GPCRs Regulate DNA Integrity

Subjects: Biochemistry & Molecular Biology

Contributor: Stuart Maudsley, Deborah Walter, Hanne Leysen, Romi Vandoren

G protein-coupled receptors (GPCRs) and their associated signaling proteins represent one of the most diverse cellular signaling systems involved in both physiological and pathophysiological processes. Aging represents perhaps the most complex biological process in humans and involves a progressive degradation of systemic integrity and physiological resilience. This is in part mediated by age-related aberrations in energy metabolism, mitochondrial function, protein folding and sorting, inflammatory activity, signal transduction fidelity and genomic stability. An increased rate of unrepaired DNA damage is considered to be one of the key 'hallmarks' of aging. Over the last two decades our appreciation of the complexity of GPCR signaling systems has expanded their functional signaling repertoire. One such example of this is the incipient role of GPCRs and GPCR-interacting proteins in DNA damage and repair (DDR) mechanisms. Emerging data now suggest that GPCRs could function as stress sensors for intracellular damage such as oxidative stress. Given this role of GPCRs in the DNA damage response process, coupled to the effective history of drug targeting of these receptors, this suggests that one important future activity of GPCR therapeutics is the rational control of DNA damage repair systems.

Keywords: G protein-coupled receptor (GPCR) ; aging ; DNA damage ; β-arrestin ; G protein-coupled receptor kinase (GRK) ; interactome ; G protein-coupled receptor kinase interacting protein 2 (GIT2) ; ataxia telangiectasia mutated (ATM) ; clock proteins ; energy metabolism

1. Introduction

With the knowledge gained about mechanisms underlying health and disease, as well as improved living standards and sanitization, there has been a major increase in the global average lifespan ^[1]. The World Health Organization reported in 2019 that an estimated 703 million people were aged 65 or older. This number is expected to double by 2050 (United Nations, Department of Economic and Social Affairs, Population Division (2019). World Population Ageing 2019: Highlights (ST/ESA/SER.A/430)). Despite this positive result of improved healthcare, a major complication incurred with this increase in the size of the worldwide elderly population is the burgeoning prevalence of aging-related diseases including neurodegenerative disorders, cardiovascular diseases and diabetes mellitus ^[2]. This has been borne out through multiple studies connecting age-related molecular pathologies and the incidence of these disorders ^{[3][4][5]}. These reports have suggested that the aging process itself is an underlying cause for multiple diseases; however, aging itself is not considered a disorder, but a normal physiological process ^[2]. Pathological aging can be defined as a progressive deterioration of physiological functions, which will eventually lead to systemic dysfunction and death ^[2]. These pathophysiological alterations include metabolic dysfunction, genome instability, telomere attrition and oxidative stress ^[6]. A greater understanding of these processes should improve our capacity to prevent or treat age-related diseases ^[8].

Transmembrane heptahelical GPCRs represent perhaps the most studied and effective drug targets to date. Their near ubiquitous role in physiological processes, coupled to their capacity to recognize a wide diversity of impinging molecules, makes them ideal targets for pharmacotherapeutic design [9|[10]]. As a testament to the functional efficacy of targeting GPCRs in disease, 475 drugs (~34% of all drugs approved by the FDA, acting on over 108 unique GPCR targets) are currently clinically employed [9]. While currently dominating the realm of therapeutics, there is still a strong impetus for future GPCR-based drug design. There are over 300 new experimental drugs that are currently in clinical trials, of which ~20% target 66 previously unexploited GPCR systems. The major disease indications for GPCR modulators have shown a trend towards diabetes, obesity and Alzheimer disease (AD), all of which are strongly age-dependent disorders. While the majority of the worldwide drug design effort has been made using the concept of exploiting and controlling the G protein-dependent signaling modality of GPCRs, there is now a growing field of more 'engineered efficacy' therapeutics that can utilize alternative modes of non-G protein-mediated GPCR signaling [11][12][13]. The emergence of these new and diverse GPCR signaling modes expands our concepts of the types of signaling systems that can be controlled through GPCR modulation. Here we will investigate one of these new target systems that may hold the key to the future treatment of multiple age-related disorders [141][151][16][17], *i.e.* the DNA damage-response (DDR) system.

As life proceeds through the individual's aging process, both endogenous (*e.g.* reactive oxygen species (ROS)) and environmental (*e.g.* ionizing radiation) stressors are constantly attacking DNA, causing structural damage [8]. Unrepaired DNA damage negatively affects genome replication and transcription, causing wide-scale chromosomal aberrations that disrupt critical cell functions such as energy metabolism and protein folding/management ^{[18][19][20]}. Given the importance of DNA-protective activity as an anti-aging strategy, coupled to the feasibility of GPCR druggability, the generation of GPCR-based DDR controlling agents holds considerable promise for improved treatments for both disorders of genomic aging such as Werner syndrome or Ataxia-Telangiectasia, as well as age-related disorders such as Metabolic Syndrome or Parkinson's disease.

2. G Protein-Coupled Receptor Systems: Intersections with aging and senescence

2.1. GPCR Signaling Diversity

The GPCR superfamily forms one of the largest and most diverse groups in the human proteome ^[21]. Different GPCRs interact with extremely divergent chemical compounds ranging from photons, neurotransmitters, small metabolites to complex hormones and exogenous animal toxins. Thus the GPCR superfamily encompasses a broad range of cellular sensory systems that allows tissues to sense and adjust to diverse endogenous and environmental perturbagens. GPCRs are unique in their structural and functional diversity and have been therapeutically exploited to effectively combat a plethora of diseases.

Even though GPCRs are most structurally distinct in the orthosteric ligand-binding site, they show the most conservation in the transducer site [22]. In other words, GPCRs interact with disparate extracellular ligands, but elicit a typical intracellular response observed across the GPCR superfamily. All GPCRs bind heterotrimeric G proteins consisting of aßy subunits ($G\alpha\beta\gamma$) and can induce a G protein signaling cascade. There are 16 G α , 5 G β and 13 Gy subunits in humans, which allows many different aggregations and signaling outputs [23]. The responses reach from activation to inhibition and are classified based on the downstream function of the α subunits into four families, G α s, G α i/o, G α q/11, and G α 12/13 ^[24]. Classical G protein signaling is induced through ligand stabilization of the heptahelical core to create an active tertiary structure and induces conformational changes in the intracellular loops and the carboxyl terminus. In a reverse manner, the binding of G proteins to receptors exerts an allosteric effect on the ligand binding region and increases the agonist binding affinity [25]. The ligand-activated receptor conformation promotes binding of the Gaßy and triggers the exchange of GDP for GTP on the a subunit. In essence GPCRs act as ligand-activated guanine nucleotide exchange factors. G protein nucleotide exchange facilitates the dissociation of the heterotrimeric G protein in its GTP-bound α subunit and the free βy dimer. Both subunit groupscan then activate intracellular downstream effectors, such as ion channels or enzymes, and thereby amplify the signal for cellular activities. The GTP-bound α subunit triggers its cognate effectors such as the cAMP catalyzing enzyme adenylyl cyclase, inositol 1,4,5-phosphate (IP3) or phospholipase C. The free β y dimer also shows modulatory activities on downstream effectors, e.g. promoting the interaction of G protein-coupled receptor kinase (GRK) with GPCRs which then phosphorylates the C-terminus or intracellular loops of the GPCR. The phosphorylation facilitates coupling to β -arrestin and directly affects GPCR function by desensitization ^[26]. The α subunit is inactivated by cleaving GTP to GDP and eventually recombines with the βy subunit and terminates its activity.

In addition to this G protein signaling mechanism data gathered in the last two decades points towards additional non-G protein signaling activities of GPCRs ^{[27][28][29][30][31][32]}. The discovery that these additional signaling pathways are therapeutically tractable, such as the β -arrestin mediated signaling cascades suggests that additional drug design avenues may be fruitful. Luttrell et. al demonstrated that β arrestin mediates SRC family kinase recruitment and is critical for the activation of extracellular signal-regulated kinase 1/2 (ERK1/2) pathways ^[11]. Apart from the primary association of β -arrestin with GPCR internalization and desensitization ^[33], they appear to form complexes with the inactivated receptor and other scaffolding proteins that evoke GPCR associated signals ^[27]. β -arrestins have been demonstrated to bind a wide variety of kinases, e.g., E3 ubiquitin ligases, phosphodiesterases and transcription factors ^[34]. More recently it has been shown that β -arrestin activation through β 2-adrenergic receptors (β 2AR) augments DNA damage, p53 degradation and apoptosis under chronic stress conditions ^[35]. Therefore, targeting β 2AR and attenuating β -arrestin signaling might be an attractive target to support DNA damage repair mechanisms.

Multiple lines of research have recently focused on the conceptual modelling of GPCRs contend with the possibility of employing signaling preferences between G protein or β -arrestin signaling, known as signaling 'bias'. GPCRs likely exist in a set of multiple conformational states (which can be then stabilized by distinct ligands) ^[36] associated with the preferential activation of certain pathways ^[31]. Shifting of the equilibrium constants between these multiple states is likely to be the basis of therapeutically relevant drug effects - an improved understanding of the complex GPCR activity landscape will hopefully augment our ability to design signal-selective therapies [37]. The topology of this activity

landscape is likely to be a product of the capacity of GPCRs to form 'higher-order' stable receptor protein complexes [30] ^[36]. These large multiprotein complexes are often described as 'receptorsomes'. It is likely that ligands evoke a range of signaling outputs between G protein and non-G protein signaling - these actions can either take place sequentially via pluridimensional signaling complexes or simultaneously via the concurrent activation of more dedicated and restrictive signaling complexes [38]. The signaling outcome is likely determined by G protein activity and the expression profile of additional proteins involved in creating stable receptorsome complexes [39]. Further extending the appreciation of GPCR signaling complexity is the additional contextual basis of signaling diversity via variance in subcellular localization. GPCR research for many years has focused on the molecular species present on the cell surface at the plasma membrane. The majority of GPCR copies in the cell however reside in multiple intracellular compartments - initially thought to act as a reserve supply only for the cell surface species. Initial research demonstrating how subcellular localization could specify discrete signaling activities emanated from investigations into receptor endocytosis [40][41][42][43]. While the endocytic modulation of GPCR signaling still involved cell surface GPCR species, multiple specifically intracellular environments have now been shown to be important sites of signaling diversification, e.g. mitochondria, endoplasmic reticulum, Golgi apparatus and nucleus [44][45]. This additional signaling capacity proposes that GPCRs interact with intracellular stimulatory factors - or even deleterious factors such as reactive oxygen species - to transduce their full signaling repertoire. This concept therefore strongly suggests that GPCRs might act as molecular sensors for compounds that directly or indirectly induce oxidative stress and/or DNA damage [32].

While many researchers often view signal transduction complexity as a hindrance to therapeutic development, in fact it is a positive aspect as the added nuances of signal transduction actually facilitate the rational creation of multiple distinct biased agents. Indicative of this capacity for exploitation of signaling complexity therapeutically-relevant signaling bias has already been shown for several receptors including, the GnRH receptor ^[46], chemokine receptors ^[47], the parathyroid hormone receptor ^[30] and the neurotensin-1 receptor ^[48].

2.2. GPCR Functionality in the Context of Molecular Gerontology

Aging is a time-dependent functional decline characterized by the accumulation of cellular damage leading to a progressive loss of physiological integrity ^[49]. Accordingly, aging is a major risk factor for chronic metabolic disorders such as obesity, type 2 diabetes and cardiovascular disease ^[50]. Part of the aging process involves loss of homeostatic balances leading to redox imbalances, impairment of oxidative phosphorylation and accumulation of dysfunctional mitochondria ^[51]. Systemic metabolic dysregulation, including mitochondrial dysfunction, is a key hallmark of aging. The dysregulation results in elevated levels of reactive oxygen species (ROS) and sustained oxidative DNA damage, requiring activation of DNA damage repair mechanisms ^{[52][53]}. This causes adverse changes in cells over time, increasing vulnerability to developing nearly all diseases ^[54].

From a signal transduction perspective GPCRs can be seen as the master regulators of somatic tissue functions - the other side of this physiological coin however implicates GPCRs in the etiopathophysiology of a wide range of human diseases. Inter-connected GPCR systems allow highly specific inter-organ crosstalk, making up a complex neuroendocrine control network that regulates 'neurometabolic' activity^[55]. Via the regulation of a diverse range of G protein-dependent and non-G protein signaling modalities GPCRs possess a potent capacity to control intermediary cell metabolism events (*e.g.* kinase activation or calcium mobilization) as well as transcriptional and translational efficacies^[55]. ^{[56][57][58][59][60]}. Therefore, as GPCR systems are likely involved (via a diverse range of activities) in the broadest range of physiological functions their importance in the regulation of the aging process is unsurprising. It is highly likely that one of the most important nexi within this GPCR-aging axis is the functional interplay at the level of genomic integrity control and the DDR process. Molecular aging causes accumulating protein and DNA damage and can also generate age-related increases in cellular senescence. Many stimuli, both intra- and extracellular, can elicit a senescence response. These include dysfunctional telomeres, DNA damage, the expression of certain oncogenes, perturbations to chromatin organization and strong mitogenic signals^{[51][61]}. Many of these stimuli are signaled by pathways that overlap in activating the tumor suppressor protein 53 (p53) and upregulate the cyclin-dependent kinase (CDK) inhibitors p21 and p16 ^{[50][62]}.

2.2.1 GPCRs and the DNA damage repair process

We have recently demonstrated that multiple components of the GPCR signaling system can modulate the activity of signaling proteins directly or indirectly involved in DNA damage and/or repair ^{[63][64][65]}. From its relatively recent inception ^[66] aging research has focused strongly on the links between energy metabolism and age-related damage ^{[67][68]}. Disruption of the glucose metabolic system is nearly universal in aging and as this primary metabolic process falters, a negative energy balance occurs, as ATP levels reduce while reactive oxygen species (ROS) levels increase. The simultaneous loss of the capacity to maintain energetic processes in the face of increasing oxidative stress will eventually overwhelm cellular antioxidant capabilities, resulting in oxidative damage of DNA. Many age-related disorders that affect our population, such as neurodegeneration, type 2 diabetes mellitus (T2DM) and cardiovascular disorders, are caused by these hallmarks ^[2].

Stress-induced DNA damage is arguably the most prominent underlying cause of aging and in turn may be one of the primary players in almost all age-related disorders [69]. This hypothesis has been supported by the investigation of accelerated aging disorders, *i.e.* Hutchinson–Gilford progeria syndrome, Ataxia Telangiectasia and Werner syndrome, which have one commonality, they are a direct effect of DNA damage response (DDR) and repair disruption ^{[70][71][72][73]} [74].

Stress-induced DNA damage can occur at the level of whole chromosome structures, as well as to exposed single- or double-strand entities. Chromosomal DNA stability is provided by nucleoprotein-DNA structures termed telomeres ^[75]. Mammalian telomeres are repetitive DNA sequences, which form a lariat-like structure by associating with the multimeric Shelterin protein complex (also known as the telosome) to shield the exposed ends of chromosomal DNA from damage ^[76]. Telomeres shorten progressively with each cell replication cycle ^[22], thus imposing a functional limit on the number of times a cell can safely divide. Significantly shortened telomeres trigger cellular senescence in normal cells, or genomic instability in pre-malignant cells, which contribute to numerous degenerative and aging-related diseases ^[78]. Multiple lines of research, from human, murine and in cellulo studies, have shown that oxidative stress is associated with accelerated telomere shortening and dysfunction ^{[79][80][81][82]}. Several mechanistic models have been proposed to explain how oxidative stress accelerates telomere shortening. Firstly, it has been proposed that oxidative stress triggers cell death and/or senescence, and as a compensation, the extant cells then undergo further recuperative divisions, leading to increased telomere shortening ^[78]. In addition to this it has been proposed that ROS induce SSBs at telomeres directly, or as intermediates in lesion repair, leading to replication fork collapse and telomere loss ^[83].

GPCRs - as well as their associated signaling proteins - have been recently the focus of considerable research with respect to their capacity to control signaling activities linked to maintenance of genomic integrity [84][85][86][87]. Interventions linked to GPCR activity have been shown to be associated with classical sites of DNA damage including the aforementioned telomeric sites [88], PARP1 functionality [89] and via the orchestration of multi-protein repair complexes [90]. In addition to these mechanistic intersections between GPCR and DDR mechanisms we have recently identified the G protein-coupled receptor (GPCR) associated protein, GIT2, as a potential keystone in aging ^[2]. Further work demonstrated that this receptor scaffolding protein also plays a role in oxidative stress responses [91], and is crucial for integrating several components of the DDR [64]. GIT2 knockout (GIT2KO) mice showed an increased vulnerability to DNA damage [64], displayed symptoms of T2DM [92], showed signs of 'inflammaging' [65][93] and most importantly, showed accelerated aging compared to their wild-type littermates [65]. While this makes GIT2 an interesting target for treating multiple age-related disorders, GIT2 is a scaffolding protein and is therefore difficult to target directly. Typically, drugs are designed to be directed at enzymes, ion channels or receptors. However, as GIT2 is a GPCR interacting protein, we employed transcriptional profiling to identify a 'preferred' GPCR partner for GIT2 signaling and found a strong link with the Relaxin-3 receptor (RXFP3). RXFP3 and GIT2 play similar roles in metabolic aging processes [32]. We also determined the connection between RXFP3 and GIT2 by investigating the role of RXFP3 in oxidative stress and DDR. Analyzing the effects of oxidizing (hydrogen peroxide) and DNA-damaging (camptothecin) stressors on the interacting partners of RXFP3 using Affinity Purification-Mass Spectrometry, we found multiple proteins linked to DDR and cell cycle control. RXFP3 expression increased in response to DNA damage, overexpression, and Relaxin 3-mediated stimulation of RXFP3 reduced phosphorylation of DNA damage marker H2AX, and of the repair protein BRCA1 - both resulting in the attenuation of DNA damage. Our data suggests an RXFP3-GIT2 system that could regulate cellular degradation after DNA damage, and could be a novel mechanism for mitigating the rate of age-related damage accumulation ^[32]. Hence, GPCR signaling systems may represent multifunctional sensors for DNA damaging insults, and their rational exploitation via novel drug design may facilitate our ability to augment DNA repair processes therapeutically. We proposed that GPCR systems may have long evolved side-by-side with emerging DDR systems to act as sensors, and ameliorative effectors, for intracellular DNA damage and age-related stresses.

2.2.2 GPCRs and cellular senescence

Cellular senescence is a multifaceted process that arrests the proliferation of cells but keeps them from going into an apoptotic state. Cell growth arrest and hyporesponsiveness to extrinsic stimuli via cell surface receptors, such as GPCRs, are hallmarks of senescent cells [94][95][96][97]. Cells that express markers of senescence have been shown to accumulate with age and at sites of certain age-related pathologies. Accumulation of senescent cells can cause local and systemic inflammation, tissue destruction, immune system inhibition and stem and progenitor cell dysfunction [62]. Recent findings

implicate p16-dependent senescence in three components of aging — decrements in neurogenesis, haematopoiesis and pancreatic function [61]. These senescent cells possess distinct functional phenotypes, compared to normal cells, with respect to chromatin remodeling and protein secretory behavior ^{[98][99][100]}.

Cellular senescence was originally described by Hayflick and Moorhead ^[101]. An internal factor that can drive cells into replicative senescence is telomeric degradation following each cell cycle ^{[50][102]}. The age-associated shortening of telomeres is commonly regarded as an important contributor to organismal aging and considered a powerful biomarker of aging and aging-associated pathological conditions ^[103]. Since telomeres are necessary to provide stability for the chromosomal DNA ^[75], telomere attenuation will be strongly associated with DNA frailty ^{[101][104]}. Rather than representing a functional 'dead end' of cell physiology, evidence gathered over recent years has demonstrated the importance of senescence-related signaling in processes such as embryonic development ^[105], wound healing/repair ^{[106][107]} and, most importantly, aging ^{[108][109]}.

In addition to telomeric degradation, cellular senescence has been shown to respond to additional stressors. These could be DNA lesions or oxidizing agents (ROS) ^{[110][111]}, both linked through the DDR signaling pathway. But metabolites, mitochondrial dysfunction and epigenetic changes can also induce growth arrest ^[50]. The unavoidable accumulation of DNA and oxidative damage, telomere shortening and the impairment of repair mechanisms and immune system function will engender a chronic accumulation of senescent cells over time as an organism grows older ^[112]. A distinctive feature of senescent cells is the increased expression of cell cycle-inhibitory proteins such as two cyclin-dependent kinase (Cdk) inhibitors, p16 and p21, that are important for senescent cell accumulation during aging. If DNA damage persists, it causes prolonged DDR signaling and protracted proliferative arrest in the form of cellular senescence. Inhibition of DDR signaling kinases like ATM or ATR (ataxia Telangiectasia Rad3 related), allows regulation of senescent cells. These kinases effectively block cell-cycle progression through the stabilization of p53 and transcriptional activation of the cyclindependent kinase (Cdk) inhibitor p21 ^{[113][114]}.

Along with cell cycle arrest, the alteration of the functional cellular 'secretome' (i.e., the range of secreted proteins from a specific cell type) of the specific cell entering a senescent state is one of the characteristic features of this aging-associated state ^[115]. Most senescent cells display profound changes in the chromatin organization which are linked to the cell-autonomous and paracrine aspects of senescence-associated proliferation arrest ^[113]. Chromatin remodeling causes a coherent cellular response, creating an inflammatory phenotype in which a cell secretes activated interleukins, chemokines, extracellular matrix components, metalloproteinases, growth factors and other signalling molecules ^{[112][116]} ^[117]. This modulatory secretory phenotype has now been codified as the senescence-associated secretory phenotype (SASP) ^{[115][118][119]}. A small number of senescent cells can cause considerable dysfunction through their SASP phenotype by releasing factors that can cause tissue necrosis, systemic inflammation, stem cell/progenitor dysfunction, fibrosis and spread of senescence to non-senescent cells ^[62]. SASP activation is a dynamic process that accompanies senescence establishment and can be dependent on DNA damage signaling. DDR can enhance SASP by feed-forward loops that are generated between DDR signaling and ROS attack ^{[119][120]}. Interestingly, it has been demonstrated that SASP-associated activity is also strongly linked to modifications in GPCR functionality ^{[113][121][122]}.

3. Conclusion

While the aging process and the accumulation of age-related damage seem inevitable facts of metabolic life, the strong involvement of GPCR-associated signaling cascades at many levels of this process provides a potentially important and effective drug-based mechanism for amelioration and/or retardation of this process [28][123][124][124][125]. Aging, as a molecular process, is clearly a slowly developing entity, coordinated by the interaction of multiple signaling systems across almost all somatic tissues over decades. This complexity makes it a troublesome process to target using conventional 'monolithic target' therapies, e.g. the failure of anti-amyloid therapies targeting age-related dementia [126]. In contrast, complex mechanistic disease systems may be more effectively targeted by therapeutics that possess multidimensional pharmacological efficacy profiles [127][128][129][130]. The discovery and development of the concept that GPCR systems can effectively target and regulate complex transcriptomic/proteomic responses via receptorsome-based non-G proteindependent signaling [36] provides a feasible platform upon which multidimensional therapeutic interventions for aging can be created [13][28][131]. In their elegant manuscript, Watts and Strogatz [132] demonstrated that an optimal level of communication between entities, within any specific complex system, is facilitated by a level of organization where some nodes within the network possess a greater degree of regulatory connectivity compared to other nodes. In the case of molecular signaling networks in the aging process, it is likely therefore that some proteins possess more profound network-regulating functions than others ^[2]. These network-controlling factors have been termed 'keystones' or 'hubs' and are thought to provide a mechanism of dimensional condensation for highly complex cellular signaling systems. This network organization facilitates the rapid transfer of coherent biological/pathological perturbations across a complex series

of nodes by making so-called 'short cuts' across the network. As such, the super-complex aging process networks can be controlled at a trophic keystone/hub level rather than by individual sensation/regulation at the individual node (protein or gene) level $\frac{92}{133}$. These keystones therefore likely connect and coordinate multiple discrete signaling cascades that synergize to regulate multifactorial somatic processes. Demonstrating the efficiency of organizing networks in this manner, it has been shown that even networks containing thousands of nodes require only the presence of surprisingly few (5–10) keystones to facilitate rapid transfer across large systems $\frac{1321}{2}$. Targeting these trophic-level proteins, potentially via the recently discovered GPCR-based transcriptomic efficacy role, facilitates regulation of such complex disorders in a rational manner as opposed to the unfeasible proposal of therapeutic aging control at every molecular point in the network $\frac{391(131)}{1341(135)}$.

References

- 1. van Dijk G, van Heijningen S, Reijne AC, Nyakas C, van der Zee EA, Eisel UL. Integrative neurobiology of metabolic diseases, neuroinflammation, and neurodegeneration. Front Neurosci. 2015; 9: 173.
- 2. Nkuipou-Kenfack E, Koeck T, Mischak H, Pich A, Schanstra JP, Zurbig P, Schumacher B. Proteome analysis in the assessment of ageing. Ageing Res Rev. 2014; 18: 74-85.
- Redman LM, Smith SR, Burton JH, Martin CK, Il'yasova D, Ravussin E. Metabolic Slowing and Reduced Oxidative Damage with Sustained Caloric Restriction Support the Rate of Living and Oxidative Damage Theories of Aging. Cell Metab. 2018; 27: 805-15 e4.
- Colman RJ, Anderson RM, Johnson SC, Kastman EK, Kosmatka KJ, Beasley TM, Allison DB, Cruzen C, Simmons HA, Kemnitz JW, Weindruch R. Caloric restriction delays disease onset and mortality in rhesus monkeys. Science. 2009; 325: 201-4.
- Colman RJ, Beasley TM, Kemnitz JW, Johnson SC, Weindruch R, Anderson RM. Caloric restriction reduces agerelated and all-cause mortality in rhesus monkeys. Nat Commun. 2014; 5: 3557.
- 6. Lopez-Otin C, Blasco MA, Partridge L, Serrano M, Kroemer G. The hallmarks of aging. Cell. 2013; 153: 1194-217.
- 7. Chadwick W, Martin B, Chapter MC, Park SS, Wang L, Daimon CM, Brenneman R, Maudsley S. GIT2 acts as a potential keystone protein in functional hypothalamic networks associated with age-related phenotypic changes in rats. PLoS One. 2012; 7: e36975.
- 8. Pan MR, Li K, Lin SY, Hung WC. Connecting the Dots: From DNA Damage and Repair to Aging. Int J Mol Sci. 2016; 17.
- 9. Hauser AS, Attwood MM, Rask-Andersen M, Schioth HB, Gloriam DE. Trends in GPCR drug discovery: new agents, targets and indications. Nat Rev Drug Discov. 2017; 16: 829-42.
- 10. Overington JP, Al-Lazikani B, Hopkins AL. How many drug targets are there? Nat Rev Drug Discov. 2006; 5: 993-6.
- Luttrell LM, Ferguson SS, Daaka Y, Miller WE, Maudsley S, Della Rocca GJ, Lin F, Kawakatsu H, Owada K, Luttrell DK, Caron MG, Lefkowitz RJ. Beta-arrestin-dependent formation of beta2 adrenergic receptor-Src protein kinase complexes. Science. 1999; 283: 655-61.
- Maudsley S, Martin B, Janssens J, Etienne H, Jushaj A, van Gastel J, Willemsen A, Chen H, Gesty-Palmer D, Luttrell LM. Informatic deconvolution of biased GPCR signaling mechanisms from in vivo pharmacological experimentation. Methods. 2016; 92: 51-63.
- Maudsley S, Martin B, Gesty-Palmer D, Cheung H, Johnson C, Patel S, Becker KG, Wood WH, 3rd, Zhang Y, Lehrmann E, Luttrell LM. Delineation of a conserved arrestin-biased signaling repertoire in vivo. Mol Pharmacol. 2015; 87: 706-17.
- 14. Madabhushi R, Pan L, Tsai LH. DNA damage and its links to neurodegeneration. Neuron. 2014; 83: 266-82.
- 15. Chow HM, Herrup K. Genomic integrity and the ageing brain. Nat Rev Neurosci. 2015; 16: 672-84.
- Ishida T, Ishida M, Tashiro S, Yoshizumi M, Kihara Y. Role of DNA damage in cardiovascular disease. Circ J. 2014; 78: 42-50.
- 17. Dobbelstein M, Sorensen CS. Exploiting replicative stress to treat cancer. Nat Rev Drug Discov. 2015; 14: 405-23.
- 18. De I, Dogra N, Singh S. The Mitochondrial Unfolded Protein Response: Role in Cellular Homeostasis and Disease. Curr Mol Med. 2017; 17: 587-97.
- 19. Chung JH. The role of DNA-PK in aging and energy metabolism. FEBS J. 2018; 285: 1959-72.

- 20. Awate S, Brosh RM, Jr. Interactive Roles of DNA Helicases and Translocases with the Single-Stranded DNA Binding Protein RPA in Nucleic Acid Metabolism. Int J Mol Sci. 2017; 18.
- 21. Vass M, Kooistra AJ, Yang D, Stevens RC, Wang MW, de Graaf C. Chemical Diversity in the G Protein-Coupled Receptor Superfamily. Trends Pharmacol Sci. 2018; 39: 494-512.
- 22. Wingler LM, Lefkowitz RJ. Conformational Basis of G Protein-Coupled Receptor Signaling Versatility. Trends Cell Biol. 2020; 30: 736-47.
- 23. Liu Y, Yang Y, Ward R, An S, Guo XX, Li W, Xu TR. Biased signalling: the instinctive skill of the cell in the selection of appropriate signalling pathways. Biochem J. 2015; 470: 155-67.
- 24. Syrovatkina V, Alegre KO, Dey R, Huang XY. Regulation, Signaling, and Physiological Functions of G-Proteins. J Mol Biol. 2016; 428: 3850-68.
- 25. Ma N, Nivedha AK, Vaidehi N. Allosteric communication regulates ligand-specific GPCR activity. FEBS J. 2021.
- 26. Gurevich VV, Gurevich EV. GPCR Signaling Regulation: The Role of GRKs and Arrestins. Front Pharmacol. 2019; 10: 125.
- 27. Luttrell LM, Gesty-Palmer D. Beyond desensitization: physiological relevance of arrestin-dependent signaling. Pharmacol Rev. 2010; 62: 305-30.
- 28. Maudsley S, Patel SA, Park SS, Luttrell LM, Martin B. Functional signaling biases in G protein-coupled receptors: Game Theory and receptor dynamics. Mini Rev Med Chem. 2012; 12: 831-40.
- 29. Luttrell LM, Maudsley S, Bohn LM. Fulfilling the Promise of "Biased" G Protein-Coupled Receptor Agonism. Mol Pharmacol. 2015; 88: 579-88.
- Luttrell LM, Maudsley S, Gesty-Palmer D. Translating in vitro ligand bias into in vivo efficacy. Cell Signal. 2018; 41: 46-55.
- 31. Seyedabadi M, Ghahremani MH, Albert PR. Biased signaling of G protein coupled receptors (GPCRs): Molecular determinants of GPCR/transducer selectivity and therapeutic potential. Pharmacol Ther. 2019; 200: 148-78.
- van Gastel J, Leysen H, Santos-Otte P, Hendrickx JO, Azmi A, Martin B, Maudsley S. The RXFP3 receptor is functionally associated with cellular responses to oxidative stress and DNA damage. Aging (Albany NY). 2019; 11: 11268-313.
- 33. Rajagopal S, Shenoy SK. GPCR desensitization: Acute and prolonged phases. Cell Signal. 2018; 41: 9-16.
- 34. Magalhaes AC, Dunn H, Ferguson SS. Regulation of GPCR activity, trafficking and localization by GPCR-interacting proteins. Br J Pharmacol. 2012; 165: 1717-36.
- 35. Hara MR, Sachs BD, Caron MG, Lefkowitz RJ. Pharmacological blockade of a beta(2)AR-beta-arrestin-1 signaling cascade prevents the accumulation of DNA damage in a behavioral stress model. Cell Cycle. 2013; 12: 219-24.
- 36. Maudsley S, Martin B, Luttrell LM. The origins of diversity and specificity in g protein-coupled receptor signaling. J Pharmacol Exp Ther. 2005; 314: 485-94.
- 37. Gurevich VV, Gurevich EV. Biased GPCR signaling: Possible mechanisms and inherent limitations. Pharmacol Ther. 2020; 211: 107540.
- Nguyen AH, Thomsen ARB, Cahill TJ, 3rd, Huang R, Huang LY, Marcink T, Clarke OB, Heissel S, Masoudi A, Ben-Hail D, Samaan F, Dandey VP, Tan YZ, et al. Structure of an endosomal signaling GPCR-G protein-beta-arrestin megacomplex. Nat Struct Mol Biol. 2019; 26: 1123-31.
- 39. van Gastel J, Leysen H, Boddaert J, Vangenechten L, Luttrell LM, Martin B, Maudsley S. Aging-related modifications to G protein-coupled receptor signaling diversity. Pharmacol Ther. 2020; 223: 107793.
- 40. Ahn S, Maudsley S, Luttrell LM, Lefkowitz RJ, Daaka Y. Src-mediated tyrosine phosphorylation of dynamin is required for beta2-adrenergic receptor internalization and mitogen-activated protein kinase signaling. J Biol Chem. 1999; 274: 1185-8.
- 41. Pierce KL, Maudsley S, Daaka Y, Luttrell LM, Lefkowitz RJ. Role of endocytosis in the activation of the extracellular signal-regulated kinase cascade by sequestering and nonsequestering G protein-coupled receptors. Proc Natl Acad Sci U S A. 2000; 97: 1489-94.
- 42. Maudsley S, Pierce KL, Zamah AM, Miller WE, Ahn S, Daaka Y, Lefkowitz RJ, Luttrell LM. The beta(2)-adrenergic receptor mediates extracellular signal-regulated kinase activation via assembly of a multi-receptor complex with the epidermal growth factor receptor. J Biol Chem. 2000; 275: 9572-80.
- 43. Ellisdon AM, Halls ML. Compartmentalization of GPCR signalling controls unique cellular responses. Biochem Soc Trans. 2016; 44: 562-7.

- 44. Irannejad R, von Zastrow M. GPCR signaling along the endocytic pathway. Curr Opin Cell Biol. 2014; 27: 109-16.
- 45. Djeungoue-Petga MA, Hebert-Chatelain E. Linking Mitochondria and Synaptic Transmission: The CB1 Receptor. Bioessays. 2017; 39.
- 46. Maudsley S, Davidson L, Pawson AJ, Chan R, Lopez de Maturana R, Millar RP. Gonadotropin-releasing hormone (GnRH) antagonists promote proapoptotic signaling in peripheral reproductive tumor cells by activating a Galphaicoupling state of the type I GnRH receptor. Cancer Res. 2004; 64: 7533-44.
- 47. Hauser MA, Legler DF. Common and biased signaling pathways of the chemokine receptor CCR7 elicited by its ligands CCL19 and CCL21 in leukocytes. J Leukoc Biol. 2016; 99: 869-82.
- 48. Slosky LM, Bai Y, Toth K, Ray C, Rochelle LK, Badea A, Chandrasekhar R, Pogorelov VM, Abraham DM, Atluri N, Peddibhotla S, Hedrick MP, Hershberger P, et al. beta-Arrestin-Biased Allosteric Modulator of NTSR1 Selectively Attenuates Addictive Behaviors. Cell. 2020; 181: 1364-79 e14.
- 49. Collis SJ, Boulton SJ. Emerging links between the biological clock and the DNA damage response. Chromosoma. 2007; 116: 331-9.
- 50. Spinelli R, Parrillo L, Longo M, Florese P, Desiderio A, Zatterale F, Miele C, Raciti GA, Beguinot F. Molecular basis of ageing in chronic metabolic diseases. J Endocrinol Invest. 2020; 43: 1373-89.
- 51. Natarajan V, Chawla R, Mah T, Vivekanandan R, Tan SY, Sato PY, Mallilankaraman K. Mitochondrial Dysfunction in Age-Related Metabolic Disorders. Proteomics. 2020; 20: e1800404.
- 52. Sharma R, Ramanathan A. The Aging Metabolome-Biomarkers to Hub Metabolites. Proteomics. 2020; 20: e1800407.
- 53. Niccoli T, Partridge L. Ageing as a risk factor for disease. Curr Biol. 2012; 22: R741-52.
- Alemany R, Perona JS, Sanchez-Dominguez JM, Montero E, Canizares J, Bressani R, Escriba PV, Ruiz-Gutierrez V. G protein-coupled receptor systems and their lipid environment in health disorders during aging. Biochim Biophys Acta. 2007; 1768: 964-75.
- 55. Janssens J, Etienne H, Idriss S, Azmi A, Martin B, Maudsley S. Systems-Level G Protein-Coupled Receptor Therapy Across a Neurodegenerative Continuum by the GLP-1 Receptor System. Front Endocrinol (Lausanne). 2014; 5: 142.
- 56. Pi M, Nishimoto SK, Quarles LD. GPRC6A: Jack of all metabolism (or master of none). Mol Metab. 2017; 6: 185-93.
- 57. Reimann F, Gribble FM. G protein-coupled receptors as new therapeutic targets for type 2 diabetes. Diabetologia. 2016; 59: 229-33.
- 58. Amisten S, Neville M, Hawkes R, Persaud SJ, Karpe F, Salehi A. An atlas of G-protein coupled receptor expression and function in human subcutaneous adipose tissue. Pharmacol Ther. 2015; 146: 61-93.
- 59. Hudson BD, Ulven T, Milligan G. The therapeutic potential of allosteric ligands for free fatty acid sensitive GPCRs. Curr Top Med Chem. 2013; 13: 14-25.
- 60. Jaana Van Gastel JJ, Harmonie Etienne, Abdelkrim Azmi and Stuart Maudsley. The synergistic GIT2-RXFP3 system in the brain and its importance in age-related disorders. Frontiers Aging Neuroscience. 2016; 8.
- 61. Campisi J, d'Adda di Fagagna F. Cellular senescence: when bad things happen to good cells. Nat Rev Mol Cell Biol. 2007; 8: 729-40.
- 62. Tchkonia T, Palmer AK, Kirkland JL. New Horizons: Novel Approaches to Enhance Healthspan Through Targeting Cellular Senescence and Related Aging Mechanisms. J Clin Endocrinol Metab. 2021; 106: e1481-e7.
- 63. Leysen H, van Gastel J, Hendrickx JO, Santos-Otte P, Martin B, Maudsley S. G Protein-Coupled Receptor Systems as Crucial Regulators of DNA Damage Response Processes. Int J Mol Sci. 2018; 19.
- 64. Lu D, Cai H, Park SS, Siddiqui S, Premont RT, Schmalzigaug R, Paramasivam M, Seidman M, Bodogai I, Biragyn A, Daimon CM, Martin B, Maudsley S. Nuclear GIT2 is an ATM substrate and promotes DNA repair. Mol Cell Biol. 2015; 35: 1081-96.
- 65. Siddiqui S, Lustig A, Carter A, Sankar M, Daimon CM, Premont RT, Etienne H, van Gastel J, Azmi A, Janssens J, Becker KG, Zhang Y, Wood W, et al. Genomic deletion of GIT2 induces a premature age-related thymic dysfunction and systemic immune system disruption. Aging (Albany NY). 2017; 9: 706-40.
- 66. Baker GT, 3rd, Achenbaum WA. A historical perspective of research on the biology of aging from Nathan W. Shock. Exp Gerontol. 1992; 27: 261-73.
- 67. Martin B, Mattson MP, Maudsley S. Caloric restriction and intermittent fasting: two potential diets for successful brain aging. Ageing Res Rev. 2006; 5: 332-53.
- 68. Kenyon CJ. The genetics of ageing. Nature. 2010; 464: 504-12.

- 69. Maynard S, Fang EF, Scheibye-Knudsen M, Croteau DL, Bohr VA. DNA Damage, DNA Repair, Aging, and Neurodegeneration. Cold Spring Harb Perspect Med. 2015; 5.
- Gonzalo S, Kreienkamp R. DNA repair defects and genome instability in Hutchinson-Gilford Progeria Syndrome. Curr Opin Cell Biol. 2015; 34: 75-83.
- 71. Musich PR, Zou Y. Genomic instability and DNA damage responses in progeria arising from defective maturation of prelamin A. Aging (Albany NY). 2009; 1: 28-37.
- 72. Hyun M, Choi S, Stevnsner T, Ahn B. The Caenorhabditis elegans Werner syndrome protein participates in DNA damage checkpoint and DNA repair in response to CPT-induced double-strand breaks. Cell Signal. 2016; 28: 214-23.
- 73. Rothblum-Oviatt C, Wright J, Lefton-Greif MA, McGrath-Morrow SA, Crawford TO, Lederman HM. Ataxia telangiectasia: a review. Orphanet J Rare Dis. 2016; 11: 159.
- 74. Shiloh Y, Lederman HM. Ataxia-telangiectasia (A-T): An emerging dimension of premature ageing. Ageing Res Rev. 2017; 33: 76-88.
- 75. Aubert G, Lansdorp PM. Telomeres and aging. Physiol Rev. 2008; 88: 557-79.
- 76. de Lange T. Shelterin: the protein complex that shapes and safeguards human telomeres. Genes Dev. 2005; 19: 2100-10.
- 77. Harley CB, Futcher AB, Greider CW. Telomeres shorten during ageing of human fibroblasts. Nature. 1990; 345: 458-60.
- 78. Barnes RP, Fouquerel E, Opresko PL. The impact of oxidative DNA damage and stress on telomere homeostasis. Mech Ageing Dev. 2019; 177: 37-45.
- 79. Graham MK, Meeker A. Telomeres and telomerase in prostate cancer development and therapy. Nat Rev Urol. 2017; 14: 607-19.
- Jurk D, Wilson C, Passos JF, Oakley F, Correia-Melo C, Greaves L, Saretzki G, Fox C, Lawless C, Anderson R, Hewitt G, Pender SL, Fullard N, et al. Chronic inflammation induces telomere dysfunction and accelerates ageing in mice. Nat Commun. 2014; 2: 4172.
- Cattan V, Mercier N, Gardner JP, Regnault V, Labat C, Maki-Jouppila J, Nzietchueng R, Benetos A, Kimura M, Aviv A, Lacolley P. Chronic oxidative stress induces a tissue-specific reduction in telomere length in CAST/Ei mice. Free Radic Biol Med. 2008; 44: 1592-8.
- 82. Shay JW. Role of Telomeres and Telomerase in Aging and Cancer. Cancer Discov. 2016; 6: 584-93.
- 83. von Zglinicki T. Oxidative stress shortens telomeres. Trends Biochem Sci. 2002; 27: 339-44.
- Nieto A, Hara MR, Quereda V, Grant W, Saunders V, Xiao K, McDonald PH, Duckett DR. betaarrestin-1 regulates DNA repair by acting as an E3-ubiquitin ligase adaptor for 53BP1. Cell Death Differ. 2020; 27: 1200-13.
- 85. Nieto Gutierrez A, McDonald PH. GPCRs: Emerging anti-cancer drug targets. Cell Signal. 2018; 41: 65-74.
- Fan Y, Huang Z, Long C, Ning J, Zhang H, Kuang X, Zhang Q, Shen H. ID2 protects retinal pigment epithelium cells from oxidative damage through p-ERK1/2/ID2/NRF2. Arch Biochem Biophys. 2018; 650: 1-13.
- 87. Hara MR, Kovacs JJ, Whalen EJ, Rajagopal S, Strachan RT, Grant W, Towers AJ, Williams B, Lam CM, Xiao K, Shenoy SK, Gregory SG, Ahn S, et al. A stress response pathway regulates DNA damage through beta2adrenoreceptors and beta-arrestin-1. Nature. 2011; 477: 349-53.
- Herbert KE, Mistry Y, Hastings R, Poolman T, Niklason L, Williams B. Angiotensin II-mediated oxidative DNA damage accelerates cellular senescence in cultured human vascular smooth muscle cells via telomere-dependent and independent pathways. Circ Res. 2008; 102: 201-8.
- 89. Adamczyk A, Jesko H, Strosznajder RP. Alzheimer's disease related peptides affected cholinergic receptor mediated poly(ADP-ribose) polymerase activity in the hippocampus. Folia Neuropathol. 2005; 43: 139-42.
- 90. Wagner W, Kania KD, Ciszewski WM. Stimulation of lactate receptor (HCAR1) affects cellular DNA repair capacity. DNA Repair (Amst). 2017; 52: 49-58.
- 91. Chadwick W, Zhou Y, Park SS, Wang L, Mitchell N, Stone MD, Becker KG, Martin B, Maudsley S. Minimal peroxide exposure of neuronal cells induces multifaceted adaptive responses. PLoS One. 2010; 5: e14352.
- 92. Martin B, Chadwick W, Janssens J, Premont RT, Schmalzigaug R, Becker KG, Lehrmann E, Wood WH, Zhang Y, Siddiqui S, Park SS, Cong WN, Daimon CM, et al. GIT2 Acts as a Systems-Level Coordinator of Neurometabolic Activity and Pathophysiological Aging. Front Endocrinol (Lausanne). 2015; 6: 191.
- 93. Li H, Guan SB, Lu Y, Wang F, Liu YH, Liu QY. Genetic deletion of GIT2 prolongs functional recovery and suppresses chondrocyte differentiation in rats with rheumatoid arthritis. J Cell Biochem. 2018; 119: 1538-47.

- 94. Yeo EJ, Jang IS, Lim HK, Ha KS, Park SC. Agonist-specific differential changes of cellular signal transduction pathways in senescent human diploid fibroblasts. Exp Gerontol. 2002; 37: 871-83.
- 95. Yeo EJ, Park SC. Age-dependent agonist-specific dysregulation of membrane-mediated signal transduction: emergence of the gate theory of aging. Mech Ageing Dev. 2002; 123: 1563-78.
- 96. Hakim MA, Buchholz JN, Behringer EJ. Electrical dynamics of isolated cerebral and skeletal muscle endothelial tubes: Differential roles of G-protein-coupled receptors and K(+) channels. Pharmacol Res Perspect. 2018; 6: e00391.
- Xiao P, Huang X, Huang L, Yang J, Li A, Shen K, Wedegaertner PB, Jiang X. G protein-coupled receptor kinase 4induced cellular senescence and its senescence-associated gene expression profiling. Exp Cell Res. 2017; 360: 273-80.
- 98. Kuilman T, Michaloglou C, Mooi WJ, Peeper DS. The essence of senescence. Genes Dev. 2010; 24: 2463-79.
- 99. Adams PD. Healing and hurting: molecular mechanisms, functions, and pathologies of cellular senescence. Mol Cell. 2009; 36: 2-14.
- 100. Tchkonia T, Zhu Y, van Deursen J, Campisi J, Kirkland JL. Cellular senescence and the senescent secretory phenotype: therapeutic opportunities. J Clin Invest. 2013; 123: 966-72.
- 101. Hayflick L, Moorhead PS. The serial cultivation of human diploid cell strains. Exp Cell Res. 1961; 25: 585-621.
- 102. Khosla S, Farr JN, Tchkonia T, Kirkland JL. The role of cellular senescence in ageing and endocrine disease. Nat Rev Endocrinol. 2020; 16: 263-75.
- 103. Vaiserman A, Krasnienkov D. Telomere Length as a Marker of Biological Age: State-of-the-Art, Open Issues, and Future Perspectives. Front Genet. 2020; 11: 630186.
- 104. Bodnar AG, Ouellette M, Frolkis M, Holt SE, Chiu CP, Morin GB, Harley CB, Shay JW, Lichtsteiner S, Wright WE. Extension of life-span by introduction of telomerase into normal human cells. Science. 1998; 279: 349-52.
- 105. Munoz-Espin D, Canamero M, Maraver A, Gomez-Lopez G, Contreras J, Murillo-Cuesta S, Rodriguez-Baeza A, Varela-Nieto I, Ruberte J, Collado M, Serrano M. Programmed cell senescence during mammalian embryonic development. Cell. 2013; 155: 1104-18.
- 106. Jun JI, Lau LF. The matricellular protein CCN1 induces fibroblast senescence and restricts fibrosis in cutaneous wound healing. Nat Cell Biol. 2010; 12: 676-85.
- 107. Krizhanovsky V, Yon M, Dickins RA, Hearn S, Simon J, Miething C, Yee H, Zender L, Lowe SW. Senescence of activated stellate cells limits liver fibrosis. Cell. 2008; 134: 657-67.
- 108. Baker DJ, Perez-Terzic C, Jin F, Pitel KS, Niederlander NJ, Jeganathan K, Yamada S, Reyes S, Rowe L, Hiddinga HJ, Eberhardt NL, Terzic A, van Deursen JM. Opposing roles for p16Ink4a and p19Arf in senescence and ageing caused by BubR1 insufficiency. Nat Cell Biol. 2008; 10: 825-36.
- 109. Baker DJ, Wijshake T, Tchkonia T, LeBrasseur NK, Childs BG, van de Sluis B, Kirkland JL, van Deursen JM. Clearance of p16lnk4a-positive senescent cells delays ageing-associated disorders. Nature. 2011; 479: 232-6.
- 110. Nardella C, Clohessy JG, Alimonti A, Pandolfi PP. Pro-senescence therapy for cancer treatment. Nat Rev Cancer. 2011; 11: 503-11.
- 111. Sedelnikova OA, Horikawa I, Zimonjic DB, Popescu NC, Bonner WM, Barrett JC. Senescing human cells and ageing mice accumulate DNA lesions with unrepairable double-strand breaks. Nat Cell Biol. 2004; 6: 168-70.
- 112. Saez-Atienzar S, Masliah E. Cellular senescence and Alzheimer disease: the egg and the chicken scenario. Nat Rev Neurosci. 2020; 21: 433-44.
- 113. Di Micco R, Krizhanovsky V, Baker D, d'Adda di Fagagna F. Cellular senescence in ageing: from mechanisms to therapeutic opportunities. Nat Rev Mol Cell Biol. 2021; 22: 75-95.
- 114. van Deursen JM. The role of senescent cells in ageing. Nature. 2014; 509: 439-46.
- 115. Coppe JP, Patil CK, Rodier F, Sun Y, Munoz DP, Goldstein J, Nelson PS, Desprez PY, Campisi J. Senescenceassociated secretory phenotypes reveal cell-nonautonomous functions of oncogenic RAS and the p53 tumor suppressor. PLoS Biol. 2008; 6: 2853-68.
- 116. Shah PP, Donahue G, Otte GL, Capell BC, Nelson DM, Cao K, Aggarwala V, Cruickshanks HA, Rai TS, McBryan T, Gregory BD, Adams PD, Berger SL. Lamin B1 depletion in senescent cells triggers large-scale changes in gene expression and the chromatin landscape. Genes Dev. 2013; 27: 1787-99.
- 117. Zhang H, Pan KH, Cohen SN. Senescence-specific gene expression fingerprints reveal cell-type-dependent physical clustering of up-regulated chromosomal loci. Proc Natl Acad Sci U S A. 2003; 100: 3251-6.

- 118. Rodier F, Coppe JP, Patil CK, Hoeijmakers WA, Munoz DP, Raza SR, Freund A, Campeau E, Davalos AR, Campisi J. Persistent DNA damage signalling triggers senescence-associated inflammatory cytokine secretion. Nat Cell Biol. 2009; 11: 973-9.
- 119. Kuilman T, Peeper DS. Senescence-messaging secretome: SMS-ing cellular stress. Nat Rev Cancer. 2009; 9: 81-94.
- 120. Passos JF, Nelson G, Wang C, Richter T, Simillion C, Proctor CJ, Miwa S, Olijslagers S, Hallinan J, Wipat A, Saretzki G, Rudolph KL, Kirkwood TB, et al. Feedback between p21 and reactive oxygen production is necessary for cell senescence. Mol Syst Biol. 2010; 6: 347.
- 121. Guo H, Liu Z, Xu B, Hu H, Wei Z, Liu Q, Zhang X, Ding X, Wang Y, Zhao M, Gong Y, Shao C. Chemokine receptor CXCR2 is transactivated by p53 and induces p38-mediated cellular senescence in response to DNA damage. Aging Cell. 2013; 12: 1110-21.
- 122. Jin HJ, Lee HJ, Heo J, Lim J, Kim M, Kim MK, Nam HY, Hong GH, Cho YS, Choi SJ, Kim IG, Shin DM, Kim SW. Senescence-Associated MCP-1 Secretion Is Dependent on a Decline in BMI1 in Human Mesenchymal Stromal Cells. Antioxid Redox Signal. 2016; 24: 471-85.
- 123. Garcia-Martinez I, Shaker ME, Mehal WZ. Therapeutic Opportunities in Damage-Associated Molecular Pattern-Driven Metabolic Diseases. Antioxid Redox Signal. 2015; 23: 1305-15.
- 124. Pearl LH, Schierz AC, Ward SE, Al-Lazikani B, Pearl FM. Therapeutic opportunities within the DNA damage response. Nat Rev Cancer. 2015; 15: 166-80.
- 125. Williams DT, Staples CJ. Approaches for Identifying Novel Targets in Precision Medicine: Lessons from DNA Repair. Adv Exp Med Biol. 2017; 1007: 1-16.
- 126. Gold M. Phase II clinical trials of anti-amyloid beta antibodies: When is enough, enough? Alzheimers Dement (N Y). 2017; 3: 402-9.
- 127. Janssens J, Lu D, Ni B, Chadwick W, Siddiqui S, Azmi A, Etienne H, Jushaj A, van Gastel J, Martin B, Maudsley S. Development of Precision Small-Molecule Proneurotrophic Therapies for Neurodegenerative Diseases. Vitam Horm. 2017; 104: 263-311.
- 128. Chadwick W, Mitchell N, Martin B, Maudsley S. Therapeutic targeting of the endoplasmic reticulum in Alzheimer's disease. Curr Alzheimer Res. 2012; 9: 110-9.
- 129. Jacobson KA. New paradigms in GPCR drug discovery. Biochem Pharmacol. 2015; 98: 541-55.
- 130. Luttrell LM. Minireview: More than just a hammer: ligand "bias" and pharmaceutical discovery. Mol Endocrinol. 2014; 28: 281-94.
- 131. Maudsley S, Devanarayan V, Martin B, Geerts H, Brain Health Modeling I. Intelligent and effective informatic deconvolution of "Big Data" and its future impact on the quantitative nature of neurodegenerative disease therapy. Alzheimers Dement. 2018; 14: 961-75.
- 132. Watts DJ, Strogatz SH. Collective dynamics of 'small-world' networks. Nature. 1998; 393: 440-2.
- Han JD, Bertin N, Hao T, Goldberg DS, Berriz GF, Zhang LV, Dupuy D, Walhout AJ, Cusick ME, Roth FP, Vidal M. Evidence for dynamically organized modularity in the yeast protein-protein interaction network. Nature. 2004; 430: 88-93.
- 134. Hendrickx JO, van Gastel J, Leysen H, Martin B, Maudsley S. High-dimensionality Data Analysis of Pharmacological Systems Associated with Complex Diseases. Pharmacol Rev. 2020; 72: 191-217.
- 135. Bakula D, Aliper AM, Mamoshina P, Petr MA, Teklu A, Baur JA, Campisi J, Ewald CY, Georgievskaya A, Gladyshev VN, Kovalchuk O, Lamming DW, Luijsterburg MS, et al. Aging and drug discovery. Aging (Albany NY). 2018; 10: 3079-88.

Retrieved from https://encyclopedia.pub/entry/history/show/20803