# **Ruminant Livestock**

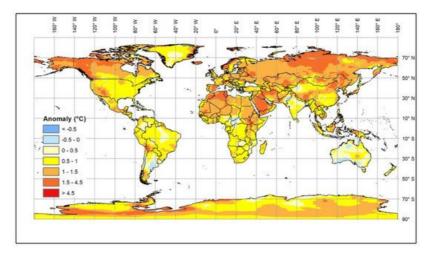
Subjects: Agriculture, Dairy & Animal Science Contributor: John Williams

After the Neolithic domestication at specific sites, animals colonized the world and adapted to live and produce in a variety of different environments. Molecular technologies permit to scan the genome of local livestock breeds in search of adaptive genes to be used in accelerated breeding schemes to mitigate the deleterious effects of climate change on livestock welfare and productivity.

Keywords: climate change ; livestock ; adaptation

### 1. Introduction

Climate change is generally causing an increase in global temperatures. The most recent estimates <sup>[1]</sup> suggest that a 1.5 °C warming compared to the 1850–1900 baseline will be reached in the second half of the current decade, but, in addition, there are longer cold periods and increased levels of solar radiation <sup>[2][3][4][5][6]</sup> (**Figure 1**).



**Figure 1.** Map of annual temperature changes in the period 1992–2020 compared to 1950–1978, created using ERA5 climate Reanalysis tools. The areas showing warming are in yellow-red and those showing cooling are in blue.

These changes affect both extensive and intensive farming systems <sup>[Z][8]</sup>. The impact of environmental changes on animals affects their health, growth, and fertility as well as the diseases to which they are exposed. In addition, availability and types of feed may change because of the impact of the climate on the production and quality of grains, pasture and forage crops <sup>[9][10]</sup>, which will affect nutrition as well as animal health and metabolism <sup>[11]</sup>. Livestock can adapt to gradual changes in environmental temperature. However, rapid changes or extended periods of extreme conditions reduce their welfare and productivity and are potentially life threatening. Therefore, the current rapid rise in global temperature is increasingly exposing livestock to stress in many countries. Some local breeds that have been kept in areas with adverse conditions, such as high temperature and humidity or drought, have become adapted over many generations; these breeds are an invaluable resource for research and breeding. It is urgent that we understand the biological mechanisms underlying their adaptability, and, in particular, to identify genomic regions and genes that control such mechanisms in order to facilitate the rapid selection of livestock resilient to climate change.

# 2. Impacts of Climate Change on Livestock

With increasing global temperatures, more productive livestock are at greater risk, because they have higher feed intake and feed consumption, which is directly related to animal heat production <sup>[12]</sup>. Animals eat less to counteract high temperatures, and nutrients are prioritized to support maintenance rather than production and reproduction. In the central U.S., for example, severe losses of beef cattle kept in feedlots have been reported because of heat waves in summer and

extreme snowstorms and wind in winter <sup>[13]</sup>. Climate related economic losses as a result of animal death and reduced performance have been seen <sup>[14]</sup>. Cattle, sheep, pigs and chickens reduce their feed intake by 3–5% for each unit increase in temperature above 30 °C <sup>[15]</sup>. Reproduction is particularly affected. Hahn <sup>[16]</sup> reported that conception rates in dairy cows are reduced by 4.6% per unit change above 70 in the temperature humidity index (THI) <sup>[17]</sup>. For beef cattle kept in range or pasture management systems, a decrease in pregnancy rates of 3.2% and 3.5% was observed for each unit increase in average THI above 70 and an increase in average temperature above 23.4 °C, respectively. Among environmental variables, temperature has the greatest effect on cow pregnancy rates <sup>[18]</sup>.

Climate change includes altered rainfall patterns that, combined with geographical factors, such as soil type, affect crop production <sup>[19][20][21]</sup>. Drought reduces biomass <sup>[22]</sup>, increases lignin accumulation in plant tissues, and reduces proteins, resulting in less digestible forages <sup>[23]</sup> and insufficient energy to meet livestock requirements <sup>[24][25]</sup>. Increased occurrence of prolonged drought is therefore of great concern to pasture-based livestock systems <sup>[23]</sup>, especially those in environments which cannot support arable production <sup>[26]</sup>.

Climate change influences the distribution of animal pathogen vectors and parasite range  $^{[22]}$  which, together with the decreased immune response of animals under stress (triggered by cortisol), exposes livestock to higher risks of disease. Early springs, warmer winters and changes in rainfall distribution affect the seasons in which pathogens, parasites and vectors are present, potentially increasing proliferation and survival of these organisms. Bluetongue recently spread northward from Africa to Europe  $^{[28]}$  as a consequence of climate-driven ecosystem changes and the associated expansion of the geographic range of the insect Culicoides imicola, the vector of the virus  $^{[29]}$ . Other vectors, such as the tick Rhipicephalus appendiculatus, which is the host for the protozoan pathogen Theileria parva, are predicted to shift their geographic range due to climate change, moving southward from central sub-Saharan Africa towards southern Africa  $^{[30]}$ . Higher temperatures in Europe have increased parasite burdens such as helminths, with a shift from species traditionally found in temperate zones such as Ostertagia ostertagi to tropically adapted species, particularly Haemonchus contortus  $^{[31][32]}$ . In addition to temperature, increased rainfall and humidity have affected the distribution of parasites. Leptospirosis in humans has been linked to transmission from livestock, with many outbreaks reported following extreme weather events around the world  $^{[33]}$ .

High resolution meteorological data are used to evaluate climate trends and variability and to predict the frequency of extreme events. Where meteorological data are not available, advanced climate modelling produces "Climate Reanalysis" datasets for a comprehensive description of the climate in three-dimensional grids. "Climate Reanalysis" has become an essential tool for modelling meteorological data to provide services to sectors dependent on climate assessments, forecasts and projections, including ecosystem management, agriculture, and livestock farming <sup>[34][35]</sup>. Climate modelling is also able to produce short- to long-term climate predictions (months to a few decades ahead), and projections extending over many decades at the global level.

# 3. Becoming Adapted

The level of heat stress experienced by an animal is the result of a combination of air temperature, relative humidity <sup>[36]</sup> and other climate factors including wind speed and solar radiation <sup>[37]</sup>. Depending on the management system, these parameters may make different contributions to the risk of thermal stress <sup>[38]</sup>. Environmental parameters can be measured and used to construct indices and set thresholds to define risk situations.

Most of the indices defining thermal stress risk have been developed for cattle, especially for dairy cows that are particularly susceptible to high temperatures. The Temperature Humidity Index (THI) <sup>[1,7]</sup> takes into account the effect of air temperature and humidity. THI was originally developed as a general indicator of heat stress for humans, but today is also applied to livestock. Over the years, the model and threshold values used to define heat stress conditions have been modified <sup>[38]</sup>, and corrections are now applied if cooling systems are used in the housing <sup>[39]</sup>. THI does not take into account the cumulative effect of high temperature <sup>[38]</sup> or the impact of wind speed and solar radiation, which are important when estimating the level of heat stress experienced by an animal. The Equivalent Temperature Index (ETI) includes air speed in the formula <sup>[40]</sup>, although solar radiation is not considered <sup>[38]</sup>. The THI adjusted (THIadj) index considers both the wind speed and the solar radiation, as well as breed and coat colour <sup>[41]</sup>. The Respiration Rate index (RR) is an extension of THIadj that also takes into account whether animals are in a shaded area or under the sun <sup>[42]</sup>.

A comprehensive review of models for predicting heat stress response in livestock is given in Rashamol et al. [43].

Senepol cattle were developed on the island of St Croix to create a breed that was polled, easily managed and tolerant of the tropical environment by crossing red polled taurine cattle with African Zebu cattle <sup>[44]</sup>. Some of these cattle have very short hair and reduced follicle density, giving the phenotype referred to as "SLICK". SLICK is controlled by a single genetic

locus and carriers of the Slick variant have lower core temperature than non-SLICK contemporaries [45]. Interestingly, the effect of SLICK is most likely through increased sweat production rather than the decrease in hair length and density [46]. The SLICK variant in Senepol cattle was initially mapped to chromosome 20 [47], and later the causative variation was identified in the prolactin receptor gene ( PRLR ). A single base deletion in exon 10 causes a frameshift that introduces a stop codon and results in the truncation of the protein [48]. Other criollo cattle breeds, such as Carora and Limonero, that were brought to the Americas from Spain 500 years ago [49] display a similar SLICK phenotype. However, these breeds do not carry the same prolactin variant that was identified in the Senepol cattle, although a genome-wide association analysis located the causative variant in or near to PLRL . DNA sequencing of SLICK Limonero cattle revealed three variants within the prolactin receptor gene that create premature stop codons in exon 11, one of which is also found in SLICK Carora cattle [50]. Recently, three novel variants were discovered in the PLRL gene in six Caribbean Basin cattle breeds. All create premature stop codons and increase heat tolerance. The occurrence of mutations in the prolactin receptor in several cattle breeds that are adapted to tropical climates and that have distinct evolutionary histories is unlikely to be by chance. Indeed, prolactin levels have been shown to be involved in thermoregulation in humans [51], showing that certain physiological processes and specific genes can be targeted by environmental pressure. The SLICK variant has now been introgressed into other breeds, including the highly productive Holstein dairy breed, creating more heat tolerant animals [<u>52</u>]

# 4. Seeking Adaptive Genes

A GWAS of cattle indigenous to Benin <sup>[53]</sup> identified several potential candidate genes associated with stress and immune response (*PTAFR, PBMR1, ADAM, TS12*), feed efficiency (*MEGF11, SLC16A4, CCDC117*), and conformation and growth (*VEPH1, CNTNAP5, GYPC*). The study of cold stress in Siberian cattle breeds identified two candidate genes (*MSANTD4* and *GRIA4*) on chromosome 15, putatively involved in cold shock response and body thermoregulation <sup>[54]</sup>. GWAS in taurine, indicine and cross-bred cattle identified *PLAG1* (BTA14), *PLRL* (BTA20) and *MSRB3* (BTA5) as candidate genes for several traits important for adaptation to extensive tropical environments <sup>[55]</sup>. A GWAS of the Frizarta dairy sheep breed, which is adapted to a high relative humidity environment, identified 39 candidate genes associated with body size traits including *TP53, BMPR1A, PIK3R5, RPL26*, and *PRKDC* <sup>[56]</sup>. An association analysis of genotype-by-environment (GxE) interactions with growth traits in Simmental cattle showed that birth weight was affected by temperature, while altitude affected weaning and yearling weight. Genes implicated in these traits included neurotransmitters (*GABRA4* and *GABRB1*), hypoxia-induced processes (*PLA2G4B, PLA2G4E, GRIN2D*, and *GRIK2*) and keratinization (KRT15, KRT31, KRT32, KRT33A, *KRT34*, and *KRT3*), all processes that play a role in physiological responses associated with adaptation to the environment <sup>[52]</sup>.

Signatures of selection related to feed adaptation have been found in sheep using an FST approach <sup>[58]</sup>. Of the seventeen genes under climatic selection, nine were related to energy metabolism. The strongest selection signal was around *TBC1D12*, on OAR22, which plays a role in GTPase regulation. The FST approach was also applied to Siberian cattle populations in order to understand the genetic basis of adaptation to cold environments <sup>[59]</sup>. Results identified several genes that have been implicated in thermal adaptation in cattle, such as *GRIA4*, *COX17*, *MAATS1*, *UPK1B*, *IFNGR1*, *DDX23*, *PPT1*, *THBS1*, *CCL5*, *ATF1*, *PLA1A*, *PRKAG1*, and *NR1I2*.

With regard to hot environments, Li and colleagues <sup>[60]</sup> investigated selection signatures of bovine heat tolerance in Dehong cattle, a Chinese indigenous zebu breed, using an F ST approach. Results indicated that genes involved in heat shock (*HSF1*), oxidative stress response (*PLCB1*, *PLCB4*), coat colour (*RAB31*), feed intake (*ATP8A1*, *SHC3*) and reproduction (*TP63*, *MAP3K13*, *PTPN4*, *PPP3CC*, *ADAMTSL1*, *SS18L1*, *OSBPL2*, *TOX*, *RREB1*, and *GRK2*) may play a role in heat adaptation.

Pairwise comparison of genetic differentiation of sheep breeds adapted to different environments identified selection signatures in the genes *MITF, FGF5, MTOR, TRHDE* and *TUBB3* that have been associated with high-altitude adaptation <sup>[61]</sup>. An FST statistic approach applied to cattle breeds reared in different environments identified several genes under positive selection for thermal tolerance <sup>[62]</sup>. HapFLK detected the Nebulin Related Anchoring Protein gene (*NRAP*) to be under selection for adaptation to cold environments <sup>[63]</sup>, *ACSS2, ALDOC, EPAS1, EGLN1* and *NUCB2* to be under selection for high-altitude adaptation in cattle <sup>[64]</sup>, and *DNAJC28, GNRH1* and *MREG* to be associated with heat stress adaptation in sheep <sup>[65]</sup>.

#### References

- Masson-Delmotte, V.; Zhai, P.; Pörtner, H.O.; Roberts, D.; Skea, J.; Shukla, P.R.; Pirani, A.; Moufouma-Okia, W.; Péan, C.; Pidcock, R.; et al. IPCC, 2018: Global Warming of 1.5 °C. An IPCC Special Report on the Impacts of Global Warming of 1.5 °C above Pre-Industrial Levels and Related Global Greenhouse Gas Emission Pathways, in the Context of Strengthening the Global Response to the Threat of Climate change, sustainable development, and efforts to eradicate poverty. 2018, in press.
- 2. Ames, D.R.; Insley, L.W. Wind-Chill Effect for Cattle and Sheep. J. Anim. Sci. 1975, 40, 161–165.
- 3. Toghiani, S.; Hay, E.H.; Roberts, A.; Rekaya, R. Impact of Cold Stress on Birth and Weaning Weight in a Composite Beef Cattle Breed. Livest. Sci. 2020, 236, 104053.
- Pezzopane, R.M.; Carlos, A.; Bernardi, C.; Azenha, M.V.; Anch, P.P.; Bosi, C.; Pedroso, F.; Esteves, N. Production and Nutritive Value of Pastures in Integrated Livestock Production Systems: Shading and Management Effects. Sci. Agric. 2020, 77, 1–9.
- 5. Belasco, E.; Cheng, Y.; Schroeder, T.C. The Impact of Extreme Weather on Cattle Feeding Profits. J. Agric. Resour. Econ. 2015, 40, 285–305.
- 6. Lees, A.M.; Sejian, V.; Wallage, A.L.; Steel, C.C.; Mader, T.L.; Lees, J.C.; Gaughan, J.B. The Impact of Heat Load on Cattle. Animals 2019, 9, 322.
- 7. Rust, W.; Holman, I.; Bloomfield, J.; Cuthbert, M.; Corstanje, R. Understanding the Potential of Climate Teleconnections to Project Future Groundwater Drought. Hydrol. Earth Syst. Sci. 2019, 23, 3233–3245.
- Thornton, P.K.; van de Steeg, J.; Notenbaert, A.; Herrero, M. The Impacts of Climate Change on Livestock and Livestock Systems in Developing Countries: A Review of What We Know and What We Need to Know. Agric. Syst. 2009, 101, 113–127.
- 9. Rotter, R.; Van de Geijn, S.C. Climate Change Effects On Plant Growth, Crop Yield and Livestock. Clim. Chang. 1999, 43, 651–681.
- 10. Wheeler, T.; Reynolds, C. Predicting the Risks from Climate Change to Forage and Crop Production for Animal Feed. Anim. Front. 2013, 3, 36–41.
- 11. Ortiz-Bobea, A.; Knippenberg, E.; Chambers, R.G. Growing Climatic Sensitivity of U.S. Agriculture Linked to Technological Change and Regional Specialization. Sci. Adv. 2018, 4, eaat4343.
- 12. Gaughan, J.B.; Cawdell-Smith, A.J. Impact of climate change on livestock production and reproduction. In Climate Change Impacts on Livestock: Adaptation and Mitigation; Sejian, V., Gaughan, J., Baumgard, L., Prasad, C., Eds.; Springer India: New Delhi, India, 2017; pp. 51–60.
- Hahn, G.; Mader, T.; Spiers, D.; Gaughan, J.; Nienaber, J.; Eigenberg, R.; Brown-Brandl, T.; Hu, Q.; Griffin, D.; Hugenford, L. Heat Wave Impacts on Feedlot Cattle: Considerations for Improved Environmental Management. In Proceedings of the Livestock Environment International Symposium Proceedings, Louisville, KY, USA, 21–23 May 2001.
- Mader, T.; Davis, S.; Gaughan, J.; Brown-brandl, T. Wind Speed and Solar Radiation Adjustments for the Temperature-Humidity Index. In Proceedings of the 16th Conference on Biometeorology and Aerobiology, Vancouver, BC, Canada, 23–27 August 2004; pp. 1–6.
- 15. National Research Council Effect of environment on utrient requirement of domestic animals. In Subcommittee on Environmental Stress; National Research Council (Ed.) The National Academies Press: Washington, DC, USA, 1981.
- 16. Hahn, G. Global Warming and Potential Impacts on Cattle and Swine in Tropical and Temperate Areas. In Proceedings of the Brazilian Congress of Biometeorology, USDA, Ljubljana, Slovenia, 1–8 September 1996.
- 17. Thom, E.C. The Discomfort Index. Weatherwise 1959, 12, 57–61.
- Amundson, J.L.; Mader, T.L.; Rasby, R.J.; Hu, Q.S. Environmental Effects on Pregnancy Rate in Beef Cattle 1. J. Anim. Sci. 2006, 84, 3415–3420.
- Shukla, P.R.; Skea, J.; Buendia, E.C.; Masson-Delmotte, V.; Pörtner, H.-O.; Roberts, D.C.; Zhai, P.; Slade, R.; Connors, S.; van Diemen, R.; et al. IPCC, 2019: Climate Change and Land: An IPCC Special Report on Climate Change, Desertification, Land Degradation, Sustainable Land Management, Food Security, and Greenhouse Gas Fluxes in Terrestrial Ecosystems. 2019, in press.
- 20. Lean, J.L.; Rind, D.H. How Will Earth's Surface Temperature Change in Future Decades? Geophys. Res. Lett. 2009, 36, 1–5.
- 21. Chapman, S.C.; Chakraborty, S.; Dreccer, F.; Howden, M. Plant Adaptation to Climate Change—Opportunities and Priorities in Breeding. Crop Pasture Sci. 2012, 63, 251–268.

- 22. Calleja-Cabrera, J.; Boter, M.; Oñate-Sánchez, L.; Pernas, M. Root Growth Adaptation to Climate Change in Crops. Front. Plant Sci. 2020, 11, 544.
- 23. Tubiello, F.N.; Soussana, J.F.; Howden, S.M. Crop and Pasture Response to Climate Change. Proc. Natl. Acad. Sci. USA 2007, 104, 19686–19690.
- 24. Hidosa, D.; Guyo, M. Journal of Fisheries & Climate Change Effects on Livestock Feed Resources: A Review. J. Fish. Livest. Prod. 2017, 5, 5–8.
- 25. Morton, J.F. The Impact of Climate Change on Smallholder and Subsistence Agriculture. Proc. Natl. Acad. Sci. USA 2007, 104, 19680–19685.
- 26. O'Mara, F.P. The Role of Grasslands in Food Security and Climate Change. Ann. Bot. 2012, 110, 1263–1270.
- Baylis, M.; Risley, C. Infectious Diseases, Climate Change Effects on. In Infectious Diseases; Kanki, P., Grimes, D., Eds.; Springer: New York, NY, USA, 2013; pp. 117–146.
- 28. Baylis, M.; Githeko, A.K. The Effects of Climate Change on Infectious Diseases of Animals; Report for the Foresight Project on Detection of Infectious Diseases; Department of Trade and Industry, UK Government: London, UK, 2006.
- 29. Wilson, A.J.; Mellor, P.S. Bluetongue in Europe: Past, Present and Future. Philos. Trans. R. Soc. London. B 2009, 364, 2669–2681.
- Olwoch, J.M.; Reyers, B.; Engelbrecht, F.A.; Erasmus, B.F.N. Climate Change and the Tick-Borne Disease, Theileriosis (East Coast Fever) in Sub-Saharan Africa. J. Arid Environ. 2008, 72, 108–120.
- Kenyon, F.; Sargison, N.D.; Skuce, P.J.; Jackson, F. Sheep Helminth Parasitic Disease in South Eastern Scotland Arising as a Possible Consequence of Climate Change. Vet. Parasitol. 2009, 163, 293–297.
- 32. Fox, N.J.; Glenn, M.; Davidson, R.S.; White, P.C.L.; Hutchings, M. Climate-Driven Tipping-Points Could Lead to Sudden, High-Intensity Parasite Outbreaks. R. Soc. Open Sci. 2015, 2, 2–14.
- Lau, C.L.; Smythe, L.D.; Craig, S.B.; Weinstein, P. Climate Change, Flooding, Urbanisation and Leptospirosis: Fuelling the Fire? Trans. R. Soc. Trop. Med. Hyg. 2010, 104, 631–638.
- Rojas-Downing, M.M.; Nejadhashemi, A.P.; Harrigan, T.; Woznicki, S.A. Climate Change and Livestock: Impacts, Adaptation, and Mitigation. Clim. Risk Manag. 2017, 16, 145–163.
- Buontempo, C.; Hutjes, R.; Beavis, P.; Berckmans, J.; Cagnazzo, C.; Vamborg, F.; Bergeron, C.; Almond, S.; Amici, A.; Ramasamy, S.; et al. Fostering the Development of Climate Services through Copernicus Climate Change Service (C3S) for Agriculture Applications. Weather Clim. Extrem. 2020, 27, 1–8.
- 36. Hill, D.; Wall, E. Dairy Cattle in a Temperate Climate: The Effects of Weather on Milk Yield and Composition Depend on Management. Animal 2015, 9, 138–149.
- 37. da Silva, V.P.R.; e Silva, R.A.; Cavalcanti, E.P.; Braga, C.C.; de Azevedo, P.V.; Singh, V.P.; Pereira, E.R.R. Trends in Solar Radiation in NCEP/NCAR Database and Measurements in Northeastern Brazil. Sol. Energy 2010, 84, 1852– 1862.
- Herbut, P.; Angrecka, S.; Walczak, J. Environmental Parameters to Assessing of Heat Stress in Dairy Cattle—A Review. Int. J. Biometeorol. 2018, 62, 2089–2097.
- St-Pierre, N.R.; Cobanov, B.; Schnitkey, G. Economic Losses from Heat Stress by US Livestock Industries. J. Dairy Sci. 2003, 86, E52–E77.
- 40. Baêta, F.C.; Meador, N.; Shanklin, M.D.; Johnson, H.D. Equivalent Temperature Index at Temperatures above the Thermoneu-Tral for Lactating Dairy Cows. In Proceedings of the Meeting of the American Society of Agricultural engineers, Chicago, IL, USA, 1987; Available online: https://agris.fao.org/agris-search/search.do?recordID=US8853966 (accessed on 21 July 2021).
- 41. Mader, T.L.; Davis, M.S.; Brown-Brandl, T. Environmental Factors Influencing Heat Stress in Feedlot Cattle. J. Anim. Sci. 2006, 84, 712–719.
- 42. Eigenberg, R.A.; Brown-Brandl, T.M.; Nienaber, J.A.; Hahn, G.L. Dynamic Response Indicators of Heat Stress in Shaded and Non-Shaded Feedlot Cattle, Part 2: Predictive Relationships. Biosyst. Eng. 2005, 91, 111–118.
- Rashamol, V.P.; Sejian, V.; Pragna, P.; Lees, A.M.; Bagath, M.; Krishnan, G.; Gaughan, J.B. Prediction Models, Assessment Methodologies and Biotechnological Tools to Quantify Heat Stress Response in Ruminant Livestock. Int. J. Biometeorol. 2019, 63, 1265–1281.
- 44. Flori, L.; Gonzatti, M.I.; Thevenon, S.; Chantal, I.; Pinto, J.; Berthier, D.; Aso, P.M.; Gautier, M. A Quasi-Exclusive European Ancestry in the Senepol Tropical Cattle Breed Highlights the Importance of the Slick Locus in Tropical Adaptation. PLoS ONE 2012, 7, e36133.

- 45. Olson, T.A.; Lucena, C.; Chase, C.C.J.; Hammond, A.C. Evidence of a Major Gene Influencing Hair Length and Heat Tolerance in Bos Taurus Cattle. J. Anim. Sci. 2003, 81, 80–90.
- Dikmen, S.; Alava, E.; Pontes, E.; Fear, J.M.; Dikmen, B.Y.; Olson, T.A.; Hansen, P.J. Differences in Thermoregulatory Ability Between Slick-Haired and Wild-Type Lactating Holstein Cows in Response to Acute Heat Stress. J. Dairy Sci. 2008, 91, 3395–3402.
- 47. Mariasegaram, M.; Chase, C.C.J.; Chaparro, J.X.; Olson, T.A.; Brenneman, R.A.; Niedz, R.P. The Slick Hair Coat Locus Maps to Chromosome 20 in Senepol-Derived Cattle. Anim. Genet. 2007, 38, 54–59.
- Littlejohn, M.D.; Henty, K.M.; Tiplady, K.; Johnson, T.; Harland, C.; Lopdell, T.; Sherlock, R.G.; Li, W.; Lukefahr, S.D.; Shanks, B.C.; et al. Functionally Reciprocal Mutations of the Prolactin Signalling Pathway Define Hairy and Slick Cattle. Nat. Commun. 2014, 5, 5861.
- 49. Martinez, A.M.; Gama, L.T.; Canon, J.; Ginja, C.; Delgado, J.V.; Dunner, S.; Al, E. Genetic Footprints of Iberian Cattle in America 500 Years after the Arrival of Columbus. PLoS ONE 2012, 7, e49066.
- Porto-neto, L.R.; Bickhart, D.M.; Landaeta-hernandez, A.J.; Utsunomiya, Y.T.; Pagan, M.; Jimenez, E.; Hansen, P.J.; Dikmen, S.; Schroeder, S.G.; Kim, E.; et al. Convergent Evolution of Slick Coat in Cattle through Truncation Mutations in the Prolactin Receptor. Front. Genet. 2018, 9, 57.
- 51. Mundel, T.; Bunn, S.J.; Hooper, P.L.; Jones, D.A. The Effects of Face Cooling during Hyperthermic Exercise in Man: Evidence for an Integrated Thermal, Neuroendocrine and Behavioural Response. Exp. Physiol. 2007, 92, 187–195.
- Dikmen, S.; Khan, F.A.; Huson, H.J.; Sonstegard, T.S.; Moss, J.I.; Dahl, G.E.; Hansen, P.J. The SLICK Hair Locus Derived from Senepol Cattle Confers Thermotolerance to Intensively Managed Lactating Holstein Cows. J. Dairy Sci. 2014, 97, 5508–5520.
- 53. Vanvanhossou, S.F.U.; Scheper, C.; Dossa, L.H.; Yin, T.; Brügemann, K.; König, S. A Multi-Breed GWAS for Morphometric Traits in Four Beninese Indigenous Cattle Breeds Reveals Loci Associated with Conformation, Carcass and Adaptive Traits. BMC Genom. 2020, 21, 783.
- 54. Igoshin, A.V.; Yurchenko, A.A.; Belonogova, N.M.; Petrovsky, D.V.; Aitnazarov, R.B.; Soloshenko, V.A.; Yudin, N.S.; Larkin, D.M. Genome-Wide Association Study and Scan for Signatures of Selection Point to Candidate Genes for Body Temperature Maintenance under the Cold Stress in Siberian Cattle Populations. BMC Genet. 2019, 20, 26.
- 55. Porto-neto, L.R.; Reverter, A.; Prayaga, K.C. The Genetic Architecture of Climatic Adaptation of Tropical Cattle. PLoS ONE 2014, 9, e113284.
- 56. Kominakis, A.; Hager-Theodorides, A.L.; Zoidis, E.; Saridaki, A.; Antonakos, G.; Tsiamis, G. Combined GWAS and 'Guilt by Association'-Based Prioritization Analysis Identifies Functional Candidate Genes for Body Size in Sheep. Genet. Sel. Evol. 2017, 49, 41.
- 57. Braz, C.U.; Rowan, T.N.; Schnabel, R.D.; Decker, J.E. Genome-wide Association Analyses Identify Genotype-byenvironment Interactions of Growth Traits in Simmental Cattle. Sci. Rep. 2021, 11, 13335.
- 58. Lv, F.; Agha, S.; Kantanen, J.; Colli, L.; Stucki, S.; Kijas, J.W.; Li, M.; Marsan, P.A. Adaptations to Climate-Mediated Selective Pressures in Sheep. Mol. Biol. Evol. 2014, 31, 3324–3343.
- 59. Igoshin, A.; Yudin, N.; Aitnazarov, R.; Yurchenko, A.A.; Larkin, D.M. Whole-Genome Resequencing Points to Candidate DNA Loci Affecting Body Temperature under Cold Stress in Siberian Cattle Populations. Life 2021, 11, 959.
- 60. Li, R.; Li, H.; Chen, R.; Chong, Q.; Xiao, H.; Chen, S. Genome-Wide Scan of Selection Signatures in Dehong Humped Cattle for Heat Tolerance and Disease Resistance. Anim. Genet. 2020, 51, 292–299.
- 61. Edea, Z.; Dadi, H.; Dessie, T.; Kim, K.S. Genomic Signatures of High-Altitude Adaptation in Ethiopian Sheep Popula-Tions. Genes Genom. 2019, 41, 973–981.
- 62. Freitas, P.H.F.; Wang, Y.; Yan, P.; Oliveira, H.R.; Schenkel, F.S.; Zhang, Y.; Brito, L.F. Genetic Diversity and Signatures of Selection for Thermal Stress in Cattle and Other Two Bos Species Adapted to Divergent Climatic Conditions. Front. Genet. 2021, 12, 102.
- Buggiotti, L.; Yurchenko, A.A.; Yudin, N.S.; Vander Jagt, C.J.; Nadezhda, V.; Kusliy, M.; Vasiliev, S.K.; Rodionov, A.N.; Boronetskaya, O.I.; Zinovieva, A.; et al. Demographic History, Adaptation, and NRAP Convergent Evolution at Amino Acid Residue 100 in the World Northernmost Cattle from Siberia. Mol. Biol. Evol. 2021, 38, 3093–3110.
- Wang, X.; Ju, Z.; Jiang, Q.; Zhong, J.; Liu, C.; Wang, J.; Hoff, J.L.; Schnabel, R.D.; Zhao, H.; Gao, Y.; et al. Introgression, Admixture and Selection Facilitate Genetic Adaptation to High-Altitude Environments in Cattle. Genomics 2021, 113, 1491–1503.
- 65. Molotsi, A.; Cloete, S.; Taylor, J.F.; Whitacre, L. Identification of Selection Signatures in South African Sheep Popula-Tions Using HAPFLK and Bayesian Fst Approaches. In Proceedings of the World Congress on Genetics Applied to

Retrieved from https://encyclopedia.pub/entry/history/show/35935