

8-HEPE against Metabolic Syndrome, Dyslipidemia, NAFLD, and Atherosclerosis

Subjects: **Food Science & Technology**

Contributor: Masamichi Hirose

Euphausia pacifica (*E. pacifica*), also called North Pacific krill, is a small, red crustacean similar to shrimp that flourishes in the North Pacific Ocean. *E. pacifica* oil contains 8-hydroxyeicosapentaenoic acid (8-HEPE) at a level more than 10 times higher than *Euphausia superba* oil. 8-HEPE can activate the transcription of peroxisome proliferator-activated receptor alpha (PPAR α), PPAR γ , and PPAR δ to levels 10, 5, and 3 times greater than eicosapentaenoic acid, respectively. 8-HEPE has beneficial effects against metabolic syndrome (reduction in body weight gain, visceral fat area, amount of gonadal white adipose tissue, and gonadal adipocyte cell size), dyslipidemia (reduction in serum triacylglycerol and low-density lipoprotein cholesterol and induction of serum high-density lipoprotein cholesterol), atherosclerosis, and nonalcoholic fatty liver disease (reduction in triglyceride accumulation and hepatic steatosis in the liver) in mice.

Euphausia pacifica

8-HEPE

N-3 polyunsaturated fatty acids

nonalcoholic fatty liver disease

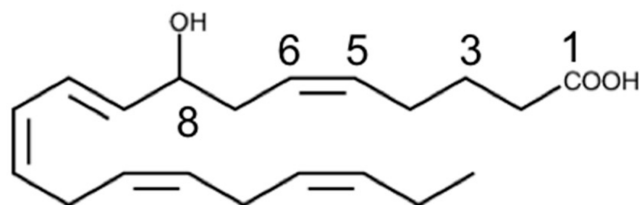
1. Introduction

Euphausia pacifica (*E. pacifica*) (**Figure 1A**), also called North Pacific krill, is a small, red crustacean similar to shrimp that flourishes in the North Pacific Ocean and is eaten in Japan. *E. pacifica* has large amounts of 8-hydroxyeicosapentaenoic acid (8-HEPE), which has physiological effects. This review discusses the chemical features of *E. pacifica* and the potential benefits of 8-hydroxyeicosapentaenoic acid (8-HEPE) extracted from *E. pacifica* against metabolic syndrome, dyslipidemia, NAFLD, and atherosclerosis in mice under the condition of a high-fat diet (HFD) or Western diet (WD).

A *Euphausia pacifica* (*E. pacifica*)



B



8-hydroxy eicosapentaenoic acid (8-HEPE)

Figure 1. (A) Photograph of *Euphausia pacifica* (*E. pacifica*), also called North Pacific krill. (B) The structural formula of 8-hydroxyeicosapentaenoic acid (8-HEPE).

2. Chemical Features of *E. pacifica*

2.1. Lipids of *E. pacifica*

At present, krill oil is generally *E. superba* (Antarctic krill) oil. There are several differences between *E. pacifica* oil and *E. superba* oil. The proportion of phospholipids in *E. pacifica* oil is higher than *E. superba* oil, and the content of oil is higher in *E. superba* than in *E. pacifica*. However, the most conspicuous difference between *E. pacifica* oil and *E. superba* oil is the content of 8-HEPE (**Figure 1B**), which is higher in *E. pacifica* oil ^[1].

2.2. 8-HEPE Extracted from *E. pacifica* and PPAR Activation

Methanol extract from *E. pacifica* activates the transcription of peroxisome proliferator-activated receptor alpha (PPAR α), PPAR γ , and PPAR δ ^[2]. In addition, 5-HEPE, 8-HEPE, 9-HEPE, 12-HEPE, and 18-HEPE (hydroxylation products of EPA) obtained from methanol extracts of *E. pacifica* act as PPAR ligands. Two of these products, 8-HEPE and 9-HEPE, enhance the transcription levels of PPARs to a significantly greater extent than 5-HEPE, 12-HEPE, 18-HEPE, EPA, or EPA ethyl ester in NIH-3T3 cells ^[2]. In fact, 8-HEPE activates the transcription of PPAR α , PPAR γ , and PPAR δ to levels 10, 5, and 3 times greater than EPA, respectively. 8-HEPE also increases the expression levels of genes regulated by PPARs, such as liver fatty-acid-binding protein, enoyl-CoA hydratase/3-hydroxyacyl CoA dehydrogenase, and carnitine palmitoyltransferase, in FaO cells. In contrast to 8-HEPE, EPA at the same concentration has weak or little effect on these gene expression levels and functions, indicating that 8-HEPE is the more potent inducer of physiological effects.

3. Potential Benefits of 8-HEPE Extracted from *E. pacifica* against NAFLD and Its Associated Diseases

3.1. Effects of 8-HEPE Extracted from *E. pacifica* on Metabolic Syndrome

Metabolic syndrome is the medical term for a combination of diabetes, hypertension, and obesity and causes dyslipidemia and fatty liver. Moreover, it is associated with greater risk of developing blood vessel diseases, such as coronary heart disease and stroke. The activation of hepatic PPAR α could ameliorate body weight gain and improve insulin sensitivity in HFD-fed obese mice ^[3]. Moreover, adipocyte hypertrophy and the functional disorder of adipose tissue, such as reduced adiponectin secretion, have been reported to be associated with obesity ^[4]. 8-HEPE (9.5 mg/kg) extracted from *E. pacifica* reduced the amount of visceral fat (**Figure 2A**) ^[5], gonadal white adipose tissue, and the size of gonadal adipocyte cells in HFD-fed mice ^[6]. PPAR α activators can increase hepatic fatty acid oxidation and decrease serum TAG levels, which are responsible for adipose cell hypertrophy and hyperplasia, leading to the regulation of obesity. Compared to EPA, 8-HEPE, which is a potent activator of PPAR α , might improve metabolic syndrome.

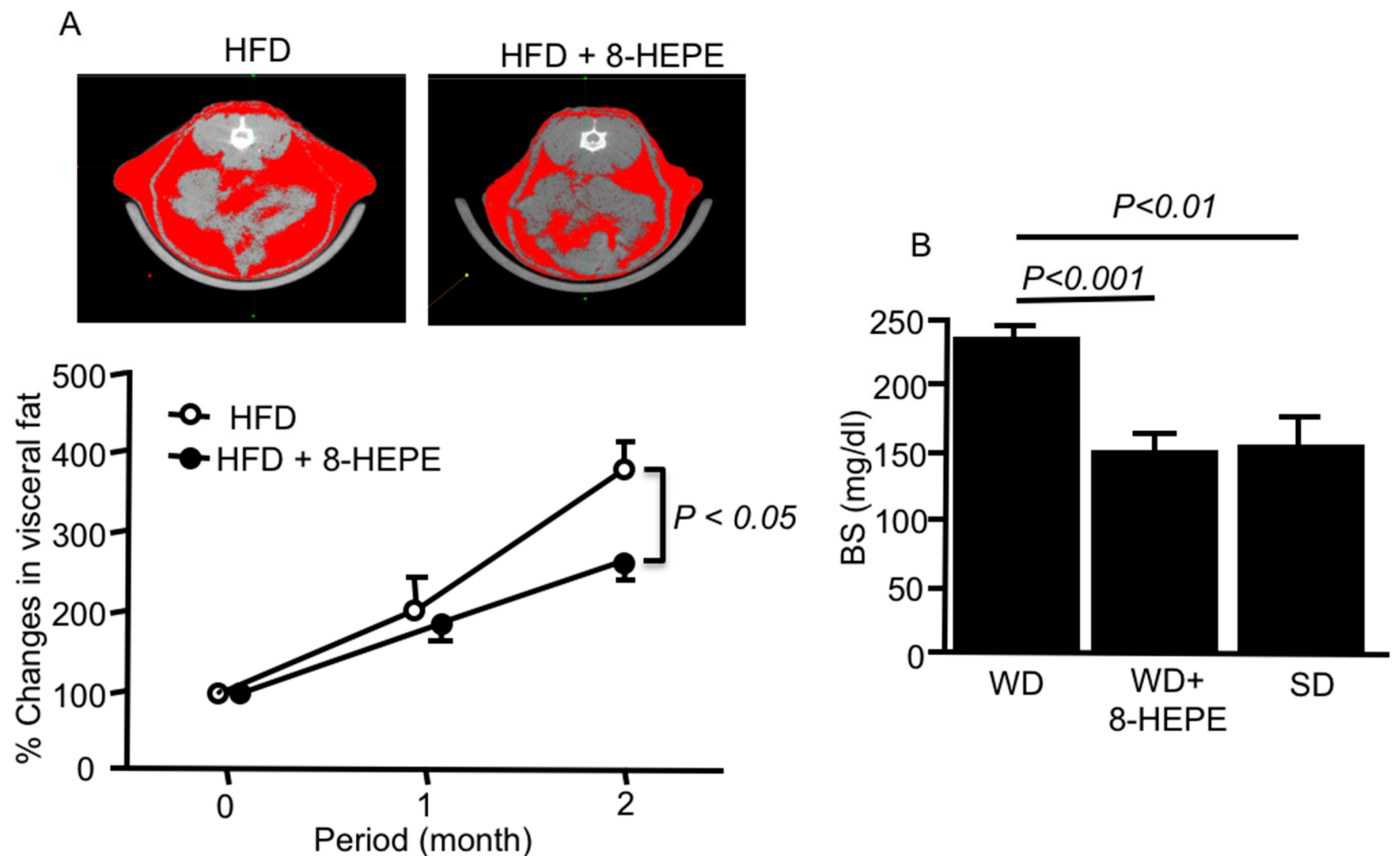


Figure 2. (A) Representative computed tomography images of the abdomen (upper) and percentage changes in visceral fat area (lower) in mice fed an HFD only or an HFD with 8-HEPE. Visceral and subcutaneous fat areas are shown in red. Data are the mean \pm SE obtained from eight mice for each group. (B) Blood sugar (BS) in apoE knock-out (apoE-KO) mice fed a WD only or WD with 8-HEPE. Data are the mean \pm SE obtained from 10 mice for each group. An analysis of variance (ANOVA) with Dunnett's test was used for the statistical analysis. HFD, high fat diet; 8-HEPE.

3.2. Effects of 8-HEPE Extracted from *E. pacifica* on Dyslipidemia

Dyslipidemia is defined by abnormal levels of plasma lipoproteins and TAG, and several types are known. Furthermore, dyslipidemia causes several complications, such as NAFLD and atherosclerosis, and its improvement is important for preventing these complications. Fibrates are an important group of drugs used to treat dyslipidemia in clinical practice. They are agonists of PPAR α , which plays important roles in the normalization of plasma lipoproteins and TAG levels. GW590735, a PPAR α agonist, increased HDL cholesterol and decreased LDL cholesterol, very low-density lipoprotein (VLDL) cholesterol, and TAG in hApoB100/hCETP mice [7]. Plasma TAG levels were significantly decreased in mice fed an HFD with 10 mg/kg 8-HEPE compared with HFD with EPA or HFD alone [6]. In contrast, plasma total cholesterol levels were similar between mice fed an HFD with 10 mg/kg 8-HEPE and HFD only [6]. Interestingly, 8-HEPE (83 mg/kg) suppressed the WD-induced increases in plasma LDL cholesterol in LDL cholesterol receptor knock-out (LDLR-KO) mice [8]. Moreover, it also increased plasma HDL cholesterol levels in WD-fed LDLR-KO mice. These results suggest that a low dose of 8-HEPE is enough to suppress plasma TAG, but a high dose of 8-HEPE might be needed to decrease plasma LDL cholesterol. Although

n-3 PUFAs, especially EPA and DHA, play a positive role in the treatment of dyslipidemia [9], these beneficial effects are considered to be mainly due to the ability of n-3 PUFAs to reduce plasma TAG levels [10]. ATP-binding cassette transporter A1 (ABCA1) transports phospholipids and free cholesterol from macrophages to lipid-free apoA-I [11][12], leading to the generation of HDL particles [13][14]. 8-HEPE, but not EPA, increased the gene expression of ABCA1 in murine OxLDL-treated J774.1 macrophages [8].

3.3. Effects of 8-HEPE Extracted from *E. pacifica* on NAFLD

Excessive calorie intake and lack of exercise have contributed to increased obesity and the prevalence of NAFLD in recent years. NAFLD is now the most important cause of chronic liver disease in the absence of excess alcohol consumption. NAFLD includes hepatic steatosis, which may progress to nonalcoholic steatohepatitis (NASH), fibrosis, and cirrhosis. Hepatic steatosis is characterized by the accumulation of TAG lipid droplets in the hepatocyte cytoplasm. Nonesterified fatty acids derived from the plasma are transported into the liver via transport proteins, such as CD36, which causes hepatic TAG accumulation. CD36 is expressed in a variety of cells, such as macrophages, and tissues [15]. Hajri et al. [16] showed that the deletion of nonhepatic CD36 gene expression causes hepatic steatosis and reduces muscle TAG contents in mice. Oil Red O histological staining, a marker of fat accumulation in the liver, was reduced in the liver of LDL-KO mice fed a WD with 83 mg/kg 8-HEPE compared with WD alone [8]. The content of TAG in the liver was also decreased in the WD with 8-HEPE than WD alone in these mice, suggesting that 8-HEPE can improve hepatic steatosis. We showed that 8-HEPE (50 μ M) significantly increased CD36 gene expression in OxLDL-treated murine J774.1 macrophages [8]. This effect may improve hepatic steatosis through the relative increase in fatty acid uptake from plasma into the macrophages compared to the liver.

3.4. Effects of 8-HEPE Extracted from *E. Pacifica* on Atherosclerosis

Several studies have highlighted the association between NAFLD and increased carotid and coronary atherosclerosis [17][18][19]. Palolini et al. [20] demonstrated that Antarctic krill oil inhibits aortic atherosclerosis in WD-fed apoE-KO mice. We elucidated the effects of 8-HEPE extracted from North Pacific krill oil on aortic atherosclerosis using apoE-KO mice. Sudan IV staining demonstrated that 8-HEPE (47 mg/kg) reduced the area of aortic atherosclerosis in WD-fed apoE-KO mice (**Figure 3**) [21], suggesting that 8-HEPE works as an inhibitor of atherosclerosis. CD36 macrophages participate in atherosclerotic arterial lesion formation by interacting with oxLDL, and CD36 deficiency reduces atherosclerotic lesion formation [22]. Moreover, plasma OxLDL levels were increased in apoE-KO mice [23]. Therefore, 8-HEPE seems to aggravate atherosclerosis by increasing CD36 gene expression in macrophages. However, Moore et al. [24] showed that the loss of CD36 in apoE-KO mice did not alleviate atherosclerotic lesions. Moreover, Zhu et al. [25] showed that the scavenger receptor activity of CD16, which is different from that of CD36, also contributed to the progression of atherosclerosis in apoE-KO mice. Therefore, increased CD36 gene expression in macrophages may not always aggravate atherosclerosis.

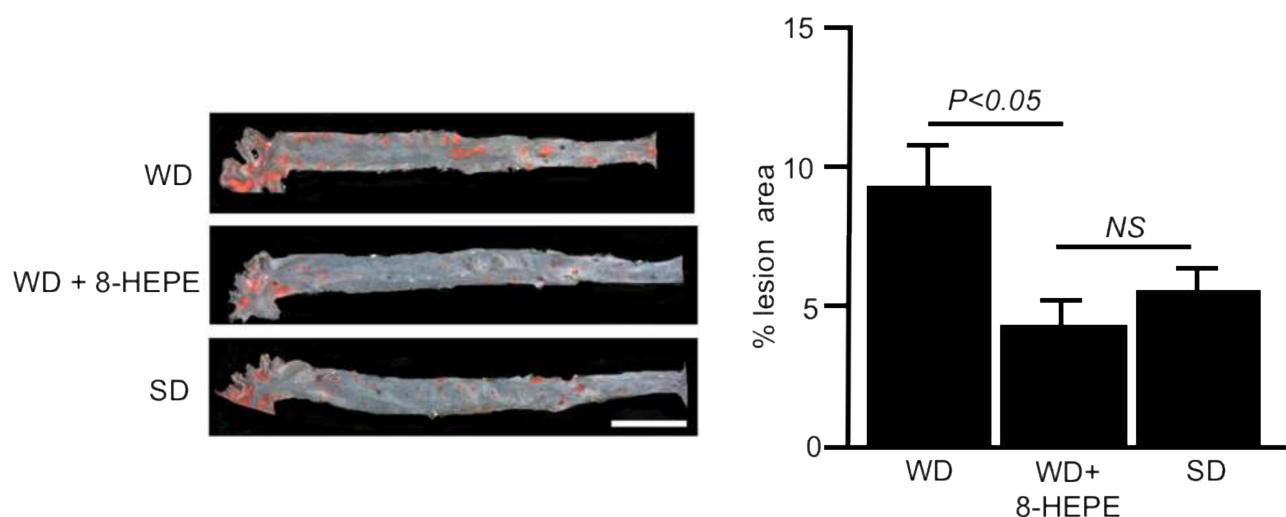


Figure 3. Representative images of the aorta (**left**) stained with Sudan IV and percentage lesion area of atherosclerosis (**right**) in apoE-KO mice fed a Western diet (WD) only or a WD with 8-HEPE (WD + 8-HEPE). Data are the mean \pm SE obtained from eight mice for each group. An analysis of variance (ANOVA) with Dunnett's test was used for the statistical analysis. SD, standard diet; NS, not significant.

References

1. Yamada, H.; Yamazaki, Y.; Koike, S.; Hakozaiki, M.; Nagahora, N.; Yuki, S.; Yano, A.; Tsurumi, K.; Okumura, T. Lipids, fatty acids and hydroxy-fatty acids of *Euphausia pacifica*. *Sci. Rep.* 2017, 7, 9944.
2. Yamada, H.; Oshiro, E.; Kikuchi, S.; Hakozaiki, M.; Takahashi, H.; Kimura, K.I. Hydroxyeicosapentaenoic acids from the Pacific krill show high ligand activities for PPARs. *J. Lipid Res.* 2014, 55, 895–904.
3. Lee, D.; Shin, Y.; Roh, J.S.; Ahn, J.; Jeong, S.; Shin, S.S.; Yoon, M. Lemon Balm Extract ALS-L1023 regulates obesity and improves insulin sensitivity via activation of hepatic PPAR α in high-fat diet-fed obese C57BL/6J mice. *Int. J. Mol. Sci.* 2020, 21, 4256.
4. Yu, Y.H.; Ginsberg, H.N. Adipocyte signaling and lipid homeostasis: Sequelae of insulin-resistant adipose tissue. *Circ. Res.* 2005, 96, 1042–1052.
5. Hirose, M.; Yamada, H.; Kamata, Y.; Yoshida, K.; Sasaki, H.; Tanji, M.; Ibi, M. Preventive Effect of New Functional Material Derived from *E. pacifica* on Visceral Fat. Society of Cerebral-Cardio-Vascular Anti-Aging 2015 Abstract. Available online: <http://www.plus-s-ac.com/ccvaa/english.html> (accessed on 28 November 2015).
6. Yamada, H.; Kikuchi, S.; Hakozaiki, M.; Motodate, K.; Nagahora, N.; Hirose, M. 8-Hydroxyeicosapentaenoic acid decreases plasma and hepatic triglycerides via activation of

- peroxisome proliferator-activated receptor alpha in high-fat diet-induced obese mice. *J. Lipids*. 2016, 7498508.
7. Hansen, M.K.; McVey, M.J.; White, R.F.; Legos, J.J.; Brusq, J.M.; Grillot, D.A.; Issandou, M.; Barone, F.C. Selective CETP inhibition and PPARalpha agonism increase HDL cholesterol and reduce LDL cholesterol in human ApoB100/human CETP transgenic mice. *J. Cardiovasc. Pharmacol. Ther.* 2010, 15, 196–202.
 8. Saito, M.; Ishida, N.; Yamada, H.; Ibi, M.; Hirose, M. 8-HEPE- concentrated materials from Pacific krill improve plasma cholesterol levels and hepatic steatosis in high cholesterol diet-fed low-density lipoprotein (LDL) receptor-deficient mice. *Biol. Pharm. Bull.* 2020, 43, 919–924.
 9. Wen, Y.; Dai, J.H.; Gao, Q. Effects of Omega-3 fatty acid on major cardiovascular events and mortality in patients with coronary heart disease: A meta-analysis of randomized controlled trials. *Nutr. Metab. Cardiovasc. Dis.* 2014, 24, 470–475.
 10. Lopez-Huertas, E. The effect of EPA and DHA on metabolic syndrome patients: A systematic review of randomised controlled trials. *Br. J. Nutr.* 2012, 107, 185–194.
 11. Lee, J.; Park, Y.; Koo, S. ATP-binding cassette transporter A1 and HDL metabolism: Effects of fatty acids. *J. Nutr. Biochem.* 2012, 23, 1–7.
 12. Landry, Y.; Denis, M.; Nandi, S.; Bell, S.; Vaughan, A.; Zha, X. ATP-binding cassette transporter A1 expression disrupts raft membrane microdomains through its ATPase-related functions. *J. Biol. Chem.* 2006, 281, 36091–36101.
 13. Phillips, M. Molecular mechanisms of cellular cholesterol efflux. *J. Biol. Chem.* 2014, 289, 24020–24029.
 14. Vedhachalam, C.; Duong, P.; Nickel, M.; Nguyen, D.; Dhanasekaran, P.; Saito, H.; Rothblat, G.H.; Lund-Katz, S.; Phillips, M.C. Mechanism of ATP-binding cassette transporter A1-mediated cellular lipid efflux to apolipoprotein A-I and formation of high-density lipoprotein particles. *J. Biol. Chem.* 2007, 282, 25123–25130.
 15. Wilson, C.G.; Tran, J.L.; Erion, D.M.; Vera, N.B.; Febbraio, M.; Weiss, E.J. Hepatocyte-specific disruption of CD36 attenuates fatty liver and improves insulin sensitivity in HFD-fed mice. *Endocrinology* 2016, 157, 570–585.
 16. Hajri, T.; Han, X.X.; Bonen, A.; Abumrad, N.A. Defective fatty acid uptake modulates insulin responsiveness and metabolic responses to diet in CD36-null mice. *J. Clin. Invest.* 2002, 109, 1381–1389.
 17. Bhatia, L.S.; Curzen, N.P.; Calder, P.C.; Byrne, C.D. Non-alcoholic fatty liver disease: A new and important cardiovascular risk factor? *Eur. Heart J.* 2012, 33, 1190–1200.

18. Gastaldelli, A.; Kozakova, M.; Hojlund, K.; Flyvbjerg, A.; Favuzzi, A.; Mitrakou, A.; Balkau, B. Fatty liver is associated with insulin resistance, risk of coronary heart disease, and early atherosclerosis in a large European population. *Hepatology* 2009, 49, 1537–1544.
19. Sung, K.C.; Wild, S.H.; Kwag, H.J.; Byrne, C.D. Fatty liver, insulin resistance, and features of metabolic syndrome: Relationships with coronary artery calcium in 10,153 people. *Diabetes Care*. 2012, 35, 2359–2364.
20. Parolini, C.; Bjorndal, B.; Busnelli, M.; Manzini, S.; Ganzetti, G.S.; Dellera, F.; Ramsvik, M.; Bruheim, I.; Berge, R.K.; Chiesa, G. Effect of Dietary Components from Antarctic Krill on Atherosclerosis in apoE-Deficient Mice. *Mol. Nutr. Food Res.* 2017, 61, 1700098.
21. Ibi, M.; Saito, M.; Ishida, N.; Yamada, Y.; Hirose, M. 8-HEPE concentrated materials from pacific krill suppresses atherosclerosis in apoE-deficient mice. *Circulation* 2019, 140 (Suppl. 1), A11153.
22. Park, Y.M. CD36, a scavenger receptor implicated in atherosclerosis. *Exp. Mol. Med.* 2014, 46, e99.
23. Kato, R.; Mori, C.; Kitazato, K.; Arata, S.; Obama, T.; Mori, M.; Takahashi, K.; Aiuchi, T.; Takano, T.; Itabe, H. Transient increase in plasma oxidized LDL during the progression of atherosclerosis in apolipoprotein E knockout mice. *Arterioscler. Thromb. Vasc. Biol.* 2009, 29, 33–39.
24. Moore, K.J.; Kunjathoor, V.V.; Koehn, S.L.; Manning, J.J.; Tseng, A.A.; Silver, J.M.; McKee, M.; Freeman, M.W. Loss of receptor-mediated lipid uptake via scavenger receptor A or CD36 pathways does not ameliorate atherosclerosis in hyperlipidemic mice. *J. Clin. Invest.* 2005, 115, 2192–2201.
25. Zhu, X.; Ng, H.P.; Lai, Y.C.; Craigo, J.K.; Nagilla, P.S.; Raghani, P.; Nagarajan, S. Scavenger receptor function of mouse Fcy receptor III contributes to progression of atherosclerosis in apolipoprotein E hyperlipidemic mice. *J. Immunol.* 2014, 193, 2483–2495.

Retrieved from <https://encyclopedia.pub/entry/history/show/38054>