinizing hormone (LH) thereafter acting on the LH/CG receptor [41]. FGF9 [42] and DHH [43] are other factors secreted by Sertoli cells and responsible for normal Leydig cell differentiation.

2.3. Ovarian Differentiation

Genes involved in ovarian differentiation of the bipotential gonad increase their expression somewhat later than those involved in testis differentiation. WNT4 and RSPO1 stabilise β -catenin, encoded by *CTNNB1*, which promotes the expression of ovarian genes, like *FST* (follistatin) and *FOXL2* (Figure 1), resulting in granulosa cell differentiation [44,45,46]. Follistatin also counteracts FGF9, SOX9 and activins A and B, thus repressing the formation of the coelomic vessel [13,47,48,49,50]. By upregulating DAX1, WNT4 also antagonizes SF1 and thus inhibits androgen biosynthesis [51]. WNT4 also acts as a germ cell survival factor in the ovary [52]. In the mouse, FOXL2 is required to continuously suppress SOX9 until adulthood, thus preventing ovarian cells from transdifferentiating into “testis-like” cells [53]. Finally, a number of factors are essential for folliculogenesis [54,55]: neurotrophins and their receptors promote follicular development, FIG α is essential for primordial follicle formation, NOBOX, SOHLH1 and SOHLH2 play a role in the differentiation to primary follicles, and BMP15 and GDF9 are involved in later stages of folliculogenesis.

2.4. The Differentiation of the Internal and External Genitalia

The ground-breaking observations made by Jost almost seventy years ago [56] taught us that the differentiation of the gonadal ridges into testes is determinant for the virilisation of the foetus. This is due to the fact that the testes have two endocrine cell populations: Leydig cells produce androgens, which induce the differentiation of the Wolffian duct into the epididymis, vas deferens and seminal vesicles and the virilisation of the primordia of the external genitalia [57], and Sertoli cells secrete anti-Müllerian hormone (AMH), responsible for the regression of the Müllerian ducts, which otherwise form the Fallopian tubes, the uterus and the upper vagina [58,59] (Figure 1).

AMH acts very early in foetal life through its interaction with the specific AMH type 2 receptor, encoded by *AMHR2* [60]. The molecular mechanisms involved in this process have recently been reviewed in detail [61]. On the other

hand, testosterone and dihydrotestosterone [62], acting through the androgen receptor, drive the differentiation of derivatives of the Wolffian duct, the urogenital sinus and the external genitalia. The extent of the regression of Müllerian ducts as well as of the male differentiation of the internal and external genitalia is commensurate with the functional capacity of the foetal testes to produce AMH and androgens during the narrow window of foetal sex differentiation [5].

3. Pathogenesis of XX Maleness

As it can be deduced from the knowledge of the normal mechanisms underlying foetal sex differentiation described above, testicular tissue can occur in individuals when Y-chromosome sequences encompassing *SRY* are present (Figure 2). These individuals, known as *SRY*-positive, may have a pure 46,XX karyotype or 46,XX/46,XY chimaerism. Nonetheless, testicular development may take place in the absence of *SRY*, these cases being described as *SRY*-negative. Finally, male development of the external genitalia can occur in the absence of testicular tissue, for instance in human patients with congenital adrenal hyperplasia, androgen-secreting adrenal or ovarian tumours, maternal use of anabolic steroids or placental aromatase deficiency. These are beyond the scope of the present review.

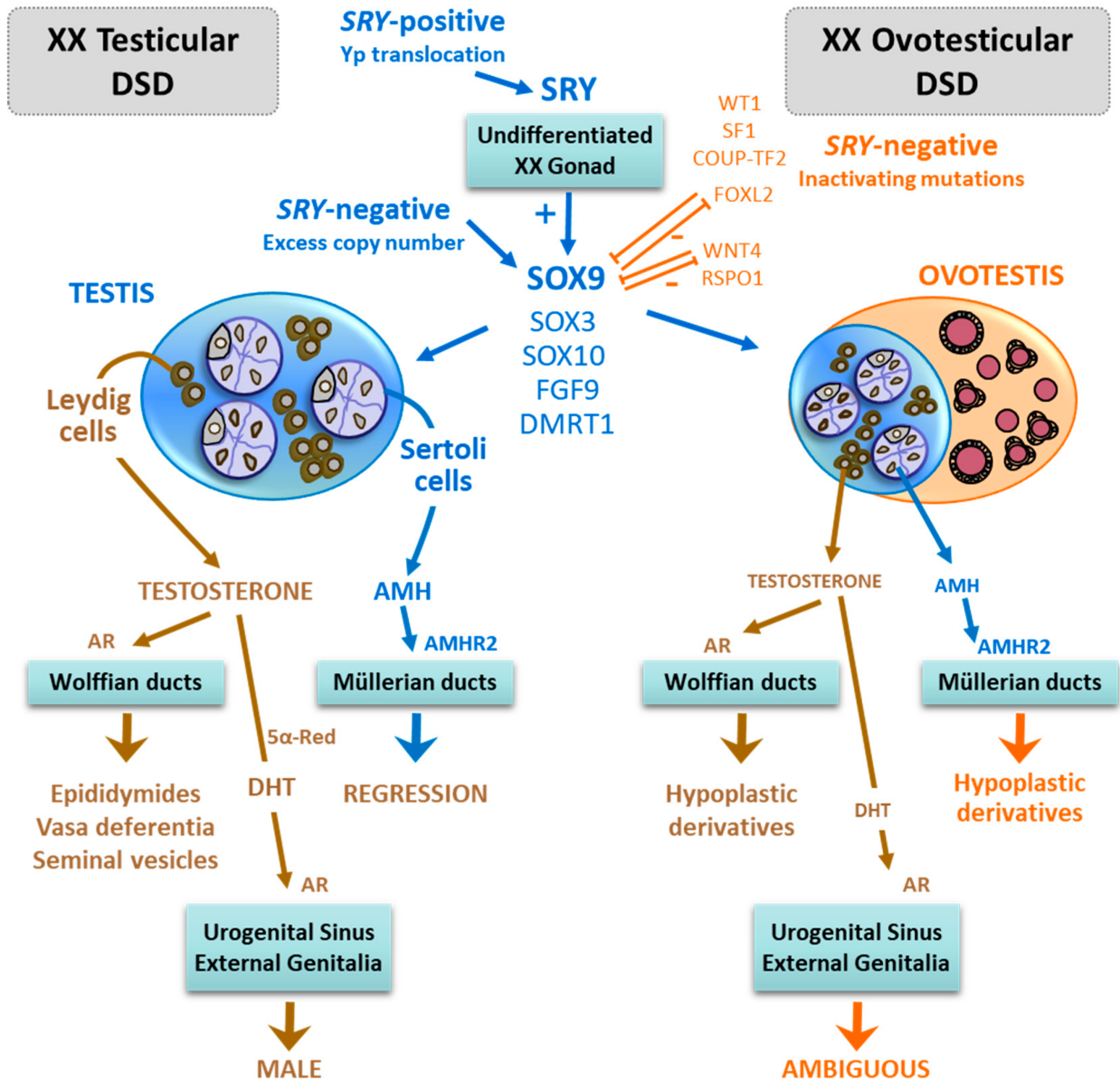


Figure 2. Testicular and ovotesticular differentiation in XX foetuses. In individuals with Yp translocations, the presence of SRY triggers testicular differentiation (SRY-positive cases). In SRY-negative individuals, testicular development may result from overexpression of “pro-testicular” factors (e.g., SOX9, SOX3, SOX10, FGF9, DMRT1) due to duplications/triplications of gene coding or regulatory sequences, or from inactivating mutations of “pro-ovarian” factors (e.g., RSPO1, WNT4, FOXL2) or factors that are believed to favour the gene dosage balance through the ovarian pathway (WT1, SF1, COUP-TF2). Generally, though not always, testicular DSD presents with male genitalia and ovotesticular DSD with ambiguous genitalia.

In both SRY-positive and SRY-negative situations, the gonads can completely differentiate into testes, and the condition is known as “XX male” or “46,XX testicular disorder of sex development (DSD)” (Figure 2). When testicular and ovarian tissues are present, as confirmed by histological studies showing the presence of seminiferous tubules and ovarian follicles with oocytes, the condition is known as “hermaphroditism” (term used widely in previous decades but now limited to animals, see below) or “ovotesticular DSD” (preferred for humans)

(Figure 2). The proportions of ovarian and testicular tissue can vary amongst patients, and the presentation may also be asymmetric, with predominance of one tissue type on one side and of the other contralaterally, or even the existence of only one tissue type on one side and only the other or both contralaterally. As discussed below, the amount of functional testicular tissue determines the anatomical aspect of the internal and external genitalia, while the amount of ovarian tissue has no effect on the genital phenotype of the new-born.

References

1. Albert Chapelle; Herman Hortling; Mikko Niemi; Johan Wennström; XX Sex Chromosomes in a Human Male. *Acta Medica Scandinavica* **2009**, *175*, 25-38, 10.1111/j.0954-6820.1964.tb04630.x.
2. Adina F. Turcu; Richard J. Auchus; Adrenal steroidogenesis and congenital adrenal hyperplasia.. *Endocrinology and Metabolism Clinics of North America* **2015**, *44*, 275-296, 10.1016/j.ecl.2015.02.002.
3. Christopher Hakim; Vasantha Padmanabhan; Arpita K. Vyas; Gestational Hyperandrogenism in Developmental Programming. *Endocrinology* **2016**, *158*, 199-212, 10.1210/en.2016-1801.
4. A. Belgorosky; G. Guercio; C. Pepe; N. Saraco; M.A. Rivarola; Genetic and Clinical Spectrum of Aromatase Deficiency in Infancy, Childhood and Adolescence. *Hormone Research in Paediatrics* **2009**, *72*, 321-330, 10.1159/000249159.
5. Rodolfo Alberto Rey; Romina P. Grinspon; Normal male sexual differentiation and aetiology of disorders of sex development. *Best Practice & Research Clinical Endocrinology & Metabolism* **2011**, *25*, 221-238, 10.1016/j.beem.2010.08.013.
6. Dorien Baetens; Hannah Verdin; Elfride De Baere; Martine Cools; Update on the genetics of differences of sex development (DSD). *Best Practice & Research Clinical Endocrinology & Metabolism* **2019**, *33*, 101271, 10.1016/j.beem.2019.04.005.
7. Ohad S. Birk; Delane E. Casiano; Christopher A. Wassif; Tiziana Cogliati; Liping Zhao; Yangu Zhao; Alexander Grinberg; SingPing Huang; Jordan A. Kreidberg; Keith L. Parker; et al. Forbes D. PorterHeiner Westphal The LIM homeobox gene Lhx9 is essential for mouse gonad formation. *Nature* **2000**, *403*, 909-913, 10.1038/35002622.
8. Dagmar Wilhelm; Christoph Englert; The Wilms tumor suppressor WT1 regulates early gonad development by activation of Sf1. *Genes & Development* **2002**, *16*, 1839-1851, 10.1101/gad.220102.
9. Rey, R.; Josso, N.; Racine, C. Sexual Differentiation. In Endotext; De Groot, L.J., Chrousos, G., Dungan, K., Feingold, K.R., Grossman, A., Hershman, J.M., Koch, C., Korbonits, M., McLachlan, R., New, M., et al., Eds.; MDTtext.com, Inc.: South Dartmouth, MA, USA, 2016

10. Nicholas D. Hastie; Wilms' tumour 1 (WT1) in development, homeostasis and disease. *Development* **2017**, *144*, 2862-2872, 10.1242/dev.153163.
11. Jenifer P. Suntharalingham; Federica Buonocore; Andrew J. Duncan; John C. Achermann; DAX-1 (NR0B1) and steroidogenic factor-1 (SF-1, NR5A1) in human disease.. *Best Practice & Research Clinical Endocrinology & Metabolism* **2015**, *29*, 607-619, 10.1016/j.beem.2015.07.004.
12. Xunrong Luo; Yayoi Ikeda; Keith L. Parker; A cell-specific nuclear receptor is essential for adrenal and gonadal development and sexual differentiation. *Cell* **1994**, *77*, 481-490, 10.1016/0092-8674(94)90211-9.
13. Yi-Tzu Lin; Blanche Capel; Cell fate commitment during mammalian sex determination.. *Current Opinion in Genetics & Development* **2015**, *32*, 144-152, 10.1016/j.gde.2015.03.003.
14. Christian Larney; Timothy L. Bailey; Peter Koopman; Switching on sex: transcriptional regulation of the testis-determining gene Sry.. *Development* **2014**, *141*, 2195-2205, 10.1242/dev.107052.
15. Monica Bullejos; Peter Koopman; Spatially dynamic expression of Sry in mouse genital ridges. *Developmental Dynamics* **2001**, *221*, 201-205, 10.1002/dvdy.1134.
16. Claude M Nagamine; Ken-Ichirou Morohashi; Cherlyn Carlisle; Dennis K Chang; Sex Reversal Caused by Mus musculus domesticus Y Chromosomes Linked to Variant Expression of the Testis-Determining Gene Sry. *Developmental Biology* **1999**, *216*, 182-194, 10.1006/dbio.1999.9436.
17. Monica Bullejos; Peter Koopman; Delayed Sry and Sox9 expression in developing mouse gonads underlies B6-YDOM sex reversal. *Developmental Biology* **2005**, *278*, 473-481, 10.1016/j.ydbio.2004.11.030.
18. S J Palmer; P S Burgoyne; In situ analysis of fetal, prepuberal and adult XX---XY chimaeric mouse testes: Sertoli cells are predominantly, but not exclusively, XY.. *Development* **1991**, *112*, 265-268.
19. Steven C. Munger; Anirudh Natarajan; Loren L. Looger; Uwe Ohler; Blanche Capel; Fine Time Course Expression Analysis Identifies Cascades of Activation and Repression and Maps a Putative Regulator of Mammalian Sex Determination. *PLOS Genetics* **2013**, *9*, e1003630, 10.1371/journal.pgen.1003630.
20. Alexander Quinn; Peter Koopman; The Molecular Genetics of Sex Determination and Sex Reversal in Mammals. *Seminars in Reproductive Medicine* **2012**, *30*, 351-363, 10.1055/s-0032-1324718.
21. Jennifer S. Colvin; Rebecca P. Green; Jennifer Schmahl; Blanche Capel; David M. Ornitz; Male-to-female sex reversal in mice lacking fibroblast growth factor 9.. *Cell* **2001**, *104*, 875-889, 10.1016/s0092-8674(01)00284-7.

22. Sara Morais Da Silva; Adam Hacker; Vince Harley; Peter Goodfellow; Amanda Swain; Robin Lovell-Badge; Vincent Harley; Sox9 expression during gonadal development implies a conserved role for the gene in testis differentiation in mammals and birds. *Nature Genetics* **1996**, *14*, 62-68, 10.1038/ng0996-62.
23. Nathalie Josso; Isabelle Lamarre; Jean-Yves Picard; Philippe Berta; Norman Davies; Nicole Morichon; Marc Peschanski; Roland Jeny; Anti-Müllerian hormone in early human development. *Early Human Development* **1993**, *33*, 91-99, 10.1016/0378-3782(93)90204-8.
24. Nathalie Josso; Rodolfo Alberto Rey; Jean-Yves Picard; Anti-Müllerian Hormone: A Valuable Addition to the Toolbox of the Pediatric Endocrinologist. *International Journal of Endocrinology* **2013**, *2013*, 674105, 10.1155/2013/674105.
25. Ralf Werner; Hartmut Merz; Wiebke Birnbaum; Louise Marshall; Tatjana Schröder; Benedikt Reiz; Jennifer M. Kavran; Tobias Bäumer; Philipp Capetian; Olaf Hiort; et al. 46,XY Gonadal Dysgenesis due to a Homozygous Mutation in Desert Hedgehog (DHH) Identified by Exome Sequencing.. *The Journal of Clinical Endocrinology & Metabolism* **2015**, *100*, E1022-E1029, 10.1210/jc.2015-1314.
26. Humphrey Hung-Chang Yao; Wendy Whoriskey; Blanche Capel; Desert Hedgehog/Patched 1 signaling specifies fetal Leydig cell fate in testis organogenesis. *Genes & Development* **2002**, *16*, 1433-1440, 10.1101/gad.981202.
27. P. Cantó; D. Söderlund; E. Reyes; J. P. Méndez; Mutations in the Desert hedgehog (DHH) Gene in Patients with 46,XY Complete Pure Gonadal Dysgenesis. *The Journal of Clinical Endocrinology & Metabolism* **2004**, *89*, 4480-4483, 10.1210/jc.2004-0863.
28. Eske Bendsen; Anne Grete Byskov; Steen B. Laursen; Hans-Peter E Larsen; Claus Y. Andersen; Lars G. Westergaard; Number of germ cells and somatic cells in human fetal testes during the first weeks after sex differentiation.. *Human Reproduction* **2003**, *18*, 13-18, 10.1093/humrep/deg057.
29. Andrea S. Cupp; Mehmet Uzumcu; Michael K. Skinner; Chemotactic Role of Neurotrophin 3 in the Embryonic Testis That Facilitates Male Sex Determination1. *Biology of Reproduction* **2003**, *68*, 2033-2037, 10.1095/biolreprod.102.012617.
30. Andrea S. Cupp; Lino Tessarollo; Michael K. Skinner; Testis developmental phenotypes in neurotrophin receptor trkA and trkC null mutations: role in formation of seminiferous cords and germ cell survival.. *Biology of Reproduction* **2002**, *66*, 1838-1845, 10.1095/biolreprod66.6.1838.
31. Jennifer Brennan; Jeannie Karl; Blanche Capel; Divergent Vascular Mechanisms Downstream of Sry Establish the Arterial System in the XY Gonad. *Developmental Biology* **2002**, *244*, 418-428, 10.1006/dbio.2002.0578.

32. Terje Svingen; Peter Koopman; Building the mammalian testis: origins, differentiation, and assembly of the component cell populations. *Genes & Development* **2013**, *27*, 2409-2426, 10.1101/gad.228080.113.
33. R. Sekido; R. Lovell-Badge; Genetic Control of Testis Development. *Sexual Development* **2013**, *7*, 21-32, 10.1159/000342221.
34. A Mayerhofer; G Lahr; K Seidl; B Eusterschulte; A Christoph; M Gratzl; The neural cell adhesion molecule (NCAM) provides clues to the development of testicular Leydig cells.. *Journal of Andrology* **1996**, *17*, 223-230.
35. Tony DeFalco; Satoru Takahashi; Blanche Capel; Two distinct origins for Leydig cell progenitors in the fetal testis.. *Developmental Biology* **2011**, *352*, 14-26, 10.1016/j.ydbio.2011.01.011.
36. Amanda Swain; Sex Determination: Time for Meiosis? The Gonad Decides. *Current Biology* **2006**, *16*, R507-R509, 10.1016/j.cub.2006.06.009.
37. Liangbiao Hu; Ana Monteiro; Heather Johnston; Peter King; Peter J O'Shaughnessy; Expression of Cyp21a1 and Cyp11b1 in the fetal mouse testis. *Reproduction* **2007**, *134*, 585-591, 10.1530/rep-07-0133.
38. Jennifer Brennan; Christopher Tilmann; Blanche Capel; Pdgfr- α mediates testis cord organization and fetal Leydig cell development in the XY gonad. *Genes & Development* **2003**, *17*, 800-810, 10.1101/gad.1052503.
39. R. Ann Word; Fredrick W. George; Jean D. Wilson; Bruce R. Carr; Testosterone Synthesis and Adenylate Cyclase Activity in the Early Human Fetal Testis Appear to Be Independent of Human Chorionic Gonadotropin Control*. *The Journal of Clinical Endocrinology & Metabolism* **1989**, *69*, 204-208, 10.1210/jcem-69-1-204.
40. K. C. Jonas; Olayiwola O Oduwole; Hellevi Peltoketo; Susana B Rulli; Ilpo T Huhtaniemi; Mouse models of altered gonadotrophin action: insight into male reproductive disorders. *Reproduction* **2014**, *148*, R63-R70, 10.1530/rep-14-0302.
41. Hannie Kremer; Robert Kraaij; Sérgio P.A. Toledo; Miriam Post; Julia B. Fridman; César Y. Hayashida; Margo Van Reen; Edwin Milgrom; Hans-Hilger Ropers; Edwin Mariman; et al. Axel P.N. Themmen Han G. Brunner Male pseudohermaphroditism due to a homozygous missense mutation of the luteinizing hormone receptor gene. *Nature Genetics* **1995**, *9*, 160-164, 10.1038/ng0295-160.
42. J S Colvin; A C White; S J Pratt; D M Ornitz; Lung hypoplasia and neonatal death in Fgf9-null mice identify this gene as an essential regulator of lung mesenchyme.. *Development* **2001**, *128*, 2095-2106.
43. Ann M. Clark; Kristin K. Garland; Lonnie D. Russell; Desert hedgehog (Dhh) gene is required in the mouse testis for formation of adult-type Leydig cells and normal development of peritubular

- cells and seminiferous tubules.. *Biology of Reproduction* **2000**, *63*, 1825-1838, 10.1095/biolreprod.63.6.1825.
44. Maëlle Pannetier; Anne-Amandine Chassot; Marie-Christine Chaboissier; Eric Pailhoux; Involvement of FOXL2 and RSPO1 in Ovarian Determination, Development, and Maintenance in Mammals. *Sexual Development* **2016**, *10*, 167-184, 10.1159/000448667.
45. Anne-Amandine Chassot; Isabelle Gillot; Marie-Christine Chaboissier; R-spondin1, WNT4, and the CTNNB1 signaling pathway: strict control over ovarian differentiation. *Reproduction* **2014**, *148*, R97-R110, 10.1530/rep-14-0177.
46. Sara Tomaselli; Francesca Megiorni; Lin Lin; Maria Cristina Mazzilli; Dianne Gerrelli; Silvia Majore; Paola Grammatico; John C. Achermann; Human RSPO1/R-spondin1 Is Expressed during Early Ovary Development and Augments β -Catenin Signaling. *PLOS ONE* **2011**, *6*, e16366, 10.1371/journal.pone.0016366.
47. E K Ungewitter; H.H.-C. Yao; E.K. Ungewitte; How to make a gonad: cellular mechanisms governing formation of the testes and ovaries.. *Sexual Development* **2012**, *7*, 7-20, 10.1159/000338612.
48. Anna Biason-Lauber; Marie-Christine Chaboissier; Ovarian development and disease: The known and the unexpected. *Seminars in Cell & Developmental Biology* **2015**, *45*, 59-67, 10.1016/j.semcd.2015.10.021.
49. Hitomi Suzuki; Masami Kanai-Azuma; Yoshiakira Kanai; From Sex Determination to Initial Folliculogenesis in Mammalian Ovaries: Morphogenetic Waves along the Anteroposterior and Dorsoventral Axes. *Sexual Development* **2015**, *9*, 190-204, 10.1159/000440689.
50. Gwenn-Aël Carre; Andy Greenfield; Characterising Novel Pathways in Testis Determination Using Mouse Genetics. *Sexual Development* **2014**, *8*, 199-207, 10.1159/000358402.
51. Minna Heikkilä; Renata Prunskaitė; Florence Naillat; Petri Itäranta; Jussi Vuoristo; Juhani Leppäluoto; Hellevi Peltoketo; Seppo Vainio; The Partial Female to Male Sex Reversal in Wnt-4-Deficient Females Involves Induced Expression of Testosterone Biosynthetic Genes and Testosterone Production, and Depends on Androgen Action. *Endocrinology* **2005**, *146*, 4016-4023, 10.1210/en.2005-0463.
52. Raphael H. Rastetter; Pascal Bernard; James S. Palmer; Anne-Amandine Chassot; Huijun Chen; Patrick S. Western; Robert G. Ramsay; Marie-Christine Chaboissier; Dagmar Wilhelm; Marker genes identify three somatic cell types in the fetal mouse ovary. *Developmental Biology* **2014**, *394*, 242-252, 10.1016/j.ydbio.2014.08.013.
53. N. Henriette Uhlenhaut; Susanne Jakob; Katrin Anlag; Tobias Eisenberger; Ryohei Sekido; Jana Kress; Anna-Corina Treier; Claudia Klugmann; Christian Klasen; Nadine I. Holter; et al. Dieter RiethmacherGünther SchützAustin J. CooneyRobin Lovell-BadgeMathias Treier Somatic Sex

- Reprogramming of Adult Ovaries to Testes by FOXL2 Ablation. *Cell* **2009**, *139*, 1130-1142, 10.1016/j.cell.2009.11.021.
54. Mark A. Edson; Ankur K. Nagaraja; Martin M. Matzuk; The mammalian ovary from genesis to revelation.. *Endocrine Reviews* **2009**, *30*, 624-712, 10.1210/er.2009-0012.
55. Katja Hummitzsch; Helen F. Irving-Rodgers; Nicholas Hatzirodos; Wendy Bonner; Laetitia Sabatier; Dieter P. Reinhardt; Yoshikazu Sado; Yoshifumi Ninomiya; Dagmar Wilhelm; Raymond J. Rodgers; A New Model of Development of the Mammalian Ovary and Follicles. *PLOS ONE* **2013**, *8*, e55578, 10.1371/journal.pone.0055578.
56. Jost, A.; Problems of fetal endocrinology: The gonadal and hypophyseal hormones. *Horm. Res.* **1953**, *8*, 379-418.
57. Berenice B. Mendonca; Elaine M.F. Costa; Alicia Belgorosky; Marco Aurelio Rivarola; Sorahia Domenice; 46,XY DSD due to impaired androgen production. *Best Practice & Research Clinical Endocrinology & Metabolism* **2010**, *24*, 243-262, 10.1016/j.beem.2009.11.003.
58. Analía V. Freire; Romina P. Grinspon; Rodolfo A. Rey; Importance of Serum Testicular Protein Hormone Measurement in the Assessment of Disorders of Sex Development. *Sexual Development* **2017**, *12*, 30-40, 10.1159/000479572.
59. Nathalie Josso; Anti-Müllerian hormone : a look back and ahead. *Reproduction* **2019**, *null*, , 10.1530/rep-18-0602.
60. Sandrine Imbeaud; Emmanuelle Faure; Isabelle Lamarre; Marie-Geneviève Mattei; Nathalie Di Clemente; Richard Tizard; Danièle Carré-Eusèbe; Corinne Belville; Lars Tragethon; Christopher Tonkin; et al. Janice Nelson Michele McAuliffe Jean-Michel Bidart Abdul Lababidi Nathalie Josso Richard L. Cate Jean-Yves Picard Marie-Geneviève [Egrave] Insensitivity to anti-Müllerian hormone due to a mutation in the human anti-Müllerian hormone receptor. *Nature Genetics* **1995**, *11*, 382-388, 10.1038/ng1295-382.
61. Malcolm M Moses; Richard R Behringer; A gene regulatory network for Müllerian duct regression.. *Environmental Epigenetics* **2019**, *5*, dvz017, 10.1093/eep/dvz017.
62. Richard J. Auchus; Walter L. Miller; Defects in Androgen Biosynthesis Causing 46,XY Disorders of Sexual Development. *Seminars in Reproductive Medicine* **2012**, *30*, 417-426, 10.1055/s-0032-1324726.

Retrieved from <https://encyclopedia.pub/entry/history/show/35515>