# Impact of Cyanobacteria Blooms on Environment and Health

Subjects: Water Resources

Contributor: Weizhen Zhang, Jing Liu, Yunxing Xiao, Yumiao Zhang, Yangjinzhi Yu, Zheng Zheng, Yafeng Liu, Qi Li

Cyanobacteria blooms are a global aquatic environment problem. Due to global warming and water eutrophication, the surface cyanobacteria accumulate in a certain area to form cyanobacteria blooms driven by wind. Cyanobacteria blooms change the physical and chemical properties of water and cause pollution. Moreover, cyanobacteria release organic matter, N (nitrogen) and P (phosphorus) into the water during their apoptosis, accelerating the eutrophication of the water, threatening aquatic flora and fauna, and affecting the community structure and abundance of microorganisms in the water. Simultaneously, toxins and carcinogens released from cyanobacteria can be enriched through the food chain/web, endangering human health.

Keywords: cyanobacteria blooms ; cyanobacteria toxins ; aquatic environment

## 1. Introduction

Cyanobacteria are widely distributed in marine and freshwater, and have more robust adaptability than most eukaryotes <sup>[1]</sup>. They have the ability to grow and reproduce in extreme environments (ice and snow, hot springs, alkaline soda lakes, brine pools, deserts, and polar regions) <sup>[2]</sup>. When organic matter, N (Nitrogen), P (Phosphorus), and other nutrients are enriched in water, cyanobacteria multiply and accumulate into dominant groups. The cyanobacteria blooms, which form green, red-brown, and red in freshwater or marine, are one of the most notable symptoms of nutrient enrichment or eutrophication <sup>[3]</sup>. Cyanobacteria blooms are becoming increasingly common worldwide and pose a serious threat to the sustainability of aquatic ecosystems, such as Taihu Lake in China, Lake Erie in the United States, Lake Winnipeg in Canada, and Lake Nieuwe Meer in the Netherlands <sup>[4]</sup>. Since the 1930s, plenty of studies have been carried out on cyanobacteria blooms, including the causes of cyanobacteria blooms <sup>[5]</sup>, the harm of cyanobacteria products and the symbiosis of algae and bacteria <sup>[6]</sup>, and the nutrient effect of cyanobacteria blooms <sup>[7]</sup>.

### 2. The Pollution of Cyanobacteria Blooms to Water

It is considered that cyanobacteria blooms form when the cyanobacteria reaches  $10^5$  cells/mL, or the chlorophyll a (Chla) concentration reaches 10 µg/L, and a visible covering layer forms on the surface of the water <sup>[8]</sup>. The cyanobacteria blooms' decay process has a more serious impact on the aquatic environment. Aerobic and anaerobic reactions exist in the degradation process of cyanobacteria, and toxins and odorous gases are released. During the decomposition of cyanobacteria blooms, a large number of organic substances and soluble nutrients will be released to water, which will lower the transparency of water, aggravate the eutrophication of water, and form "black spots" <sup>[9]</sup>. Cyanobacteria blooms will lead to the acidity of the water, the rising trend of conductivity, the continuous increase in chemical oxygen demand, and the increase in organic matter concentration in the water <sup>[10]</sup>. In addition, organic debris formed by cyanobacteria accumulation has a high decomposition rate in the water, which can be decomposed by 41.9% within 48 h <sup>[11]</sup>, which will harm the ecosystem of the water <sup>[8][12]</sup>. A large amount of dissolved organic matter (DOM) is released during the decline of cyanobacteria, and with the progress of the reaction, dissolved organic carbon (DOC) is converted into dissolved inorganic carbon (DIC), and most of them are, lastly, transformed into humus, which is challenging to degrade <sup>[13]</sup>.

#### 2.1. Impacts of Cyanobacteria Blooms on Aquatic Fauna

During cyanobacteria blooms, a large number of dead cyanobacteria will sink to the bottom and decompose, consuming oxygen, which will reduce the dissolved oxygen (DO) in water, thus affecting the living conditions of aquatic fauna, causing the disappearance of some fish, shellfish, and invertebrates, and decreasing the species diversity of the aquatic ecosystem <sup>[14]</sup>. In the apoptosis of cyanobacteria, secondary metabolites such as toxins, odorous substances, and other substances were released into the water, and the concentration of ammonia ( $NH_4^+$ ) and microcystins (MCs) will increase simultaneously, causing acute or chronic adverse effects on aquatic organisms <sup>[15]</sup>.

Exposure to MCs leads to lipid peroxidation, DNA damage, and changes in antioxidant enzymes, such as superoxide dismutase (SOD), peroxidase (POD), and catalase (CAT) in different aquatic organisms. MCs can cause damage to their circulatory system, digestive system, and immune system. Simultaneously, they will induce changes in detoxification enzymes such as glutathione S-transferase (GST) and glutathione peroxidase (GPx) <sup>[16]</sup>. Studies have demonstrated that the liver is the main target of MCs <sup>[17]</sup>. By studying the bioaccumulation law of MCs in two snails in a cyanobacteria-bloom plateau lake, it was found that the hepatopancreas was the main target of two snails <sup>[18]</sup>. Some reports indicate that the concentration of MCs in the intestines, gonads, and muscles of *Cyprinus carpio* was lower but higher in hepatopancreas <sup>[19]</sup>. MCs are enriched in the hepatopancreas of *Macrobrachium rosenbergii*, destroying the structure and function of hepatopancreas, causing dose-dependent and time-dependent toxic effects <sup>[20]</sup>. Andersen et al. found that a high dose of microcystin-LR (MC-LR) could lead to diffuse necrosis and hepatic megalocytosis in the whole liver of *Atlantic salmon* <sup>[21]</sup>. Previous studies have demonstrated that MCs can be transferred to more sensitive organisms through the food chain/network <sup>[22]</sup>.

 $NH_4^+$  can induce the antioxidant defense of juvenile crucian carp. High concentration  $NH_4^+$  has toxic effects on CAT, SOD, and glutathione (GSH) in the fish liver <sup>[23]</sup>. Histopathological changes in the gills, liver, and kidney of *Oreochromis niloticus* are caused by different concentrations of  $NH_4^+$ , and include gill congestion, telangiectasia, turbid swelling, edema degeneration of liver tissue, kidney congestion, and glomerulonephritis <sup>[24]</sup>.  $NH_4^+$  significantly affects the plasma and hematological parameters of juvenile *Megalabrama amblycephala*, demonstrating histopathological changes in the gills, liver, and kidney of fish. The severity of the lesions is different, with the liver exhibiting the most extensive damage, followed by the gills and kidneys <sup>[25]</sup>.

In addition, it is reported that MCs and  $NH_4^+$  have synergistic effects on the immunotoxicity of aquatic organisms. After combined poisoning, the peripheral interspace of the lymphocytes of *Megalabrama amblycephala* is broadened, the nucleus is atrophied, and the mitochondria are swollen. Moreover, the exposure to algae toxin and  $NH_4^+$  has a significant interaction with macrophage phagocytosis activity, respiratory burst activities, a total number of white blood cells and the transcriptional levels of *slgM*, *mlgD*, and *slgZ* genes of *Megalabrama amblycephala* [15].

#### 2.2. Impacts of Cyanobacteria Blooms on Aquatic Flora

The cyanobacteria blooms have strong inhibitory effect on the photosynthetic activity of aquatic flora, leading to leaf death and irreversible inhibition of photosynthesis <sup>[26]</sup>. Long-term and high-concentration aggregation of cyanobacteria will shade, consume oxygen, and release allelochemicals and MCs, resulting in the disappearance of submerged vegetation <sup>[27]</sup>. Cyanobacteria blooms lead to the Chla of *Potamongeton malaianus* and *Stuckenia pectinata* decreasing by 50% and 56%, respectively <sup>[28]</sup>.

MCs can induce the reactive oxygen species (ROS) production and an increase in malondialdehyde (MDA), exacerbating the oxidative damage for aquatic flora <sup>[29]</sup>. MCs can bind irreversibly with phosphatase-1 (PP1) and phosphatase-2A (PP2A) covalently, causing a series of biochemical reactions in cells to be disordered and changing chlorophyll contents and pigment composition in plants <sup>[30]</sup>.

The anatoxin-a produced by cyanobacteria can cause the disorder of oxidative stress reaction in aquatic flora <sup>[31]</sup>. Treatment with 0.01–0.2 µg/mL MC-LR for 96 h can inhibit the growth of *Spirodela oligorrhiza* <sup>[32]</sup>. MC-LR concentration of 1.0 µg/L can significantly impede the development of the roots of *Lepidium sativum*, and a concentration of 10 µg/L can inhibit the growth of the whole plant <sup>[33]</sup>. It has also been found that 0.12–3 µg/mL MCs can hinder the growth of *Oryza sativa* L. <sup>[34]</sup>. In addition, MCs can cause the gap of aeration tissue in the rhizomes of *Phragmites australis* to be blocked by callus-like tissue, resulting in the gangrene of outer skin tissue in the reed root. When exposed to 10–40 µg/mL of MC-LR for 120 h, the cytoskeleton of reed root changes (microtubule degradation), and its roots swell and deform <sup>[35]</sup>.

MCs can damage DNA and produce genotoxicity. Nuclear shrinkage and chromatin condensation can be observed in the root tip meristem cells of *Phragmites australis* treated with MCs, and chromatin condensation is often accompanied by nuclear shrinkage and apoptosis <sup>[36]</sup>. DNA damage effect of MCs on *Oryza sativa* root cells by DNA fragmentation and random amplified polymorphic DNA (RAPD) <sup>[37]</sup>. Furthermore, the affected biochemical processes involved protein folding and stress response, protein biosynthesis, regulation of cell signal and gene expression, and energy and carbohydrate metabolism <sup>[38]</sup>.

The high concentrations of  $NH_4^+$  and nitrate nitrogen ( $NO_3^--N$ ) released by cyanobacteria decay have toxic effects on aquatic plants, resulting in the yellowing of plant leaves, inhibition of growth, and root morphological changes <sup>[39]</sup>. A high concentration of  $NH_4^+$  can also inhibit the absorption of K<sup>+</sup>,  $Ca^{2+}$  and  $Mg^{2+}$  by plant cells, resulting in a disturbance of ion

balance [40]. Studies have also demonstrated that a high concentration of NH<sub>4</sub><sup>+</sup> leads to the destruction of the antioxidant system balance of aquatic flora, and the accumulation of ROS, which leads to the damage of plasma membrane [41].

#### 2.3. Impact of Cyanobacteria Blooms on Microorganisms in the Aquatic Environment

Studies on the effects of cyanobacteria blooms on microorganisms in water mainly focus on the community structure and activity of microorganisms, especially at the genus level <sup>[42]</sup>. Cyanobacteria blooms in the summer, and the abundance of *Proteobacteria* in the water and sediment of Zhushan Bay is the highest at the phylum level, followed by Actinomycetes. At the genus level, the dominant bacteria in the water are *GpXI* and *GpIIa*, and the predominant bacteria in the sediment are *Gp6* and *GpIIa* <sup>[43]</sup>. Meanwhile, the different stages of cyanobacteria blooms will lead to changes in DO, N, and P in surface sediments <sup>[44]</sup>. Studies have studied and analyzed the bacterial community diversity in Poyang Lake waters and found a specific correlation between DO, Cond, salinity, mineralization, nutrients, and bacterial community diversity index <sup>[45]</sup>. In addition, debris formed during the degradation of cyanobacteria will precipitate into the surface sediments, stimulating the growth of microorganisms. Studies have demonstrated that the total bacterial diversity of water decreases during cyanobacteria blooms <sup>[46]</sup>. The decomposition of cyanobacteria will increase the diversity and abundance of ammoniated bacteria in sediments, among which the relative abundance of *Nitrosomonas oligotropha* is as high as 75% <sup>[47]</sup>.

Studies have demonstrated that the accumulation of cyanobacteria will lead to a change in microbial community structure and a decrease in diversity in the chironomid larvae gut. The relative abundance of  $\beta$ -proteobacteria increased to 40.6%, and the relative abundance of  $\delta$ -proteobacteria decreased to 4.1%. Moreover, cyanobacteria blooms can promote the expression of the *nosZ* gene and increase the abundance of *nirK* denitrifying bacteria <sup>[48]</sup>. The occurrence of cyanobacteria blooms will lead to the decrease in  $\alpha$ -diversity of the bacterial community <sup>[49]</sup>.

## 3. Impacts of Cyanobacteria Blooms on Human Health

Cyanobacteria blooms directly affect drinking water. In 1996, in Caruaru, Brazil, 50 dialysis clinic patients died because of using water contaminated with MCs <sup>[50]</sup>. In 1999, the cyanobacteria blooms in Dianchi Lake covered an area of 20 km<sup>2</sup>. In May 2007, a massive cyanobacteria bloom in Taihu Lake (Wuxi, China) led to a drinking water crisis for 2 million people in the city of Wuxi <sup>[51]</sup>. In August 2014, cyanobacteria blooms in Lake Erie increased the concentration of MCs in the drinking water, threatening the drinking water safety of nearly half a million people <sup>[52]</sup>.

When cyanobacteria blooms decompose, releasing many odor substances and cyanotoxins, it has been found that 2methylisoborneol (MIB) and geosmin are the most common substances that cause odor (musty smell) in drinking water, and their odor threshold concentrations are only 9 and 4 ng/L, respectively <sup>[53]</sup>. Among the eight kinds of odor in the table, except the chemical taste, chloride taste, and medicinal taste, the other five kinds of odor substances are related to odor compounds produced by algae. Excessive odor content in water affects the quality of drinking water and human health <sup>[51]</sup>.

Cyanobacteria can release toxins such as the hepatotoxin class, neurotoxin, and endotoxin. MCs is the most widely distributed in water, which is a cyclic heptapeptide composed of seven amino acids, mainly produced by *Microcystis* and *Anabaena* <sup>[15]</sup>. *Microcystis* is the dominant species of cyanobacteria blooms in Taihu Lake, and its biomass can account for 40–98% of the total algae biomass <sup>[54]</sup>. *Anabaena* is the most common species in cyanobacteria blooms and the only species with hepatotoxic and neurotoxic secondary metabolites <sup>[55]</sup>. Turner et al. analyzed the MCs of cyanobacteria in freshwater ecosystems in the United Kingdom and found that more than 50% of the water bodies had MCs, and of which about 13% exceeded the World Health Organization (WHO) medium health threshold (20  $\mu$ g/L) <sup>[56]</sup>.

The toxins can be accumulated by organisms and transferred through the food chain/network. Cyanotoxins are chronically toxic to humans, which lead to acute gastroenteritis, respiratory adverse reaction, eye and ear irritation, skin rash, mouth ulcers, and other diseases <sup>[57]</sup>. In addition, algal toxins can inhibit the synthesis of protein phosphatase, resulting in hyperphosphorylation of critical regulatory proteins in the signal transduction process that controls cytoskeleton tissues <sup>[58]</sup>.

MCs are hydrophilic and soluble in the blood of organisms. They cannot penetrate the lipid membrane through passive diffusion <sup>[59]</sup>. Therefore, most ingested toxins cannot pass through the ileal epithelium, stay in the digestive tract, and are most likely excreted through feces <sup>[60]</sup>. However, some studies have demonstrated that ingested MCs can be transported by bile acid membrane transporters (such as organic anion transporters (OATPs)) through the ileum into the venous blood flow and from the portal vein into hepatocytes <sup>[61]</sup>. The liver is the main target organ for the accumulation and detoxification of MCs. At the same time, MCs can also be detected in other organs (such as the intestine, kidney, brain,

lung, and heart), though to a much lesser extent. High doses of cyanobacterial toxins can cause acute liver damage, hepatomegaly, liver hemorrhage, loss of liver cell structure and function, and even biological respiratory arrest <sup>[62]</sup>.

#### References

- Mohr, K.I.; Brinkmann, N.; Friedl, T. Cyanobacteria. In Encyclopedia of Geobiology; Springer: Dordrecht, The Netherlands, 2011; pp. 306–311.
- 2. Stewart, I.; Carmichael, W.W.; Backer, L.C. Toxic Cyanobacteria. Water Sanit.-Relat. Dis. Environ. 2011, 8, 95–109.
- Paerl, H.W. Nuisance phytoplankton blooms in coastal, estuarine, and inland waters. Limnol. Oceanogr. 1988, 33, 823– 843.
- 4. Qian, K.; Chen, Y.; Song, X. Long-term development of phytoplankton dominant species related to eutrophicarion in Lake Taihu. Ecol. Sci. 2008, 27, 65–70.
- 5. Bai, M.; Zhou, S.; Zhao, M. Cyanobacterial bloom control in Taihu basin: Analysis of cost-risk analysis framework based on cooperative game. J. Clean. Prod. 2018, 195, 318–327.
- Guo, Y.; Liu, M.; Liu, L.; Liu, X.; Chen, H.; Yang, J. The antibiotic resistome of free-living and particle-attached bacteria under a reservoir cyanobacterial bloom. Environ. Int. 2018, 117, 107–115.
- Aubriot, L.; Bonilla, S. Regulation of phosphate uptake reveals cyanobacterial bloom resilience to shifting N: P ratios. Freshwater Biol. 2018, 63, 318–329.
- Kong, F.; Ma, R.; Gao, J.; Wu, X. The theory and practice of prevention, forecast and warning on cyanobacteria bloom in Lake Taihu. J. Lake Sci. 2009, 21, 314–328.
- Liu, G.; Zhong, J.; He, J.; Zhang, L.; Fan, C. Effects of Black Spots of Dead-Cyanobacterial Mats on Fe-S-P Cycling in Sediments of Zhushan Bay, Lake Taihu. J. Environ. Sci. 2009, 30, 2520–2526.
- 10. Shang, L.; Ke, F.; Li, W.; Xu, X.; Song, Y.; Feng, M. Laboratory research on the contaminants release during the anaerobic decomposition of high-density cyanobacteria. J. Lake Sci. 2013, 25, 47–54.
- 11. Li, K.; Guan, B.; Liu, Z. Experiments on decomposition rate and release forms of nitrogen and phosphorus from the decomposing cyanobacterial detritus. J. Lake Sci. 2011, 23, 919–925.
- 12. Krivtsov, V.; Bellinger, E.G.; Sigee, D.C. Elemental composition of Microcystis aeruginosa under conditions of lake nutrient depletion. Aquat. Ecol. 2005, 39, 123–134.
- Li, X.; Li, Z.; Wang, X.; Zhang, S.; Wang, H.; Li, R.; Wang, G.; Li, Q. Characteristics of Dissolved Organic Matter in Overlying Water During Algal Bloom Decay. Environ. Sci. 2021, 42, 3281–3290.
- 14. Li, D. Ecological threshold for prevention and control of cyanobacterial blooms in Baiyangdian Lake. Chin. Res. Acad. Environ. Sci. 2021.
- Xia, H.; Song, T.; Wang, L.; Jiang, L.; Zhou, Q.; Wang, W.; Liu, L.; Yang, P.; Zhang, X. Effects of dietary toxic cyanobacteria and ammonia exposure on immune function of blunt snout bream (Megalabrama amblycephala). Fish Shellfish Immunol. 2018, 78, 383–391.
- Ferrão-Filho, A.D.S.; Kozlowsky-Suzuki, B. Cyanotoxins: Bioaccumulation and effects on aquatic animals. Mar. Drugs 2011, 9, 2729–2772.
- 17. Yang, J.; Hu, L.; Zhou, W.; Chen, J.; Shi, Z. Bioaccumulation of microcystin and antioxidative response in Carassius auratus L. Ecol. Environ. Sci. 2009, 18, 2044–2050.
- Zhang, J.; Wang, Z.; Song, Z.; Xie, Z.; Li, L.; Song, L. Bioaccumulation of microcystins in two freshwater gastropods from a cyanobacteria-bloom plateau lake, Lake Dianchi. Environ. Pollut. 2012, 164, 227–234.
- 19. Fischer, W.J.; Dietrich, D.R. Pathological and biochemical characterization of microcystin-induced hepatopancreas and kidney damage in carp (Cyprinus carpio). Toxicol. Appl. Pharmacol. 2000, 164, 73–81.
- 20. Cao, Q.; Wang, L.; Yang, H.; Wei, W.; Zhang, Y. Low-dose microcystins MC-LR induced hepatopancreas injury and apoptosis in Macrobrachium rosenbergii. Asian J. Ecotox. 2020, 2, 171–179.
- 21. Andersen, R.J.; Luu, H.A.; Chen, D.Z.X.; Holmes, C.F.B.; Kent, M.L.; Blanc, M.L.; Taylor, F.J.R.; Williams, D.E. Chemical and biological evidence links microcystins to salmon 'netpen liver disease'. Toxicon 1993, 31, 1315.
- 22. Xie, L.; Yokoyama, A.; Nakamura, K.; Park, H. Accumulation of microcystins in various organs of the freshwater snail sinotaia histrica and three fishes in a temperate lake, the eutrophic lake suwa, japan. Toxicon 2007, 49, 646–652.

- 23. Yang, W.; Xiang, F.; Liang, L.; Zhou, Y. Toxicity of Ammonia and Its Effects on Oxidative Stress Mechanisms of Juvenile Crucian Carp (Carassius auratus). J. Freshw. Ecol. 2010, 25, 297–302.
- 24. Benli, A.Ç.K.; Özkul, A. Acute toxicity and histopathological effects of sublethal fenitrothion on Nile tilapia, Oreochromis niloticus. Pestic. Biochem. Phys. 2010, 97, 32–35.
- 25. Zhang, W.; Sun, S.; Ge, X.; Xia, S.; Zhu, J.; Miao, L.; Lin, Y.; Liang, H.; Pen, W.; Su, Y.; et al. Acute effects of ammonia exposure on the plasma and haematological parameters and histological structure of the juvenile blunt snout bream, Megalobrama amblycephala, and post-exposure recovery. Aquac. Res. 2017, 49, 1008–1019.
- 26. Li, D.; Li, G.; Chen, W.; Liu, Y. Interactions between a cyanobacterial bloom (Microcystis) and the submerged aquatic plant Ceratophyllum oryzetorum. Kom. Chin. J. Oceanol. Limn. 2009, 27, 38–42.
- 27. Shang, Y.; Guan, B.; Zheng, J.; Kang, Y. Effect of Cyanobacteria Accumulation on Water Environment and Submerged Plant Growth. Chin. Agric. Sci. Bull. 2015, 31, 195–198.
- 28. Chen, K.; Li, W.; Wu, Q.; Qiang, S. Impacts of Cycanobacteria on the Growth of Submerged Macrophytes, Dianchi Lake. J. Lake Sci. 2003, 15, 364–368.
- 29. Jiang, M.; Zhou, Y.; Wang, N.; Xu, L.; Zheng, Z.; Zhang, J. Allelopathic effects of harmful algal extracts and exudates on biofilms on leaves of Vallisneria natans. Sci. Total. Environ. 2018, 655, 823–830.
- 30. Saqrane, S.; Oudra, B. Cyanobacterial toxins: A short review on phytotoxic effect in an aquatic environment. Afr. J. Environ. Sci. Technol. 2011, 5, 1146–1151.
- Ha, M.H.; Contardo-Jara, V.; Pflugmacher, S. Uptake of the cyanobacterial neurotoxin, anatoxin-a, and alterations in oxidative stress in the submerged aquatic plant Ceratophyllum demersum. Ecotox. Environ. Safe 2014, 101, 205–212.
- 32. Romanowska-Duda, Z.; Tarczyńska, M. The influence of microcystin-LR and hepatotoxic cyanobacterial extract on the water plant Spirodela oligorrhiza. Environ. Toxicol. 2010, 17, 434–440.
- 33. Gehringer, M.M.; Kewada, V.; Coates, N.; Downing, T.G. The use of Lepidium sativum in a plant bioassay system for the detection of microcystin-LR. Toxicon 2003, 41, 871–876.
- 34. Chen, J.; Song, L.; Dai, J.; Gan, N.; Liu, Z. Effects of microcystins on the growth and the activity of superoxide dismutase and peroxidase of rape (Brassica napus L.) and rice (Oryza sativa L.). Toxicon 2004, 43, 393–400.
- 35. Máthé, C.; Beyer, D.; Erdődi, F.; Serfőző, Z.; Székvölgyi, L.; Vasas, G.; M-Hamvas, M.; Jámbrik, K.; Gonda, S.; Kiss, A.; et al. Microcystin-LR induces abnormal root development by altering microtubule organization in tissue-cultured common reed (Phragmites australis) plantlets. Aquat. Toxicol. 2009, 92, 122–130.
- Jámbrik, K.; Máthé, C.; Vasas, G.; Beyer, D.; Molnár, E.; Borbély, G.; M-Hamvas, M. Microcystin-LR induces chromatin alterations and modulates neutral single-strand-preferring nuclease activity in Phragmites australis. J. Plant Physiol. 2011, 168, 678–686.
- 37. Chen, J.; Ye, J.; Zhang, H.; Jiang, X.; Zhang, Y.; Liu, Z. Freshwater toxic cyanobacteria induced DNA damage in apple (Malus pumila), rape (Brassica napus) and rice (Oryza sativa). J. Hazard. Mater. 2011, 190, 240–244.
- Azevedo, C.C.; Azevedo, J.; Osório, H.; Vasconcelos, V.; Campos, A. Early physiological and biochemical responses of rice seedlings to low concentration of microcystin-LR. Ecotoxicology 2014, 23, 107–121.
- 39. Jampeetong, A.; Brix, H. Nitrogen nutrition of Salvinia natans: Effects of inorganic nitrogen form on growth, morphology, nitrate reductase activity and uptake kinetics of ammonium and nitrate. Aquat. Bot. 2009, 90, 67–73.
- 40. Wang, C.; Zhang, S.; Wang, P.; Hou, J.; Li, W.; Zhang, W. Metabolic adaptations to ammonia-induced oxidative stress in leaves of the submerged macrophyte Vallisneria natans (Lour.) Hara. Aquat. Toxicol. 2008, 87, 88–98.
- 41. Zhu, Z.; Chen, C.; Jia, H.; Wei, L.; Yin, D. Effects of different nitrogen forms on growth and physiological indexes of Vallisneria natans. J. Plant Resour. Environ. 2006, 15, 48–51.
- 42. Ma, Q.; Wang, Y.; Li, C.; Fang, P.; Zhou, J.; Fang, W. Influence of cyanobacterial bloom dominated by Planktothrix sp. and Cylindrospermopsis raciborskii on microflora structure of intestine, gill and culture enviroment of cultured Eriocheir sinensis. Mar. Fish 2021, 43, 595–606.
- 43. Xue, Y.; Jiang, C.; Geng, J.; Xie, W.; Zhang, H.; Chen, X. Profiles of Bacterioplankton Based on qPCR and 16S rDNA High-throughput Sequencing during a Heavy Cyanobacterial Bloom in Zhushan Bay, Taihu Lake. Environ. Monit. Forewarn. 2017, 9, 19–23.
- 44. Shao, K.; Gao, G.; Wang, Y.; Tang, X.; Qin, B. Vertical diversity of sediment bacterial communities in two different trophic states of the eutrophic Lake Taihu, China. J. Environ. Sci. China 2013, 25, 1186–1194.
- 45. Zheng, G.; Huang, H.; Tu, Z.; Zhang, L.; Jin, L.; Bai, Y.; Sun, R.; Zhang, Z. An Analysis of Correlation Between Bacterial Diversity and Water Environment in Poyang Lake. Acta Agric. Univ. Jiangxiensis 2017, 39, 549–558.

- Zhang, W.; Gu, P.; Zhu, W.; Wang, N.; Jiang, M.; He, J. Phenotype changes of cyanobacterial and microbial distribution characteristics of surface sediments in different periods of cyanobacterial blooms in taihu lake. Aquat. Ecol. 2020, 54, 591–607.
- 47. Huang, R.; Shen, F.; Luo, J.; Wang, S.; Tang, Q.; Xu, M.; Wu, Y.; Zhao, D. Effects of Withering of Cyanobacteria Bloom on Abundance and Community Composition of Ammonia-Oxidizing Bacteria in Surface Lake Sediments. J. Ecol. Rural Environ. 2015, 31, 334–339.
- 48. Sun, X. The Influence of Zoobenthos on Community Structure and Function of Nitrogen Transformation Microbe in Sediment from Eutrophic Lakes. Ph.D. Thesis, Nanjing University, Nanjing, China, 2015.
- 49. Peng, Y.; Lu, J.; Chen, H.; Xiao, L. Dynamic Changes of Nitrogen-Transforming and Phosphorus-Accumulating Bacteria Along with the Formation of Cyanobacterial Blooms. Environ. Sci. 2018, 39, 4938–4945.
- 50. Jochimsen, E.M.; Carmichael, W.W.; An, J.; Cardo, D.M.; Cookson, S.T.; Holmes, C.E. Liver failure and death after exposure to microcystins at a hemodialysis center in Brazil. N. Engl. J. Med. 1998, 338, 873–878.
- 51. Qin, B.; Zhu, G.; Gao, G.; Zhang, Y.; Li, W.; Paerl, H.W.; Carmichael, W.W. A drinking water crisis in Lake Taihu, China: Linkage to climatic variability and lake management. Environ. Manag. 2010, 45, 105–112.
- 52. Levy, S. Microcystis Rising: Why Phosphorus Reduction Isn't Enough to Stop CyanoHABs. Environ. Health Perspect. 2017, 125, A34.
- 53. Watson, S.B. Aquatic taste and odor: A primary signal of drinking-water integrity. J. Toxicol. Environ. Health 2004, 67, 1779–1795.
- 54. Chen, Y.; Qin, B.; Teubner, K.; Dokulil, M.T. Long-term dynamics of phytoplankton assemblages: Microcystisdomination in Lake Taihu, a large shallow lake in China. J. Plankton. Res. 2003, 25, 445–453.
- 55. Sivonen, K.; Niemelä, S.I.; Niemi, R.M.; Lepistö, L.; Luoma, T.H.; Räsänen, L.A. Toxic cyanobacteria (blue-green algae) in Finnish fresh and coastal waters. Hydrobiologia 1990, 190, 267–275.
- 56. Turner, A.D.; Dhanji-Rapkova, M.; O'Neill, A.; Coates, L.; Lewis, A.; Lewis, K. Analysis of microcystins in cyanobacterial blooms from freshwater bodies in England. Toxins 2018, 10, 39.
- 57. Gallitelli, M.; Ungaro, N.; Addante, L.M.; Procacci, V.; Silveri, N.G.; Sabbà, C. Respiratory Illness as a Reaction to Tropical Algal Blooms Occurring in a Temperate Climate. Jama 2005, 293, 2599–2600.
- 58. Meng, G.; Sun, Y.; Fu, W.; Guo, Z.; Xu, K. Microcystin-LR induces cytoskeleton system reorganization through hyperphosphorylation of tau and hsp27 via pp2a inhibition and subsequent activation of the p38 mapk signaling pathway in neuroendocrine (pc12) cells. Toxicology 2011, 290, 219–230.
- 59. Vesterkvist, P.S.M.; Meriluoto, J.A.O. Interaction between microcystins of different hydrophobicities and lipid monolayers. Toxicon 2003, 41, 349–355.
- 60. Ernst, B. Investigations on the Impact of Toxic Cyanobacteria on Fish as Exemplified by the Coregonids in Lake Ammersee. Ph.D. Thesis, University of Konstanz, Konstanz, Germany, 2008.
- Fischer, W.; Altheimer, S.; Cattori, V.; Meier, P.J.; Dietrich, D.R.; Hagenbuch, B. Organic anion transporting polypeptides expressed in liver and brain mediate uptake of microcystin—Science Direct. Toxicol. App. Pharm. 2005, 203, 257–263.
- Nishiwaki-Matsushima, R.; Ohta, T.; Nishiwaki, S.; Suganuma, M.; Kohyama, K.; Ishikawa, T.; Carmichael, W.W.; Fujiki, H. Liver tumor promotion by the cyanobacterial cyclic peptide toxin microcystin-LR. J. Cancer Res. Clin. 1992, 118, 420–424.

Retrieved from https://encyclopedia.pub/entry/history/show/70625