

CD44

Subjects: Anatomy & Morphology

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CD44, a non-kinase cell surface transmembrane glycoprotein, has been widely implicated as a cancer stem cell (CSC) marker in several cancers. Cells overexpressing CD44 possess several CSC traits, such as self-renewal and epithelial-mesenchymal transition (EMT) capability, as well as a resistance to chemo- and radiotherapy.

Keywords: CD44 ; regulation ; tumourigenesis

1. CD44 Structure and Isoforms

The full-length CD44 gene comprises 20 exons, with the constant exons 1–5 and 16–20 encoding the N-terminal and C-terminal domains respectively, which are homologous domains shared by all CD44 family members ^[1]. The smallest and the most expressed CD44 isoform is the CD44 standard (CD44s), constructed of ten constant exons with no variant exons ^[2]. The other isoform, the CD44 variant (CD44v), differs from CD44s by the insertion or excision of alternatively spliced exons between the N-terminal and C-terminal domains ^[3]. Tolg et al. ^[4] confirmed that besides ten constant exons, the mouse and rat genome has at least ten variant exons, all of which can be combined alternatively into CD44 mRNA. They suggested that the variant exons be numbered by the exon code v1 to v10. Screaton et al. ^[5] described the structure of the human CD44 gene, reporting that it contains 19 exons crossing some 50 kilobases of DNA with ten constant exons and nine variant exons coded v2–v10 ^{[6][7]}. CD44v isoforms may contain a single variant exon such as CD44v3 or CD44v6, or a combination of variant exons such as CD44v3–v7 and CD44v8–v10. Individual cells can continually alter the splicing of CD44 pre-mRNA, resulting in many possible combinations of these variant exons, giving the potential for great diversity ^[3].

The CD44 protein has four primary characteristic regions: the extracellular region, the stem region (standard stem region and/or variable stem region), the transmembrane region (TM), and the short C-terminal intracellular/cytoplasmic (CP) region ^[8]. The extracellular part consists of seven extracellular domains (1–5, 6 and 7 of the constant exons) including N-terminal domains (ligand-binding region). The stem region (alternative splicing area) has an insertion of one or more of the variant exons between exon 5 and exon 6. The transmembrane region is encoded by a single exon (exon 8), whereas the cytoplasmic region is encoded by exon 10 or exon 9. However, exon 9 is spliced out in almost all CD44 cDNA isoforms ^[3] ^[9]. Several isoforms of the human CD44 molecule are associated with tumour progression and stemness in various cancers, such as breast cancer ^[10], gliomas ^{[11][12]}, head and neck squamous cell carcinoma ^[13], pancreatic cancer ^{[14][15]}, prostate cancer ^[16] and colorectal cancer ^{[17][18]} (**Figure 1** and **Table 1**). The complexity of the CD44 protein is further augmented by post-translational modifications including variance glycosylation with O-glycans, N-glycans and glycosaminoglycans, such as chondroitin sulphate and heparan sulphate ^[8]. Due to these side-chain attachments, the conserved format of CD44 (37 kDa) is enlarged to 80–100 kDa with some isoforms surpassing 200 kDa due to a high level of glycosylation ^[3]. An illustration of CD44 protein structure is shown in **Figure 2**.

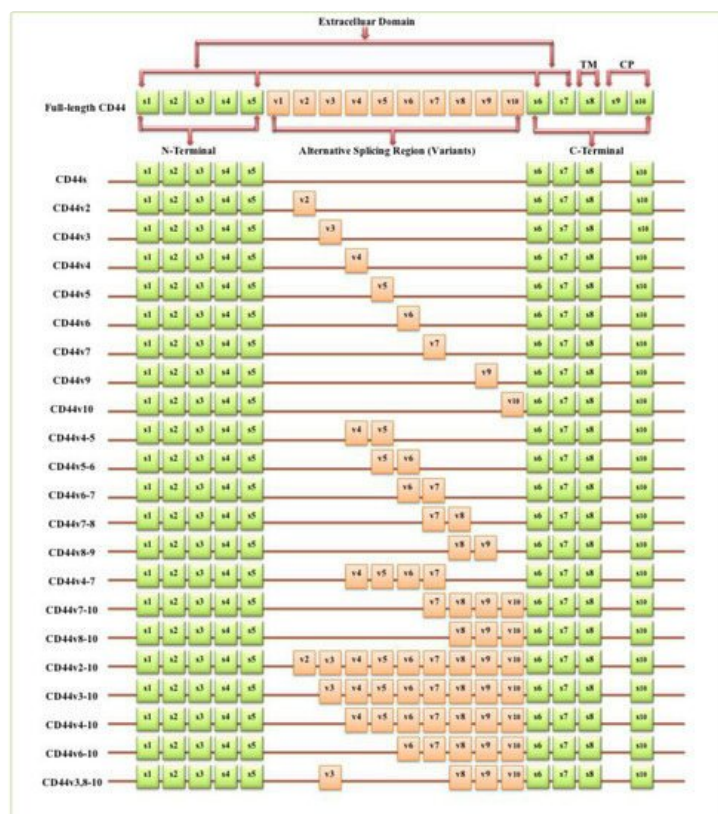


Figure 1. Schematic diagram of the mouse CD44 gene and most CD44 isoforms involved in cancer progression. The full-length CD44 gene contains 20 exons in mice and 19 exons in humans, with the constant exons 1–5 and 16–20 encoding the N-terminal and the C-terminal domains. CD44 standard (CD44s) is encoded by these ten constant exons and contains no variant exons, whereas the CD44 variant (CD44v) is produced by the alternative splicing of a variable insertion of nine extra exons in humans or ten extra exons in mice. These extra exons are exons 6–15, typically identified as (v1 to v10) in mice and the exons 7–15 identified as (v2 to v10) in humans and are located between the N-terminal and C-terminal domains. CD44v can contain one or multiple variant exons and exon 19 is spliced out in all CD44 isoforms. Abbreviations: CD44s, CD44 standard; CD44v, CD44 variant; s, standard; v, variant; TM, transmembrane; CP, cytoplasmic. Green boxes refer to the constant/standard exons. Orange boxes refer to the variant exons.

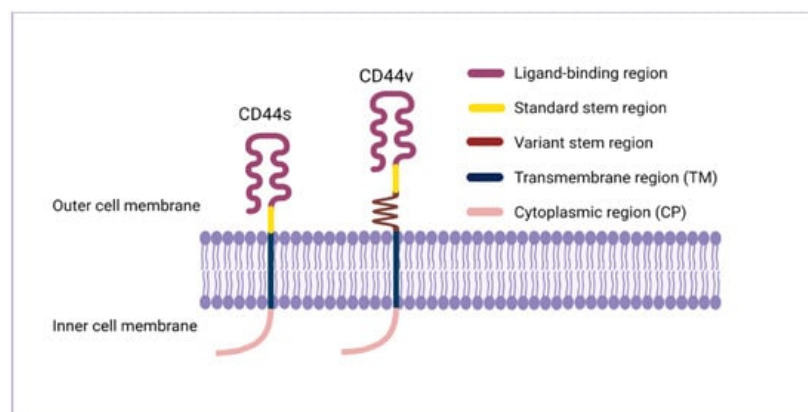


Figure 2. CD44 protein structure. The CD44 protein has four primary regions: the extracellular region consists of seven extracellular domains including N-terminal domains (ligand-binding region), the stem region (variable stem region and/or standard stem region) which is the alternative splicing area containing an insertion of one or more variant exons, the transmembrane region (TM), and the C-terminal cytoplasmic (CP) region.

Table 1. CD44 isoforms relevant to cancer progression. Abbreviations: CSCs, cancer stem cells; EMT, epithelial–mesenchymal transition; DFS, disease-free survival; OS, overall survival; TNM stage, tumour (T), node (N), and metastasis (M) stage; FIGO stage, the international federation of gynaecology and obstetrics stage; NHL, Non-Hodgkin's lymphoma; HPV, human papillomavirus; MAPK, mitogen-activated protein kinase.

CD44 Isoform	Association in Cancer Progress	Cancer Type	Ref
CD44, non-specified	Tumour cell aggregation, metastasis	Breast cancer	[10]

CD44 Isoform	Association in Cancer Progress	Cancer Type	Ref
CD44, non-specified	Adhesion, migration, invasion	Glioblastoma	[11] [12]
CD44, non-specified	Angiogenesis	Head and neck squamous carcinoma	[13]
CD44, non-specified	Invasion, metastasis, EMT, cancer progression, poor prognosis	Pancreatic cancer	[14] [15]
CD44, non-specified	Proliferation, migration, invasion	Prostate Cancer	[16]
CD44, non-specified	Metastasis, poor differentiation, invasion	Colorectal cancer	[17] [18]
CD44s	Tumour initiation, CSCs traits induction	Breast cancer	[19]
CD44s	Metastasis	Breast cancer	[20]
CD44s	EMT regulation, cancer progression	Breast cancer	[21]
CD44s	Poor DFS, poor OS, invasion, EMT	Hepatocellular carcinoma	[22]
CD44s	Invasion, metastasis, EMT, poor differentiation, chemotaxis	Gallbladder cancer	[23]
CD44s	Proliferation, invasion, migration, EMT, stemness	Prostate cancer	[24]
CD44s	EMT, invasion, metastasis, chemoresistance	Pancreatic ductal adenocarcinoma	[25]
CD44s	EMT, radio-resistance	Pancreatic cancer	[26]
CD44v2	Poor OS, advanced cancer stage	Colorectal cancer	[27]
CD44v2	Poor OS, invasion	Pancreatic cancer	[28]
CD44v3	Poor OS, invasion, metastasis	Oral squamous carcinoma	[29]
CD44v3	Stem cells self-renewal	Myeloid leukaemia	[30]
CD44v3	Metastasis	Colorectal adenocarcinoma	[31]
CD44v4	Proliferation, migration, radio-resistance	Head and neck squamous carcinoma	[32]
CD44v5	High histological grade, poor differentiation, poor OS	Hepatocellular carcinoma	[33]
CD44v6	Tumour budding, invasion, metastasis	Oral squamous carcinoma	[34]
CD44v6	Proliferation, invasion, adhesion, metastasis, EMT, chemo/radio-resistance	Prostate cancer	[35]
CD44v6	Local recurrence, invasion, metastasis	Tongue squamous carcinoma	[36]
CD44v6	Tumour budding, locoregional failure (metastasis, local recurrence)	Colorectal cancer	[37]
CD44v6	Proliferation, migration, radio-resistance	Head and neck squamous carcinoma	[32]
CD44v6	Metastasis	Colorectal adenocarcinoma	[31]
CD44v6	Poor OS, invasion	Pancreatic cancer	[28]
CD44v6	High histological grade, poor differentiation, poor OS	Hepatocellular carcinoma	[33]
CD44v6	Invasion, metastasis, poor OS, TNM stage	Pancreatic cancer	[38]
CD44v6	FIGO stage, poor prognosis	Cervical cancer	[39]
CD44v6	Metastasis, self-adhesion of aggressive NHL cells	Non-Hodgkin's lymphoma	[40]
CD44v6	Infiltration, metastasis	Oesophageal squamous carcinoma	[41]

CD44 Isoform	Association in Cancer Progress	Cancer Type	Ref
CD44v6	Proliferation, myofibroblastic differentiation	Gastric cancer	[42]
CD44v7	Proliferation, migration, radio-resistance	Head and neck squamous carcinoma	[32]
CD44v9	Increased tumourigenicity	Gallbladder cancer	[23]
CD44v9	Invasion, metastasis, poor OS, TNM stage	Pancreatic cancer	[38]
CD44v9	Proliferation, invasion, migration, EMT	Cholangiocarcinoma	[43]
CD44v9	Invasion, migration, worse prognosis	Bladder cancer	[44]
CD44v10	High histological grade, poor differentiation, poor OS	Hepatocellular carcinoma	[33]
CD44v10	Histological grade, clinical and pathological stage, poor survival	Renal carcinoma	[45]
CD44v10	Migration, metastasis, promote tumourigenesis	Breast cancer	[46] [47]
CD44v4-5	Infiltration, metastasis	Oesophageal squamous carcinoma	[41]
CD44v4-5	Poor differentiation	Non-small cell lung carcinoma	[48]
CD44v5-6	Proliferation, KRAS/MAPK signalling, promoting tumour development	Lung adenocarcinoma	[49]
CD44v6-7	Metastasis	Pancreatic adenocarcinoma	[2]
CD44v7-8	High histological grade, poor differentiation, poor OS	Hepatocellular carcinoma	[33]
CD44v7-8	FIGO stage, poor prognosis	Cervical cancer	[39]
CD44v7-8	Invasion, high-risk HPV infection	Uterine cervical squamous carcinoma	[50]
CD44v8-9	Proliferation, KRAS/MAPK signalling, promoting tumour development	Lung adenocarcinoma	[49]
CD44v4-7	Metastasis	Pancreatic adenocarcinoma	[2]
CD44v7-10	Invasion	Prostate cancer	[51]
CD44v8-10	Migration, metastasis, sphere formation	Breast cancer	[52]
CD44v8-10	Tumour initiation, CSCs traits induction	Gastric cancer	[53]
CD44v8-10	Metastasis	Lung cancer	[54]
CD44v8-10	Metastasis, relapse	Gastric cancer	[55]
CD44v8-10	Poor prognosis, chemo/radio-resistance	Oesophageal squamous carcinoma	[56]
CD44v8-10	Chemoresistance	Urothelial cancer	[57]
CD44v2-10	CSCs traits induction, tumour subtype, oncogenic signalling pathways	Breast cancer	[58]
CD44v3-10	CSCs traits induction, tumour subtype, oncogenic signalling pathways	Breast cancer	[58]
CD44v3-10	Metastasis, self-adhesion of aggressive NHL cells	Non-Hodgkin's lymphoma	[40]
CD44v4-10	Tumour initiation, wild-type phenotype	Intestinal cancer	[6]
CD44v6-10	Metastasis, self-adhesion of aggressive NHL cells	Non-Hodgkin's lymphoma	[40]
CD44v6-10	Metastasis, relapse	Gastric cancer	[55]
CD44v3, 8-10	Metastasis, relapse	Gastric cancer	[55]
CD44v3, 8-10	Metastasis, migration	Breast cancer	[59]

2. CD44 Expression in Normal Cells

CD44 is significantly expressed in lymphocytes, smooth muscle, fibroblasts and various types of epithelia and is involved in lymphocyte homing, cell adhesion and aggregation, cell migration, leukocyte activation, lymphopoiesis and myelopoiesis, angiogenesis and cytokine release [3][60]. CD44s was initially isolated from haematopoietic cells even though it is expressed in several other tissues including the liver, lung, pancreas, skin and central nervous system [3]. CD44s is expressed in adult tissues and embryo tissues from day 9.5 post coitum, whereas numerous isoforms of CD44v show a highly specialised expression pattern and are already in the egg cylinder at day 6.5 of development [61]. In contrast to CD44s, CD44v isoforms distribution is more restricted to a selected range of cells during specific stages of activation, maturation or development including macrophages, activated lymphocytes, keratinocytes and some epithelial cells such as in the stomach, bladder and uterine cervix [3] and many carcinomas. In normal tissues, CD44 isoforms play a role in hyaluronic acid (HA) metabolism regulation, whereby loss of CD44 expression disrupts HA metabolism and impairs hair regrowth, wound healing and keratinocyte proliferation [62].

3. CD44 Expression in Tumours

Numerous studies indicated that lymphoma, breast, colon and endometrial cancer have elevated levels of CD44 mRNA [60]. Increasing evidence also suggests that CD44 is extensively overexpressed in other cancer types including gallbladder, prostate, ovarian, oral squamous cell carcinoma and gastric cancer, correlating with aggressive biological behaviour and a poor prognosis [63]. The role of CD44 in tumours is not well defined, however, elevated levels of CD44 are associated with numerous malignant tumours. The physiological functions of CD44 indicate that it is involved in the metastasis of tumours [3]. For instance, lung adenocarcinoma cells show a high expression of CD44v, which correlates with enhanced CSCs characteristics, proliferation and resistance to chemotherapeutics [64], whereas these variants, especially CD44v6, are closely related to metastasis of pancreatic carcinoma cells [60]. Many studies have investigated CD44 expression levels in several cancers in comparison to their adjacent normal tissues and explored the relationship with tumour progression and clinicopathological outcomes by mining various publicly available databases, including The Cancer Genome Atlas (TCGA), Tumour Immune Estimation Resource (TIMER) database, Oncomine database, Gene Expression Profiling Interactive Analysis (GEPIA), In silico Transcriptomics (IST) database, R2 online database, SAGE Genie tools, and Human Gene Expression Map (HGEM) (Table 2 and Figure 3).



Figure 3. CD44 distribution in normal versus cancerous tissues and its correlation with clinical outcomes.

Table 2. Low and high CD44 expression in normal and tumour tissues respectively and association with clinical outcomes.

Cancer Type	Correlation with Clinical Outcomes	Public Database	Reference
Gallbladder cancer, hepatocellular carcinoma, cholangiocarcinoma	Poor prognosis, advanced TNM stage, poor OS, aggressive tumour behaviour (proliferation, migration, invasion, clonogenicity)	TCGA database	[63]
Colon cancer, gastric cancer, brain cancer, stomach cancer, pancreatic cancer, liver cancer	Benign OS rate in gastric cancer, poor OS in colon cancer, TNM staging, differentiation degree, and poor survival in pancreatic cancer	SAGE Genie and Oncomine database	[65][66]
Head and neck squamous carcinoma	Poor OS, poor differentiation, angiogenesis, immune regulation, invasion	TCGA database	[67]
Head and neck squamous carcinoma	Pro-angiogenetic phenotype	TCGA database	[13]
Prostate cancer	Advanced T stage, higher Gleason score, poor differentiation	TCGA database	[68]
Colon adenocarcinoma	Therapy resistance	TCGA database and GEPIA	[69]
Head and neck squamous carcinoma, acute myeloid leukaemia (AML), lung carcinoma	Not specified	IST database and HGEM database	[70]
Glioblastoma	Poor OS, hypoxia-induced gene signature	TCGA database	[71]
Glioblastoma	Poor OS, therapy resistance	R2 online database	[72]
Invasive ductal breast carcinoma	Invasion, metastasis	TCGA database	[73]
Brain and CNS cancer, colorectal cancer, melanoma, sarcoma, gastric cancer, head and neck carcinoma, kidney cancer, oesophageal cancer, cholangiocarcinoma, pancreatic cancer	EMT, drug resistance, metastasis, immune infiltration and suppression features, poor survival, higher mutation burden, afflict older patients	Oncomine database and TIMER database	[74]

References

- Loh, T.J.; Moon, H.; Cho, S.; Jang, H.; Liu, Y.C.; Tai, H.; Jung, D.W.; Williams, D.R.; Kim, H.R.; Shin, M.G.; et al. CD44 alternative splicing and hnRNP A1 expression are associated with the metastasis of breast cancer. *Oncol. Rep.* 2015, 34, 1231–1238.
- Rall, C.J.; Rustgi, A.K. CD44 isoform expression in primary and metastatic pancreatic adenocarcinoma. *Cancer Res.* 1995, 55, 1831–1835.
- Mishra, M.N.; Chandavarkar, V.; Sharma, R.; Bhargava, D. Structure, function and role of CD44 in neoplasia. *J. Oral Maxillofac. Pathol.* 2019, 23, 267–272.
- Tölg, C.; Hofmann, M.; Herrlich, P.; Ponta, H. Splicing choice from ten variant exons establishes CD44 variability. *Nucleic Acids Res.* 1993, 21, 1225–1229.
- Screaton, G.R.; Bell, M.V.; Jackson, D.G.; Cornelis, F.B.; Gerth, U.; Bell, J.I. Genomic structure of DNA encoding the lymphocyte homing receptor CD44 reveals at least 12 alternatively spliced exons. *Proc. Natl. Acad. Sci. USA* 1992, 89, 12160–12164.
- Zeilstra, J.; Joosten, S.P.; van Andel, H.; Tölg, C.; Berns, A.; Snoek, M.; Van De Wetering, M.; Spaargaren, M.; Clevers, H.; Pals, S.T. Stem cell CD44v isoforms promote intestinal cancer formation in Apc(min) mice downstream of Wnt signaling. *Oncogene* 2014, 33, 665–670.
- Goodison, S.; Urquidí, V.; Tarín, D. CD44 cell adhesion molecules. *Mol. Pathol.* 1999, 52, 189–196.
- Chen, K.L.; Li, D.; Lu, T.X.; Chang, S.W. Structural Characterization of the CD44 Stem Region for Standard and Cancer-Associated Isoforms. *Int. J. Mol. Sci.* 2020, 21, 336.
- Misra, S.; Hascall, V.C.; Markwald, R.R.; Ghatak, S. Interactions between Hyaluronan and Its Receptors (CD44, RHAMM) Regulate the Activities of Inflammation and Cancer. *Front. Immunol.* 2015, 6, 201.
- Liu, X.; Taftaf, R.; Kawaguchi, M.; Chang, Y.F.; Chen, W.; Entenberg, D.; Zhang, Y.; Gerratana, L.; Huang, S.; Patel, D. B.; et al. Homophilic CD44 Interactions Mediate Tumor Cell Aggregation and Polyclonal Metastasis in Patient-Derived Breast Cancer Models. *Cancer Discov.* 2019, 9, 96–113.

11. Wolf, K.J.; Shukla, P.; Springer, K.; Lee, S.; Coombes, J.D.; Choy, C.J.; Kenny, S.J.; Xu, K.; Kumar, S. A mode of cell adhesion and migration facilitated by CD44-dependent microtentacles. *Proc. Natl. Acad. Sci. USA* 2020, 117, 11432–11443.
12. Hassn Mesrati, M.; Behrooz, A.B.; Abuhamad, A.Y.; Syahir, A. Understanding Glioblastoma Biomarkers: Knocking a Mountain with a Hammer. *Cells* 2020, 9, 1236.
13. Ludwig, N.; Szczepanski, M.J.; Gluszko, A.; Szafarowski, T.; Azambuja, J.H.; Dolg, L.; Gellrich, N.-C.; Kampmann, A.; Whiteside, T.L.; Zimmerer, R.M. CD44(+) tumor cells promote early angiogenesis in head and neck squamous cell carcinoma. *Cancer Lett.* 2019, 467, 85–95.
14. Gzil, A.; Zarębska, I.; Bursiewicz, W.; Antosik, P.; Grzanka, D.; Szyłberg, Ł. Markers of pancreatic cancer stem cells and their clinical and therapeutic implications. *Mol. Biol. Rep.* 2019, 46, 6629–6645.
15. Durko, L.; Włodarski, W.; Stasikowska-Kanicka, O.; Wagrowska-Danilewicz, M.; Danilewicz, M.; Hogendorf, P.; Strzelczyk, J.; Malecka-Panas, E. Expression and Clinical Significance of Cancer Stem Cell Markers CD24, CD44, and CD133 in Pancreatic Ductal Adenocarcinoma and Chronic Pancreatitis. *Dis. Markers* 2017, 2017, 3276806.
16. Li, W.; Qian, L.; Lin, J.; Huang, G.; Hao, N.; Wei, X.; Wang, W.; Liang, J. CD44 regulates prostate cancer proliferation, invasion and migration via PDK1 and PFKFB4. *Oncotarget* 2017, 8, 65143–65151.
17. Wang, Z.; Tang, Y.; Xie, L.; Huang, A.; Xue, C.; Gu, Z.; Wang, K.; Zong, S. The Prognostic and Clinical Value of CD44 in Colorectal Cancer: A Meta-Analysis. *Front. Oncol.* 2019, 9, 309.
18. Holah, N.S.; Aiad, H.A.; Asaad, N.Y.; Elkhoully, E.A.; Lasheen, A.G. Evaluation of the role of CD44 as a cancer stem cell marker in colorectal carcinoma: Immunohistochemical study. *Menoufia Med. J.* 2017, 30, 174–183.
19. Zhang, H.; Brown, R.L.; Wei, Y.; Zhao, P.; Liu, S.; Liu, X.; Deng, Y.; Hu, X.; Zhang, J.; Gao, X.D.; et al. CD44 splice isoform switching determines breast cancer stem cell state. *Genes Dev.* 2019, 33, 166–179.
20. Zhao, P.; Xu, Y.; Wei, Y.; Qiu, Q.; Chew, T.L.; Kang, Y.; Cheng, C. The CD44s splice isoform is a central mediator for invadopodia activity. *J. Cell Sci.* 2016, 129, 1355–1365.
21. Brown, R.L.; Reinke, L.M.; Damerow, M.S.; Perez, D.; Chodosh, L.A.; Yang, J.; Cheng, C. CD44 splice isoform switching in human and mouse epithelium is essential for epithelial-mesenchymal transition and breast cancer progression. *J. Clin. Investig.* 2011, 121, 1064–1074.
22. Mima, K.; Okabe, H.; Ishimoto, T.; Hayashi, H.; Nakagawa, S.; Kuroki, H.; Watanabe, M.; Beppu, T.; Tamada, M.; Nagano, O.; et al. CD44s regulates the TGF- β -mediated mesenchymal phenotype and is associated with poor prognosis in patients with hepatocellular carcinoma. *Cancer Res.* 2012, 72, 3414–3423.
23. Miwa, T.; Nagata, T.; Kojima, H.; Sekine, S.; Okumura, T. Isoform switch of CD44 induces different chemotactic and tumorigenic ability in gallbladder cancer. *Int. J. Oncol.* 2017, 51, 771–780.
24. Chen, Q.; Gu, M.; Cai, Z.k.; Zhao, H.; Sun, S.C.; Liu, C.; Zhan, M.; Chen, Y.B.; Wang, Z. TGF- β 1 promotes epithelial-to-mesenchymal transition and stemness of prostate cancer cells by inducing PCBP1 degradation and alternative splicing of CD44. *Cell. Mol. Life Sci.* 2021, 78, 949–962.
25. Zhao, S.; Chen, C.; Chang, K.; Karnad, A.; Jagirdar, J.; Kumar, A.P.; Freeman, J.W. CD44 Expression Level and Isoform Contributes to Pancreatic Cancer Cell Plasticity, Invasiveness, and Response to Therapy. *Clin. Cancer Res.* 2016, 22, 5592–5604.
26. Tsubouchi, K.; Minami, K.; Hayashi, N.; Yokoyama, Y.; Mori, S.; Yamamoto, H.; Koizumi, M. The CD44 standard isoform contributes to radioresistance of pancreatic cancer cells. *J. Radiat. Res.* 2017, 58, 816–826.
27. Ozawa, M.; Ichikawa, Y.; Zheng, Y.W.; Oshima, T.; Miyata, H.; Nakazawa, K.; Guan, H.B.; Shiozawa, M.; Akaike, M.; Watanabe, K.; et al. Prognostic significance of CD44 variant 2 upregulation in colorectal cancer. *Br. J. Cancer* 2014, 111, 365–374.
28. Gotoda, T.; Matsumura, Y.; Kondo, H.; Saitoh, D.; Shimada, Y.; Kosuge, T.; Kanai, Y.; Kakizoe, T. Expression of CD44 variants and its association with survival in pancreatic cancer. *Jpn. J. Cancer Res.* 1998, 89, 1033–1040.
29. Todoroki, K.; Ogasawara, S.; Akiba, J.; Nakayama, M.; Naito, Y.; Seki, N.; Kusukawa, J.; Yano, H. CD44v3+/CD24- cells possess cancer stem cell-like properties in human oral squamous cell carcinoma. *Int. J. Oncol.* 2016, 48, 99–109.
30. Holm, F.; Hellqvist, E.; Mason, C.N.; Ali, S.A.; Delos-Santos, N.; Barrett, C.L.; Chun, H.J.; Minden, M.D.; Moore, R.A.; Marra, M.A.; et al. Reversion to an embryonic alternative splicing program enhances leukemia stem cell self-renewal. *Proc. Natl. Acad. Sci. USA* 2015, 112, 15444–15449.
31. Bánky, B.; Rásó-Barnett, L.; Barbai, T.; Tímár, J.; Becságh, P.; Rásó, E. Characteristics of CD44 alternative splice pattern in the course of human colorectal adenocarcinoma progression. *Mol. Cancer* 2012, 11, 83.

32. Spiegelberg, D.; Kuku, G.; Selvaraju, R.; Nestor, M. Characterization of CD44 variant expression in head and neck squamous cell carcinomas. *Tumour Biol.* 2014, 35, 2053–2062.
33. Endo, K.; Terada, T. Protein expression of CD44 (standard and variant isoforms) in hepatocellular carcinoma: Relationships with tumor grade, clinicopathologic parameters, p53 expression, and patient survival. *J. Hepatol.* 2000, 32, 78–84.
34. Okuyama, K.; Fukushima, H.; Naruse, T.; Yanamoto, S.; Tsuchihashi, H.; Umeda, M. CD44 Variant 6 Expression and Tumor Budding in the Medullary Invasion Front of Mandibular Gingival Squamous Cell Carcinoma Are Predictive Factors for Cervical Lymph Node Metastasis. *Pathol. Oncol. Res.* 2019, 25, 603–609.
35. Ni, J.; Cozzi, P.J.; Hao, J.L.; Beretov, J.; Chang, L.; Duan, W.; Shigdar, S.; Delprado, W.J.; Graham, P.H.; Bucci, J.; et al. CD44 variant 6 is associated with prostate cancer metastasis and chemo-/radioresistance. *Prostate* 2014, 74, 602–617.
36. Yanamoto, S.; Yamada, S.; Takahashi, H.; Naruse, T.; Matsushita, Y.; Ikeda, H.; Shiraishi, T.; Seki, S.; Fujita, S.; Ikeda, T.; et al. Expression of the cancer stem cell markers CD44v6 and ABCG2 in tongue cancer: Effect of neoadjuvant chemotherapy on local recurrence. *Int. J. Oncol.* 2014, 44, 1153–1162.
37. Masaki, T.; Goto, A.; Sugiyama, M.; Matsuoaka, H.; Abe, N.; Sakamoto, A.; Atomi, Y. Possible contribution of CD44 variant 6 and nuclear beta-catenin expression to the formation of budding tumor cells in patients with T1 colorectal carcinoma. *Cancer* 2001, 92, 2539–2546.
38. Li, Z.; Chen, K.; Jiang, P.; Zhang, X.; Li, X.; Li, Z. CD44v/CD44s expression patterns are associated with the survival of pancreatic carcinoma patients. *Diagn. Pathol.* 2014, 9, 79.
39. Kainz, C.; Kohlberger, P.; Tempfer, C.; Sliutz, G.; Gitsch, G.; Reinthaller, A.; Breiteneker, G. Prognostic value of CD44 splice variants in human stage III cervical cancer. *Eur. J. Cancer* 1995, 31, 1706–1709.
40. Yakushijin, Y.; Steckel, J.; Kharbanda, S.; Hasserjian, R.; Neuberger, D.; Jiang, W.; Anderson, I.; Shipp, M.A. A directly spliced exon 10-containing CD44 variant promotes the metastasis and homotypic aggregation of aggressive non-Hodgkin's lymphoma. *Blood* 1998, 91, 4282–4291.
41. Li, D.M.; Li, S.S.; Zhang, Y.H.; Zhang, H.J.; Gao, D.L.; Wang, Y.X. Expression of human chorionic gonadotropin, CD44v6 and CD44v4/5 in esophageal squamous cell carcinoma. *World J. Gastroenterol.* 2005, 11, 7401–7404.
42. Lourenço, B.N.; Springer, N.L.; Ferreira, D.; Oliveira, C.; Granja, P.L.; Fischbach, C. CD44v6 increases gastric cancer malignant phenotype by modulating adipose stromal cell-mediated ECM remodeling. *Integr. Biol.* 2018, 10, 145–158.
43. Suwannakul, N.; Ma, N.; Midorikawa, K.; Oikawa, S.; Kobayashi, H.; He, F.; Kawanishi, S.; Murata, M. CD44v9 Induces Stem Cell-Like Phenotypes in Human Cholangiocarcinoma. *Front. Cell Dev. Biol.* 2020, 8, 417.
44. Kobayashi, K.; Matsumoto, H.; Matsuyama, H.; Fujii, N.; Inoue, R.; Yamamoto, Y.; Nagao, K. Clinical significance of CD44 variant 9 expression as a prognostic indicator in bladder cancer. *Oncol. Rep.* 2016, 36, 2852–2860.
45. Li, N.; Tsuji, M.; Kanda, K.; Murakami, Y.; Kanayama, H.; Kagawa, S. Analysis of CD44 isoform v10 expression and its prognostic value in renal cell carcinoma. *BJU Int.* 2000, 85, 514–518.
46. Iida, N.; Bourguignon, L.Y. Coexpression of CD44 variant (v10/ex14) and CD44S in human mammary epithelial cells promotes tumorigenesis. *J. Cell. Physiol.* 1997, 171, 152–160.
47. Iida, J.; Clancy, R.; Dorchak, J.; Somiari, R.I.; Somiari, S.; Cutler, M.L.; Mural, R.J.; Shriver, C.D. DNA aptamers against exon v10 of CD44 inhibit breast cancer cell migration. *PLoS ONE* 2014, 9, e88712.
48. Tran, T.A.; Kallakury, B.V.; Sheehan, C.E.; Ross, J.S. Expression of CD44 standard form and variant isoforms in non-small cell lung carcinomas. *Hum. Pathol.* 1997, 28, 809–814.
49. Zhao, P.; Damerow, M.S.; Stern, P.; Liu, A.H.; Sweet-Cordero, A.; Siziopikou, K.; Neilson, J.R.; Sharp, P.A.; Cheng, C. CD44 promotes Kras-dependent lung adenocarcinoma. *Oncogene* 2013, 32, 5186–5190.
50. Dall, P.; Hekele, A.; Ikenberg, H.; Göppinger, A.; Bauknecht, T.; Pfeleiderer, A.; Moll, J.; Hofmann, M.; Ponta, H.; Herrlich, P. Increasing incidence of CD44v7/8 epitope expression during uterine cervical carcinogenesis. *Int. J. Cancer* 1996, 69, 79–85.
51. Omara-Opyene, A.L.; Qiu, J.; Shah, G.V.; Iczkowski, K.A. Prostate cancer invasion is influenced more by expression of a CD44 isoform including variant 9 than by Muc18. *Lab. Invest.* 2004, 84, 894–907.
52. Hiraga, T.; Nakamura, H. Comparable roles of CD44v8-10 and CD44s in the development of bone metastases in a mouse model. *Oncol. Lett.* 2016, 12, 2962–2969.
53. Lau, W.M.; Teng, E.; Chong, H.S.; Lopez, K.A.; Tay, A.Y.; Salto-Tellez, M.; Shabbir, A.; So, J.B.; Chan, S.L. CD44v8-10 is a cancer-specific marker for gastric cancer stem cells. *Cancer Res.* 2014, 74, 2630–2641.
54. Yae, T.; Tsuchihashi, K.; Ishimoto, T.; Motohara, T.; Yoshikawa, M.; Yoshida, G.J.; Wada, T.; Masuko, T.; Mogushi, K.; Tanaka, H.; et al. Alternative splicing of CD44 mRNA by ESRP1 enhances lung colonization of metastatic cancer cell. *Nat.*

55. Choi, E.S.; Kim, H.; Kim, H.P.; Choi, Y.; Goh, S.H. CD44v8-10 as a potential theranostic biomarker for targeting disseminated cancer cells in advanced gastric cancer. *Sci. Rep.* 2017, 7, 4930.
56. Kagami, T.; Yamade, M.; Suzuki, T.; Uotani, T.; Tani, S.; Hamaya, Y.; Iwaizumi, M.; Osawa, S.; Sugimoto, K.; Baba, S.; et al. High expression level of CD44v8-10 in cancer stem-like cells is associated with poor prognosis in esophageal squamous cell carcinoma patients treated with chemoradiotherapy. *Oncotarget* 2018, 9, 34876–34888.
57. Hagiwara, M.; Kikuchi, E.; Tanaka, N.; Kosaka, T.; Mikami, S.; Saya, H.; Oya, M. Variant isoforms of CD44 involves acquisition of chemoresistance to cisplatin and has potential as a novel indicator for identifying a cisplatin-resistant population in urothelial cancer. *BMC Cancer* 2018, 18, 113.
58. Olsson, E.; Honeth, G.; Bendahl, P.O.; Saal, L.H.; Gruvberger-Saal, S.; Ringnér, M.; Vallon-Christersson, J.; Jönsson, G.; Holm, K.; Lövgren, K.; et al. CD44 isoforms are heterogeneously expressed in breast cancer and correlate with tumor subtypes and cancer stem cell markers. *BMC Cancer* 2011, 11, 418.
59. Bourguignon, L.Y.; Zhu, H.; Shao, L.; Zhu, D.; Chen, Y.W. Rho-kinase (ROK) promotes CD44v (3,8-10)-ankyrin interaction and tumor cell migration in metastatic breast cancer cells. *Cell Motil. Cytoskeleton* 1999, 43, 269–287.
60. Fujita, N.; Yaegashi, N.; Ide, Y.; Sato, S.; Nakamura, M.; Ishiwata, I.; Yajima, A. Expression of CD44 in normal human versus tumor endometrial tissues: Possible implication of reduced expression of CD44 in lymph-vascular space involvement of cancer cells. *Cancer Res.* 1994, 54, 3922–3928.
61. Ruiz, P.; Schwärzler, C.; Günthert, U. CD44 isoforms during differentiation and development. *BioEssays* 1995, 17, 17–24.
62. Yu, Q.; Stamenkovic, I. Localization of matrix metalloproteinase 9 to the cell surface provides a mechanism for CD44-mediated tumor invasion. *Genes Dev.* 1999, 13, 35–48.
63. He, Y.; Xue, C.; Yu, Y.; Chen, J.; Chen, X.; Ren, F.; Ren, Z.; Cui, G.; Sun, R. CD44 is overexpressed and correlated with tumor progression in gallbladder cancer. *Cancer Manag. Res.* 2018, 10, 3857–3865.
64. Nishino, M.; Ozaki, M.; Hegab, A.E.; Hamamoto, J.; Kagawa, S.; Arai, D.; Yasuda, H.; Naoki, K.; Soejima, K.; Saya, H.; et al. Variant CD44 expression is enriching for a cell population with cancer stem cell-like characteristics in human lung adenocarcinoma. *J. Cancer* 2017, 8, 1774–1785.
65. Xia, P.; Xu, X.Y. Prognostic significance of CD44 in human colon cancer and gastric cancer: Evidence from bioinformatic analyses. *Oncotarget* 2016, 7, 45538–45546.
66. Li, X.P.; Zhang, X.W.; Zheng, L.Z.; Guo, W.J. Expression of CD44 in pancreatic cancer and its significance. *Int. J. Clin. Exp. Pathol.* 2015, 8, 6724–6731.
67. Mishra, A.; Sriram, H.; Chandarana, P.; Tanavde, V.; Kumar, R.V.; Gopinath, A.; Govindarajan, R.; Ramaswamy, S.; Sasivasam, S. Decreased expression of cell adhesion genes in cancer stem-like cells isolated from primary oral squamous cell carcinomas. *Tumour Biol.* 2018, 40, 1010428318780859.
68. Carneiro, I.; Quintela-Vieira, F.; Lobo, J.; Moreira-Barbosa, C.; Menezes, F.D.; Martins, A.T.; Oliveira, J.; Silva, R.; Jerónimo, C.; Henrique, R. Expression of EMT-Related Genes CAMK2N1 and WNT5A is increased in Locally Invasive and Metastatic Prostate Cancer. *J. Cancer* 2019, 10, 5915–5925.
69. Dzobo, K.; Senthelane, D.A.; Ganz, C.; Thomford, N.E. The significance of Cancer stem cell markers' gene expression and Relevance for Survival Outcomes. *OMICS J. Integr. Biol.* 2020. Preprint.
70. Rajarajan, A.; Stokes, A.; Bloor, B.K.; Ceder, R.; Desai, H.; Grafström, R.C.; Odell, E.W. CD44 expression in oro-pharyngeal carcinoma tissues and cell lines. *PLoS ONE* 2012, 7, e28776.
71. Pietras, A.; Katz, A.M.; Ekström, E.J.; Wee, B.; Halliday, J.J.; Pitter, K.L.; Werbeck, J.L.; Amankulor, N.M.; Huse, J.T.; Holland, E.C. Osteopontin-CD44 signaling in the glioma perivascular niche enhances cancer stem cell phenotypes and promotes aggressive tumor growth. *Cell Stem Cell* 2014, 14, 357–369.
72. Wang, F.; Zheng, Z.; Guan, J.; Qi, D.; Zhou, S.; Shen, X.; Wang, F.; Wenkert, D.; Kirmani, B.; Solouki, T.; et al. Identification of a panel of genes as a prognostic biomarker for glioblastoma. *EBioMedicine* 2018, 37, 68–77.
73. Liu, M.Y.; Su, H.; Huang, H.L.; Chen, J.Q. Cancer stem-like cells with increased expression of NY-ESO-1 initiate breast cancer metastasis. *Oncol. Lett.* 2019, 18, 3664–3672.
74. Dzobo, K.; Sinkala, M. The Cancer Stem Cell Marker CD44 Play Key Roles in Immune Suppression/Evasion, Drug Resistance, Epithelial-Mesenchymal Transition, and Metastasis in Several Human Cancers. *OMICS J. Integr. Biol.* 2021, 20 21, 2021010490.

