

MSC-Secretome for Autoimmune and Immune-mediated Inflammatory Diseases

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Immune-mediated inflammatory diseases (IMIDs) encompass several entities such as “classic” autoimmune disorders or immune-mediated diseases with autoinflammatory characteristics. Adult stem cells including mesenchymal stem cells (MSCs) are by far the most commonly used type in clinical practice. However, due to the possible side effects of MSC-based treatments, there is an increase in interest in the MSC-secretome (containing large extracellular vesicles, microvesicles, and exosomes) as an alternative therapeutic option in IMIDs. A wide spectrum of MSC-secretome-related biological activities has been proven including anti-inflammatory, anti-apoptotic, and immunomodulatory properties. In comparison with MSCs, the secretome is less immunogenic but exerts similar biological actions, so it can be considered as an ideal cell-free therapeutic alternative.

mesenchymal stem cells

Systemic Lupus Erythematosus

Autoimmune disease

Rheumatoid arthritis

Multiple sclerosis

Type 1 diabetes mellitus

1. Systemic Lupus Erythematosus

Systemic lupus erythematosus (SLE), an idiopathic, multifactorial chronic autoimmune disease, is defined mainly by immune dysregulation, antibodies to nuclear and cytoplasmic antigens, systemic inflammation with a wide spectrum of clinical manifestations, and a relapsing-remitting course [1].

Although the use of autologous MSCs in autoimmune and inflammatory diseases appears promising [2][3], there is evidence for obvious MSC deficiency in SLE [4]. MSCs derived from SLE patients exhibit a variety of structural and functional failures compared to healthy MSCs [5][6]. SLE-originated MSCs display abnormalities in their growth, proliferation, differentiation, apoptosis, migration, cytokine production, and immunomodulatory properties. The theoretical basis for using the MSC-secretome in SLE is partly due to the MSC limitations and partly because of the fact that the secretome could affect almost all types of immunocompetent cells.

In SLE, the effect of the MSC-secretome on innate and adaptive immune cells acts against the mechanisms of immune dysregulation. Cargoes of extracellular vesicles such as DNAs, mRNAs, miRNAs, lncRNAs, and circRNAs have been shown to regulate autoimmunity and are associated with SLE activity [7][8]. MSC-exosomes from the human umbilical cord were found to increase the fraction of M2 macrophage polarization both in murine and human lupus [9][10]. Moreover, a recent study confirmed that exosomal tsRNA-21109 from MSCs alleviates SLE by

inhibiting macrophage M1 polarization [11]. Human MSC-EVs inhibit T cell differentiation toward Th1 via glycolysis and cytokine pathway regulation [12].

Accumulating evidence suggests that non-coding RNAs, miRNAs, and circRNAs derived from MSC-EV play a crucial role in the pathogenesis of SLE [7][13][14]. Despite the fact that MSC therapies have demonstrated promise in models of SLE-related pathologies, the function and mechanism of MSC-exos remain unknown. However, it is clear that the MSC-secretome possesses potent anti-inflammatory and immunomodulatory effects [13]. Therefore, the application of the MSC-secretome could be considered as an ideal therapeutic option for severe, refractory SLE. Additionally, MSC-secretome may represent a cell-free, low-immunogenicity alternate solution to disadvantaged MSC therapies [13].

2. Inflammatory Arthritides

Rheumatoid arthritis (RA) is a systemic autoimmune disease of unknown origin that is primarily characterized by chronic, progressive polyarthritis, but also frequently involves a broad spectrum of extra-articular organs by terms of inflammation [15]. In RA, patients lose self-tolerance, and start to produce autoantibodies.

There are numerous conventional and novel disease-modifying anti-rheumatic drugs available for the treatment of RA. Along with their immunosuppressive and immunomodulatory effects, however, several adverse effects must also be considered [15]. Thus, the secretome derived from MSCs with immunomodulatory properties could serve as a suitable therapeutic option.

Many intriguing results have already been obtained from animal experiments within that direction. Not only was the secretome from MSC found to have a significant anti-inflammatory effect in a murine model of collagen-induced arthritis (CIA), but this effect varied among the different secretome fractions [16]. In this experiment, the MSC-derived exosomes were more effective than the microvesicles in increasing the number of regulatory T cells. Additionally, MSC-derived microvesicles induced significantly less IL-10 and TGF β production by the T and B cells than MSCs per se [17].

Proliferation, invasion, and inflammatory response abnormalities of rheumatoid fibroblast-like synoviocytes (RA-FLS) are critical in the progression of RA. Exosomes derived from modified MSCs (Exo-150; containing miR-150-5p) were found to target matrix metalloproteinase 14 (MMP14), and thus inhibit the migration and invasion of RA-FLS [18]. Exo-150 injections decreased the hind paw thickness and clinical arthritic scores in mice with CIA. Exo-150 also inhibited synoviocyte hyperplasia and angiogenesis, thereby reducing joint destruction.

Human umbilical cord MSC-derived exosomes (hUCMSCEs) have also been shown to beneficially influence bone damage in CIA rats [19]. The exosomes inhibited bone destruction by rebalancing the receptor activator of the NF κ B ligand/osteoprotegerin (RANKL/OPG) ratio in a concentration-dependent manner. Furthermore, the intra-articular administration of hUCMSCEs markedly suppressed the serum and synovial fluid levels of the C-C motif chemokine ligand 2 (CCL2) and C-X-C motif chemokine ligand 12 (CXCL12) in the experimental animals [20].

Exosomal circRNAs derived from MSCs could also be a promising therapeutic approach for RA treatment. MSC-derived exosomal circFBXW7 treatment inhibited RA-FLS proliferation, migration, and inflammatory response as well as the RA rat model damage by sponging miR-216a-3p and activating histone deacetylase 4 (HDAC4) [21].

Although animal experiments have been extremely encouraging and no significant adverse effects associated with the use of secretome have been reported, human trials have not begun yet. It should also be borne in mind that some new research [22][23] suggests that epigenetically modified bone-marrow-derived MSCs (and presumably also their secretome) producing more IL-6 may promote the progression of RA by migrating to the synovium. In any case, more research is required to investigate MSC-based and cell-free therapeutic options for human RA.

Spondyloarthritis (SpA) is defined by synovial and enthesal involvement, resulting in spinal and oligoarticular peripheral arthritis, most commonly in genetically predisposed (HLA-B27+) individuals. Additionally, extra-musculoskeletal, systemic inflammatory organ manifestations may also occur. SpA may be listed as an autoinflammatory disease, with a predominant involvement of innate immunity. Adult spondyloarthritis can be classified into several well-defined subtypes; (one of the most known is Bechterew's disease) [24].

Interestingly, the use of MSCs and MSC-secretome in the SpA group can cause not only beneficial but also detrimental effects. In mice with a low expression of A20 (a modulator of signal transducer and activator of transcription 1/STAT1-dependent gene transcription) in myeloid cells, early enthesitis has been found [25]. In parallel, the inhibition of the JAK/STAT1 pathway resulted in clinical and histological improvement of enthesitis [25]. It would not be unexpected if a similar defect in human BM-MSCs contributed to the development of enthesitis in response to TNF stimulation. In a TNF transgenic mouse model, the selective expression of TNF receptor I in mesenchymal cells was detected, and enthesitis occurred regardless of the presence or absence of mature T and B cells. This indicates that only mesenchymal cells are crucial for the development of joint inflammation [26][27]. Because reactive arthritis sometimes precedes spondyloarthritis, it is hypothesized that transient or persistent silent infections affecting the bone marrow may contribute to the adverse phenotyping of bone marrow-derived MSCs [28]. Likewise, the role of EVs has been found to promote the initiation and spread of the underlying disease (including spondyloarthritides), especially in obese people, through an interaction between adipocytes and immune cells mediated by EVs [29][30]. In spondyloarthritides, the study of positive or negative therapeutic effects of the MSC-secretome is essential. In addition to animal models and cell culture experiments, detailed human studies must be introduced.

3. Multiple Sclerosis

Multiple sclerosis (MS) is an inflammatory demyelinating autoimmune disease of the central nervous system in which the protective myelinated axons of the nerve fibers in the brain and spinal cord are targeted, along with parallel disruption of the blood–brain barrier. MS can be present in several forms such as a progressive or a fluctuating, relapsing-remitting type [31][32][33].

The success of using MSCs in MS has fallen short of the expectations. In patients with the relapsing-remitting type, disease-modifying treatment altered the activity of MSCs [34]. Additionally, it was established that the patients' own MSCs were unsuitable for disease prevention [35]. MSC differentiation into neurons, however, has a significant neuroprotective effect [36]. MSC-exosomes, as nanoscale vesicles coated with a lipid membrane, readily cross the blood–brain barrier. Exosomes can be used to modify autoreactive T cells in a pro-tolerance direction and deliver factors to the central nervous system that promote neuronal survival and remyelination [36]. Thus, their utility in MS is almost self-evident.

By modifying exosomes, receptors may be added to their surface, allowing for targeted drug delivery to cells. Exosome-encapsulated curcumin or JSI124 (a selective STAT3 inhibitor) were delivered to microglia cells via an intranasal route in mice [37]. The results of the experimental autoimmune encephalitis (EAE) demonstrated that mice treated intranasally with modified exosomes are protected from the progression of the inflammatory disorder induced by the myelin oligodendrocyte glycoprotein peptide. Intranasal administration of the modified exosomes resulted in the rapid delivery of the exosome-encapsulated drug to the brain, where it was selectively taken up by microglial cells and subsequently induced their apoptosis [37]. MSC-derived EVs also have an effect on splenic mononuclear cells in EAE mice, increasing their IL-10 secretion. Moreover, they can exert immunoregulatory effects by delivering potent biosubstances to immunocompetent cells via EVs (e.g., TGF β , programmed death-ligand 1, Gal-1) [38]. These findings may help to explain the efficacy of the MSC-secretome in the EAE mice model. Additionally, in a mouse model of progressive MS (i.e., Theiler's murine encephalomyelitis virus /TMEV/-induced demyelinating disease) it was demonstrated that EVs exert potent immunoregulatory actions. IV administration of EVs derived from human adipose-derived MSCs ameliorated motor deficits in infected mice via immunomodulatory effects, reducing brain atrophy and promoting remyelination [39]. In contrast, however, it was demonstrated that exosomes secreted by microglia during early TMEV infection contained viral RNA and were capable of activating uninfected bystander CNS cells, thereby promoting an inflammatory immune response. Thus, exosomes secreted by microglia during virus infection could contribute to viral persistence and neuroinflammation, both of which contribute to the development of demyelinating disease [40].

The therapeutic efficacy of MSC-exosomes is also supported by the fact that MSC-derived exosomes injected into the tail vein of EAE rats exhibited immunoregulatory capacity, indicating that exosomes from MSCs could regulate microglia polarization from the M1 to M2 phenotype [41]. Additional cell-specific therapies are also possible via exosome manipulation. Recently, covalent conjugation of the carboxylic acid-functionalized LJM-3064 aptamer to the amine groups on the MSC-exosome surface was performed. The aptamer-exosome bioconjugate was found to promote the oligodendroglia cell line proliferation in vitro. Furthermore, upon in vivo administration of the prepared bioconjugate to female EAE-mice, the inflammatory response was suppressed along with a decrease in the demyelinating CNS lesions, thus resulting in a reduction in the disease severity [42].

Exosomes loaded selectively and made their membrane cell specific enables the development of additional targeted treatment options. Exosomes isolated from rhesus monkey MSCs were utilized in two mouse models of demyelination (EAE and a toxic demyelination model using cuprizone; CPZ) [43]. In the CPZ model, MSC-Exos significantly improved the neurological outcome, decreased the amyloid precursor protein density, decreased

neuroinflammation by increasing the M2 phenotype of microglia and their associated cytokines, and inhibited the TLR2/interleukin 1 receptor associated kinase 1/NF κ B pathway. Moreover, the MSC-Exo treatment considerably promoted cognitive function, facilitated remyelination, increased M2 phenotypic polarization, and inhibited TLR2 signaling [43].

Exosomes derived from neural stem cells were targeted using a lentivirus-armed PDGFR-alpha ligand capable of anchoring the membrane [44]. Exosomes modified in this manner appeared to be capable of targeting oligodendrocyte progenitor cells in the neural lesion area. Following this, the exosomes were loaded with bryostatin-1, and the targeted exosomes were administered to the CPZ-treated mice. The data demonstrated that loaded-armed exosomes have a potent therapeutic effect. This novel exosome-based delivery method significantly enhanced the myelin sheath's ability to protect integrity and promote remyelination. Additionally, it inhibited astrogliosis and axon damage as well as pro-inflammatory microglia [44].

Although these procedures may represent a paradigm shift in the treatment of MS, their clinical utility remains unknown. In a recent study, the suppressive activity of hUCMSCEs was investigated in vitro in co-culture with peripheral blood mononuclear cells (PBMCs) [45]. It was demonstrated that MSCs inhibited PBMC proliferation and that MSC-derived exosomes were more effective at suppressing proliferation with a low alloreaction rate. However, after extensive sub-culturing, the suppression capacity of MSCs and their exosomes decreased significantly [45]. Regrettably, additional evaluations are necessary to progress toward a clinically applicable functional approach. According to www.clinicaltrials.gov (accessed on 22 May 2022), only one study is planned in relapsing MS, in which, as a sub-target, the analysis of molecular and biochemical parameters including circulating exosomes are planned to perform and compare responder and non-responder patients.

4. Uveitis

Uveitis is an intraocular inflammatory condition, a form of potentially blinding diseases that can affect either one or both eyes. Different kinds of uveitis may affect various areas of the eye [46]. Uveitis can arise as a result of the immune system countering an eye infection, but it can also happen as a consequence of the immune system, attacking healthy tissue in the eyes, and some forms are considered due to aberrant autoimmune responses [46]. Conventionally, systemic and local methods have been used to treat the disease, though their long-term side effects are frequently severe.

Animal models of experimental autoimmune uveitis are frequently used to study the disease [47]. Upon a rat model of experimental autoimmune uveitis (EAU) [48], it has been demonstrated that the topical application of MSC-Exos can inhibit the autoimmune response in vivo. It was discovered that periocular injections of human MSC-Exo can decrease immune cell infiltration, preserve retinal structure, and restore retinal function in EAU rats. This suggests that MSC-Exos may be useful in treating autoimmune uveitis in humans. Significantly, xenogeneic MSC-Exo from humans improved the autoimmune reaction in rats [48].

The post-uveitis macular hole is a potentially severe complication [49]. In a recent pilot study, the efficacy of the intravitreal injection of MSC-Exos following standard pars plana vitrectomy (PPV) in conjunction with inverted internal limiting membrane (ILM) flap technique and ILM peeling was evaluated for the treatment of massive and resistant macular holes [50]. The findings suggest that intravitreal injection of MSC-Exos at the end of standard PPV may improve the anatomical and visual outcomes of surgery for macular holes that have been resistant to treatment. MSC-Exo therapy was found to be safer than MSC therapy in terms of the cell proliferation risk, and it was less invasive since no additional surgery was required [50]. In posterior uveitis, cells of the retinal pigment epithelium (RPE) may be destroyed by inflammatory activities [50]. RPE cells may release immunosuppressive exosomes that suppress Th17 and Th22 cells [51]. In light of the immunoregulatory effect of RPE-derived exosomes, they just might be used as a treatment for uveitis. It has been found that MSCs-derived exosomes from bone marrow can mitigate autoimmune uveoretinitis [52]. Since it is evident that only a few studies have examined the efficacy of MSC-secretome in uveitis, the need for additional research is unquestionable.

5. Type 1 Diabetes Mellitus

Type 1 (or insulin-dependent) diabetes mellitus (T1DM) is a chronic autoimmune disease that results in the destruction of insulin-producing β -cells in the pancreas [53]. Consequently, the reduced level of insulin finally leads to hyperglycemia. Repeated insulin injections indicate that insulin levels are not stable in affected subjects and that T1DM patients require long-lasting treatment.

Immunotherapies and cell replacement, which include islet transplantation and stem cell differentiation into β -cells, are the primary treatment strategies for T1DM [54][55]. The simultaneous restorative and immunomodulatory properties of EVs aligned with new hypotheses regarding the nature of T1DM could make them a favorable treatment option. In general, therapies for T1DM should be able to restore B cells while modulating the immune system [56]. Autoreactive T cells have been recognized as the primary attackers due to immune dysregulation. In contrast, regulatory T cells have lost their ability to generate peripheral tolerance to β -cells [57]. EVs derived from distinct MSCs frequently exert their immunomodulatory effects in different ways.

In murine models of T1DM, EVs that originated from bone marrow-derived MSCs resulted in a delayed onset of T1DM, the preservation of islet cells, the reduction in insulitis and T cell infiltration, the decrease in antigen-presenting cell (APC) and T cell activation, lessening the Th1/Th17 population, and a decline in the production of inflammatory cytokines including IL-17, IL-6, IFNy, TNF α , and IL-12 [52]. It was detected in mice that β (MIN6)-cell-derived EVs reduced the number of pro-inflammatory macrophages in pancreatic islets and enhanced islet angiogenesis [58].

EVs from human adipose tissue-derived MSCs increased the number of regulatory T cells in the spleen, raised the levels of immunomodulatory cytokines (like IL-4, IL-10, and TGF β), and decreased the production of inflammatory cytokines such as IFNy and IL-17 [59]. Accordingly, EV therapy stabilized blood glucose and eradicated diabetes symptoms.

The total exosome fraction isolated from Psh-Fas-anti-miR375 transfected human bone marrow-derived MSCs co-cultured with peripheral blood MSCs was found to reduce islet cell apoptosis and stimulate insulin release. Moreover, exosomes increased the number of regulatory T cells, reduced the levels of inflammatory cytokines (IL-2 and IFNy), and diminished immune-rejection after islet transplantation [60].

It was found that human bone marrow-derived MSC-EVs induced the immature IL-10-producing phenotype of dendritic cells, increased the IL-10 and IL-6 levels, and the number of regulatory T cells, while it decreased the number of Th17 cells [61]. Last but not least, EVs derived from human cord blood stem cells induced monocyte differentiation into anti-inflammatory M2 macrophages [62].

6. Inflammatory Bowel Diseases

The term inflammatory bowel disease (IBD) refers to a group of disorders that are characterized by chronic, progressive immune-mediated inflammation of the digestive tract with frequent extra-intestinal, systemic organ involvement. Ulcerative colitis (UC) and Crohn's disease (CD) are listed as the prototype disorders [63]. The GI-tract plays a crucial role in maintaining the host's immune homeostasis, so a disrupted balance between immune tolerance and inflammation may lead to IBD [63].

Based on animal models of IBD, the use of MSCs has been shown to have effective anti-inflammatory and tissue repair actions. The efficacy and safety of the MSC-secretome have also been demonstrated in animal studies. It is hypothesized that if MSCs are injected locally into the therapy-refractory fistulas of Crohn's disease patients, the closure of fistulas is mainly due to the immunomodulatory activity of the secretome, instead of the MSCs per se [64].

The local or systemic administration of MSC-derived EVs in mice IBD models induced by dextran sodium sulfate (DSS) has been shown to ameliorate the clinical severity of colitis, resulting in improved survival rates and reduced gut inflammation in histology [65][66][67]. These positive effects have been linked to the polarization of macrophages toward the M2 phenotype as well as the reduction in oxidative stress in the affected tissues [67][68]. It has also been demonstrated that IFNy- and TNF α -preconditioned MSCs enrich EVs in HGF, PGE2, or TGF β [65]. In mice models of DSS-induced colitis, MSC-derived conditioned medium (MSC-CM) has been found to downregulate the expression of TNF α or IL-1 but upregulate IL-10 [69].

In vitro exposure of macrophages to the MSC-derived secretome, followed by their implantation in a 2,4,6-trinitrobenzenesulfonic acid (TNBS)-induced colitis mouse model, demonstrated that preconditioned macrophages enhanced the survival rate by decreasing weight loss and bloody stool, thereby preventing colitis progression and disease recurrence [70].

Although animal and cell culture researches in IBD have indicated the favorable effect of the MSC-secretome, human studies are still needed.

7. Graft-Versus-Host Disease

Graft versus host disease (GvHD) represents a potentially fatal multiorgan complication of allogeneic bone marrow or peripheral blood HSC transplantation. It is based on the fact that the host appears to non-self to the graft and so it can stimulate antigenically, employing a broad spectrum of immune mechanisms. GvHD is classified into acute and chronic types [21]. The chronic form frequently mimics autoimmune diseases.

In 2004, it was reported that the immunomodulatory qualities of MSCs were effective in curing GvHD caused by HSCT [22]. Since then, extensive preclinical and clinical trials in humans have investigated the use of MSCs to treat GvHD. However, the outcomes are inconsistent and contentious [23][24]. This has been primarily attributed to technical issues, but it may also be due to the ability of each patient's unique microenvironment to polarize the anti-inflammatory MSC2 phenotype. In this situation, cell-free therapies could be a suitable option.

Undoubtedly, EVs have demonstrated promising results in the prevention and treatment of GvHD [25]. Using MSC-EVs to treat acute and chronic GvHD has been shown to reduce the severity of clinical manifestations (e.g., symptoms of liver, gastrointestinal tract, skin) [26]. Recently, in an acute GvHD mouse model, the effects of bone marrow-MSC-derived EVs administered intraperitoneally were investigated. The results underscored that EVs could not only lessen the severity of the disease but also stop it from getting worse, and so increase the number of animals that survived [27]. Despite all of this, human studies have not begun yet.

8. Psoriasis

Psoriasis refers to a chronic, immune-mediated, non-infectious inflammatory skin disease with keratinocyte hyperproliferation in the epidermis that is characterized clinically by the development of reddish, scaly papules and plaques. Several subtypes can be distinguished. Systemic manifestations such as painful (spondylo)arthritis or ocular involvement could also occur [28]. Psoriasis is considered as an autoimmune disease.

Until recently, there have been no applicable animal models of psoriasis, so the xenotransplantation of psoriatic skin pieces from human to mice with severe combined immunodeficiency was utilized [29]. The occurrence of animal models that have been genetically modified has altered this circumstance. Mice with an epidermis-specific deletion of the inhibitor of nuclear factor-B kinase 2 can develop a psoriasis-like skin disease independent of T cells [30]. It has been recently demonstrated that the subcutaneous injection of hUCMSCEs significantly suppressed the proliferation of epidermis and reduced the psoriasis area and severity index scores in imiquimod-induced mice [31]. Moreover, the expression of IL-17, IL-23, and CCL20 was reduced, and the phosphorylation of STAT3 was inhibited both in the skin of mice and human keratinocytes. In addition, the maturation and activation of dendritic cells were inhibited when co-cultured with hUCMSCEs in vitro, and the expression level of IL-23 was also reduced. Furthermore, in a rat model of psoriasis induced by imiquimod, the topical application of a cream containing bone marrow-derived MSC-secretome ameliorated the skin lesions [30].

In a recent and only human study [32], adipose tissue-derived MSCs were collected, multiplied, and then the MSC-CM was extracted and concentrated. The conditioned medium was topically applied to the patients' affected skin

areas once a day over one month. The clearance of silvery scales and the severity of plaques were completely abolished within the treatment period. Moreover, the beneficial effect of the treatment was observed after 6 months.

The results so far are encouraging for skin diseases, but further studies are required to determine the route and duration of application for human therapy.

9. Atopic Dermatitis

Atopic dermatitis (AD) is a common, heterogeneous, chronic, itchy, inflammatory skin disease frequently with increased serum IgE. Defects in terminal keratinocyte differentiation and altered immune responses are cardinal disease determinants. AD can be the initial manifestation of a spectrum of allergic diseases (the “atopic march” theory) [83].

It has been reported that exosomes derived from adipose tissue-derived MSCs (AT-MSCs) inhibit the lipopolysaccharide-induced nitric oxide production of RAW264.7 macrophages [83]. In a murine model of AD, the subcutaneous and intravenous administration of the AT-MSC-exosomes mitigated the symptoms and decreased the serum IgE levels in a dose-dependent manner. In AD-like lesions, the AT-MSC-exosomes reduced the number of CD86+ cells, mast cells, and CD206+ cells. After systemic administration, these exosomes alleviated AD-like symptoms, and in skin lesions, the mRNA levels of IL-4, IL-31, IL-23, and TNF α were reduced [84][85].

Therefore, it seems that AT-MSC-exosomes are able to favorably influence the outcome of AD, but this hypothesis demands further investigations.

10. Asthma

Asthma is a heterogeneous chronic inflammatory disease of the lungs' airways provoked by exogenous (like air pollution, cigarette smoke) and endogenous factors (such as oxidative stress, chronic inflammation), and are mainly driven by a “type 2-high” immune signature [86]. Most current medications focus on regulating the inflammatory cascade, but cannot reverse tissue remodeling.

Some years ago, in an ovalbumin-induced murine asthma model (OVA-mice) [87], it was demonstrated that adipose-tissue derived EVs were superior to MSCs in terms of decreasing pro-inflammatory mediators and inflammatory cell infiltration. Both therapeutic interventions also reduced the number of eosinophils, collagen fiber content, and TGF β level in a comparable manner [87]. Furthermore, in OVA-mice [88], hypoxic hUCMSCEs significantly ameliorated eosinophils and pro-inflammatory mediators (IL-4 and IL-13). Moreover, hypoxic-EVs were generally more potent in suppressing airway inflammation, preventing airway remodeling, and decreasing the expression of pro-fibrogenic markers [88]. Additionally, in OVA-mice, it was also found that EVs from MSCs in adipose tissue helped reduce allergic airway inflammation and airway hyperresponsiveness caused by the activation of regulatory T cells to expansion [89]. Despite the encouraging results, to identify the EV components responsible for the suppression of allergic airway inflammation in asthmatic mice, additional investigations are

required. Moreover, human experiments provide greater biological complexity than model experiments with mice. Thus, human models are needed to figure out what kind of role adipose-tissue MSC-derived EVs might play in allergic airway inflammation.

References

1. Aringer, M.; Alarcón-Riquelme, M.E.; Clowse, M.; Pons-Estel, G.J.; Vital, E.M.; Dall'Era, M. A glimpse into the future of systemic lupus erythematosus. *Ther. Adv. Musculoskelet Dis.* 2022, 14, 1–21.
2. Wang, L.T.; Liu, K.J.; Sytwu, H.K.; Yen, M.L.; Yen, B.L. Advances in mesenchymal stem cell therapy for immune and inflammatory diseases: Use of cell-free products and human pluripotent stem cell-derived mesenchymal stem cells. *Stem Cells Transl. Med.* 2021, 10, 1288–1303.
3. Zhuang, W.Z.; Lin, Y.H.; Su, L.J.; Wu, M.S.; Jeng, H.Y.; Chang, H.C.; Huang, Y.H.; Ling, T.Y. Mesenchymal stem/stromal cell-based therapy: Mechanism, systemic safety and biodistribution for precision clinical applications. *J. Biomed. Sci.* 2021, 28, 28.
4. Popis, M.; Konwerska, A.; Partyka, M.; Wieczorkiewicz, M.; Ciesiółka, S.; Stefańska, K.; Spaczyńska, J.; Golkar-Narenji, A.; Jeseta, M.; Bukowska, D.; et al. Mesenchymal stem cells and their secretome—Candidates for safe and effective therapy for systemic lupus erythematosus. *Med. J. Cell Biol.* 2021, 9, 110–122.
5. Sun, L.Y.; Zhang, H.Y.; Feng, X.B.; Hou, Y.Y.; Lu, L.W.; Fan, L.M. Abnormality of bone marrow-derived mesenchymal stem cells in patients with systemic lupus erythematosus. *Lupus* 2007, 16, 121–128.
6. Fathollahi, A.; Gabalou, N.B.; Aslani, S. Mesenchymal stem cell transplantation in systemic lupus erythematosus, a mesenchymal stem cell disorder. *Lupus* 2018, 27, 1053–1064.
7. Perez-Hernandez, J.; Martinez-Arroyo, O.; Ortega, A.; Galera, M.; Solis-Salguero, M.A.; Chaves, F.J.; Redon, J.; Forner, M.J.; Cortes, R. Urinary exosomal miR-146a as a marker of albuminuria, activity changes and disease flares in lupus nephritis. *J. Nephrol.* 2021, 34, 1157–1167.
8. Cortes, R.; Forner, M.J. Circular RNAs: Novel biomarkers of disease activity in systemic lupus erythematosus? *Clin. Sci.* 2019, 133, 1049–1052.
9. Chen, X.; Wei, Q.; Sun, H.; Zhang, X.; Yang, C.; Tao, Y.; Nong, G. Exosomes Derived from Human Umbilical Cord Mesenchymal Stem Cells Regulate Macrophage Polarization to Attenuate Systemic Lupus Erythematosus-Associated Diffuse Alveolar Hemorrhage in Mice. *Int. J. Stem Cells.* 2021, 14, 331–340.

10. Shi, D.; Wang, D.; Li, X.; Zhang, H.; Che, N.; Lu, Z.; Sun, L. Allogeneic transplantation of umbilical cord-derived mesenchymal stem cells for diffuse alveolar hemorrhage in systemic lupus erythematosus. *Clin. Rheumatol.* 2012, 31, 841–846.
11. Dou, R.; Zhang, X.; Xu, X.; Wang, P.; Yan, B. Mesenchymal stem cell exosomal tsRNA-21109 alleviate systemic lupus erythematosus by inhibiting macrophage M1 polarization. *Mol. Immunol.* 2021, 139, 106–114.
12. Xu, H.; Chen, W.; Zheng, F.; Tang, D.; Liu, D.; Wang, G.; Xu, Y.; Yin, L.; Zhang, X.; Dai, Y. Reconstruction and analysis of the aberrant lncRNA-miRNA-mRNA network in systemic lupus erythematosus. *Lupus* 2020, 29, 398–406.
13. Yang, C.; Sun, J.; Tian, Y.; Li, H.; Zhang, L.; Yang, J.; Wang, J.; Zhang, J.; Yan, S.; Xu, D. Immunomodulatory Effect of MSCs and MSCs-Derived Extracellular Vesicles in Systemic Lupus Erythematosus. *Front. Immunol.* 2021, 12, 714832.
14. Perez-Hernandez, J.; Forner, M.J.; Pinto, C.; Chaves, F.J.; Cortes, R.; Redon, J. Increased Urinary Exosomal MicroRNAs in Patients with Systemic Lupus Erythematosus. *PLoS ONE* 2015, 10, e0138618.
15. Wei, K.; Jiang, P.; Zhao, J.; Jin, Y.; Zhang, R.; Chang, C.; Xu, L.; Xu, L.; Shi, Y.; Guo, S.; et al. Biomarkers to Predict DMARDs Efficacy and Adverse Effect in Rheumatoid Arthritis. *Front. Immunol.* 2022, 13, 865267.
16. Cosenza, S.; Toupet, K.; Maumus, M.; Luz-Crawford, P.; Blanc-Brude, O.; Jorgensen, C.; Noël, D. Mesenchymal stem cells-derived exosomes are more immunosuppressive than microparticles in inflammatory arthritis. *Theranostics* 2018, 8, 1399–1410.
17. Conforti, A.; Scarsella, M.; Starc, N.; Giorda, E.; Biagini, S.; Proia, A.; Carsetti, R.; Locatelli, F.; Bernardo, M.E. Microvescicles derived from mesenchymal stromal cells are not as effective as their cellular counterpart in the ability to modulate immune responses in vitro. *Stem Cells Dev.* 2014, 23, 2591–2599.
18. Chen, Z.; Wang, H.; Xia, Y.; Yan, F.; Lu, Y. Therapeutic Potential of Mesenchymal Cell-Derived miRNA-150-5p-Expressing Exosomes in Rheumatoid Arthritis Mediated by the Modulation of MMP14 and VEGF. *J. Immunol.* 2018, 201, 2472–2482.
19. Gao, J.; Xu, K.; Zhang, G.; Han, J.; Liu, Y.; Zhang, L. FRI0510 the effect and mechanism of human umbilical cord mesenchymal stem cell-derived exosomes on bone destruction of collagen induced arthritis rats. *Ann. Rheum. Dis.* 2019, 78, 950.
20. He, P. AB0291E the effect of human umbilical cord mesenchymal stem cell-derived exosomes on chemokines in collagen-induced arthritis rats. *Ann. Rheum. Dis.* 2019, 78, 1606.
21. Chang, L.; Kan, L. Mesenchymal Stem Cell-Originated Exosomal Circular RNA circFBXW7 Attenuates Cell Proliferation, Migration and Inflammation of Fibroblast-Like Synoviocytes by

Targeting miR-216a-3p/HDAC4 in Rheumatoid Arthritis. *J. Inflamm. Res.* 2021, **14**, 6157–6171.

22. Berthelot, J.M.; Sibilia, J. Rampant infections of bone marrow stem cell niches as triggers for spondyloarthropathies and rheumatoid arthritis. *Clin. Exp. Rheumatol.* 2016, **34**, 329–336.

23. Berthelot, J.M.; Sibilia, J. Trained Immunity and Autoimmune Disease: Did Eve Sin before Adam? *Jt. Bone Spine* 2019, **86**, 293–295.

24. Mease, P.J. Suspecting and Diagnosing the Patient with Spondyloarthritis and What to Expect from Therapy. *Rheum. Dis. Clin. North Am.* 2022, **48**, 507–521.

25. De Wilde, K.; Martens, A.; Lambrecht, S.; Jacques, P.; Drennan, M.B.; Debusschere, K.; Govindarajan, S.; Coudenys, J.; Verheugen, E.; Windels, F.; et al. A20 inhibition of STAT1 expression in myeloid cells: A novel endogenous regulatory mechanism preventing development of enthesitis. *Ann. Rheum. Dis.* 2017, **76**, 585–592.

26. Armaka, M.; Apostolaki, M.; Jacques, P.; Kontoyiannis, D.L.; Elewaut, D.; Kollias, G. Mesenchymal cell targeting by TNF as a common pathogenic principle in chronic inflammatory joint and intestinal diseases. *J. Exp. Med.* 2008, **205**, 331–337.

27. Jacques, P.; Lambrecht, S.; Verheugen, E.; Pauwels, E.; Kollias, G.; Armaka, M.; Verhoye, M.; Van der Linden, A.; Achten, R.; Lories, R.J.; et al. Proof of concept: Enthesitis and new bone formation in spondyloarthritis are driven by mechanical strain and stromal cells. *Ann. Rheum. Dis.* 2014, **73**, 437–445.

28. Berthelot, J.M.; Le Goff, B.; Maugars, Y. Bone marrow mesenchymal stem cells in rheumatoid arthritis, spondyloarthritis, and ankylosing spondylitis: Problems rather than solutions? *Arthritis Res. Ther.* 2019, **21**, 239.

29. Kumar, V.; Kiran, S.; Kumar, S.; Singh, U.P. Extracellular vesicles in obesity and its associated inflammation. *Int. Rev. Immunol.* 2022, **41**, 30–44.

30. Wang, Y.Y.; Wang, Y.D.; Qi, X.Y.; Liao, Z.Z.; Mai, Y.N.; Xiao, X.H. Organokines and Exosomes: Integrators of Adipose Tissue Macrophage Polarization and Recruitment in Obesity. *Front. Endocrinol.* 2022, **13**, 839849.

31. Duarte-Silva, E.; Meuth, S.G.; Peixoto, C.A. Microbial Metabolites in Multiple Sclerosis: Implications for Pathogenesis and Treatment. *Front. Neurosci.* 2022, **16**, 885031.

32. Sandi, D.; Kokas, Z.; Biernacki, T.; Bencsik, K.; Klivényi, P.; Vécsei, L. Proteomics in Multiple Sclerosis: The Perspective of the Clinician. *Int. J. Mol. Sci.* 2022, **23**, 5162.

33. Travers, B.S.; Tsang, B.K.; Barton, J.L. Multiple sclerosis: Diagnosis, disease-modifying therapy and prognosis. *Aust. J. Gen. Pract.* 2022, **51**, 199–206.

34. Emamnejad, R.; Sahraian, M.; Shakiba, Y.; Salehi, Z.; Masoomi, A.; Imani, D.; Najafi, F.; Laribi, B.; Shirzad, H.; Izad, M. Circulating mesenchymal stem cells, stromal derived factor (SDF)-1 and IP-

10 levels increased in clinically active multiple sclerosis patients but not in clinically stable patients treated with beta interferon. *Mult. Scler. Relat. Disord.* 2019, 35, 233–238.

35. Sargent, A.; Shano, G.; Karl, M.; Garrison, E.; Miller, C.; Miller, R.H. Transcriptional Profiling of Mesenchymal Stem Cells Identifies Distinct Neuroimmune Pathways Altered by CNS Disease. *Int. J. Stem Cells* 2018, 11, 48–60.

36. Baharloo, H.; Azimi, M.; Salehi, Z.; Izad, M. Mesenchymal Stem Cell-Derived Exosomes: A Promising Therapeutic Ace Card to Address Autoimmune Diseases. *Int. J. Stem Cells* 2020, 13, 13–23.

37. Zhuang, X.; Xiang, X.; Grizzle, W.; Sun, D.; Zhang, S.; Axtell, R.C.; Ju, S.; Mu, J.; Zhang, L.; Steinman, L.; et al. Treatment of brain inflammatory diseases by delivering exosome encapsulated anti-inflammatory drugs from the nasal region to the brain. *Mol. Ther.* 2011, 19, 1769–1779.

38. Mokarizadeh, A.; Delirezh, N.; Morshedi, A.; Mosayebi, G.; Farshid, A.A.; Mardani, K. Microvesicles derived from mesenchymal stem cells: Potent organelles for induction of tolerogenic signaling. *Immunol. Lett.* 2012, 147, 47–54.

39. Laso-García, F.; Ramos-Cejudo, J.; Carrillo-Salinas, F.J.; Otero-Ortega, L.; Feliú, A.; Gómez-de Frutos, M.; Mecha, M.; Díez-Tejedor, E.; Guaza, C.; Gutiérrez-Fernández, M. Therapeutic potential of extracellular vesicles derived from human mesenchymal stem cells in a model of progressive multiple sclerosis. *PLoS ONE* 2018, 13, e0202590.

40. Luong, N.; Olson, J.K. Exosomes Secreted by Microglia During Virus Infection in the Central Nervous System Activate an Inflammatory Response in Bystander Cells. *Front. Cell Dev. Biol.* 2021, 9, 661935.

41. Li, Z.; Liu, F.; He, X.; Yang, X.; Shan, F.; Feng, J. Exosomes derived from mesenchymal stem cells attenuate inflammation and demyelination of the central nervous system in EAE rats by regulating the polarization of microglia. *Int. Immunopharmacol.* 2019, 67, 268–280.

42. Hosseini Shamsili, F.; Alibolandi, M.; Rafatpanah, H.; Abnous, K.; Mahmoudi, M.; Kalantari, M.; Taghdisi, S.M.; Ramezani, M. Immunomodulatory properties of MSC-derived exosomes armed with high affinity aptamer toward myelin as a platform for reducing multiple sclerosis clinical score. *J. Control Release* 2019, 299, 149–164.

43. Zhang, J.; Buller, B.A.; Zhang, Z.G.; Zhang, Y.; Lu, M.; Rosene, D.L.; Medalla, M.; Moore, T.L.; Chopp, M. Exosomes derived from bone marrow mesenchymal stromal cells promote remyelination and reduce neuroinflammation in the demyelinating central nervous system. *Exp. Neurol.* 2022, 347, 113895.

44. Wu, X.Y.; Liao, B.Y.; Xiao, D.; Wu, W.C.; Xiao, Y.; Alexander, T.; Song, S.J.; Zhao, Z.H.; Zhang, Y.; Wang, Z.H.; et al. Encapsulation of bryostatin-1 by targeted exosomes enhances remyelination

and neuroprotection effects in the cuprizone-induced demyelinating animal model of multiple sclerosis. *Biomater. Sci.* 2022, **10**, 714–727.

45. Baharloo, H.; Nouraei, Z.; Azimi, M.; Moghadasi, A.N.; Tavassolifar, M.J.; Moradi, B.; Sahraian, M.A.; Izad, M. Umbilical cord mesenchymal stem cells as well as their released exosomes suppress proliferation of activated PBMCs in multiple sclerosis. *Scand. J. Immunol.* 2021, **93**, e13013.

46. Miller, J.R.; Hanumunthadu, D. Inflammatory eye disease: An overview of clinical presentation and management. *Clin. Med.* 2022, **22**, 100–103.

47. Liang, D.; Shao, H.; Born, W.K.; O'Brien, R.L.; Kaplan, H.J.; Sun, D. Connection between $\gamma\delta$ T-cell- and Adenosine- Mediated Immune Regulation in the Pathogenesis of Experimental Autoimmune Uveitis. *Crit. Rev. Immunol.* 2018, **38**, 233–243.

48. Bai, L.; Shao, H.; Wang, H.; Zhang, Z.; Su, C.; Dong, L.; Yu, B.; Chen, X.; Li, X.; Zhang, X. Effects of Mesenchymal Stem Cell-Derived Exosomes on Experimental Autoimmune Uveitis. *Sci. Rep.* 2017, **7**, 4323.

49. Shukla, D.; Dhawan, A. Evolution and Management of a Post-Uveitis Macular Hole. *Ophthalmic Surg. Lasers Imaging* 2010, **1–3**, online ahead of print.

50. Zhang, X.; Liu, J.; Yu, B.; Ma, F.; Ren, X.; Li, X. Effects of mesenchymal stem cells and their exosomes on the healing of large and refractory macular holes. *Graefes. Arch. Clin. Exp. Ophthalmol.* 2018, **256**, 2041–2052.

51. Shi, Q.; Wang, Q.; Li, J.; Zhou, X.; Fan, H.; Wang, F.; Liu, H.; Sun, X.; Sun, X. A2E Suppresses Regulatory Function of RPE Cells in Th1 Cell Differentiation Via Production of IL-1 β and Inhibition of PGE2. *Investig. Ophthalmol. Vis. Sci.* 2015, **56**, 7728–7738.

52. Shigemoto-Kuroda, T.; Oh, J.Y.; Kim, D.K.; Jeong, H.J.; Park, S.Y.; Lee, H.J.; Park, J.W.; Kim, T.W.; An, S.Y.; Prockop, D.J.; et al. MSC-derived Extracellular Vesicles Attenuate Immune Responses in Two Autoimmune Murine Models: Type 1 Diabetes and Uveoretinitis. *Stem Cell Rep.* 2017, **8**, 1214–1225.

53. Syed, F.Z. Type 1 Diabetes Mellitus. *Ann. Intern. Med.* 2022, **175**, ITC33–ITC48.

54. Warshauer, J.T.; Bluestone, J.A.; Anderson, M.S. New Frontiers in the Treatment of Type 1 Diabetes. *Cell Metab.* 2020, **31**, 46–61.

55. Brovkina, O.; Dashinimaev, E. Advances and complications of regenerative medicine in diabetes therapy. *PeerJ* 2020, **8**, e9746.

56. Roep, B.O.; Thomaidou, S.; van Tienhoven, R.; Zaldumbide, A. Type 1 Diabetes Mellitus as a Disease of the β -Cell (do Not Blame the Immune System)? *Nat. Rev. Endocrinol.* 2021, **17**, 150–161.

57. Soltani, S.; Mansouri, K.; Emami Aleagha, M.S.; Moasefi, N.; Yavari, N.; Shakouri, S.K.; Notararigo, S.; Shojaeian, A.; Pociot, F.; Yarani, R. Extracellular Vesicle Therapy for Type 1 Diabetes. *Front. Immunol.* 2022, 13, 865782.

58. Sun, Y.; Mao, Q.; Shen, C.; Wang, C.; Jia, W. Exosomes From β -Cells Alleviated Hyperglycemia and Enhanced Angiogenesis in Islets of Streptozotocin-Induced Diabetic Mice. *Diabetes Metab. Syndr. Obes.* 2019, 12, 2053–2064.

59. Nojehdehi, S.; Soudi, S.; Hesampour, A.; Rasouli, S.; Soleimani, M.; Hashemi, S.M. Immunomodulatory Effects of Mesenchymal Stem Cell-Derived Exosomes on Experimental Type-1 Autoimmune Diabetes. *J. Cell Biochem.* 2018, 119, 9433–9443.

60. Wen, D.; Peng, Y.; Liu, D.; Weizmann, Y.; Mahato, R.I. Mesenchymal Stem Cell and Derived Exosome as Small RNA Carrier and Immunomodulator to Improve Islet Transplantation. *J. Control Release* 2016, 238, 166–175.

61. Favaro, E.; Carpanetto, A.; Caorsi, C.; Giovarelli, M.; Angelini, C.; Cavallo-Perin, P.; Tetta, C.; Camussi, G.; Zanone, M.M. Human Mesenchymal Stem Cells and Derived Extracellular Vesicles Induce Regulatory Dendritic Cells in Type 1 Diabetic Patients. *Diabetologia* 2016, 59, 325–333.

62. Hu, W.; Song, X.; Yu, H.; Sun, J.; Zhao, Y. Differentiation of Monocytes into Phenotypically Distinct Macrophages After Treatment With Human Cord Blood Stem Cell (CB-SC)-Derived Exosomes. *J. Vis. Exp.* 2020, 165, e61562.

63. Peixoto, R.D.; Ferreira, A.R.; Cleary, J.M.; Fogacci, J.P.; Vasconcelos, J.P.; Jácome, A.A. Risk of Cancer in Inflammatory Bowel Disease and Pitfalls in Oncologic Therapy. *J. Gastrointest. Cancer* 2022, 1–11, online ahead of print.

64. Buscaill, E.; Le Cosquer, G.; Gross, F.; Lebrin, M.; Bugarel, L.; Deraison, C.; Vergnolle, N.; Bournet, B.; Gilletta, C.; Buscaill, L. Adipose-Derived Stem Cells in the Treatment of Perianal Fistulas in Crohn's Disease: Rationale, Clinical Results and Perspectives. *Int. J. Mol. Sci.* 2021, 22, 9967.

65. An, J.H.; Li, Q.; Bhang, D.H.; Song, W.J.; Youn, H.Y. TNF- α and INF- γ primed canine stem cell-derived extracellular vesicles alleviate experimental murine colitis. *Sci. Rep.* 2020, 10, 2115.

66. Liu, H.; Liang, Z.; Wang, F.; Zhou, C.; Zheng, X.; Hu, T.; He, X.; Wu, X.; Lan, P. Exosomes from mesenchymal stromal cells reduce murine colonic inflammation via a macrophage-dependent mechanism. *JCI Insight* 2019, 4, e131273.

67. Duan, L.; Huang, H.; Zhao, X.; Zhou, M.; Chen, S.; Wang, C.; Han, Z.; Han, Z.C.; Guo, Z.; Li, Z.; et al. Extracellular vesicles derived from human placental mesenchymal stem cells alleviate experimental colitis in mice by inhibiting inflammation and oxidative stress. *Int. J. Mol. Med.* 2020, 46, 1551–1561.

68. Cao, L.; Xu, H.; Wang, G.; Liu, M.; Tian, D.; Yuan, Z. Extracellular vesicles derived from bone marrow mesenchymal stem cells attenuate dextran sodium sulfate-induced ulcerative colitis by promoting M2 macrophage polarization. *Int. Immunopharmacol.* 2019, 72, 264–274.

69. Legaki, E.; Roubelakis, M.G.; Theodoropoulos, G.E.; Lazaris, A.; Kollia, A.; Karamanolis, G.; Marinos, E.; Gazouli, M. Therapeutic Potential of Secreted Molecules Derived from Human Amniotic Fluid Mesenchymal Stem/Stroma Cells in a Mice Model of Colitis. *Stem Cell Rev. Rep.* 2016, 12, 604–612.

70. Anderson, P.; Souza-Moreira, L.; Morell, M.; Caro, M.; O’Valle, F.; Gonzalez-Rey, E.; Delgado, M. Adipose-derived mesenchymal stromal cells induce immunomodulatory macrophages which protect from experimental colitis and sepsis. *Gut* 2013, 62, 1131–1141.

71. Sobkowiak-Sobierajska, A.; Lindemans, C.; Sykora, T.; Wachowiak, J.; Dalle, J.H.; Bonig, H.; Gennery, A.; Lawitschka, A. Management of Chronic Graft-vs.-Host Disease in Children and Adolescents With ALL: Present Status and Model for a Personalised Management Plan. *Front. Pediatr.* 2022, 10, 808103.

72. Le Blanc, K.; Rasmusson, I.; Sundberg, B.; Götherström, C.; Hassan, M.; Uzunel, M.; Ringdén, O. Treatment of severe acute graft-versus-host disease with third party haploidentical mesenchymal stem cells. *Lancet* 2004, 363, 1439–1441.

73. Ferrara, J.L.M.; Chaudhry, M.S. GVHD: Biology matters. *Hematology Am. Soc. Hematol. Educ. Program.* 2018, 2018, 221–227.

74. Zhang, L.; Chu, J.; Yu, J.; Wei, W. Cellular and molecular mechanisms in graft-versus-host disease. *J. Leukoc. Biol.* 2016, 99, 279–287.

75. Gupta, M.; Tieu, A.; Slobodian, M.; Shorr, R.; Burger, D.; Lalu, M.M.; Allan, D.S. Preclinical Studies of MSC-Derived Extracellular Vesicles to Treat or Prevent Graft Versus Host Disease: A Systematic Review of the Literature. *Stem Cell Rev. Rep.* 2021, 17, 332–340.

76. Wang, L.; Gu, Z.; Zhao, X.; Yang, N.; Wang, F.; Deng, A.; Zhao, S.; Luo, L.; Wei, H.; Guan, L.; et al. Extracellular Vesicles Released from Human Umbilical Cord-Derived Mesenchymal Stromal Cells Prevent Life-Threatening Acute Graft-Versus-Host Disease in a Mouse Model of Allogeneic Hematopoietic Stem Cell Transplantation. *Stem Cells Dev.* 2016, 25, 1874–1883.

77. Dal Collo, G.; Adamo, A.; Gatti, A.; Tamellini, E.; Bazzoni, R.; Takam Kamga, P.; Tecchio, C.; Quaglia, F.M.; Krampera, M. Functional dosing of mesenchymal stromal cell-derived extracellular vesicles for the prevention of acute graft-versus-host-disease. *Stem Cells*. 2020, 38, 698–711.

78. Branisteanu, D.E.; Cojocaru, C.; Diaconu, R.; Porumb, E.A.; Alexa, A.I.; Nicolescu, A.C.; Brihan, I.; Bogdanici, C.M.; Branisteanu, G.; Dimitriu, A.; et al. Update on the etiopathogenesis of psoriasis (Review). *Exp. Ther. Med.* 2022, 23, 201.

79. Nickoloff, B.J. Animal models of psoriasis. *Expert Opin. Investig. Drugs.* 1999, 8, 393–401.

80. Sarycheva, M.V.; Nadezhina, N.A.; Nadezhdin, S.V.; Bondarev, V.P.; Mycic, A.V.; Burda, Y.E.; Pokrovskiy, M.V.; Danilenko, L.M.; Peresypkina, A.A. Effect of Multipotent Mesenchymal Stromal Cells Secretome on Imiquimod-Induced Psoriasis in Rats. *J. Int. Pharm. Res.* 2019, 46, 296–301.

81. Zhang, Y.; Yan, J.; Li, Z.; Zheng, J.; Sun, Q. Exosomes Derived from Human Umbilical Cord Mesenchymal Stem Cells Alleviate Psoriasis-like Skin Inflammation. *J. Interferon Cytokine Res.* 2022, 42, 8–18.

82. Seetharaman, R.; Mahmood, A.; Kshatriya, P.; Patel, D.; Srivastava, A. Mesenchymal Stem Cell Conditioned Media Ameliorate Psoriasis Vulgaris: A Case Study. *Case Rep. Dermatol. Med.* 2019, 2019, 8309103.

83. Chovatiya, R.; Silverberg, J.I. The Heterogeneity of Atopic Dermatitis. *J. Drugs Dermatol.* 2022, 21, 172–176.

84. Cho, B.S.; Kim, J.O.; Ha, D.H.; Yi, Y.W. Exosomes derived from human adipose tissue-derived mesenchymal stem cells alleviate atopic dermatitis. *Stem Cell Res. Ther.* 2018, 9, 187.

85. Wang, W.M.; Wu, C.; Jin, H.Z. Exosomes in chronic inflammatory skin diseases and skin tumors. *Exp. Dermatol.* 2019, 28, 213–218.

86. Rodriguez Del Rio, P.; Liu, A.H.; Borres, M.P.; Södergren, E.; Iachetti, F.; Casale, T.B. Asthma and Allergy: Unravelling a Tangled Relationship with a Focus on New Biomarkers and Treatment. *Int. J. Mol. Sci.* 2022, 23, 3881.

87. de Castro, L.L.; Xisto, D.G.; Kitoko, J.Z.; Cruz, F.F.; Olsen, P.C.; Redondo, P.A.G.; Ferreira, T.P.T.; Weiss, D.J.; Martins, M.A.; Morales, M.M.; et al. Human adipose tissue mesenchymal stromal cells and their extracellular vesicles act differentially on lung mechanics and inflammation in experimental allergic asthma. *Stem Cell Res. Ther.* 2017, 8, 151.

88. Dong, L.; Wang, Y.; Zheng, T.; Pu, Y.; Ma, Y.; Qi, X.; Zhang, W.; Xue, F.; Shan, Z.; Liu, J.; et al. Hypoxic hUCMSC-derived extracellular vesicles attenuate allergic airway inflammation and airway remodeling in chronic asthma mice. *Stem Cell Res. Ther.* 2021, 12, 4.

89. Mun, S.J.; Kang, S.A.; Park, H.K.; Yu, H.S.; Cho, K.S.; Roh, H.J. Intranasally Administered Extracellular Vesicles from Adipose Stem Cells Have Immunomodulatory Effects in a Mouse Model of Asthma. *Stem Cells Int.* 2021, 2021, 6686625.

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