

Sarcopenia and Approaches

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Sarcopenia, an age-related decline in skeletal muscle mass and function, dramatically affects the quality of life. Although there is a consensus that sarcopenia is a multifactorial syndrome, the etiology and underlying mechanisms are not yet delineated. Moreover, research about nutritional interventions to prevent the development of sarcopenia is mainly focused on the amount and quality of protein intake. The impact of several nutrition strategies that consider timing of food intake, anti-inflammatory nutrients, metabolic control, and the role of mitochondrial function on the progression of sarcopenia is not fully understood. This narrative review summarizes the metabolic background of this phenomenon and proposes an integral nutritional approach (including dietary supplements such as creatine monohydrate) to target potential molecular pathways that may affect reduce or ameliorate the adverse effects of sarcopenia. Lastly, miRNAs, in particular those produced by skeletal muscle (MyomiR), might represent a valid tool to evaluate sarcopenia progression as a potential rapid and early biomarker for diagnosis and characterization.

mitochondria

aging

protein

muscle

1. Introduction

According to the World Health Organization (WHO), life expectancy has increased by 5 years since 2000 in developed countries ^[1]. This increase in life expectancy leads to aging and, in turn, to physical and cognitive decline. Current research shows that physical exercise can be a decisive protective factor for both functional decline and negative body composition changes during aging ^{[2][3]}. Resistance and endurance exercise have been shown to contribute to the improvement of cognition ^[4], as well as related psychological and social factors ^[5]. However, nutrition and eating strategies also play an essential role in preventing and treating functional limitations in the elderly population. For example, muscle mass declines by approximately 3–8% for each decade after 30 years, and this percentage increases significantly in people older than 60 years ^[6].

Sarcopenia is a condition characterized by a progressive and generalized loss of skeletal muscle mass and function with an increased risk of adverse outcomes such as disability, metabolic dysfunction, poor quality of life, and death ^[7]. Even though sarcopenia is mainly associated with the aging process seen in the elderly, there are several other populations at risk due to lifestyle decisions or pathological states. These include sedentary, immobilization, malnutrition, diabetes, obesity, and other acute or chronic inflammatory diseases that could promote the loss of muscle mass ^[8]. Moreover, the loss of muscle mass could also negatively affect the outcomes of those conditions ^{[9][10]}. In the last decade, three different consensus papers were published, giving both a definition of sarcopenia and diagnostic criteria. According to the European Working Group on Sarcopenia in Older

People (EWGSOP), the presence of low skeletal muscle mass (e.g., DXA, Anthropometry) and either low muscle strength (e.g., handgrip, isokinetic) or low muscle performance (e.g., walking speed, muscle power), which is often related to the most advanced stages of sarcopenia, are the criteria for the diagnosis of sarcopenia [9]. For both the European Society for Clinical Nutrition and Metabolism Special Interest Groups (ESPEN-SIG) and the International Working Group on Sarcopenia (IWGS), only the loss of muscle concomitant with a loss of muscle strength (which could also be assessed by walking speed) represents the recommended parameters. The IWGS also highlights that the loss of muscle mass could be alone or in conjunction with an increase in fat mass [10][11].

Thus, the diagnosis of sarcopenia can then be carried out by assessing the following parameters in the elderly (>65 years): (i) if walking speed is below 0.8 m/s at the 4 m walking test, and there is a low amount of muscle mass (i.e., a percentage of muscle mass divided by height squared is below two standard deviations of the normal young mean (<7.23 kg/m² in men and <5.67 kg/m² in women) as defined using dual-energy X-ray absorptiometry [12], or (ii) if the walking speed at the 4 m walking test is higher than 0.8 m/s, the hand-grip strength should be tested. If this last value is lower than 20 kg in women and 30 kg in men, the muscle mass must be analyzed [12].

While sarcopenia is mainly observed in the elderly, it can also develop in young adults [10]. A clear cause of sarcopenia cannot always be identified. Thus, the category of primary and secondary sarcopenia may be useful for choosing the best strategy to delay the progression. Primary sarcopenia is considered when there is no other evident cause rather than the aging process itself, while sarcopenia can be considered secondary when one or more of the following causes are evident [10]: (i) activity-related causes (bed rest, sedentary lifestyle, reconditioning, or even zero-gravity conditions seen in astronauts); (ii) nutrition-related causes, as a result of an inadequate dietary intake either energy or protein, malabsorption of nutrients or gastrointestinal disorders, and anorexigenic medication; (iii) disease-related causes, where there is a cross-talk between muscle mass and other organs that could lead to sarcopenia, such as inflammation, endocrine diseases, advanced organ failure, and malignancy. In this sense, sarcopenia is linked to the prognostic state of some pathologies, e.g., in those that affect the intestinal function and, as aforementioned, the absorption of nutrients as reported in abdominal hernias [13] and inflammatory bowel disease (IBD) [14]. Furthermore, sarcopenia has been recently related to the prognosis of various tumors [15][16][17][18], which leads to a monitoring status at the initial stage, as well as during and after chemotherapy.

2. The Importance of MicroRNAs

Low-grade inflammation is defined as an inflammatory state that cannot be determined by classical standards such as CRP or where an increase in IL-6 and TNF α is sometimes noted. It is established in sarcopenia [19], as well as in various pathological states such as obesity [20], cancer [21], polycystic ovary syndrome [22], and osteoarthritis [23]. In addition to the afflicted tissue, the latent and chronic inflammation state makes the whole organism less efficient by triggering chain reactions through crosstalk between tissues. For instance, Wang et al. [24] proposed that, during insulin resistance (associated with obesity), there is a phenomenon of latent inflammation that aggravates the extent and progression of sarcopenia. It is reasonable to think that an inflammatory state is also present in sarcopenia; therefore, if it is a secondary state or there are chronic comorbidities, there will be a further contribution to the inflammatory state. Hence, mitigating inflammation could be beneficial in the management of sarcopenia.

Kwang Byun et al. [25] correlated systemic inflammation present in chronic obstructive pulmonary disease (COPD) with sarcopenia. Similarly, Dalle et al. [26] demonstrated that three apparently different pathological states (i.e., type II diabetes, osteoarthritis, and COPD) are associated with an inflammatory state and lead to sarcopenia. Interestingly, a latent inflammatory state such as IBD also correlates to a greater onset of sarcopenia [14][27]. This status of low-grade inflammation has been a constant focus of research in order to identify new potential biomarkers.

MicroRNAs (miRNAs) are a unique class of short endogenous nucleotides sequences (around 15–30 bases). They are single-stranded noncoding RNAs capable of modulating gene expression by binding to the complementary regions of the 3'UTR sequence of specific mRNA targets, resulting in the inhibition of protein synthesis (translation) and/or mRNA degradation. This peculiar regulatory capability makes them crucial for normal development in all living beings [28][29]. miRNAs are present in all tissues and body fluids [30][31]. One important characteristic of the skeletal muscle is a group of miRNAs, identified as myomiRs [32][33][34], which seem to have a central role in the regulation of skeletal muscle plasticity by coordinating changes in fiber type and muscle mass in response to different contractile activity. Like every tissue, skeletal muscle also expresses miRNAs. In particular, the pool of these molecules is defined as myomiRs and is related to the differentiation of satellite cells, the maintenance of physiological trophism, the switch between fibers, and the development and conservation of muscle mass in response to physical exercise [32]. For example, Soares and colleagues [33] demonstrated an important regulatory action of a group of miRNAs on the progression of muscle atrophy. Brown et al. [35] showed that a pool of miRNAs (miR-23a, miR-182, miR-486, miR-206, miR-21, miR-27, and miR-128) are strong regulators of muscle size via the FOXO1 pathway, PTEN genes and translational regulation, and myostatin signaling. MyomiRs are secreted via exosomal vesicles, circulate in the bloodstream, and serve as regulators/communicators in proximal muscle tissue and even fat cells [33][34].

As aforementioned, sarcopenia is certainly the result of several factors, and its etiopathogenesis is still not well identified. For this reason, the identification of miRNAs might contribute to better understand this phenomenon, although the description of the myomiR profile is in its infancy. This group of miRNAs is potentially involved in the regulation of the satellite cell differentiation, the general proteostasis, the structure and type of muscle fibers, mitochondria and oxidative stress metabolism, the neurodegeneration process, and the infiltration of adipocytes into skeletal muscle tissue [34][36].

2.1. Satellite Cell Regulation

The differentiation of satellite cells is a fundamental process for the maintenance of muscle trophism. In this sense, certain miRNAs (miR-1, miR-206, and miR-486) have been identified to regulate cell survival and proliferation [37][38]. miRNAs have been shown to downregulate the MyoD and paired-box transcription factor (e.g., Pax3) pathways, which result in an inhibition of apoptosis, thereby increasing or maintaining muscle mass. These results have been seen in preclinical research; hence, it is highly plausible they have similar action mechanisms in humans (considering well-conserved metabolic signatures) [39]. It is noteworthy that both endurance and resistance training impact myomiRs, particularly those involved in skeletal muscle allostasis. For example, certain miRNAs regulate

the expression of growth factors (miR-29, miR-422-5p, and miR-143-3p), cell-cycle regulation (Let-7b and Let-7e), and myocyte differentiation (miR-139, miR-155, miR-501-3p, and miR-29) [\[40\]](#).

2.2. Proteostasis

At the moment, there are not many studies in humans. For example, Connors et al. [\[41\]](#) reported an increase in miR-424-5p during a decrease in protein synthesis at the skeletal muscle level with a consequent loss of muscle mass. It is well known that the control of protein metabolism is mediated by several miRNAs: miR-199, miR-125b, and miR-195 regulate hormones such as insulin and IGF-1; miR-432, miR-675-3p, miR-26a, miR-29, and miR-199-3p regulate signal transduction within the muscle cell; miR-27 and miR-128 serve as myostatin regulators; miR-129c, miR-23c, miR-27a, and miR-35 regulate protein catabolism [\[36\]](#).

2.3. Size and Type of Muscle Fiber

Some miRNAs seem to play a decisive role in the structure of muscle fibers. In particular, miR-23a and miR-182 regulate what are called iatrogenic genes (i.e., those who oversee the muscle atrophy program), as they appear to be able to restore drug-induced atrophy. Similarly, miR-21 and miR-206 are capable of acting on the regulation of atrophy. On the other hand, miR-27a seems to downregulate myostatin, thus favoring muscle turnover in a positive balance [\[34\]](#)[\[42\]](#)[\[43\]](#).

3. Counteracting Strategies

3.1. Physical Exercise with Emphasis on Resistance Training

The fundamental role of physical exercise in countering the progression of sarcopenia, associated or not with obesity, is now evident. Clinical research has confirmed the effectiveness of physical exercise, both cardiovascular and resistance training [\[44\]](#). As we described before, myomiRs are closely related to an optimal condition of muscle tissue and show an important role as signaling molecules that mediate physiological adaptations to exercise training. Furthermore, these miRNAs change differently in response to cardiovascular, resistance, or combined exercise (e.g., miR-1, miR-133a/b, miR-206, miR-499a-5p, and miR-486), but with no apparent difference in response between young and old men [\[45\]](#). There is strong evidence that strength training is one of, if not the most, effective interventional strategy to enhance muscle mass and strength in the elderly; thus, it can be used for treating, slowing, and/or preventing sarcopenia and dynapenia [\[46\]](#). Resistance training enhances physiological adaptations of the neuromuscular system, which positively affects the muscle strength. Maximal motor unit discharge rates increased 49% in older adults that followed only 6 weeks of a high-intensity progressive strength training program [\[47\]](#). Moreover, muscular factors independent of muscle mass, such as fascicle length and tendon stiffness, have also been observed to improve (10% and 64%, respectively) following resistance training in older adults [\[48\]](#). Moreover, resistance training is also a powerful stimulus for muscle protein synthesis, which leads to an increase in muscle mass. In this sense, an increase in the cross-sectional area of the thigh muscle (+4.6%) has been reported in mobility-limited older adults after 24 weeks of a resistance training program in conjunction with

protein supplementation [49]. Thus, there is a strong consensus in this regard. A review of 121 trials including over 6700 participants concluded that *'progressive resistance training is an effective intervention for improving physical functioning in older people, including improving strength and the performance of some simple and complex activities'* [50]. The authors reported a large positive effect on muscle mass, strength, and functionality. Additionally, high-intensity resistance training is associated with greater benefits in muscle strength with an average improvement of 5.3% after each incremental in exercise intensity from low intensity (<60% 1-RM), to low/moderate intensity (60–69% 1-RM), moderate/high intensity (70–79% 1-RM), and high intensity ($\geq 80\%$ 1-RM) [47]. High-intensity resistance training has been reported to be well tolerated in older adults, particularly when a proper progression is applied [51][52], although intensities between 65% and 75% 1-RM can be sufficient to promote significant adaptations [53]. Higher resistance training volumes are associated with greater improvements in lean body mass after controlling for a variety of confounders (e.g., age, study duration, sex, and training intensity and frequency) [54]. With regard to strength training frequency, 2–4 days per week are commonly recommended with training typically being performed on alternating days (e.g., Monday, Wednesday, and Friday) [53]. A well-prescribed resistance training program should also include exercises targeting all major muscle groups, but emphasis on lower limbs is recommended. Significant improvements in muscle strength and size have been reported in training programs that include 1–3 sets per exercise [55] with an adjustment of the numbers of repetitions that considers the maximum number that can be performed with a given intensity (max effort) [56]. Even in delicate conditions such as osteoarthritis or spondylarthritis, strength training can give excellent results [57][58]. Lastly, in order to continually reach improvements in mass, strength, and functional capacity, it is key to consistently incorporate progression and variation into the program. Every training variable can be adjusted over time considering the training experience of the subject and the adaptation rate on a case-by-case basis [53][59]. It is worth mentioning that it is not always possible to practice physical exercise, particularly strength training, for bedridden subjects and/or long-term patients due to chronic diseases such as cirrhosis, COPD, or severe renal insufficiency up to dialysis.

3.2. Nutrition and Supplementation

Maintenance of energy balance is crucial during a period of muscle disuse, but simply overfeeding does not further attenuate muscle atrophy since this merely increases adipose tissue. The key factor behind an accelerated loss of muscle tissue during a period of reduced food intake may not be the lower energy intake per se but more specifically the reduction in protein intake [60]. Barbera et al. [61] suggested some types of nutrients that would be able to influence the expression of myomiRs, regardless of the physical activity. Importantly, the intake of essential amino acids could positively impact miR-1, miR23a, miR208b, miR-499, and miR-27a, which have a positive effect on myocyte regeneration, proliferation, and differentiation [62]. In addition, resveratrol could regulate the differentiation of muscle cells and the activation of the PGC-1 α through the positive modulation of miR-21 and miR27b and the downregulation of miR-133b, miR30b, and miR-149. Other nutrients have also been found to modulate these molecules such as albumin, palmitic acid, vitamin D, and fructose [63]. PGC-1 α is a critical cofactor for mitochondrial biogenesis that it is mainly activated by the AMP-activated protein kinase (AMPK) pathway. AMPK is one of the main energy sensors (perturbations in ATP/ADP ratio) that regulate energy metabolism (e.g., protein synthesis, as an energy cost process) [64]. High levels of PGC-1 α are associated with muscle mass sparing during sarcopenia, possibly by means of a reduction in the protein breakdown via FOXO inhibition (with no changes in

protein synthesis) [65]. It has been shown that FOXO induces the expression of atrogin-1 and MuRF1 under conditions of energy stress in myofibers, but activation of PGC-1 α could attenuate the negative regulation of proteostasis [65][66].

Starvation and aggressive hypocaloric diets have been reported as deleterious to the muscle mass and function, especially when the protein needs are not achieved [67][68]. This might be due to inhibition of the mTORC1 pathway, as demonstrated after some weeks of low-carbohydrate high-fat (LCHF) diets [69]. Even though extreme nutrient and energy deprivation induces autophagy, a mild carbohydrate restriction may result in a favorable impact on sarcopenia outcomes [65][70][71]. In fact, caloric restriction might confer lifespan and health benefits and, therefore, it is not surprising that intermittent and periodized caloric restrictions (e.g., alternate-day fasting or intermittent fasting) might be suitable as a counteracting strategy for sarcopenia [72][73]. Thus, certain biological elements might prevent the excessive activation of UPS via negative regulation of pro-atrophy transcription factors without modifying the translational process. From a nutritional standpoint, adequate protein intake and certain antioxidants (e.g., secondary metabolites) could modulate the muscle protein synthesis and breakdown. The subsections below summarize relevant findings in this regard.

3.2.1. High-Protein Diet

Older people have a diminished myofibrillar protein synthesis response to protein intake, which may have a strong influence on the progression of sarcopenia, and it is exacerbated in elderly population with obesity. This age-related muscle 'anabolic resistance' is more evident in response to low or moderate protein intake which is common in the diet of older individuals [74]. In addition, the current recommended dietary protein intake of 0.8 g/kg/day might not be sufficient for preserving muscle mass and quality on a long-term basis [75][76][77]. In a recent review and meta-analysis, carried out on older subjects with overweight and obesity, it was concluded that protein intakes ≥ 1.0 g/kg/day have a greater protective effect on the loss of lean tissue than lower intakes. It is worth mentioning that subjects were 50 years old, considering that, as age advances, the amount of protein intake may become more essential [78][79]. Moreover, feeding is a critical modulator of the inner biological clock; therefore, both the timing and the type of food can be important [80][81]. Erratic eating patterns can disrupt the temporal coordination of metabolism and physiology, which is associated with chronic diseases that are also characteristic of aging such as sarcopenia [82][83]. Therefore, timing is crucial to regulate autophagy and mitophagy in more metabolically sensitive populations such as older adults.

References

1. Available online: <https://www.who.int/data/gho/data/themes/topics/indicator-groups/indicator-group-details/GHO/life-expectancy-and-healthy-life-expectancy> (accessed on 24 May 2021).
2. Hunter, G.R.; McCarthy, J.P.; Bamman, M.M. Effects of Resistance Training on Older Adults. *Sports Med.* 2004, 34, 329–348.

3. Latham, N.K.; Bennett, D.A.; Stretton, C.M.; Anderson, C.S. Systematic Review of Progressive Resistance Strength Training in Older Adults. *J. Gerontol. Ser. A Biol. Sci. Med. Sci.* 2004, 59, M48–M61.
4. Cassilhas, R.C.; Tufik, S.; de Mello, M.T. Physical exercise, neuroplasticity, spatial learning and memory. *Cell Mol. Life Sci.* 2016, 73, 975–983.
5. Marcos-Pardo, P.J.; Martínez-Rodríguez, A.; Gil-Arias, A. Impact of a motivational resistance-training programme on adherence and body composition in the elderly. *Sci. Rep.* 2018, 8, 1370.
6. Burgos Peláez, R. Enfoque terapéutico global de la sarcopenia. *J. Nutr. Hosp.* 2006, 21, 51–60.
7. Rosenberg, I.H. Sarcopenia: Origins and Clinical Relevance. *Clin. Geriatr. Med.* 2011, 27, 337–339.
8. Biolo, G.; Fleming, R.Y.D.; Wolfe, R.R. Physiologic hyperinsulinemia stimulates protein synthesis and enhances transport of selected amino acids in human skeletal muscle. *J. Clin. Investig.* 1995, 95, 811–819.
9. Cruz-Jentoft, A.J.; Baeyens, J.P.; Bauer, J.M.; Boirie, Y.; Cederholm, T.; Landi, F.; Martin, F.C.; Michel, J.-P.; Rolland, Y.; Schneider, S.M. Sarcopenia: European consensus on definition and diagnosis: Report of the European Working Group on Sarcopenia in Older People. *Age Aging* 2010, 39, 412–423.
10. Muscaritoli, M.; Anker, S.; Argiles, J.M.; Aversa, Z.; Bauer, J.; Biolo, G.; Boirie, Y.; Bosaeus, I.; Cederholm, T.; Costelli, P.; et al. Consensus definition of sarcopenia, cachexia and pre-cachexia: Joint document elaborated by Special Interest Groups (SIG) “cachexia-anorexia in chronic wasting diseases” and “nutrition in geriatrics”. *Clin. Nutr.* 2010, 29, 154–159.
11. Fielding, R.A.; Vellas, B.; Evans, W.J.; Bhasin, S.; Morley, J.E.; Newman, A.B.; van Kan, G.A.; Andrieu, S.; Bauer, J.; Breuille, D.; et al. Sarcopenia: An Undiagnosed Condition in Older Adults. Current Consensus Definition: Prevalence, Etiology, and Consequences. International Working Group on Sarcopenia. *J. Am. Med. Dir. Assoc.* 2011, 12, 249–256.
12. Santilli, V.; Bernetti, A.; Mangone, M.; Paoloni, M. Clinical definition of sarcopenia. *Clin. Cases Miner. Bone Metab.* 2014, 11, 177–180.
13. Clark, S.T.; Malietzis, G.; Grove, T.N.; Jenkins, J.T.; Windsor, A.C.J.; Kontovounisios, C.; Warren, O. The emerging role of sarcopenia as a prognostic indicator in patients undergoing abdominal wall hernia repairs: A systematic review of the literature. *Hernia* 2020, 24, 1361–1370.
14. De Andrade, M.I.S.; Maio, R.; Dourado, K.F.; de Macêdo, P.F.C.; Neto, A.C.B. Excessive Weight—Muscle Depletion Paradox And Cardiovascular Risk Factors In Outpatients With Inflammatory Bowel Disease. *Arq. Gastroenterol.* 2015, 52, 37–45.

15. Shachar, S.S.; Williams, G.; Muss, H.B.; Nishijima, T.F. Prognostic value of sarcopenia in adults with solid tumours: A meta-analysis and systematic review. *Eur. J. Cancer* 2016, 57, 58–67.
16. Ubachs, J.; Ziemons, J.; Minis-Rutten, I.J.; Kruitwagen, R.F.; Kleijnen, J.; Lambrechts, S.; Damink, S.O.; Rensen, S.S.; Van Gorp, T. Sarcopenia and ovarian cancer survival: A systematic review and meta-analysis. *J. Cachex-Sarcopenia Muscle* 2019, 10, 1165–1174.
17. Strassmann, D.; Hensen, B.; Grünwald, V.; Stange, K.; Eggers, H.; Länger, F.; Omar, M.; Zardo, P.; Christiansen, H.; Reuter, C.W.; et al. Impact of sarcopenia in advanced and metastatic soft tissue sarcoma. *Int. J. Clin. Oncol.* 2021, 1–10.
18. Tieland, M.; Trouwborst, I.; Clark, B.C. Skeletal muscle performance and ageing. *J. Cachex-Sarcopenia Muscle* 2017, 9, 3–19.
19. Villarroja, F.; Cereijo, R.; Gavalda-Navarro, A.; Villarroja, J.; Giralt, M. Inflammation of brown/beige adipose tissues in obesity and metabolic disease. *J. Intern. Med.* 2018, 284, 492–504.
20. Iyengar, N.M.; Gucalp, A.; Dannenberg, A.J.; Hudis, C.A. Obesity and Cancer Mechanisms: Tumor Microenvironment and Inflammation. *J. Clin. Oncol.* 2016, 34, 4270–4276.
21. Ojeda-Ojeda, M.; Murri, M.; Insenser, M.; Escobar-Morreale, H. Mediators of Low-Grade Chronic Inflammation in Polycystic Ovary Syndrome (PCOS). *Curr. Pharm. Des.* 2013, 19, 5775–5791.
22. Scanzello, C.R. Role of low-grade inflammation in osteoarthritis. *Curr. Opin. Rheumatol.* 2017, 29, 79–85.
23. Beyer, I.; Mets, T.; Bautmans, I. Chronic low-grade inflammation and age-related sarcopenia. *Curr. Opin. Clin. Nutr. Metab. Care* 2012, 15, 12–22.
24. Wang, M.; Tan, Y.; Shi, Y.; Wang, X.; Liao, Z.; Wei, P. Diabetes and Sarcopenic Obesity: Pathogenesis, Diagnosis, and Treatments. *Front. Endocrinol.* 2020, 11, 568.
25. Byun, M.K.; Na Cho, E.; Chang, J.; Ahn, C.M.; Kim, H.J. Sarcopenia correlates with systemic inflammation in COPD. *Int. J. Chronic Obstr. Pulm. Dis.* 2017, 12, 669–675.
26. Dalle, S.; Koppo, K. Is inflammatory signaling involved in disease-related muscle wasting? Evidence from osteoarthritis, chronic obstructive pulmonary disease and type II diabetes. *Exp. Gerontol.* 2020, 137, 110964.
27. Dhaliwal, A.; Quinlan, J.; Overthrow, K.; Greig, C.; Lord, J.; Armstrong, M.; Cooper, S. Sarcopenia in Inflammatory Bowel Disease: A Narrative Overview. *Nutrients* 2021, 13, 656.
28. Zhang, W.; Peng, P.; Shen, K. Role of Exosome Shuttle RNA in Cell-to-Cell Communication. *Zhongguo Yi Xue Ke Xue Yuan Xue Bao* 2016, 38, 480–483.

29. Moran, Y.; Agron, M.; Praher, D.; Technau, U. The evolutionary origin of plant and animal microRNAs. *Nat. Ecol. Evol.* 2017, 1, 0027.
30. McCall, M.N.; Kim, M.-S.; Adil, M.; Patil, A.H.; Lu, Y.; Mitchell, C.J.; Leal-Rojas, P.; Xu, J.; Kumar, M.; Dawson, V.L.; et al. Toward the human cellular microRNAome. *Genome Res.* 2017, 27, 1769–1781.
31. Kovanda, A.; Režen, T.; Rogelj, B. MicroRNA in skeletal muscle development, growth, atrophy, and disease. *Wiley Interdiscip. Rev. RNA* 2014, 5, 509–525.
32. McCarthy, J.J. The MyomiR Network in Skeletal Muscle Plasticity. *Exerc. Sport Sci. Rev.* 2011, 39, 150–154.
33. Soares, R.J.R.; Cagnin, S.; Chemello, F.; Silvestrin, M.; Musaro, A.; De Pitta, C.; Lanfranchi, G.; Sandri, M. Involvement of MicroRNAs in the Regulation of Muscle Wasting during Catabolic Conditions. *J. Biol. Chem.* 2014, 289, 21909–21925.
34. Mitchelson, K.R.; Qin, W.Y. Roles of the canonical myomiRs miR-1, -133 and -206 in cell development and disease. *World J Biol Chem.* 2015, 6, 162–208.
35. Mytidou, C.; Koutsoulidou, A.; Katsioloudi, A.; Prokopi, M.; Kapnisis, K.; Michailidou, K.; Anayiotos, A.; Phylactou, L.A. Muscle-derived exosomes encapsulate myomiRs and are involved in local skeletal muscle tissue communication. *FASEB J.* 2021, 35, e21279.
36. Brown, D.M.; Goljanek-Whysall, K. microRNAs: Modulators of the underlying pathophysiology of sarcopenia? *Ageing Res. Rev.* 2015, 24, 263–273.
37. Yin, J.; Qian, Z.; Chen, Y.; Li, Y.; Zhou, X. MicroRNA regulatory networks in the pathogenesis of sarcopenia. *J. Cell. Mol. Med.* 2020, 24, 4900–4912.
38. Chen, J.-F.; Tao, Y.; Li, J.; Deng, Z.; Yan, Z.; Xiao, X.; Wang, D.-Z. microRNA-1 and microRNA-206 regulate skeletal muscle satellite cell proliferation and differentiation by repressing Pax7. *J. Cell Biol.* 2010, 190, 867–879.
39. Dey, B.K.; Gagan, J.; Dutta, A. miR-206 and -486 Induce Myoblast Differentiation by Downregulating Pax7. *Mol. Cell. Biol.* 2011, 31, 203–214.
40. Hirai, H.; Verma, M.; Watanabe, S.; Tastad, C.; Asakura, Y.; Asakura, A. MyoD regulates apoptosis of myoblasts through microRNA-mediated down-regulation of Pax3. *J. Cell Biol.* 2010, 191, 347–365.
41. Fochi, S.; Giuriato, G.; De Simone, T.; Gomez-Lira, M.; Tamburin, S.; Del Piccolo, L.; Schena, F.; Venturelli, M.; Romanelli, M. Regulation of microRNAs in Satellite Cell Renewal, Muscle Function, Sarcopenia and the Role of Exercise. *Int. J. Mol. Sci.* 2020, 21, 6732.
42. Connolly, M.; Paul, R.; Garros, R.F.; Natanek, S.A.; Bloch, S.; Lee, J.; Lorenzo, J.P.; Patel, H.; Cooper, C.; Sayer, A.A.; et al. miR-424-5p reduces ribosomal RNA and protein synthesis in

- muscle wasting. *J. Cachex-Sarcopenia Muscle* 2017, 9, 400–416.
43. McFarlane, C.; Vajjala, A.; Arigela, H.; Lokireddy, S.; Ge, X.; Bonala, S.; Manickam, R.; Kambadur, R.; Sharma, M. Negative Auto-Regulation of Myostatin Expression is Mediated by Smad3 and MicroRNA-27. *PLoS ONE* 2014, 9, e87687.
 44. Gan, Z.; Fu, T.; Kelly, D.P.; Vega, R.B. Skeletal muscle mitochondrial remodeling in exercise and diseases. *Cell Res.* 2018, 28, 969–980.
 45. Trouwborst, I.; Verreijen, A.; Memelink, R.; Massanet, P.; Boirie, Y.; Weijs, P.; Tieland, M. Exercise and Nutrition Strategies to Counteract Sarcopenic Obesity. *Nutrients* 2018, 10, 605.
 46. Burton, L.A.; Sumukadas, D. Optimal management of sarcopenia. *Clin. Interv. Aging* 2010, 5, 217–228.
 47. Kamen, G.; Knight, C.A. Training-Related Adaptations in Motor Unit Discharge Rate in Young and Older Adults. *J. Gerontol. Ser. A Boil. Sci. Med. Sci.* 2004, 59, 1334–1338.
 48. Reeves, N.D.; Maganaris, C.N.; Narici, M.V. Effect of strength training on human patella tendon mechanical properties of older individuals. *J. Physiol.* 2003, 548, 971–981.
 49. Chale, A.; Cloutier, G.J.; Hau, C.; Phillips, E.M.; Dallal, G.E.; Fielding, R.A. Efficacy of Whey Protein Supplementation on Resistance Exercise-Induced Changes in Lean Mass, Muscle Strength, and Physical Function in Mobility-Limited Older Adults. *Journals Gerontol. Ser. A Boil. Sci. Med. Sci.* 2012, 68, 682–690.
 50. Liu, C.-J.; Latham, N.K. Progressive resistance strength training for improving physical function in older adults. *Cochrane Database Syst. Rev.* 2009, 2009, CD002759.
 51. Peterson, M.D.; Rhea, M.R.; Sen, A.; Gordon, P. Resistance exercise for muscular strength in older adults: A meta-analysis. *Ageing Res. Rev.* 2010, 9, 226–237.
 52. Singh, N.A.; Quine, S.; Clemson, L.M.; Williams, E.J.; Williamson, D.A.; Stavrinos, T.M.; Grady, J.N.; Perry, T.J.; Lloyd, B.D.; Smith, E.; et al. Effects of High-Intensity Progressive Resistance Training and Targeted Multidisciplinary Treatment of Frailty on Mortality and Nursing Home Admissions after Hip Fracture: A Randomized Controlled Trial. *J. Am. Med. Dir. Assoc.* 2012, 13, 24–30.
 53. Nelson, M.E.; Rejeski, W.J.; Blair, S.N.; Duncan, P.; Judge, J.O.; King, A.C.; Macera, C.A.; Castaneda-Sceppa, C. Physical activity and public health in older adults: Recommendation from the American College of Sports Medicine and the American Heart Association. *Med. Sci. Sports Exerc.* 2007, 39, 1435–1445.
 54. Peterson, M.D.; Sen, A.; Gordon, P. Influence of Resistance Exercise on Lean Body Mass in Aging Adults: A meta-analysis. *Med. Sci. Sports Exerc.* 2011, 43, 249–258.

55. Starkey, D.B.; Pollock, M.L.; Ishida, Y.; Welsch, M.A.; Brechue, W.F.; Graves, J.E.; Feigenbaum, M.S. Effect of resistance training volume on strength and muscle thickness. *Med. Sci. Sports Exerc.* 1996, 28, 1311–1320.
56. Law, T.D.; Clark, L.A.; Clark, B.C. Resistance Exercise to Prevent and Manage Sarcopenia and Dynapenia. *Annu. Rev. Gerontol. Geriatr.* 2016, 36, 205–228.
57. Cannataro, R.; Di Maio, L.; Malorgio, A.; Micheli, M.L.; Cione, E. Spondyloarthritis and Strength Training: A 4-Year Report. *J. Funct. Morphol. Kinesiol.* 2021, 6, 58.
58. Malorgio, A.; Malorgio, M.; Benedetti, M.; Casarosa, S.; Cannataro, R. High intensity resistance training as intervention method to knee osteoarthritis. *Sports Med. Health Sci.* 2021, 3, 46–48.
59. Kakehi, S.; Wakabayashi, H.; Inuma, H.; Inose, T.; Shioya, M.; Aoyama, Y.; Hara, T.; Uchimura, K.; Tomita, K.; Okamoto, M.; et al. Rehabilitation Nutrition and Exercise Therapy for Sarcopenia. *World J. Men's Health* 2021, 39, e13.
60. Wall, B.T.; van Loon, L.J. Nutritional strategies to attenuate muscle disuse atrophy. *Nutr. Rev.* 2013, 71, 195–208.
61. Barbiera, A.; Pelosi, L.; Sica, G.; Scicchitano, B.M. Nutrition and microRNAs: Novel Insights to Fight Sarcopenia. *Antioxidants* 2020, 9, 951.
62. Drummond, M.J.; Glynn, E.L.; Fry, C.S.; Dhanani, S.; Volpi, E.; Rasmussen, B.B. Essential Amino Acids Increase MicroRNA-499, -208b, and -23a and Downregulate Myostatin and Myocyte Enhancer Factor 2C mRNA Expression in Human Skeletal Muscle. *J. Nutr.* 2009, 139, 2279–2284.
63. Lançon, A.; Kaminski, J.; Tili, E.; Michaille, J.-J.; Latruffe, N. Control of MicroRNA Expression as a New Way for Resveratrol To Deliver Its Beneficial Effects. *J. Agric. Food Chem.* 2012, 60, 8783–8789.
64. Kjørbsted, R.; Hingst, J.R.; Fentz, J.; Foretz, M.; Sanz, M.; Pehmøller, C.; Shum, M.; Marette, A.; Mounier, R.; Treebak, J.T.; et al. AMPK in skeletal muscle function and metabolism. *FASEB J.* 2018, 32, 1741–1777.
65. Schiaffino, S.; Dyar, K.; Ciciliot, S.; Blaauw, B.; Sandri, M. Mechanisms regulating skeletal muscle growth and atrophy. *FEBS J.* 2013, 280, 4294–4314.
66. Canto, C.; Auwerx, J. PGC-1 α , SIRT1 and AMPK, an energy sensing network that controls energy expenditure. *Curr. Opin. Lipidol.* 2009, 20, 98–105.
67. Hector, A.J.; McGlory, C.; Damas, F.; Mazara, N.; Baker, S.K.; Phillips, S.M. Pronounced energy restriction with elevated protein intake results in no change in proteolysis and reductions in skeletal muscle protein synthesis that are mitigated by resistance exercise. *FASEB J.* 2017, 32, 265–275.

68. Murphy, C.H.; Churchward-Venne, T.A.; Mitchell, C.J.; Kolar, N.M.; Kassis, A.; Karagounis, L.G.; Burke, L.M.; Hawley, J.A.; Phillips, S.M. Hypoenergetic diet-induced reductions in myofibrillar protein synthesis are restored with resistance training and balanced daily protein ingestion in older men. *Am. J. Physiol. Metab.* 2015, 308, E734–E743.
69. Ferretti, R.; Moura, E.G.; Dos Santos, V.C.; Caldeira, E.J.; Conte, M.; Matsumura, C.Y.; Pertille, A.; Mosqueira, M. High-fat diet suppresses the positive effect of creatine supplementation on skeletal muscle function by reducing protein expression of IGF-PI3K-AKT-mTOR pathway. *PLoS ONE* 2018, 13, e0199728.
70. McDaniel, S.S.; Rensing, N.R.; Thio, L.L.; Yamada, K.A.; Wong, M. The ketogenic diet inhibits the mammalian target of rapamycin (mTOR) pathway. *Epilepsia* 2011, 52, e7–e11.
71. Paoli, A.; Bianco, A.; Damiani, E.; Bosco, G. Ketogenic Diet in Neuromuscular and Neurodegenerative Diseases. *BioMed Res. Int.* 2014, 2014, 474296.
72. Jiao, J.; Demontis, F. Skeletal muscle autophagy and its role in sarcopenia and organismal aging. *Curr. Opin. Pharmacol.* 2017, 34, 1–6.
73. Webster, B.R.; Scott, I.; Traba, J.; Han, K.; Sack, M.N. Regulation of autophagy and mitophagy by nutrient availability and acetylation. *Biochim. Biophys. Acta (BBA)-Mol. Cell Biol. Lipids* 2014, 1841, 525–534.
74. Smeuninx, B.; McKendry, J.; Wilson, D.; Martin, U.; Breen, L. Age-Related Anabolic Resistance of Myofibrillar Protein Synthesis Is Exacerbated in Obese Inactive Individuals. *J. Clin. Endocrinol. Metab.* 2017, 102, 3535–3545.
75. Beasley, J.M.; Shikany, J.M.; Thomson, C.A. The Role of Dietary Protein Intake in the Prevention of Sarcopenia of Aging. *Nutr. Clin. Pract.* 2013, 28, 684–690.
76. Courtney-Martin, G.; Ball, R.O.; Pencharz, P.B.; Elango, R. Protein Requirements during Aging. *Nutrients* 2016, 8, 492.
77. Phillips, S.; Chevalier, S.; Leidy, H.J. Protein “requirements” beyond the RDA: Implications for optimizing health. *Appl. Physiol. Nutr. Metab.* 2016, 41, 565–572.
78. Witard, O.C.; Wardle, S.L.; Macnaughton, L.S.; Hodgson, A.B.; Tipton, K.D. Protein Considerations for Optimising Skeletal Muscle Mass in Healthy Young and Older Adults. *Nutrients* 2016, 8, 181.
79. Kim, J.E.; O’Connor, L.E.; Sands, L.; Slebodnik, M.B.; Campbell, W.W. Effects of dietary protein intake on body composition changes after weight loss in older adults: A systematic review and meta-analysis. *Nutr. Rev.* 2016, 74, 210–224.
80. Potter, G.D.M.; Cade, J.; Grant, P.J.; Hardie, L.J. Nutrition and the circadian system. *Br. J. Nutr.* 2016, 116, 434–442.

81. Ribas-Latre, A.; Eckel-Mahan, K. Interdependence of nutrient metabolism and the circadian clock system: Importance for metabolic health. *Mol. Metab.* 2016, 5, 133–152.
 82. Albrecht, U. Timing to Perfection: The Biology of Central and Peripheral Circadian Clocks. *Neuron* 2012, 74, 246–260.
 83. Mohawk, J.A.; Green, C.B.; Takahashi, J.S. Central and Peripheral Circadian Clocks in Mammals. *Annu. Rev. Neurosci.* 2012, 35, 445–462.
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