# Long Non-Coding RNAs in Triple-Negative Breast Cancer

Subjects: Biochemistry & Molecular Biology

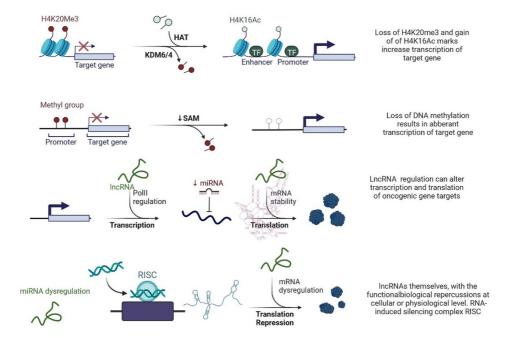
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Triple-negative breast cancer (TNBC) is a more aggressive type of breast cancer due to its heterogeneity and complex molecular mechanisms. TNBC has a high risk for metastasis, and it is difficult to manage clinical conditions of the patients. Long non-coding RNAs (lncRNAs) have emerged as a novel target to treat the multistep process of TNBC. LncRNAs regulate epigenetic expression levels, cell proliferation and apoptosis, and tumour invasiveness and metastasis. Thus, lncRNA-based early diagnosis and treatment options could be helpful, especially for patients with severe TNBC.

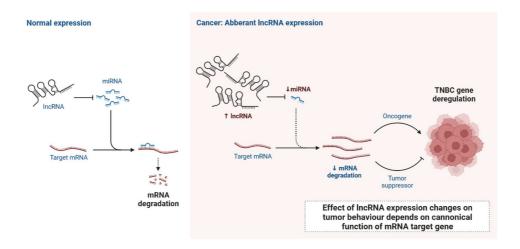
Keywords: triple-negative breast cancer; IncRNA; diagnosis; targeted drug development and resistance

### 1. LncRNAs

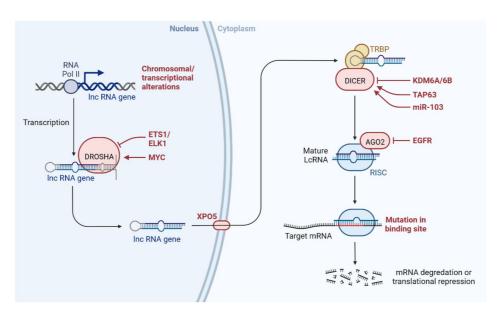
IncRNAs are actively involved in gene expression, epigenetic deregulation, chromatin remodelling, DNA methylation, translation of oncogenic gene targets, and biogenesis (Figure 1). They are transcribed by RNA polymerase II, after which most transcripts are spliced, and are mainly found in the nucleus and chromatin, being expressed in cells and tissues in a specific manner [1][2][3]. Transcriptional regulation and various molecular processes in the cytoplasm are controlled by IncRNAs; various circulating IncRNAs are transmitted via exosomes and bind to various transcription factors, chromatinregulated complexes, RNA-binding proteins, nascent RNA transcripts, and chromatin [3]. The normal expression of IncRNAs and the effect of their expression changes on tumour behaviour depends on the canonical function of the mRNA target genes (Figure 2). IncRNAs can bind to the active site of proteins and regulate molecular processes at the posttranscriptional level. They are involved in functional biological processes at the cellular or physiological levels. RNAinduced silencing complexes (RISCs) are formed with the help of lysine-specific demethylase 5B (KDM5B, also known as histone demethylase JARID1B), trimethylation of lysine 4 on the histone H3 protein subunit (H3K4me3), monomethylation of lysine 4 on the histone H3 protein subunit (H3K4me1), hsa-miR-448 (also known as miRNA448), breast cancer 1/2 (BRCA1/2), retinoblastoma protein (pRB), caveolin-1 (CAV-1), Homeobox protein Hox-A5 (HOXA5), Stratifin (SFN), methyl groups (CH3), and Ras homolog gene family, member A (RhoA) (Figure 1 and Figure 3) [4]. In 2019, it was found that the IncRNA MIR100HG regulates proliferation in TNBC and the expression of the p27 gene after formation of an RNA-DNA triplex at the promoter [5]. Moreover, MIR100HG silencing leads to reduced transcription and translation of p27 [5][6]. Three triplex-forming oligonucleotides (TFOs) have been observed on the IncRNA of p27, which binds to the triplextargeting ability (TTA) site at the 5'UTR; this event has been observed in TNBC cell lysates [7]. The binding of TFO1 and TTA is a unique mechanism by which MIR100HG regulates the transcription factors at the promoter region of p27 [7][8]. Plasmacytoma variant translocation 1 (PVT1) is another type of lncRNA that is transcribed by a gene situated at the 8q24 chromosomal region and plays and important role in TNBC development. It contains 12 exons that when spliced generate IncRNAs [9]. PVT1 binds to Krüppel-like factor 5 (KLF5) and generates a BAP1 deubiquitinase that induces TNBC via beta-catenin upregulation. Furthermore, the PVT1 promoter also acts as a regulator of the expression of the MYC protooncogene and BHLH transcription factor (c-MYC) [10]. These findings show that IncRNAs also mediate regulation at the transcriptional level.



**Figure 1.** Epigenetic deregulation in cancer including chromatin remodelling, DNA methylation, and non-coding RNA regulation that alters transcription and translation of oncogenic gene targets.



**Figure 2.** Normal expression of lncRNA and effect of lncRNA expression changes on tumour behaviour depends on canonical function of mRNA target gene.



**Figure 3.** IncRNAs are involved with the functional repercussions at the cellular and physiological level. RNA-induced silencing complex (RISC): KDM5B (lysine-specific demethylase 5B also known as histone demethylase JARID1B), H3K4me3 (trimethylation of lysine 4 on the histone H3 protein subunit), H3K4me1 (monomethylation of lysine 4 on the histone H3 protein subunit), hsa-miR-448 (also known miRNA448), BRCA1/2 (breast cancer 1/2), pRB (retinoblastoma

## 2. Clinical Updates on IncRNAs in TNBC

Recently, IncRNA expression in patients with TNBC was investigated; 1034 IncRNAs were identified using NGS technologies and microarrays, out of which, 537 IncRNAs regulate 451 protein-coding genes [11]. These genes are also detected in TNBC cells and are involved in cell signalling pathways such as the MAPK and PI3K-Akt pathways, which may lead to heterogeneity [10][11]. IncRNAs also act as miRNAs, binding to miRNA-targeted mRNAs and dysregulated miRNAs [12]. This crosstalk forms a complex post-transcriptional regulatory network including mRNAs and IncRNAs that is called the competing endogenous RNA (ceRNA) network [13]. ceRNA-mediated regulatory mechanisms constitute an important pathway in IncRNA-modulated post-transcriptional regulation in TNBC [14]. A microarray-based ceRNA network analysis revealed that 4852 IncRNAs are related to the diagnosis and treatment outcome of TNBC [15]. Another study using the TCGA database found that 150 IncRNAs are expressed at the tissue level and 823 in serum and these IncRNAs could act as prognostic factors in TNBC [16]. Furthermore, the study found that the IncRNA OSTN-AS1 is a novel immune-related prognostic marker [16]. An integrated ceRNA network involving three miRNAs (CHRDL1, FCGR1A, and RSAD2) and two IncRNAs (HIF1A-AS2 and AK124454) was developed using microarray analysis [17]. These findings demonstrate that IncRNAs play major roles in the regulation of cell signalling, genetic heterogeneity, TNBC development, and pathological features (Figure 4) shown in Table 1.

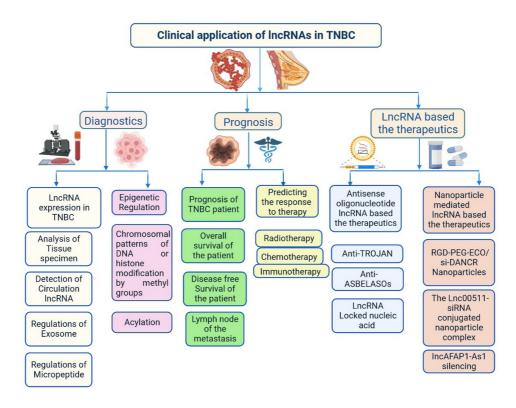


Figure 4. Clinical importance of IncRNA in triple-negative breast cancer.

**Table 1.** Important IncRNAs associated with triple-negative breast cancer.

S. N.	IncRNAs	Regulation of Expression	Clinical Importance	Potential Targets	Reference
1	HOTAIR	Upregulation	Increase cell invasion and migration	LEF1/TCF4	[18]
2	DRHC	Downregulation	Reduce cell proliferation	HOTAIR	[ <u>19</u> ]
3	LINC01133	Upregulation	Promote phenotypic features like cell stem cells (CSCs)	KLF4	[20]
4	LINC01096	Upregulation	Encourage cell invasion	miR-3130-3p	[21]
5	HEIH	Upregulation	Increase cell proliferation and prevent cell death	miR-4458/SOCS1	[ <u>22</u> ]
6	ARNILA	Downregulation	Invasion and metastasis	miR-204/SOX4	[23]
7	LINC02095	Upregulation	Promote cell proliferation	SOX9	[24]

S. N.	IncRNAs	Regulation of Expression	Clinical Importance	Potential Targets	Reference
8	WT1-AS	Downregulation	Inhibit cell migration and invasion	TGF-β1	[25]
9	GAS5	Downregulation	Promote cell apoptosis	miR-378a-5p/SUFU	[26]
10	CCAT1	Upregulation	Encourage cell division	miR-218/ZFX	[27]
11	ASRPS	Downregulation	Inhibit angiogenesis and cell proliferation	STAT3	[28]
12	AND2-AS1	Downregulation	Inhibit angiogenesis inhibit cell division	RUNX2	[29]
13	POU3F3	Upregulation	Promote cell proliferation and inhibit cell apoptosis	Caspase-9	[30]
14	NEF	Downregulation	Inhibit cell migration and invasion	miR-155	[ <u>31</u> ]
15	ZEB2-AS1	Upregulation	Promote cell proliferation, metastasis, and EMT	ZEB2	[32]
16	LINC0009	Upregulation	Increase cell proliferation and invasion	miR-383-5p/RBM3	[33]
17	ANRIL	Upregulation	Increase cell proliferation and apoptosis	miR-448/KDM5B	[ <u>34]</u>
18	SNHG12	Upregulation	Induce cell proliferation, migration, and apoptosis	MMP13	[35]
19	LUCAT1	Upregulation	Encourage cell division, movement, and invasion	miR-5702	[36]
20	PCAT6	Upregulation	Radiotherapy resistance	miR-185-5p/TPD52	[ <u>37]</u>
22	HULC	Upregulation	Promote metastasis	MMP-2, MMP-9	[38]
23	PAPAS	Upregulation	Induce cell migration and invasion	miR-34a	[ <u>39]</u>
24	HCP5	Upregulation	Increase cell proliferation; reduce cell apoptosis	miR-219a-5p/BIRC3	[40]
25	NRAD1	Upregulation	Stimulate cell proliferation and CSC-like phenotypic traits	miR-219a-5p/BIRC3	[ <u>41</u> ]
26	SNAR	Upregulation	Stimulate cell division		[ <u>42]</u>
27	AWPPH	Upregulation	Activate cell proliferation	miR-21; FZD7	<u>[43]</u>
28	sONE	Downregulation	Prevent cell proliferation	TP53/c-Myc	[44]
29	DANCR	Upregulation	Promote cell proliferation and invasion	miR-216a-5p	<u>[45]</u>
30	LINK-A	Upregulation	Increase resistance to immunotherapy, AKT inhibitors, and glycolysis reprogramming	PI3K/GPCR	<u>[46]</u>
31	MIR503HG	Downregulation	Reduce cell migration and invasion	miR-103/OLFM4	<u>[47]</u>
32	NEAT1	Upregulation	Increase cell apoptosis		[48]
33	PTCSC3	Downregulation	Prevent cell proliferation	H19	<u>[49]</u>
34	NRON	Downregulation	Inhibit cell proliferation	snaR	[ <u>50]</u>
35	TROJAN	Upregulation	Promote cell proliferation and invasion	ZMYND8	[ <u>51</u> ]
36	NAMPT-AS	Upregulation	Increase cell metastasis	miR-548b-3p/NAMPT	[11]
37	MANCR	Upregulation	Promote cell proliferation; inhibit DNA damage		[ <u>52]</u>
38	RMST	Downregulation	Prevent cell proliferation		[53]
39	SK AI1BC	Upregulation	Increase cell migration and invasion	K AI1	<u>[54]</u>
40	ROR	Upregulation	Promote cell invasion and metastasis	miR-145/ARF6	[ <u>55]</u>
41	AIRN	Downregulation	Inhibit cell migration and invasion	Wnt/β- catenin/mTOR/PI3K	<u>[56]</u>
42	LINC- ZNF469-3	Upregulation	Promote cell invasion	miR-574-5p/ZEB1	[ <u>57]</u>

S. N.	IncRNAs	Regulation of Expression	Clinical Importance	Potential Targets	Reference
43	PDCD4-AS1	Downregulation	Inhibit cell proliferation and migration	PDCD4	[ <u>58]</u>
44	HOST2	Downregulation	Inhibit cell proliferation	et-7 b/CDK6	[59]
45	BORG	Upregulation	Promote doxorubicin resistance	RPA1	[60]
46	PVT1	Upregulation	Promote cell proliferation and migration, and EMT	p21, KLF5/β-catenin	[ <u>10]</u>
47	H19	Upregulation	Promote paclitaxel resistance and CSC-like phenotypic traits	Akt	[ <u>49]</u>
48	TP73-AS1	Downregulation	Promote cell vasculogenic mimicry	miR-490-3p/TWIST1	[ <u>61</u> ]
49	TUG1	Downregulation	Enhance cisplatin sensitivity	miR-197/NLK	[62]
50	MIR100HG	Upregulation	Promote cell proliferation	p27	[63]
51	LINC01638	Upregulation	Promote cell proliferation	с-Мус	[64]

#### 2.1. Importance of IncRNAs in Tumour Invasiveness and Metastasis

Tumour invasion and metastasis explain the severity and mortality rate in patients with TNBC (**Figure 4**) [65][66]. GAS5 overexpression induces the expression of miR-196a-5p, which activates the FOXO1/PI3K/Akt signalling pathway [67]. TROJAN is a drug that reduces the metastasis burden. Degradation of TROJAN is regulated by ZMYND8, and the ubiquitin–proteasome pathway is involved in this process [68]. CCAT1 activates the migration of TNBC cells via miR-218/ZFX signalling [27]. Various ncRNAs are involved in cell migration and invasion via specific regulatory pathways, including MIR503HG through the miR-103/OLFM4 axis [47], CCAT1 through the dysregulation of the miR-218/ZFX axis [27], AFAP1-AS1 through the activation of Wnt/β-catenin signalling [69], miR-34a through the activation of EMT-associated signalling pathways [70], PAPAS through miR-34a.83 downregulation [39], soNE through soNE/NOS3/NO signalling activation [40], LINC-ZNF469-3 by activating the miR-574-5p/ZEB1 axis [58][65], ZEB2 through the activation of PI3K/Akt/GSK3β/ZEB2 signalling [32], PVT1 by regulating p21 and KLF5/β-catenin signalling [10], ARNILA by mimicking ceRNA for miR-204, AIRN by downregulating Wnt/β-catenin/mTOR/PI3K signalling [12], RMST by downregulating Wnt/β-catenin/mTOR/PI3K signalling [54], and MALAT1 by upregulating miR-129-5p and miR-1/Slug expression [71]. Furthermore, miR-448 and some other lncRNAs play very important roles in invasion and metastasis, including SKAI1BC, HULC, HOTAIR, SNHG12, SNAR, WT1-AS, LINC01096, DANCR, NEF, HIF1A-AS2, LncKLHDC7B, and ROR [17][18][19][25][35][42] [45][46][47][48][49][50][51][52][53][54][55][56][72][73]

#### 2.2. Importance of IncRNAs in Clinical Diagnosis

Several studies have found that IncRNAs are involved in the regulation of various transcription factors, epigenetic changes, chromatin remodelling, DNA methylation patterns, alternative splicing, post-translational modifications, and interaction with small peptides. All these events have great importance in the early diagnosis and treatment of patients with TNBC [11][73]. IncRNA expression levels in the blood and tissues of patients with TNBC at different stages has been investigated [11]. Based on reverse transcription quantitative PCR analysis data, the IncRNAs HIF1A-AS2, UCA1, and ANRIL can be used for TNBC detection, with areas under the curve in the range of 0.827-0.840, and a diagnostic accuracy of 0.962 for ANRIL [74]. ANRIL, SOX2OT, and ANRASSF1 are used to differentiate between healthy and TNBC cells. TINCR expression is used to differentiate various histological subtypes of BC, as it is highly expressed in TNBC cells [75]. UCA1 is associated with TNBC, acting as a specific marker for TNBC diagnosis. EZH2 is highly expressed in TNBC tissues and prevents apoptosis by activating the miR-4458/SOCS1 axis  $\frac{[76]}{}$ . LINC00299 expression is increased in TNBC. Several IncRNAs bind to mRNAs, protecting them and increasing their stability. The oncogenic transcription factor SOX9 is activated by LINC02095 [TZ]. DANCR interacts with RXRA and activates PI3K/Akt signalling in TNBC [45]. LINC00152 enhances NEDD4-1-facilitated ubiquitination and dysregulation of PTEN protein in TNBC [78]. Cell cycle arrest at the G1 phase is induced by MIR100HG, with p27 binding to RNA-DNA; p27 is a cyclin-dependent kinase (CDK) inhibitor. Cell cycle arrest at the G0/G1 phase is induced by LINC00339 and RMST in TNBC through the miR-377-3p/HOXC6 signalling pathway [5][6][64][79]. GAS5 is actively involved in the inhibition of TNBC cells through its action on miR-196a-5p and miR-378a-5p/SUFU signalling [80]. Further understanding of the roles of all these IncRNAs in TNBC is needed to improve early diagnosis and clinical management of patients. Various genes are targeted by ncRNAs, including LARP7, CDKN1A, KLF2, TIA1, DDX3X, CDK, and QKI [81][82][83][84][85]. An analysis of the TCGA database showed that 1097 IncRNAs are expressed in BC, with 1510 differentially expressed IncRNAs in TNBC cells, 35 plasma IncRNAs in TNBC, and 672 in non-TNBC cells [11]. Some lncRNAs are directly linked to prognosis in TNBC, including FOXCUT,

LINC00299, AP000924.1, AC091043.1, AL354793.1, AC010343.3, and FGF10-AS1 [11]. Plasma-specific IncRNAs are also used for diagnosis of TNBC, such as UCA1, ANRIL, and HIF1A-AS2 [17]. IncRNAs associated with lymph node metastasis, such as LINC000173, LINC00096, ZEB2-AS1, HIF1A-AS2, HULC, LUCAT1, SNHG12, MALAT1, HOTAIR, HIF1A-AS2, LINC00096, ADPGK-AS1, and ZEB2-AS1, have also shown importance in diagnosis and prognosis [11][17][36] [86]

#### 2.3. Importance of IncRNAs in Treatment

IncRNAs affect the response to treatments such as chemotherapy, immunotherapy, and radiotherapy [87]. H19 is expressed in patients with TNBC during neoadjuvant chemotherapy and is related to effective clinical outcomes. LINK-A expression is linked to response to pembrolizumab treatment in patients with TNBC because its decreased expression reduces CD8<sup>+</sup> T-cell infiltration [46]. These IncRNAs act as biomarkers for treatment response in patients with TNBC. LncAFAP1-AS1 expression has been observed in patients with TNBC who received radiotherapy after surgery, and this IncRNA acts as biomarker for radiotherapy [69]. Moreover, IncRNAs are involved in angiogenesis. LINC01133 expression is induced by mesenchymal stem/stromal cells that adjoin TNBC cells  $\frac{[20]}{}$ . IncRNAs are actively involved in the regulation of cell proliferation and apoptosis as well as drug resistance in TNBC [31][34][48][87][88]. DRHC and HOTAIR inhibit TNBC growth and development  $\frac{[18]}{}$ . HOTAIR plays a role in the invasion and migration of TNBC cells and is used as a biomarker for TNBC metastasis in circulation and tissues, indicating poor survival and response [18][19]. DRHC inhibits TNBC cell proliferation by downregulating the expression of HOTAIR, whereas HOTAIR does not affect the expression level of DRHC. H19 expression is reduced in TNBC cells, whereas PTCSC3 expression is not altered by H19 overexpression [48]. HIST2H2BC and SNRPEP4 were identified in 165 frozen tissue samples by transcriptome microarrays; these IncRNAs are involved in taxane chemotherapy in patients with TNBC. Increased miR-377-3p expression delays TNBC progression by regulating the inc00339/miR-377-3p/HOXC6 axis and inhibits TNBC proliferation and apoptosis. Therefore, it is used as therapeutic target. HIF1A-AS2 expression is upregulated in TNBC mammary tissue, which is linked to overall survival. HOTAIR is closely associated with androgen receptor expression and used as a therapeutic strategy to prevent metastasis. The miR-199a/FOXP2 pathway is induced by LINC01133 and triggers the proliferation of TNBC cells. Various IncRNAs act as stem cell markers, such as DANCR, LINC01638, LINC-ZNF469-3, NEAT1, NRAD1, and ASRPS [62][74]. Some IncRNAs promote vasculogenic mimicry, providing growth supplementation for tumour formation in TNBC. TP73-AS1, which is activated by the miR-490-3p/TWIST1 pathway, is one example. LINK-A alters glycolysis by mediating HIF1α phosphorylation at Tyr565 and Ser7 [31][34][88][89]. MANCR inhibits DNA damage and prevents disease progression [53]. AWPPH is involved in the prevention of tumourigenesis upon treatment with carboplatin; AWPPH small interfering RNA (siRNA) silencing leads to increased chemosensitivity in TNBC [43][90]. TUG1 induces the expression of miR-197, reduces the activation of WNT signalling, and enhances TNBC cell sensitivity to cisplatin [62]. These findings demonstrate the importance of IncRNAs in the prevention of tumourigenesis. More studies are required to explore IncRNA treatment options. Early studies showed that HOTAIR recruits the polycomb repressive complex 2 to its target genes through the Corest/Rest H3K4 demethylase complex [62].

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