

Aerobic Granular Sludge

Subjects: Engineering, Environmental

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The aerobic granular sludge (AGS) method, which has been developed for several years, may represent an alternative to traditional technologies. One of the barriers to AGS deployment is the limited knowledge on the determinants and efficiency of the anaerobic digestion (AD) of AGS, as little research has been devoted to it.

Keywords: aerobic granular sludge ; anaerobic digestion ; wastewater treatment ; biogas

1. Introduction

The ever-more stringent quality standards for wastewater treatment effluents call for the development and deployment of efficient, commercially viable, and environmentally friendly wastewater treatment methods ^{[1][2]}. These criteria are largely met by established methods for microbial biodegradation and removal of pollutants, whether via conventional activated sludge (CAS) suspended in the wastewater or biofilm deposited on packing elements ^{[3][4]}. Nevertheless, in many cases, these technologies can be substituted with the aerobic granular sludge (AGS) method, which has been developed for several years ^{[5][6]}. Cases in which the implementation of AGS technology should be considered include the following possibilities: simplifying the technology by resigning from a multi-reactor sewage treatment line ^[7], changing the qualitative characteristics of the sewage flowing into the treatment plant to which AGS is relatively resistant ^[8], shortening the work cycle reactors ^[9], elimination of variable oxygen conditions in order to remove biogenic compounds ^[10], resignation from sludge separation devices ^[11], shortening the sedimentation phase ^[12], resignation from the system of recirculation pumps and agitators ^[13], reducing energy consumption ^[14].

AGS sewage treatment systems are now widely accepted as promising and forward- looking solutions due to their high technical readiness level, optimized processes for cultivating stable granules, as well as established and verified pollutant biodegradation parameters ^[15]. AGS has a number of clear advantages over CAS. These include, in particular: versatility, well-established treatment performance across different types of effluent, better and faster pollutant removal, improved settleability (and, thus, shorter retention times in AGS separation systems), and reduced bioreactor area/size ^[16]. This translates to lower investment and operating costs for wastewater treatment, bolstering the commercial competitiveness of AGS technology. Given these benefits, as well as the processing advantages, it is no wonder that AGS has been growing in popularity among researchers and municipal plant operators ^[9]. The method has been proven to be effective in full-scale systems for municipal/urban wastewater treatment, as well in biodegradation of various industrial pollutants. Large-scale systems have been designed and commissioned all around the world in Nereda® technology (Royal HaskoningDHV, Amersfoort, The Netherlands) ^{[17][18]}.

Recent years have seen a rapid spread of AGS wastewater treatment systems, which help identify and fully explore the strengths of the technology, as well as find solutions to existing technical issues and emergent operational hurdles ^[19]. By mid-2021, the number of AGS wastewater treatment plants was close to 90 facilities, meaning that it had more than doubled since 2018 ^[20]. The facilities ranged from 100 to 600,000 m³ in size and were designed primarily for full biodegradation of organic matter or nitrogen/phosphorus removal from household sewage and from mixed household–industrial sewage ^[21]. An analysis of data provided by AGS-deploying companies shows there were 13 new full-scale wastewater treatment plants being constructed and commissioned in 2020–2021. A further 11 are being discussed and designed, to be commissioned by 2025 ^[21], which speaks to the rapidly growing take-up of the technology. Many commissioned AGS installations are actually retrofitted CAS systems, modified to improve pollutant removal and ensure compliance with stringent quality requirements for wastewater-treatment effluent ^[22]. The number of full-scale AGS installations is provided in **Figure 1** ^[21].

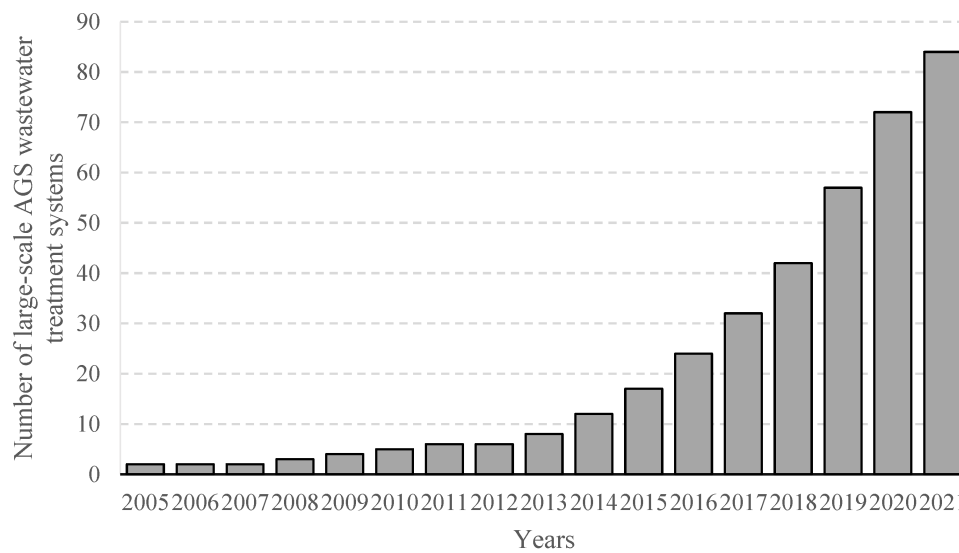


Figure 1. Worldwide number of large-scale AGS wastewater treatment systems from 2005 to 2021.

AGS is an advanced, optimized and well-explored technology incorporated into numerous large-scale installations. Nevertheless, it does have certain well-documented drawbacks that preclude its large-scale deployment and take-up. Researchers and operators have reported issues with granulated biomass instability, especially at longer running times [23] [24]. This very often leads to diminished performance of AGS-separation systems, and may also cause the plant effluent to become re-contaminated with the dispersed bacterial suspension. This also leads to higher levels of pollutants detected during the treatment process [25].

Another barrier to the competitiveness of AGS is the limited knowledge on how to manage and, ultimately, neutralize the resultant surplus sludge. One of the most popular ways of militating or fully eliminating the nuisance-inducing sludge is by using anaerobic digestion (AD) [26]. This is a very well explored technology, commonly used as one of the steps in CAS processing to stabilize sludge, remove organic matter, reduce sludge volume, improve dewaterability, reduce sanitary indicators, limit nuisance smells, improve fertilizing properties and capture methane-rich biogas [27]. Due to the different properties and characteristics of AGS, the processes currently in use need to be tested for suitability and effectiveness in anaerobic treatment of sewage sludge. This means that the underpinnings and technological parameters of the AD process need to be validated and adapted to a substrate with a different chemical composition and properties. Relatively little research to date has focused on analyzing and optimizing anaerobic digestion of AGS, so there is a real need to review the existing findings and find prospective avenues for future research and practical efforts that could further the scientific understanding and practical applicability of the process.

2. AGS Characteristics and Applications

AGS, as defined by the International Water Association (IWA), is made up of aggregates of microbial origin, which do not coagulate under reduced hydrodynamic shear, and which settle significantly faster than activated sludge flocs [28]. AGS granules are spherical or elliptical in shape. Their morphology is determined by the technological parameters of the wastewater treatment process (Table 1), including the pollutant load, the age of the sludge, the intensity of aeration and stirring, the type of feedstock, as well as any alternations to the design and operation cycle of the biological reactor [29]. The influence of operating conditions on the AGS characteristics, as well as graphics and photos of granules, have been presented in many scientific publications [30][31][32].

Table 1. The influence of operating conditions on AGS characteristics.

| Operating Conditions | Impacts on Granulation Process | References |
|----------------------------|---|------------|
| Additives Metal Cations | Ca^{2+} , Mg^{2+} , $\text{Fe}^{2+}/\text{Fe}^{3+}$ intensify granulation by neutralizing negatively charged sludge particles and enhancing adsorption/bridging interactions. | [23][33] |
| Aerobic starvation | Granulation is initiated by the lack of nutrients, increasing shear force and an increase in the hydrophobicity of the bacteria. | [34] |
| Coagulant or inert carrier | Effect on the neutralization of negatively charged particles, which promotes aggregation and adsorption of the flocs. The large surface area of coagulants and inert carriers increases the granulation efficiency. | [21][33] |

| Operating Conditions | Impacts on Granulation Process | References |
|--|---|------------|
| Extracellular polymeric substances (EPS) | EPS aggregates bacterial cells and other solid particles to a granule precursor. The high content of EPS in the system allows the granules to withstand high values of hydraulic and pollutant loading. | [23][35] |
| Food to microorganism F/M ratio | A high F/M ratio facilitates the formation of large granules. Finding the right F/M ratio is essential for achieving a fast and stable granulation. | [36][37] |
| Hydraulic retention time (HRT) | Increasing HRT reduces OLR, which limits the granulation efficiency, hinders sedimentation and leads to a decrease in biomass concentration in the technological system, as well as the size and stability of granules. | [36][38] |
| Hydrodynamic shear force | It regulates the growth of fibres, the porosity and density of the granules as well as the stability of granulation. Higher hydrodynamic shear provides better compaction and density of the granules. | [21][39] |
| Organic loading rate (OLR) | High OLR allows for quick and efficient granulation, while delayed and difficult granulation formation was observed with low OLR. | [36][40] |
| Seeding sludge | Type of seed pellets may contain cations and other properties that can help speed up the granulation process. It also acts as a nucleus that promotes the attraction of sludge flocs. | [21][33] |
| Settling period | Removes poorly settling, flocculent sludge, enabling the deposition of appropriate granules and the selection of appropriate species of microorganisms. | [21][33] |
| Sludge retention time SRT | Prolonged SRT causes deterioration of aerobic granulation, discharge of aging granular sludge and retention of appropriate newly synthesized granules is required for the stability of the aerobic granular sludge process, while shorter SRT results in a reduction of the size of the sludge flocs. | [41][37] |
| Temperature | Granulation was successfully carried out in the temperature range of 8–30 °C. It was proven that low temperatures caused an increase in fiber content, causing leaching of bacterial cells and instability of granules. | [41][41] |
| Volumetric exchange ratio | High volumetric exchange rates increase the granulation, facilitating the formation and improving the sedimentation properties of the granules. | [36][40] |

Growing AGS takes time, which can be up to three months in extreme cases, and requires the maintenance of optimal conditions, including suitable hydraulic parameters and organic loads [42]. Granule formation is facilitated by the hydrophobic surfaces of the microorganisms. According to thermodynamics, an increase in bacterial hydrophobicity leads to a reduction in the Gibbs energy of the cell, which causes microbial cells to join together and form aggregates [43]. AGS have been shown to be up to three times more hydrophobic than inoculated CAS. The cell hydrophobicity is an important inducing and maintaining force for cell-to-cell immobilization and cell-to-carrier surface attachment [44][45]. The specific gravity of the oxygen granules is positively related to the hydrophobicity of the cell. High cell hydrophobicity intensifies the intercellular interaction, results in more efficient sedimentation and favors a more stable structure of the AGS community [46]. There are two biological causes that could potentially be responsible for the increase in cell hydrophobicity during bio-granulation [47]: one of them is the community variability in multispecies aggregation of microorganisms, and the other is the change of properties on the cell surface (mainly EPS closely related to the cell surface). It has been shown that the composition of EPS, especially extracellular protein, correlates with the hydrophobicity of cells in the aggregation of microorganisms [48]. The more hydrophobic proteins in AGS are responsible for increasing the adhesion potential, keeping adjacent microbial cells together, accelerating granulation, and maintaining granule stability [49].

The extracellular polymeric substances (EPS) excreted by microorganisms play a major role in AGS formation [48]. EPS are mostly composed of proteins and polysaccharides that form a matrix which immobilizes the activated sludge bacteria within [30]. One of the roles of EPS is that it changes the surface charge of cells, thus reducing their electrostatic repulsive force [50]. Filamentous microorganisms play a huge role in the first phase of AGS formation by serving as a scaffold upon which individual bacterial cells can settle [51]. A diagram showing how a single AGS granule is formed is presented in **Figure 2**.

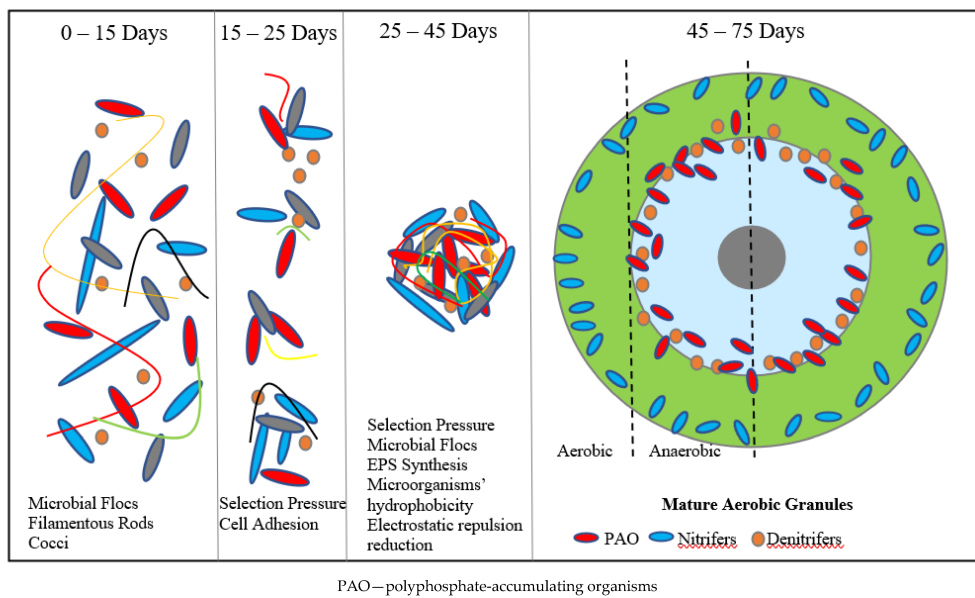


Figure 2. Illustrative chart of AGS granulation process.

The color of the AGS depends on the chemical composition of the wastewater, the process parameters, the reactor design and the taxonomic composition of the granule-forming microorganisms [52]. Granule sizes range between 0.2 and 16.0 mm [53]. Higher pollutant loads in the biomass have been shown to produce larger AGS sizes, whereas smaller granules are formed in reactors with limited availability of organic feedstock over a long period of time [54]. The specific gravity of AGS ranges from 1.004 to 1.100 kg dry matter (DM)/m³, which is much more than that of CAS, with water content between 94 and 97%, compared with 99% for CAS [55]. The volume index of AGS is less than 50 cm³/g DM, and can even reach 20 cm³/g DM in some cases. The surface structures of AGS are peppered with pores, fissures, cracks and channels, allowing feedstocks and metabolites to be carried between the waste and AGS interior [56]. AGS porosity is lower than CAS, and can decrease further as the individual granules become bigger. Pore blockage can hamper feedstock diffusion, which limits microbial activity in the AGS [57]. AGS can be kept in storage for several weeks to several months. The properties of the stored sludge are determined by temperature and the type of medium [58]. However, storage leads to biomass liquefaction and progressive disintegration of AGS [59]. A comparison of AGS and CAS properties is presented in **Table 2**.

Table 2. Comparison of AGS and CAS characteristics.

| Parameter | Unit | Value | | Reference |
|-------------------------|------|---|---|-----------|
| | | AGS | CAS | |
| Shape | - | Compact and spherical granular structure | Irregular and flocculent structure | [39] |
| Size | µm | >200 | 50–300 | [41] |
| Settling velocity | m/h | 10–130 | 2–10 | [60] |
| Specific gravity | - | 1.010–1.017 | 0.997–1.01 | [7] |
| | | 1.004–1.100 | 1.002–1.106 | [55] |
| Water content | % | 94–97 | 99 | [55] |
| Sludge Volume Index | | | | |
| 5 min | mL/g | 30–60 | – | [39][61] |
| 30 min | | 30–60 | 110–160 | |
| Redox microenvironments | - | Aerobic, anoxic, and anaerobic microbial layers | Minimum feasibility for anaerobic zones | [7][62] |
| EPS synthesis | - | High EPS content in aerobic granules as compared to CAS | Lower EPS content | [7][63] |
| OLR | - | Capable of withstanding high OLR | Poor removal performance at high OLR | [7][63] |

| Parameter | Unit | Value | CAS | Reference |
|---|------|--|---|-----------|
| | | AGS | | |
| Resistance to shock and fluctuating OLR | - | Able to remove pollutants under shock or fluctuating OLR | Poor removal under shock or fluctuating OLR | [7] |
| Tolerance to toxic compounds | - | Higher tolerance to toxic pollutants | Lower tolerance to toxic pollutants | [7][63] |

The AGS structure can house all kinds of organisms: heterotrophic and autotrophic, aerobic, facultative anaerobic, and obligate anaerobic at the deeper layers of the granule. *Accumulibacter* spp. has been shown to be the primary species at depths up to 200 μm , whereas *Competibacter* spp. is the predominant species in the core [64]. In another experiment, the ammonia-oxidizing *Nitrosomonas* spp. was found at a depth of 70–100 μm from the edge of the granule, whereas anaerobic bacteria resided at a depth of 800 μm and below [65].

The taxonomic structures in the granules change depending on wastewater composition. Granules grown on brewery/malthouse wastewater were colonized mainly by the filamentous *Thiothrix* sp. and *Sphaerotilus natans* [66]. Other researchers have noted that reactors fed with glucose had more filamentous bacteria, whereas acetate grown granules mostly contained bacilliforms [65]. The species composition may be shaped by temperature and dissolved oxygen in the reactor. Song et al. [67] have found *Thermomonas* to be present in the granules at 25 °C, whereas *Curtobacterium ammoniigenes* and *Ottowia* prevailed at 30 °C. *Thiothrix* sp. proliferates in low-oxygen conditions [68], *Microthrix parvicella* can colonize granules at a wide range of oxygen levels [69]. Thanks to the anaerobic and low-oxygen zones, the granules support simultaneous nitrification, denitrification, partial oxygenation of ammoniacal nitrogen into nitrate nitrogen (III), and Anammox processes [70]. Granules are more stable with the presence of slow-growing Anammox bacteria, as well as first- and second-step nitrifying bacteria [71].

AGS has been grown and used to treat dairy [72] and brewery wastewater [73], landfill leachate [74], and municipal sewage [7]. AGS can also be used to treat toxic wastewater. Adapted granules have been demonstrated to effectively degrade phenol at levels of up to 5000 mg/dm³ [75]. After a long acclimatization period, acetate-grown granules are able to completely degrade 4-chlorophenol [76]. Phenol derivatives can also be removed when labile carbon is provided as a secondary feedstock [77]. Aerobic granules have been demonstrated to successfully degrade methyl tert-butyl ether with ethanol as a co-feedstock. Ester removal can reach 100%, with the removal rates positively correlating with its levels in the wastewater [78]. Example uses of AGS in the treatment of wastewater of various origins and characteristics are presented in **Table 3**.

Table 3. Avenues of AGS use in wastewater treatment.

| Type of Sewage/Waste | Initial Concentration [mg/dm ³] | Reactor Configuration and Operation Conditions | Removal Efficiency [%] | Final Concentration [mg/dm ³] * | Reference |
|--|---|--|--|--|-----------|
| Dairy wastewater | COD: 2800; TN: 40; TP: 30 | 12 L SBR; Cycle period: 8 h | COD: 90; TN: 80; TP: 67 | COD: 28; TN: 8; TP: 9.9 | [79] |
| Household wastewater | COD: 506; BOD ₅ : 224; TN: 49.4; AN: 39 TP: 6.7; | Two SBR tanks with height of 7.5 m and working volume of 9600 m ³ each; Cycle: 6.5 h in dry season, 3 h in rainy season; VER: 65% | COD: 88; BOD ₅ : 96; TN: 86; AN: 97; TP: 87 | COD: 60.7; BOD ₅ : 9; TN: 6.9; AN: 1.2; TP: 0.9 | [13] |
| Household sewage (40%) + industrial wastewater (60%) | COD: 1000; AN: 60 | SBR; Height: 100 cm and Diameter: 20 cm; Cycle period: 4 h; VER: 50% | COD: 80; TN: 98 | COD: 200; AN: 1.2 | [80] |
| Livestock wastewater | COD: 3600; TN: 650; TP: 380 | 4 L SBR; Cycle period: 4 h; VER: 50%; 27–30 °C | COD: 74; TN: 73; TP: 70 | COD: 93.6; TN: 175.5; TP: 114 | [81] |
| Palm oil mill effluents | COD: 69500; AN: 45 | 3 L SBR; Cycle period: 3 h; VER: 50% | COD: 91.1; AN: 97.6 | COD: 69500; AN: 45 | [82] |
| Septic wastewater | COD: 971; AN: 80; TP: 19; SS: 670 | Three SBR tanks (7 m height) with maximum capacity of 5000 m ³ d ⁻¹ | COD: 94; AN: 99; TP: 83.5; SS: 98 | COD: 58.3; AN: 0.8; TP: 3.1; SS: 13.4 | [61] |

| Type of Sewage/Waste | Initial Concentration [mg/dm ³] | Reactor Configuration and Operation Conditions | Removal Efficiency [%] | Final Concentration [mg/dm ³] * | Reference |
|--|---|---|-------------------------------------|---|-----------|
| Slaughter house wastewater | COD: 1250 ± 150; AN: 120 ± 20; TP: 30 ± 5 | 20 L volume SBR; Cycle period: 6 h; VER: 50%; 18–22 °C | COD: 95.1; AN: 99.3; TP: 83.5 | COD: 6.1; AN: 0.8; TP: 5.0 | [83] |
| Rubber industry wastewater | COD: 1850; TN: 248; AN: 49 | 0.6 L SBR; Cycle period: 3 h; VER: 50%; 27 ± 1 °C | COD: 96.5; TN: 89.4 AN: 94.7 | COD: 64.75; TN: 2.6 AN: 94.7 | [84] |
| Synthetic wastewater with phenol | 200 1000 | 1 L SBR, 12 h cycle, 2 cycles per day, 30 °C, 200 rpm | 100 97 | - 30 | [85] |
| Synthetic wastewater with p-Nitrophenol (PNP) | 200 | 1 L SBR, 75% VER; 24 h cycle with 23.5 h aeration, 1.6 cm/s SAV, 30 °C | 100 | - | [86] |
| Synthetic wastewater with 2,4-Dinitrotoluene (2,4-DNT) | 10 | 1 L SBR, 70% VER; 24 h cycle with 23 h aeration, 1.2 cm/s SAV, 30 °C, 100 rpm | 90 | 1 | [87] |

AN—ammonia nitrogen, BOD₅—biochemical oxygen demand, COD—chemical oxygen demand, rpm—rotation speed, SAV—superficial air velocity, SBR—sequencing biological reactor, TN—total nitrogen, TP—total phosphorus, VER—volumetric exchange ratio, * own calculations.

The first pilot AGS-growing plant was constructed in Ede (Netherlands) in 2003 and consisted of two parallel batch reactors with a volume of approx. 1.5 m³ [88]. Another pilot-scale installation for treating municipal wastewater was established at the Zhuzhuanjing sewage treatment plant in Hefei (China), consisting of two 1 m³ batch reactors [89]. The first aerobic-granule sewage treatment plant was commissioned in Gansbaai. It was designed for a 4000 m³/d flow capacity and consisted of three parallel batch reactors, 7 m high and 8 m in diameter. A full-scale aerobic-granule system also operates in Epe (Netherlands). The facility is supplied with municipal and food-industry effluent at 8000 m³/h [20][39]. Li et al. [22] have described an aerobic-granule municipal wastewater treatment plant in Yanchang. The system is fed with 50,000 m³/d wastewater, achieving full biomass granulation after a year's operation. Carbon, ammoniacal nitrogen and total nitrogen removal efficiencies were reported at 85%, ~96% and ~60%, respectively. Selected AGS wastewater treatments systems are presented in **Table 4**.

Table 4. Examples of full-scale AGS wastewater treatment systems.

| Location | Start-Up | Flow Rate [m ³ /d] | Wastewater | Removal Efficiency [%] | Reference |
|--------------------------|----------|-----------------------------------|--|--|--------------|
| Deodoro, Brazil | 2016 | Phase I—64,800 Phase II—86,400 | Municipal | COD: >90; TN: >60; TP: >50 | [39][90] |
| Epe, Netherlands | 2011 | 8000 | Municipal and food-industry | COD: 96.9; BOD ₅ : >99.4; TN: >94.7; AN: 99.8; TP: 97.2; total suspended solids (TSS): >98.5 | [20][39][91] |
| Gansbaai, South Africa | 2009 | 4000 | Municipal (with a high proportion of industrial slaughterhouse effluent) | COD: 94; TN: 90; TP: >80 | [39][90] |
| Garmerwolde, Netherlands | 2013 | 30,000 | Municipal | COD: 89.2; BOD ₅ : 96.0; TN: 86.0; TP: 90.3; TSS: 96.4 | [92] |
| Kingaroy, Australia | 2016 | 2625 | Municipal | COD: >90; TN: 95; TP: >90 | [39][90] |

| Location | Start-Up | Flow Rate [m ³ /d] | Wastewater | Removal Efficiency [%] | Reference |
|----------------|----------|-------------------------------|--|--|-----------|
| Lubawa, Poland | 2017 | 3200 | Mainly municipal, 30–40% dairy effluent | COD: 97.0; BOD ₅ : 98.2; TN: 87.0; AN: 99.4 TP: 95.4; | [93] |
| Ryki, Poland | 2015 | 5320 | Municipal | COD: >90; TN: >90; TP: >90 | [39][90] |
| Yancang, China | 2008 | 50,000 | Urban (30% domestic sewage and 70% industrial wastewater from printing and dyeing, chemical, textile and beverage) | COD: 85; TN: 59.6; AN: 95.8 | [22] |

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